

Supplementary Figure Legends

Supplementary Figure S1. The effects of RASSF1C on cell-cycle distribution and p53 activity

(A) HCT116 cells (p53^{+/+}, p53^{-/-} and p53^{-/-} complemented with HA-p53) were transfected with pcDNA-CD4 and HA-RASSF1C. The cell-cycle profile of CD4-positive cells was determined 36 h after transfection. Data are expressed as means \pm standard errors of values from three independent experiments. (B) Lysates of cells in (A) were analyzed by immunoblotting for the indicated proteins. (C) U2OS cells were co-transfected with pGL3-p53-responsive reporter and pcDNA3- β -Gal reporter constructs and the indicated combinations of pcDNA3-HA-p53 and HA-RASSF1A or HA-RASSF1C. Cell lysates were subjected to immunoblot and luciferase assays. Luciferase activity was normalized to β -galactosidase activity and expressed relative to that in cells transfected with the empty vector. Data are expressed as means \pm standard errors of values from three independent experiments. Asterisks indicate nonspecific bands.

Supplementary Figure S2. The effects of RASSF1C on p53 stability

(A) U2OS cells transfected with HA-RASSF1C were incubated for the indicated times with cycloheximide (50 μ g/ml) and analyzed by immunoblotting. (B) U2OS cells transfected with His₆-tagged ubiquitin were co-transfected with a combination of Flag-MDM2, HA-p53, and HA-RASSF1C, as indicated. Cells were treated with MG132 (20 μ M) for 6 h prior to harvesting. The His₆-purified fractions were analyzed for ubiquitinated p53. Molecular weights are in kDa.

Supplementary Figure S3. Regulation of p53 stability by RASSF1A and interaction of RASSF1A with MDM2

(A) *Mdm2*^{+/+}*p53*^{-/-} and *Mdm2*^{-/-}*p53*^{-/-} MEFs transiently transfected with HA-p53 and HA-RASSF1A, or the corresponding empty vector (control), were incubated for the indicated times with cycloheximide (50 µg/ml) and then analyzed by immunoblotting for the indicated proteins. (B) RASSF1A directly interacts with MDM2 but not with p53 *in vitro*. RASSF1A, MDM2, and p53 proteins were *in vitro* translated (IVT) in the presence of [³⁵S] methionine and mixed. Cell lysates were immunoprecipitated with anti-RASSF1A and analyzed by SDS-PAGE and autoradiography. (C) *Mdm2*^{-/-}*p53*^{-/-} MEFs transiently transfected with the indicated expression plasmids were immunoprecipitated with an anti-RASSF1A antibody and analyzed by immunoblotting.

Supplementary Figure S4. Interaction of RASSF1A with MDM2 and DAXX in the nucleus

(A) 0.2% (w/v) Triton X-100-permeabilized U2OS cells were fixed and stained with anti-RASSF1A (*green*) or anti-MDM2 (*red*) antibodies. Scale bar, 10 µm. (B) U2OS cell lysates were fractionated into nuclear and cytoplasmic fractions and analyzed by immunoblotting for the indicated proteins. β-tubulin and HDAC1 were used as cytoplasmic and nuclear markers, respectively (*left*). Both nuclear and cytoplasmic extracts were immunoprecipitated (IP) with an anti-RASSF1A antibody and the resulting precipitates were analyzed by immunoblotting for the indicated proteins (*right*). Asterisks indicate nonspecific bands. (C) 0.2% (w/v) Triton X-100-permeabilized U2OS cells were fixed and stained with anti-RASSF1A (*green*) or anti-DAXX (*red*) antibodies. Scale bar, 10 µm.

Supplementary Figure S5. Effects of the N-terminal region of RASSF1A on MDM2 and p53 stability. (A) *Mdm2*^{-/-}*p53*^{-/-} MEFs were transfected with MDM2 and the cell lysates were analyzed by immunoblotting for the indicated proteins. Fold induction of Rassf1a level was calculated using the densities of Rassf1a/Actin. (B) U2OS cells were co-transfected with expression vectors for HA-p53, pCMV-MDM2, or HA-RASSF1A-N, as indicated. The cell lysates were analyzed by immunoblotting for the indicated proteins. (C) U2OS cells were co-transfected with expression vectors for HA-p53, pCMV-MDM2, or HA-RASSF1A-C, as indicated. The cell lysates were analyzed by immunoblotting for the indicated proteins.

