

Remote Control of Diastereoselectivity in Intramolecular Reactions of Chiral Allylsilanes

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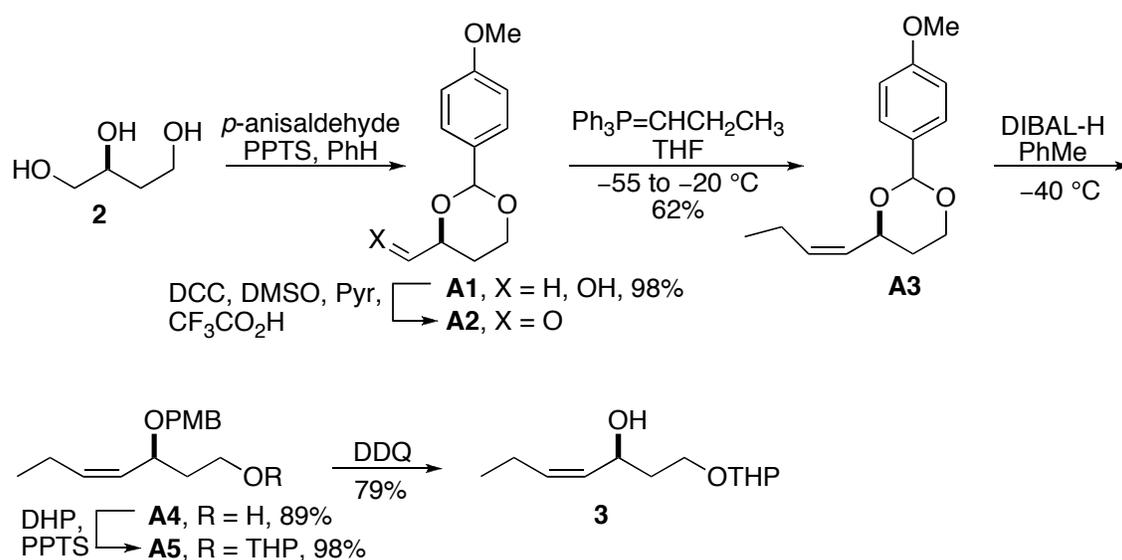
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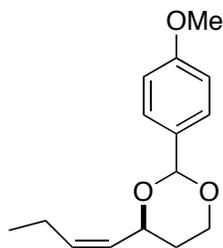
List of Known Compounds

The following compounds are known: (*E*)-5-(trimethylsilyl)pent-3-en-1-ol (**D4**),¹ 1-chloro-2,2,2-trimethyl-1,1-diphenyldisilane,² (4*S*)-2-(4'-methoxyphenyl)-1,3-dioxan-4-ylmethanol (**A1**),³ (4*S*)-2-(4'-methoxyphenyl)-1,3-dioxane-4-carbaldehyde (**A2**),³ but-3-enyl pivalate (**1.30**),⁴ 1-benzyloxy-3-butyne (**6**),⁵ 1-benzyloxy-5-hexyne (**B2**),⁶ 1-tert-butyl-diphenylsilyloxy-3-butyne (**B3**),⁷ 4-methoxy-benzyloxy-3-butyne (**B4**),⁸ and ethyl 4-(*N*-tert-butoxycarbonylamino)-3-formylbenzoate.¹

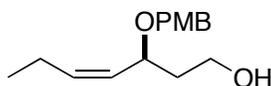
Preparation of Chiral Allylic Alcohols

Allylic alcohol **3** was prepared from the *p*-methoxybenzyl acetonide of (*S*)-(-)-1,2,4-butanetriol (**A1**)³ (Scheme A). Moffatt oxidation⁹ of **A1** afforded aldehyde **A2**,³ which was used without further purification. The olefin moiety was then installed via a Wittig reaction to give a 12.5:1 mixture of *Z/E* isomers that were separable by chromatography. DIBAL-H reduction of the major *Z* isomer **A3** followed by protecting group manipulations afforded allylic alcohol **3**.

Scheme A. Synthesis of allylic alcohol **3**.

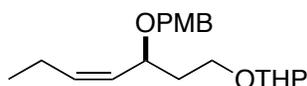


(3*S,Z*)-4-But-1-enyl-2-(4'-methoxyphenyl)[1,3]dioxane (A3). *n*-BuLi (1.0 M, 22.7 mL) was added to a solution of propyltriphenylphosphonium bromide (11.6 g, 30.3 mmol) in THF (60 mL) at 0 °C. After stirring for 20 min, the reaction mixture was cooled to -78 °C, and aldehyde **A2** was added as a solution in THF (20 mL). The reaction mixture was allowed to slowly warm to -20 °C, and was stirred for an additional 6 h. After warming to 0 °C, the reaction was quenched with water (50 mL) and extracted with ether (3 × 50 mL). The combined organic extracts were washed with brine, dried (Na₂SO₄), filtered, and concentrated. The residue obtained was triturated with hexanes and filtered. The filtrate was concentrated to give a crude oil containing ca. 12.5:1 (*Z/E*) mixture of olefin isomers. The crude product mixture was purified by chromatography (20:1 hexanes:EtOAc) to give 2.4 g (64%) of (*Z*)-**A3** as a colorless oil. $[\alpha]_D^{25} +61.6$ (*c* 1.00, CDCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.03 (t, *J* = 7.5 Hz, 3H), 1.50 (m, 1H), 1.98 (m, 1H), 2.17 (m, 2H), 3.81 (s, 3H), 4.02 (dt, *J* = 2.4, 12.4 Hz, 1H), 4.28 (ddd, *J* = 1.2, 5.0, 11.4 Hz, 1H), 4.69 (m, 1H), 5.49 (dd, *J* = 7.7, 11.0 Hz, 1H), 5.55 (m, 2H), 6.91 (d, *J* = 8.8 Hz, 2H), 7.44 (d, *J* = 8.7 Hz, 2H); ¹³C NMR (100.6 MHz, CDCl₃) δ 14.7, 21.7, 31.4, 55.7, 67.3, 74.0, 101.5, 114.0, 127.8, 129.4, 131.7, 134.4, 160.3; IR (film) 1615 cm⁻¹; MS (CI) *m/z* 249 (M⁺ + H), 152, 95; HRMS calcd for C₁₅H₂₁O₃ (M⁺ + H) 249.1491, found 249.1509.



