Using a 3-O-sulfated Heparin Octasaccharide to Inhibit the Entry of Herpes Simplex Virus 1

(Supplementary Materials)

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The 3-OH octasaccharide contained four glucosamine residues, thereby giving it four potential sites for 3-O-sulfation by 3-OST-3 as shown in Suppl Fig 1. These four positions include residue G2, G4, G6, and G8. Here we describe the results that served to identify which residue carried the 3-O-[35 S]sulfo group. The structural characterization was accomplished using a combination of chemical and enzymatic degradations from both non-reducing and reducing ends. Non-reducing end analysis permitted the identification of whether the 3-O-[35 S]sulfo group was present on either residue G2 or G4. Reducing end analysis permitted the identification of whether residue G6 or G8 carried the 3-O-[35 S]sulfo group.

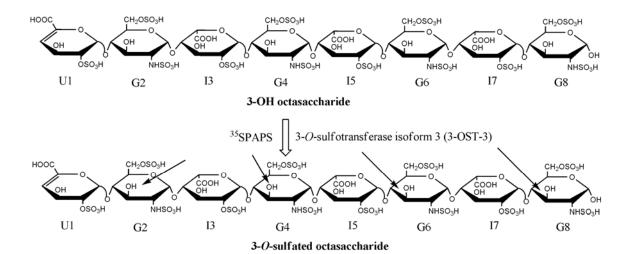
Reducing end analysis (disaccharide portion)-The results of reducing end analysis for the tetrasaccharide analysis have been presented in the main text. The 2-AB labeled 3-*O*-sulfated octasaccharide was digested with heparin lyases. The products were analyzed by PANM-HPLC, and yielded a ³⁵S-labeled component that was eluted at 55 min. We believed that this component is a disaccharide (Supplementary Fig 2A). We found that the ³⁵S-labeled component coeluted with ΔUA2S-[3-³⁵S]GlcNS3S6S, but not with ΔUA2S-[3-³⁵S]GlcNS3S6S on PAMN-HPLC (Supplementary Fig 2). Thus, our results demonstrated that the ³⁵S-labeled component that was eluted at 55 min is ΔUA2S-GlcNS3S6S. Our results also confirmed that the 3-O-sulfo group is not present at G8 residue.

Non-reducing end analysis-The non-reducing end analysis takes advantage of the substrate specificity of $\Delta^{4,5}$ glycuronate-2-sulfatase (2ase), an exolytic sulfatase. This enzyme is known to specifically remove the 2-O-sulfo group from $\Delta UA2S$, which is present at the non-reducing end of the 3-O-[^{35}S]sulfated octasaccharide (Suppl Fig 3,

U1). By subjecting the 3-*O*-[³⁵S]sulfated octasaccharide to heparin lyases digestion with or without pretreatment with 2ase permitted the determination of whether the 3-*O*-[³⁵S]sulfo group was on residue 2 as illustrated in Suppl Fig 2.

The treatment of 3-*O*-sulfated octasaccharide with 2ase was completed as determined by the elution profile by DEAE-NPR-HPLC (Suppl Fig 4B). Before the treatment, 3-*O*-sulfated octasaccharide was eluted as a major peak at 63 min (Suppl Fig 4A). The treatment with 2ase resulted in the shift of the major ³⁵S-peak to 57 min, consistent with the loss of a negative charge due to the action of sulfates (Suppl Fig 4B). We estimated that the 2ase reaction of the 3-*O*-[³⁵S]sulfated octasaccharide was close to being 95% complete.

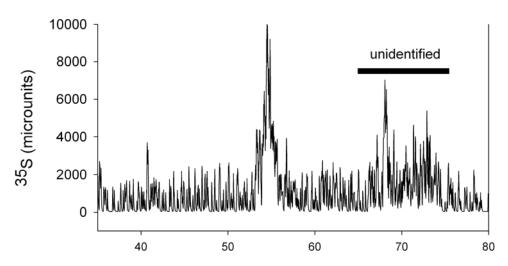
We then digested the 3-O-[35 S]sulfated octasaccharide and 2ase pretreated octasaccharide with a mixture of heparin lyases, including heparin lyase I, II and III. The resultant disaccharides were analyzed by PAMN-HPLC. As shown in Suppl. Fig 4C and 4D, both preparations gave an identical 35 S-labeled disaccharide, coeluting with the disaccharide standard, Δ UA2S-GlcNS3S6S. To this point, we concluded that the 3-O-[35 S]sulfo group was not present at residue 2. Although whether or not residue 4 carried the 3-O-[35 S]sulfo group could be proved by conducting partial digestion of the 3-O-[35 S]sulfated octasaccharide, we decided not to pursue those experiments as the data from reducing end analysis had concluded that the 3-O-sulfo group is on G6 residue as described in the main text.