Supplementary Figure S6. RASSF1A disrupts MDM-DAXX-HAUSP interactions *in vivo*. *Mdm2*^{-/-}*p53*^{-/-} MEFs co-transfected with MDM2 and Flag-DAXX or Myc-HAUSP and increasing amounts of HA-RASSF1A were treated with MG132 for 6 h and Cell lysates were analyzed by immunoblotting as in **Figure 6A**.

Supplementary Figure S7. Effects of RASSF1C on the MDM2-DAXX-HAUSP interactions *in vivo*

(A, B, C) *Mdm2*^{-/-}*p53*^{-/-} MEFs transfected with MDM2, Flag-DAXX, Myc-HAUSP or HA-RASSF1C as indicated were treated with MG132 for 6 h and immunoprecipitated (IP) with anti-MDM2 or anti-Flag antibodies. The resulting precipitates were analyzed by immunoblotting.

Supplementary Figure S8. Inhibition of HAUSP-mediated MDM2 deubiquitination by RASSF1A

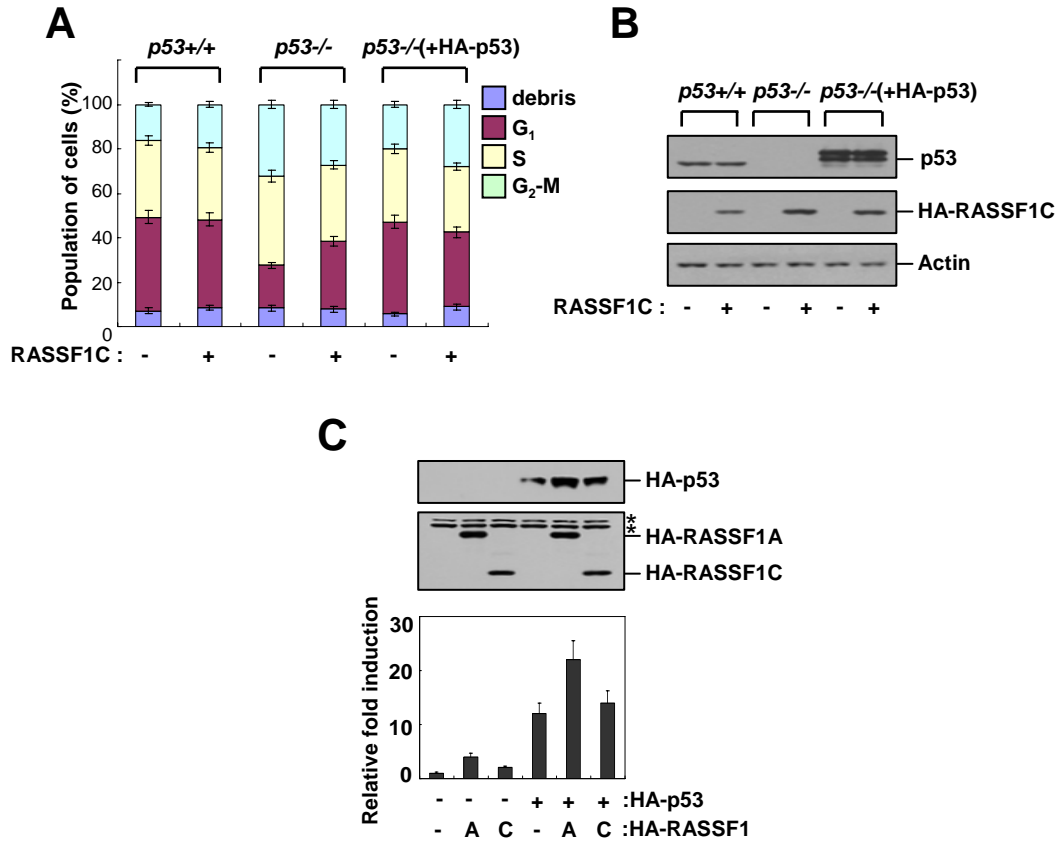
(A) U2OS cells transfected with Myc-HAUSP with or without HA-RASSF1A were incubated for the indicated times with cycloheximide (50 μ g/ml) and analyzed by immunoblotting. (B) U2OS cells co-transfected with His₆-tagged ubiquitin, Myc-HAUSP, and HA-RASSF1A were treated with MG132 (20 μ M) for 6 h and then harvested. The His₆-purified fractions were analyzed for ubiquitinated MDM2.