(3*S,Z*)-3-(4'-Methoxybenzyloxy)hept-4-en-1-ol (A4). Diisobutylaluminum hydride (37 mL, 1.5 M solution in toluene) was added to a solution of **A3** (2.73 g, 11.0 mmol) in anhydrous toluene (50 mL) at -40 °C. After stirring at -40 °C for 2h, the

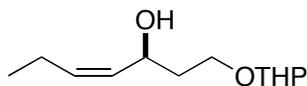
reaction was quenched by the slow addition of MeOH (20 mL). The resulting mixture was subsequently stirred in the presence of saturated aqueous potassium sodium tartrate (50 mL) while warming to room temperature for 10 h. The phases thus formed were separated and the aqueous layer was extracted with CH₂Cl₂ (3 × 50 mL). The combined organic layers were dried (Na₂SO₄), filtered, and concentrated. The crude material was purified by chromatography (3:1 hexanes:EtOAc) to afford 2.5 g (91%) of a colorless oil. $[\alpha]_D -45.0$ (c 1.25, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.01 (t, *J* = 7.5 Hz, 3H), 1.69 (m, 1H), 1.89 (m, 1H), 2.08 (m, 2H), 2.64 (br, s, 1H), 3.76 (m, 2H), 3.82 (s, 3H), 4.28 (d, *J* = 11.4 Hz, 1H), 4.40 (dt, *J* = 4.3, 8.6 Hz, 1H), 4.55 (d, *J* = 11.4 Hz, 1H), 5.35 (m, 1H), 5.64 (dt, *J* = 11.0, 7.4 Hz, 1H), 6.89 (dt, *J* = 8.7, 2.0 Hz, 2H), 7.25 (d, *J* = 8.6 Hz, 2H); ¹³C NMR (100.6 MHz, CDCl₃) δ 14.7, 21.5, 38.3, 55.7, 61.4, 70.0, 74.1, 114.2, 129.7, 129.8, 130.8, 135.8, 159.6; IR (film) 3413, 1613, 1514 cm⁻¹; MS (EI) *m/z* 268 (M⁺ + NH₄), 137, 121; HRMS calcd for C₁₅H₂₆NO₃ (M⁺ + NH₄) 268.1913, found 268.1930.



2-[(3*S*,*Z*)-3'-(4'-Methoxybenzyloxy)hept-4'-enyloxy]tetrahydro-2*H*-pyran

(A5). DHP (1.0 g, 12.0 mmol) was added to a solution of **A4** (1.88 g, 7.51 mmol) and PPTS (0.09 g, 0.38 mmol) in THF (35 mL) at 0 °C. The reaction mixture was allowed to warm to room temperature and stirred for 16 h. A saturated aqueous NaHCO₃ solution (25 mL) was then added, and the aqueous layer was extracted with ether (3 × 30 mL). The combined organic layers were washed with brine, dried (Na₂SO₄), filtered, and concentrated. The crude material was purified by chromatography (10:1 hexanes:EtOAc) to give 2.5 g (98%) of a colorless oil. ¹H NMR (400 MHz, CDCl₃) *Mixture of THP ether diastereomers*: δ 1.00 (dt, *J* = 1.4, 7.5 Hz, 3H), 1.50-2.10 (m, 11H), 3.5 (m, 2H), 3.82 (m, 4H), 4.31 (m, 2H), 4.52 (dd, *J* = 4.6, 4.2 Hz, 2H), 5.31 (m, 1H), 5.64 (m, 1H), 6.88 (dd, *J* = 0.4, 8.5 Hz, 2H), 7.27 (m, 2H); ¹³C NMR (100.6 MHz, CDCl₃) *Mixture of THP ether diastereomers; all peaks listed*: δ 14.8, 20.0, 20.1, 21.4, 21.5, 25.9, 25.9, 31.1, 31.2, 36.3, 36.4, 55.7, 62.6, 62.7, 64.5, 64.5, 69.8, 70.0, 71.1, 71.7, 99.1, 99.5, 114.1, 114.1, 129.7,

129.8, 130.3, 130.3, 131.4, 131.4, 135.6, 135.7, 159.4, 159.4; IR (film) 1613, 1512 cm^{-1} ; MS (CI) m/z 352 ($\text{M}^+ + \text{NH}_4$), 335, 121; HRMS calcd for $\text{C}_{20}\text{H}_{34}\text{NO}_4$ ($\text{M}^+ + \text{NH}_4$) 352.2488, found 352.2499.