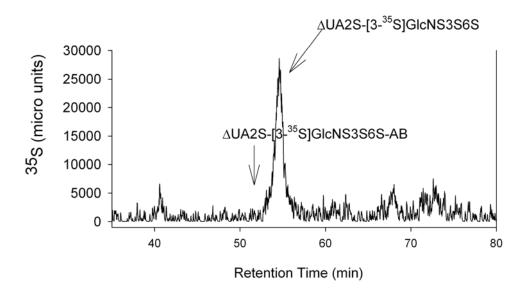
Supplementary Figure 1. Potential 3-*O***-sulfation sites in the 3-***O***-sulfated octasaccharide.** There are four N-sulfoglucosamine 6-*O*-sulfate residues can accept the 3-*O*-sulfo group. These sites are indicated by arrows.

Supplementary Figure 2

A. Heparin lyases digested 2-AB-labeled 3-O-sulfated octasaccharide

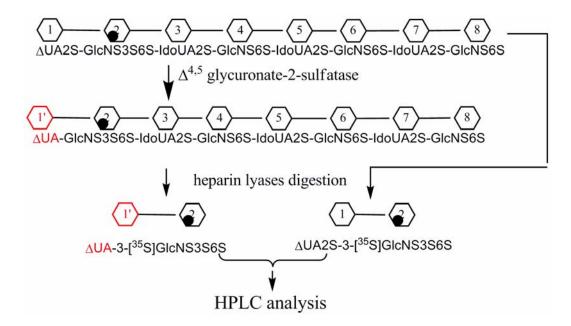


B. Coinjection of ΔUA2S-[3-35S]GlcNS3S6S with the digested octasaccharide



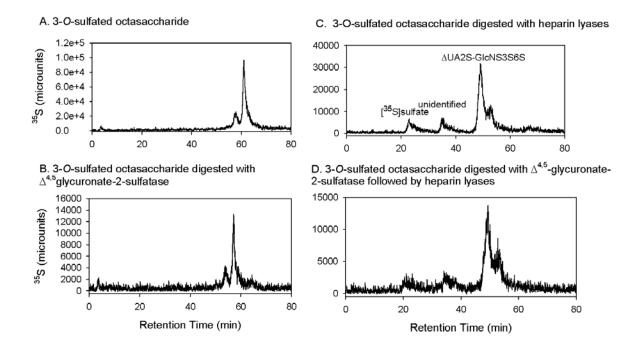
Supplementary Figure 2. Reducing end sequence analysis of 3-O-sulfated octasaccharide (disaccharide portion only). Purified 3-O-sulfated octasaccharide was digested with heparin lyases, including lyase I, II and III. The products were analyzed by PAMN-HPLC. The identities of the digested products were confirmed by coinjecting with appropriate disaccharide standards. *Panel A* shows the digestion of the 2-AB-labeled 3-O-sulfated octasaccharide alone in the region where trisulfated disaccharides are eluted. *Panel B* shows the digested octasaccharide coinjected with the Δ UA2S-[3- 35 S]GlcNS6S3S standard.

Supplementary Figure 3



Supplementary Figure 3. Strategy for nonreducing end analysis of the 3-O-sulfated octasaccharide. The octasaccharide was digested with $\Delta^{4,5}$ glycuronate-2-sulfatase, which selectively removes the 2-O-sulfo group from Residue 1. The resultant octasaccharide was then digested to a disaccharide by a mixture of heparin lyases. The structure of the disaccharide was then compared to the disaccharide from heparin lyases digested 3-O-sulfated octasaccharide that was not treated with $\Delta^{4,5}$ -glycuronate-2-sulfatase. The structural comparison was carried out by polyamine-based anion exchange (PAMN)-HPLC. *Black circle* represents potential location of 3-O-[35 S]sulfo group on residue 2.

Supplementary Figure 4



Supplementary Figure 4. HPLC chromatograms of the nonreducing end analysis of 3-O-[35 S]sulfated octasaccharide. Panels A and B show the chromatograms of 3-O-[35 S]sulfated octasaccharide and $\Delta^{4,5}$ -glycuronate-2-sulfatase digested 3-O-[35 S]sulfated octasaccharide on DEAE-NPR-HPLC, respectively. Panels C and D show the chromatograms of heparin lyases-digested 3-O-[35 S]sulfated octasaccharide and 3-O-[35 S]sulfated octasaccharide pretreated with $\Delta^{4,5}$ glycuronate-2-sulfatase on polyamine-based anion exchange (PAMN)-HPLC. The conditions for the enzyme digestions and HPLC elution conditions are described under "Supplementary Methods".