Molecular weights are in kDa.

Supplementary Figure S9. RASSF1A promotes p53 activation by decreasing MDM2 levels upon γ -irradiation treatment.

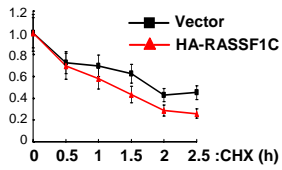
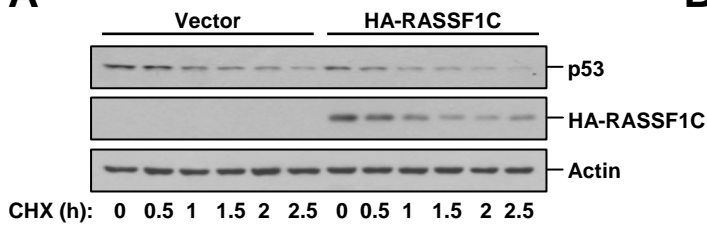
U2OS cells stably expressing pcDNA4/TO-HA-RASSF1A were incubated without (mock-treated) or with tetracycline (1 μ M) for 24 h to induce RASSF1A expression. The cells were subsequently exposed to 10 Gy of γ -irradiation, incubated for the indicated times, and then analyzed by immunoblotting for the indicated proteins.

Supplementary Figure S1

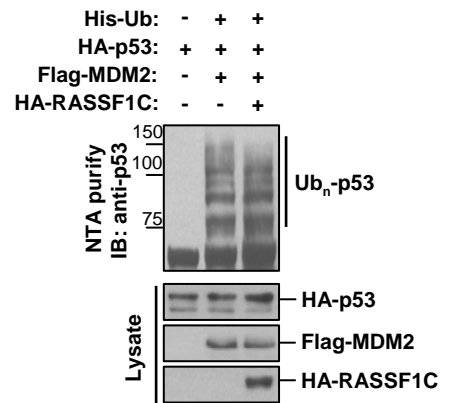


Supplementary Figure S2

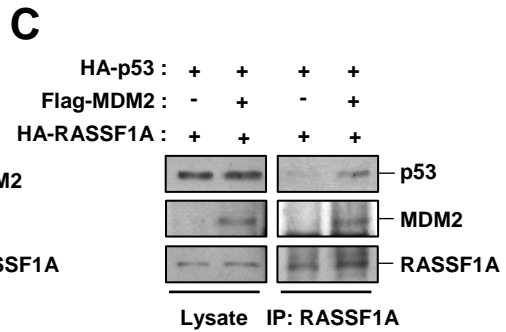
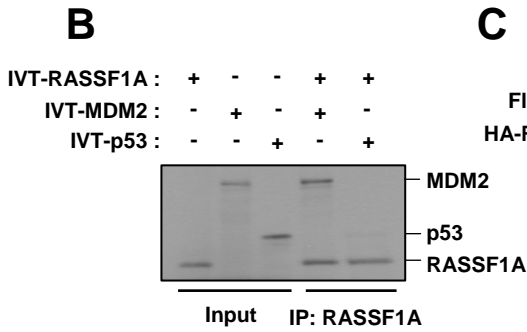
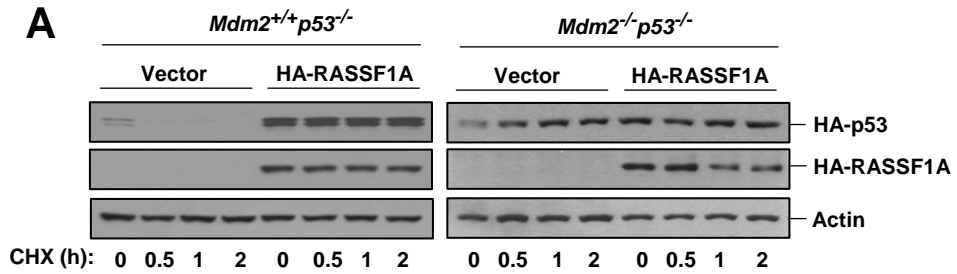
A