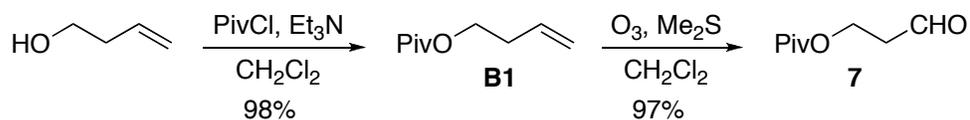


(3S,Z)-1-(Tetrahydro-2H-pyran-2'-yloxy)hept-4-en-3-ol (3). To a mixture of **A5** (1.44 g, 4.30 mmol) in CH_2Cl_2 /water (60 mL: 3mL) at room temperature was added DDQ (1.19 g, 5.25 mmol). The reaction mixture was stirred vigorously for 1.5 h, and then a saturated aqueous solution of NaHCO_3 (50 mL) was added. The layers were separated and the aqueous layer was extracted with CH_2Cl_2 (3×50 mL). The combined organic layers were washed with brine (1×50 mL), dried (Na_2SO_4), filtered, and concentrated. Chromatography (5:1 to 2:1 hexanes:EtOAc) gave 0.73 g (79%) of a colorless oil. ^1H NMR (500 MHz, CDCl_3) *Mixture of THP ether diastereomers*: δ 1.01 (dt, $J = 1.7, 7.5$ Hz, 3H), 1.56-1.64 (complex, 4H), 1.74-1.95 (complex, 4H), 2.11 (m, 2H), 2.67 (br, s, 1H), 3.55 (m, 2H), 3.94 (m, 2H), 4.61 (m, 1H), 4.69 (m, 1H), 5.45 (m, 2H); ^{13}C NMR (100.6 MHz, CDCl_3) *Mixture of THP ether diastereomers; all peaks listed*: δ 14.8, 19.8, 20.0, 21.4, 25.7, 30.9, 31.1, 37.4, 37.4, 62.6, 63.0, 65.8, 66.0, 66.8, 67.4, 99.3, 99.5, 131.9, 131.9, 133.8, 133.9; IR (film) 3431 cm^{-1} ; MS (CI) m/z 215 ($\text{M}^+ + \text{H}$), 197, 85; HRMS calcd for $\text{C}_{12}\text{H}_{23}\text{O}_3$ ($\text{M}^+ + \text{H}$) 215.1647, found 215.1639.

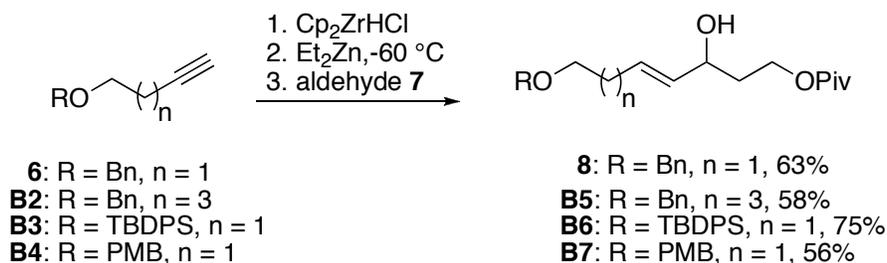
Ether-substituted allylic alcohols were prepared via a zirconium-mediated coupling of aldehyde **7** with benzyloxy or *tert*-butyldiphenylsilyloxy alkynes. The racemic allylic alcohol **8** was subjected to a kinetic resolution under Sharpless conditions¹⁰ to furnish the enantioenriched allylic alcohol (*R*)-**8** in 42% yield and $\geq 99\%$ ee as determined by proton NMR of the Mosher's ester derivative¹¹ All other ether-substituted allylic alcohols were prepared and used in racemic form.

Scheme B. Synthesis of ether-substituted allylic alcohols.

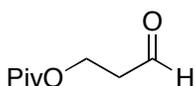
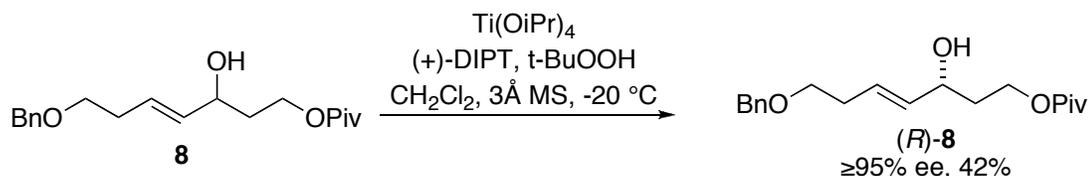
Preparation of the aldehyde coupling component:



Synthesis of racemic allylic alcohols:



Generation of enantiopure allylic alcohol:

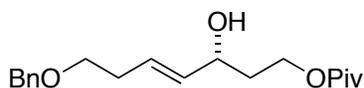


3-Oxopropyl pivalate (7). Ozone was bubbled through a solution of 3-butenyl pivalate (**B1**)⁴ (12.87 g, 82.37 mmol) in CH₂Cl₂ (150 mL) at –78 °C until a persistent blue color resulted (ca. 2 h). The reaction was quenched with dimethyl sulfide (65 mL) and allowed to warm to room temperature with stirring for 16 h. The reaction mixture was concentrated under reduced pressure and the resulting residue was dissolved in ether and filtered through a pad of silica gel. The filtrate was concentrated to afford 12.70 g (97%) of the title compound as a colorless oil. ¹H NMR (500 MHz, CDCl₃) δ 1.21 (s, 9H), 2.78 (td, *J* = 6.1, 1.5, Hz, 2H), 4.42 (t, *J* = 6.1 Hz, 2H), 9.81 (t, *J* = 1.5 Hz, 1H); ¹³C NMR

(125.8 MHz, CDCl₃) δ 27.0, 38.6, 57.9, 178.2, 199.3; IR (neat) 1728, 1483 cm⁻¹; MS (CI) *m/z* 159 (M⁺ + H), 85, 57; HRMS calcd for C₈H₁₅O₃ (M⁺ + H) 159.1021, found 159.1020.

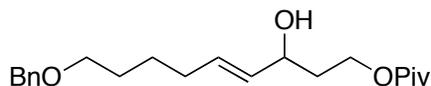
General procedure for the synthesis of ether-substituted allylic alcohols.

According to the procedure of Wipf and Xu,¹² bis(cyclopentadienyl)zirconium chloride hydride (Schwartz's reagent) (5.0 g, 19.4 mmol) was added to a solution of the alkyne (16.2 mmol) in dry CH₂Cl₂ (60 mL) at 0 °C. The resulting mixture was then warmed to room temperature and stirred until a homogeneous solution formed. The resulting yellow solution was stirred at room temperature for an additional 20 min, and then cooled to -60 °C. Diethylzinc (1.0 M in hexanes, 19.4 mmol) was then added dropwise over 45 min at -60 °C. After the addition was complete, the resulting solution was stirred at -60 °C for an additional 10 min. The reaction flask was immersed in an ice bath, and a solution of aldehyde **7** (3.1 g, 19.4 mmol) in CH₂Cl₂ (10 mL) was added dropwise over 45 min. The resulting solution was stirred at 0 °C for an additional 6 h. The light-yellow reaction mixture was then slowly poured into an ice-cold solution of aq 5% NaHCO₃ (200 mL) and stirring was continued at room temperature until gas evolution ceased. The resulting mixture was extracted with Et₂O (3 × 200 mL) and the combined organic extracts were washed with brine (1 × 200 mL), dried (Na₂SO₄), and filtered through a pad of Florisil[®]. The filtrate was concentrated, and the resulting residue was purified by chromatography to afford the allylic alcohol.



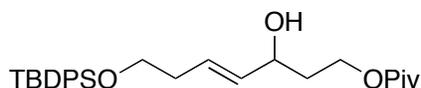
(3*R*,*E*)-7-Benzyloxy-3-hydroxyhept-4-enyl pivalate (8). The racemic allylic alcohol **8** was prepared according to the general procedure from alkyne **6**⁵ (3.09 g, 19.3 mmol) and aldehyde **7** (3.66 g, 23.2 mmol). Chromatography (5:1 hexanes:EtOAc) gave 3.9 g (63%) of a colorless oil. The racemic material was subjected to a kinetic resolution using the procedure of Sharpless.¹⁰ Accordingly, a mixture of the allylic alcohol *rac*-**8** (4.06 g, 12.7 mmol), (+)-diisopropyl tartrate (0.89 g, 3.8 mmol), and 3 Å powdered

molecular sieves (1.0 g) in CH₂Cl₂ (50.0 mL) was cooled to -20 °C and then treated with Ti(O-*i*-Pr)₄ (0.72 g, 2.5 mmol). After stirring for 30 min at -20 °C, a solution of TBHP (5.0 M, 1.9 mL) was added to the reaction mixture and stirring at -20 °C was continued for 12-14 h. After >50% conversion was obtained (monitored by ¹H NMR), the reaction was quenched with an aqueous solution (60 mL) of FeSO₄ (1.57 g) and citric acid (0.53 g) and stirred at room temperature for 1 h. The phases were separated and the aqueous phase was extracted with CH₂Cl₂ (2 × 75 mL). The combined organic layers were concentrated to approximately 50 mL and then stirred for 30 min with 30% NaOH (100 mL). The phases were separated and extracted as before, and the combined organic layers were washed with brine, dried (Na₂SO₄), filtered, and concentrated. The crude product was purified by chromatography (5:1 hexanes:EtOAc) to afford 1.72 g (42%) of the allylic alcohol (*R*)-**8** in ≥97% er (determined by ¹H NMR of the (-)-MTPA chloride-derived ester.)¹⁰ [α]_D -3.18 (*c* 0.98, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.22 (s, 9H), 1.81 (q, *J* = 6.3 Hz, 2H), 2.36 (q, *J* = 6.6 Hz, 2H), 3.52 (t, *J* = 6.7 Hz, 2H), 4.13 (m, 2H), 4.26 (dt, *J* = 11.2, 6.6 Hz, 1H), 4.52 (s, 2H), 5.57 (dd, *J* = 15.5, 6.7 Hz, 1H), 5.70 (dt, *J* = 15.5, 6.6 Hz, 1H), 7.32 (m, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ 27.6, 33.0, 36.6, 39.1, 61.7, 70.0, 70.1, 73.3, 128.0, 128.1, 128.8, 128.8, 134.5, 138.7, 179.2; IR (film) 3434, 1724 cm⁻¹; MS (CI) *m/z* 338 (M⁺ + NH₄), 320 (M⁺), 108, 91; HRMS calcd for C₁₉H₃₂NO₄ (M⁺ + NH₄) 338.2331, found 338.2326.



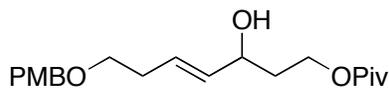
(±)-(E)-9-(Benzyloxy)-3-hydroxynon-4-enyl pivalate (B5). Prepared according to the general procedure from 1-benzyloxy-5-hexyne (**B2**)¹³ (3.10 g, 16.2 mmol) and aldehyde **7** (3.10 g, 19.4 mmol). The crude product was purified by chromatography (5:1 hexanes:EtOAc) to give 3.30 g (58%) of a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 1.22 (s, 9H), 1.48 (m, 2H), 1.64 (m, 2H), 1.83 (m, 2H), 2.01 (br, s, 1H), 2.07 (q, *J* = 7.0 Hz, 2H), 3.48 (t, *J* = 6.4 Hz, 2H), 4.15 (m, 2H), 4.27 (m, 1H), 4.52 (s, 2H), 5.49 (dd, *J* = 6.8, 15.4 Hz, 1H), 5.68 (td, *J* = 6.6, 15.3 Hz, 1H), 7.31 (m, 5H); ¹³C NMR (100.6 MHz,

CDCl₃) δ 26.1, 27.6, 29.7, 32.4, 36.7, 39.1, 61.8, 70.3, 70.6, 73.3, 127.9, 128.0, 128.8, 132.5, 132.7, 139.0, 179.2; IR (film) 3442, 1726, 1478, 1450 cm⁻¹; MS (FAB+) m/z 349 (M⁺ + H), 331, 229, 137; HRMS calcd for C₂₁H₃₆N₀O₄ (M⁺ + NH₄) 366.2644, found 366.2634.



(±)-(E)-7-(tert-Butyldiphenylsilyloxy)-3-hydroxyhept-4-enyl pivalate (B6).