B

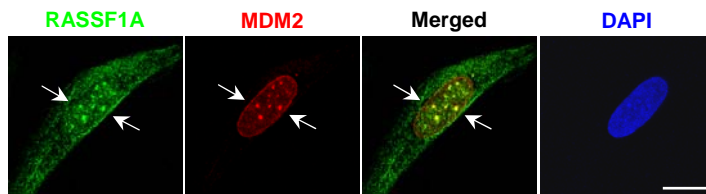


Supplementary Figure S3

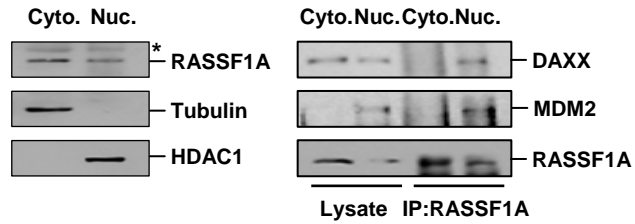


Supplementary Figure S4

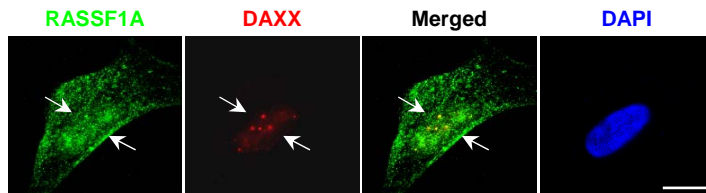
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B