Prepared according to the general procedure from alkyne **B3**⁷ (5.00 g, 16.2 mmol) and aldehyde **7** (3.34 g, 21.1 mmol). The crude product was purified by chromatography (6:1 hexanes:EtOAc) to give 5.71 g (75%) of a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 1.07 (s, 10H), 1.22 (s, 9H), 1.83 (q, J = 6.3 Hz, 2H), 2.31 (q, J = 6.6 Hz, 2H), 3.73 (t, J = 6.6 Hz, 2H), 4.14 (m, 2H), 4.28 (td, J = 6.6, 11.2 Hz, 1H), 5.54 (dd, J = 6.7, 15.4 Hz, 1H), 5.69 (td, J = 6.7, 15.5 Hz, 1H), 7.44 (m, 6H), 7.68 (m, 4H); ¹³C NMR (100.6 MHz, CDCl₃) δ 19.6, 27.2, 27.6, 36.0, 36.6, 39.2, 61.7, 63.8, 70.2, 128.0, 129.2, 130.0, 134.3, 134.4, 136.0, 179.2; IR (film) 3466, 1728, 1710 cm⁻¹; MS (FAB+) m/z 486 (M⁺ + H), 307, 154; HRMS calcd for C₂₈H₄₄N₀O₄Si (M⁺ + NH₄) 486.3040, found 486.3034.



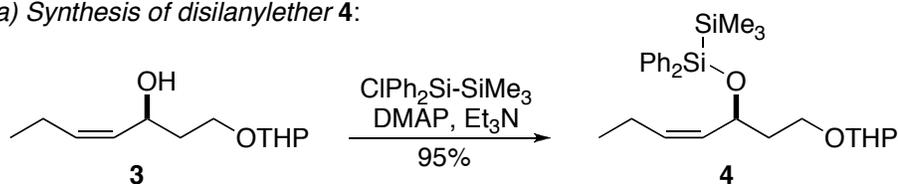
(±)-(E)-3-Hydroxy-7-(4-methoxybenzyloxy)hept-4-enyl pivalate (B7).

Prepared according to the general procedure from alkyne **B4** (2.82 g, 16.17 mmol) and aldehyde **7** (3.60 g, 22.64 mmol). The crude product was purified by chromatography (5:1 to 1:1 hexanes:EtOAc) to give 3.2 g (56%) of a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 1.20 (s, 9H), 1.84 (q, J = 6.3 Hz, 2H), 2.35 (q, J = 6.7 Hz, 2H), 3.49 (t, J = 6.8 Hz, 2H), 3.82 (s, 3H), 4.13 (m, 1H), 4.27 (m, 1H), 4.45 (s, 2H), 5.58 (dd, J = 15.5, 6.6 Hz, 1H),

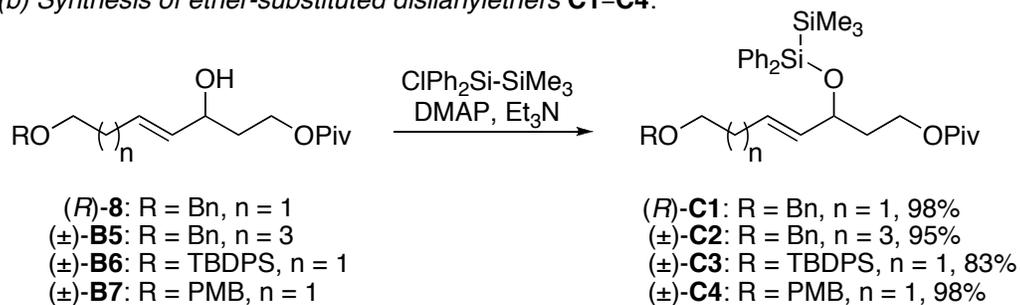
5.70 (dt, $J = 15.5, 6.7$ Hz, 1H), 6.89 (d, $J = 8.6$ Hz, 2H), 7.27 (d, $J = 8.7$ Hz, 2H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 27.2, 32.6, 36.2, 38.7, 55.3, 61.3, 69.3, 69.7, 72.6, 113.8, 128.6, 129.3, 130.4, 134.0, 159.2, 178.8; IR (film) 3410, 1680, 1570, 1480 cm^{-1} ; MS (ES+) m/z 373 ($\text{M}^+ + \text{NH}_4$), 335, 319; HRMS calcd for $\text{C}_{20}\text{H}_{34}\text{NO}_5$ ($\text{M}^+ + \text{NH}_4$) 368.2437, found 368.2416.

Scheme C. Synthesis of disilanyl ethers.

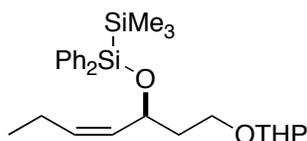
(a) Synthesis of disilanylether **4**:



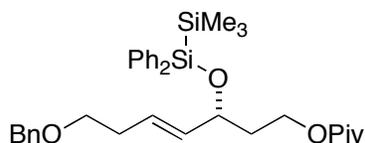
(b) Synthesis of ether-substituted disilanylethers **C1-C4**:



General procedure for the preparation of disilanyl ethers. To a mixture of the allylic alcohol (1.0 equiv), triethylamine (1.5 equiv), and a spatula tip of DMAP in THF was added a solution of 1-chloro-2,2,2-trimethyl-1,1-diphenyl disilane² (1.5 equiv) in THF at 0 °C. The reaction mixture was allowed to warm to room temperature and stirred for 16 h. Upon completion of the reaction, saturated aq NH₄Cl was added and the layers were separated. The aqueous layer was extracted with ether (3 × 60 mL) and the combined organic extracts were washed with brine, dried (Na₂SO₄), filtered, and concentrated. The crude material was purified by chromatography to afford the product disilanylether.



(3*S,Z*)-Tetrahydropyran-2-yloxy-3-(2',2',2'-trimethyl-1',1'-diphenyl-disilyloxy)hept-4-ene (4). Prepared according to the general procedure for the preparation of disilanyl ethers from allylic alcohol **3** (0.72 g, 3.36 mmol). The crude product was purified by chromatography (25:1 hexanes:EtOAc) to give 1.50 g (95%) of a colorless oil. ¹H NMR (400 MHz, CDCl₃) *Mixture of THP ether diastereomers*: δ 0.21 (s, 9H), 0.81 (td, *J* = 6.9, 10.4 Hz, 3H), 1.47-2.03 (m, 10H), 3.46 (m, 2H), 3.82 (m, 2H), 4.48 (td, *J* = 3.7, 44.4 Hz, 1H), 4.72 (m, 1H), 5.28 (qd, *J* = 11.0, 6.9 Hz, 1H), 5.41 (m, 1H), 7.39 (m, 6H), 7.59 (m, 4H); ¹³C NMR (100.6 MHz, CDCl₃) *Mixture of THP ether diastereomers; all peaks listed*: δ -0.8, 14.4, 14.6, 19.8, 20.0, 21.3, 23.1, 25.9, 31.1, 32.0, 38.9, 39.1, 62.4, 62.6, 64.2, 64.3, 67.8, 68.1, 98.9, 99.1, 128.1, 129.7, 132.3, 132.4, 132.7, 132.9, 135.3, 135.4, 137.6; IR (film) 2947, 1428 cm⁻¹; MS (EI) *m/z* 311, 255, 85; HRMS calcd for C₂₇H₄₄Si₂O₃N 486.2860 (M⁺ + NH₄), found 486.2877.



(3*R*,*E*)-7-Benzyloxy-3-(2',2',2'-trimethyl-1',1'-diphenyldisilyloxy)hept-4-enyl pivalate (C1). Prepared according to the general procedure for the preparation of disilanyl ethers from allylic alcohol (*R*)-**8** (4.10 g, 12.8 mmol). Chromatography (25:1 hexanes:EtOAc) afforded 7.33 g (ca. 100%) of a colorless oil. $[\alpha]_D^{+12.4}$ (*c* 0.98, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 0.19 (s, 9H), 1.14 (s, 9H), 1.78 (m, 1H), 1.93 (m, 1H), 2.24 (q, *J* = 6.7 Hz, 2H), 3.35 (m, 2H), 4.10 (t, *J* = 6.5 Hz, 2H), 4.30 (q, *J* = 6.5 Hz, 1H), 4.48 (s, 2H), 5.41 (dt, *J* = 15.4, 6.4 Hz, 1H), 5.48 (dd, *J* = 15.5, 7.0 Hz, 1H), 7.30-7.55 (m, 15H); ¹³C (100.6 MHz, CDCl₃) δ -0.85, 27.6, 33.0, 37.6, 39.0, 61.5, 70.0, 72.6, 73.3, 128.0, 128.1, 128.2, 128.4, 128.8, 129.8, 134.6, 135.3, 135.3, 137.3, 138.8, 178.8; IR (film) 1727 cm⁻¹; MS (FAB+) *m/z* 581.3, 255.1, 154.1; HRMS calcd for C₃₄H₅₀NO₄Si₂ (M⁺ + NH₄) 592.3278, found 592.3257.

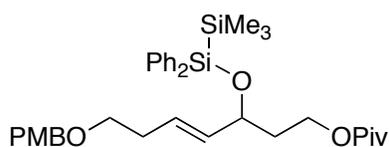


(±)-(E)-9-(Benzyloxy)-3-(2',2',2'-trimethyl-1',1'-diphenyldisilyloxy)non-4-enyl pivalate (C2). Prepared according to the general procedure for the preparation of disilanyl ethers from allylic alcohol **B5** (2.29 g, 3.42 mmol). The crude product was purified by chromatography (25:1 hexanes:EtOAc) to give 3.77 g (95%) of a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 0.20 (s, 9H), 1.15 (s, 9H), 1.34 (m, 2H), 1.56 (m, 2H), 1.78 (m, 1H), 1.94 (m, 3H), 3.45 (t, *J* = 6.5 Hz, 2H), 4.09 (t, *J* = 6.4 Hz, 2H), 4.27 (q, *J* = 6.1 Hz, 1H), 4.52 (s, 2H), 5.39 (m, 2H), 7.36 (m, 10H), 7.55 (m, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ -0.8, 26.0, 27.6, 29.7, 32.3, 37.7, 39.0, 61.6, 70.6, 72.7, 73.3, 127.9, 128.0, 128.1, 128.8, 132.2, 132.8, 135.3, 135.3, 137.4, 139.1, 178.8; IR (film) 1731, 1481

cm⁻¹; MS (FAB+) *m/z* 620 (M⁺ + NH₄), 370, 255; HRMS calcd for C₃₆H₅₄NO₄Si₂ (M⁺ + NH₄) 620.3591, found 620.3617.



(±)-(E)-7-(tert-Butyldiphenylsilyloxy)-3-(2',2',2'-trimethyl-1,1-diphenyl-disilyloxy)hept-4-enyl pivalate (C3). Prepared according to the general procedure for the preparation of disilanyl ethers from allylic alcohol **B6** (4.0 g, 5.07 mmol). Chromatography (20:1 hexanes:EtOAc) gave 5.13 g (83%) of a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ □□□□□(s, 9H), 1.05 (s, 10H), 1.14 (s, 9H), 1.77 (m, 1H), 1.91 (m, 1H), 2.17 (q, *J* = 6.6 Hz, 2H), 3.51 (q, *J* = 7.0 Hz, 2H), 4.09 (t, *J* = 6.4 Hz, 2H), 4.26 (q, *J* = 6.4 Hz, 1H), 5.42 (m, 2H), 7.33–7.67 (m, 20H); ¹³C NMR (125.8 MHz, CDCl₃) δ -1.3, 15.2, 19.1, 26.7, 27.1, 35.4, 37.2, 38.5, 61.0, 63.3, 65.8, 72.1, 127.5, 127.6, 128.0, 129.3, 129.5, 133.8, 134.0, 134.8, 135.5, 136.8, 178.3; IR (film) 1730, 1427 cm⁻¹; HRMS calcd for C₄₃H₆₂NO₄Si₃ (M⁺ + NH₄) 740.3987, found 740.4001.