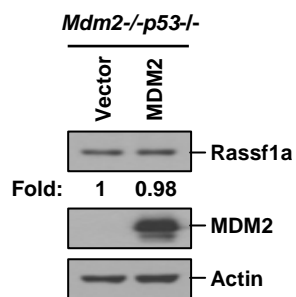


C

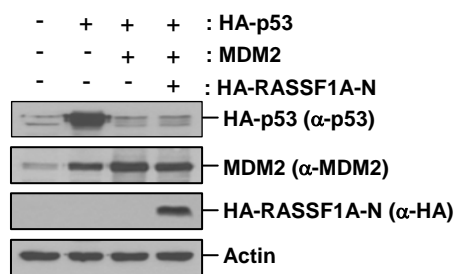


Supplementary Figure S5

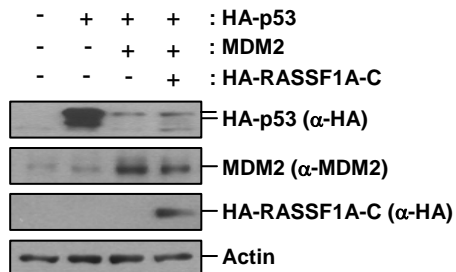
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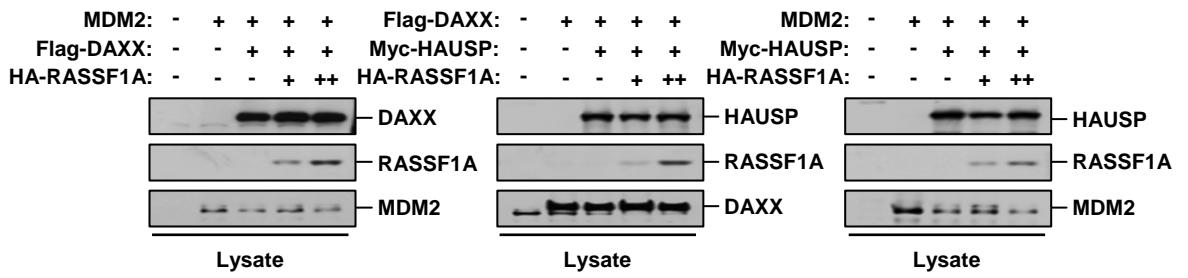
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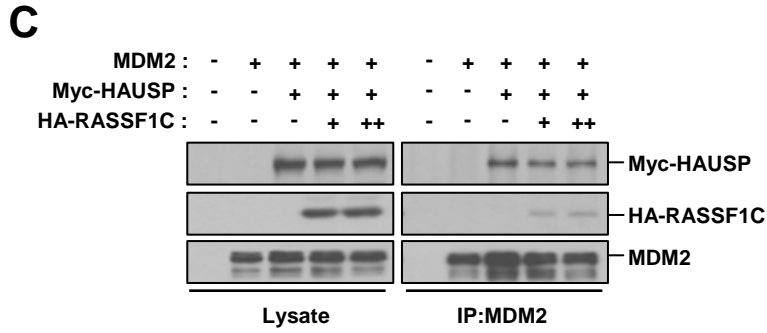
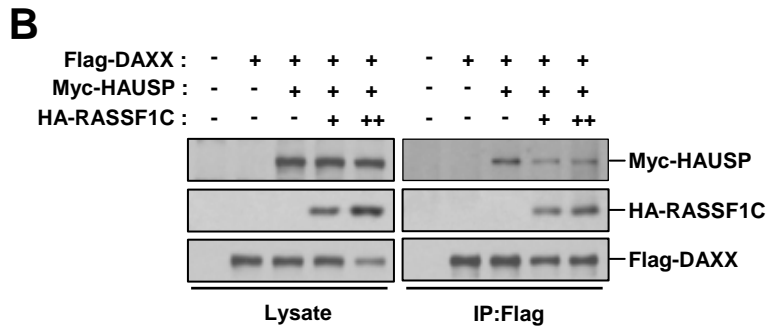
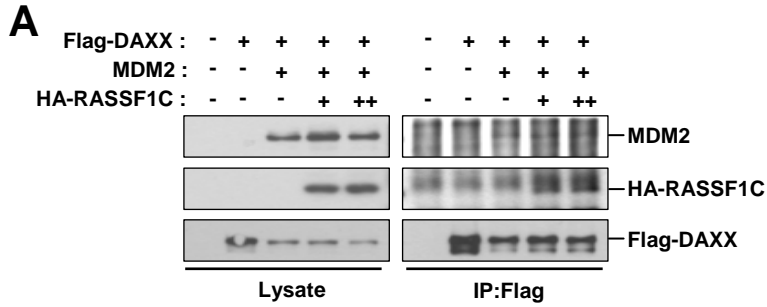
C



Supplementary Figure S6

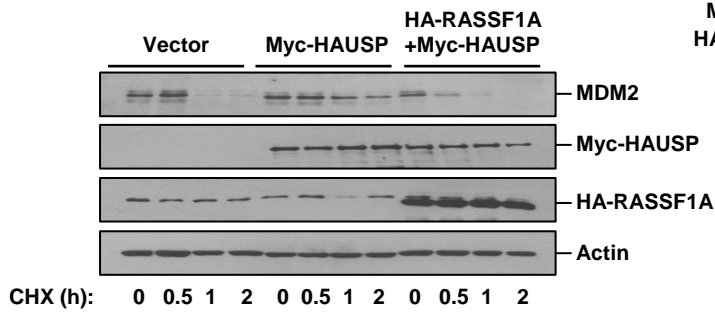


Supplementary Figure S7

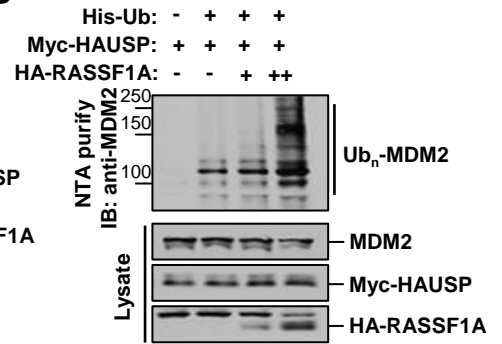


Supplementary Figure S8

A



B



Supplementary Figure S9