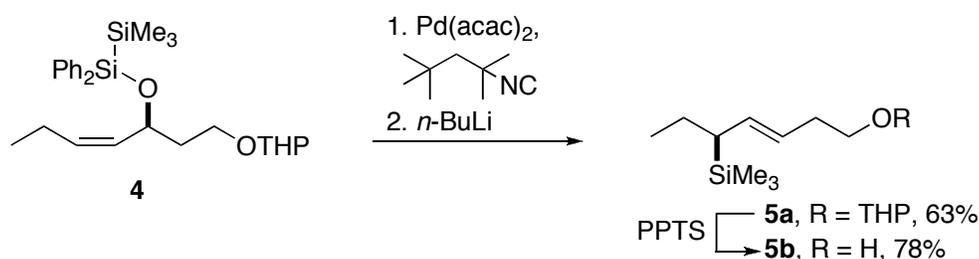
(±)-(E)-7-(4'-Methoxybenzyloxy)-3-(2',2',2'-trimethyl-1,1-diphenyl-disilyloxy)hept-4-enyl pivalate (C4). Prepared according to the general procedure for the preparation of disilanyl ethers from allylic alcohol **B7** (2.32 g, 6.62 mmol). The crude product was purified by chromatography (25:1 hexanes:EtOAc) to give 3.94 g (98%) of a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 0.19 (s, 9H), 1.14 (s, 9H), 1.29 (t, *J* = 7.0 Hz, 2H), 1.78 (m, 1H), 1.92 (m, 1H), 2.22 (q, *J* = 6.8 Hz, 2H), 3.34 (m, 2H), 3.32 (m, 2H), 3.83 (s, 3H), 4.09 (t, *J* = 6.4 Hz, 2H), 4.28 (q, *J* = 6.4 Hz, 1H), 4.41 (s, 2H), 5.41(dt, *J* = 15.5, 6.2 Hz, 1H), 5.47 (dd, *J* = 15.5, 6.9 Hz, 1H), 6.89 (d, *J* = 8.7 Hz, 2H), 7.26 (d, *J* =

8.6 Hz, 2H), 7.40–7.56 (m, 10 H); ^{13}C NMR (100.6 MHz, CDCl_3) δ -1.2, 27.2, 32.6, 37.2, 38.6, 55.3, 61.1, 69.4, 72.2, 72.6, 113.8, 127.76, 128.0, 128.1, 129.3, 129.4, 129.5, 134.0, 134.9, 159.2, 178.4; IR (film) 1710, 1440 cm^{-1} ; MS (ES+) m/z 622 ($\text{M}^+ + \text{NH}_4$), 389, 374, 373; HRMS calcd for $\text{C}_{35}\text{H}_{52}\text{NO}_5\text{Si}_2$ ($\text{M}^+ + \text{NH}_4$) 622.3384, found 622.3376.

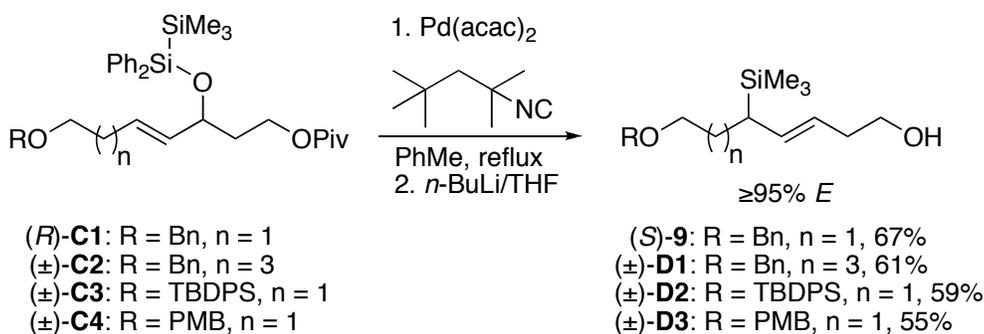
Preparation of Chiral Allylsilanes

Scheme D. Synthesis of chiral allylsilanes.

(a) Synthesis of ethyl-substituted allylsilane **5b**:



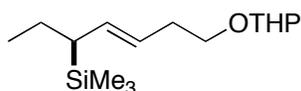
(b) Synthesis of ether-substituted allylsilanes **9**, **D1**, **D2**:



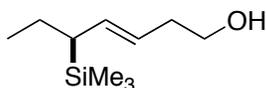
General procedure for the conversion of disilyl ethers to chiral allylsilanes.

According to the procedure of Ito,¹⁴ a mixture of 1,1,3,3-tetramethylbutyl isocyanide (0.41 equiv) and $\text{Pd}(\text{acac})_2$ (0.10 equiv) in toluene was stirred at room temperature under Ar for 5 min, resulting in a red-brown solution. A solution of the disilyl ether (1.0 equiv) in toluene (ca. 5 mL) was then added and the resulting mixture was heated at reflux for 3.5–6 h. After cooling to room temperature, the reaction mixture was filtered through Florisil[®] and concentrated under reduced pressure. The residue obtained was

dissolved in THF and cooled to 0 °C. The resulting solution was charged with *n*-BuLi (2 equiv for pivalate esters; 1.5 equiv for THP ethers), and stirred at 0 °C for 30 min. The reaction was quenched with water (20 mL) and the layers were separated. The aqueous layer was extracted with ether (3 × 30 mL), and the combined organic extracts were washed with brine, dried (Na₂SO₄), filtered, and concentrated. Chromatography of the crude material afforded the *E*-allylsilane product.

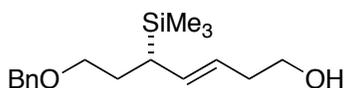


(5*S*,*E*)-1-(Tetrahydropyran-2'-yloxy)-5-trimethylsilylhept-3-ene (5a). Prepared according to the general procedure from disilanyl ether **4** (2.00 g, 4.27 mmol). The crude material was purified by chromatography (30:1 hexanes:EtOAc) to give 0.88 g (76%) of an orange-brown oil. ¹H NMR (500 MHz, CDCl₃) *Mixture of THP ether diastereomers*: δ -0.03 (s, 9H), 0.91 (t, *J* = 6.9 Hz, 3H), 1.32 (m, 2H), 1.58 (m, 5H), 1.72 (m, 1H), 1.85 (m, 1H), 2.32 (q, *J* = 6.7 Hz, 2H), 3.42 (m, 1H), 3.51 (m, 1H), 3.75 (td, *J* = 7.1, 10.8 Hz, 1H), 3.90 (m, 1H), 4.62 (s, 1H), 5.27 (m, 2H); ¹³C NMR (125.8 MHz, CDCl₃) *Mixture of THP ether diastereomers; all peaks listed*: δ -3.22, 14.1, 19.4, 19.5, 21.8, 21.9, 25.1, 30.6, 33.4, 35.3, 62.0, 62.1, 67.7, 67.8, 98.5, 98.6, 124.1, 124.2, 133.5; IR (film) 2956, 1456 cm⁻¹; MS (CI) *m/z* 271 (M⁺ + H), 197, 102; HRMS calcd for C₁₅H₃₄NO₂Si (M⁺ + NH₄) 288.2359, found 288.2362.

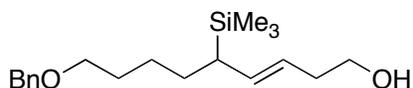


(5*S*, *E*)-5-Trimethylsilylhept-3-en-1-ol (5b). A solution of **5a** (0.55 g, 2.02 mmol) and PPTS (0.10 g, 0.41 mmol) in EtOH (15 mL) was stirred at 55 °C for 4.5 h. After cooling to room temperature, water (5.0 mL) was added to the reaction mixture,

which was extracted with ether (3 × 30 mL). The combined organic extracts were washed with brine, dried (Na₂SO₄), filtered, and concentrated. Chromatography (20:1 to 10:1 hexanes:EtOAc) gave 0.30 g (78%) of a pale yellow oil. $[\alpha]_D +7.8$ (*c* 0.83, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ -0.02 (s, 9H), 0.91 (t, *J* = 6.3 Hz, 3H), 1.30 (m, 2H), 1.52 (m, 2H), 2.30 (q, *J* = 6.6 Hz, 2H), 3.61 (t, *J* = 6.3 Hz, 2H), 5.20 (td, 6.8, 15.2 Hz, 1H), 5.40 (dd, *J* = 7.2, 15.3 Hz, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ -3.2, 14.2, 21.8, 35.5, 36.3, 62.3, 123.4, 135.5; IR (film) 3334, 1451 cm⁻¹; MS (CI) *m/z* 187 (M⁺ + H), 147, 90; HRMS calcd for C₁₀H₂₆NOSi (M⁺ + NH₄) 204.1784, found 204.1779.

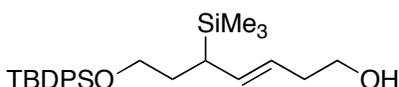


(5*S*,*E*)-7-Benzyloxy-5-trimethylsilylhept-3-en-1-ol (9). Prepared according to the general procedure using disilanyl ether (*R*)-**C1** (1.80 g, 3.18 mmol). Chromatography of the crude product (5:1 hexanes:EtOAc) gave 0.62 g (67%) of an orange-brown oil. $[\alpha]_D +28.4$ (*c* 1.22, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ 0.01 (s, 9H), 1.53 (br s, 1H), 1.64 (m, 2H) 1.84 (dt, *J* = 10.6, 7.7 Hz, 1H), 2.27 (q, *J* = 6.4 Hz, 2H) 3.44 (m, 1H), 3.54 (m, 1H), 3.59 (t, *J* = 6.3 Hz, 2H), 4.52 (AB q, *J* = 12.0, Δ*v* = 31.4 Hz, 2H), 5.22 (dt, *J* = 15.3, 7.0 Hz, 1H), 5.36 (dd, *J* = 15.1, 9.2 Hz, 1H), 7.4 (m, 5H); ¹³C NMR (125.8 MHz, CDCl₃) δ -3.4, 28.7, 29.9, 36.2, 62.2, 69.9, 72.8, 123.8, 127.4, 127.6, 128.3, 134.7, 138.5; IR (film) 3399 cm⁻¹; MS (FAB +) *m/z* 293 (M⁺ + H); HRMS calcd for C₁₇H₂₉SiO₂ (M⁺ + H) 293.1937, found 293.1933.



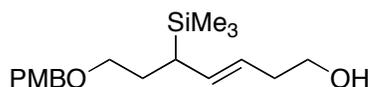
(±)-(*E*)-9-Benzyloxy-5-(trimethylsilyl)non-3-en-1-ol (D1). Prepared according to the general procedure from disilanyl ether **C2** (1.15 g, 1.90 mmol). The crude product was purified by chromatography (6:1 to 3:1 hexanes:EtOAc) to give 0.70 g (61%) of an

orange-brown oil. ^1H NMR (500 MHz, CDCl_3) δ -0.01 (s, 9H), 1.34 (m, 2H), 1.50 (m, 4H), 1.66 (m, 1H), 2.30 (q, $J = 6.5$ Hz, 2H), 3.48 (m, 2H), 3.61 (t, $J = 6.3$ Hz, 2H), 4.52 (s, 2H), 5.21 (td, $J = 7.0, 15.3$ Hz, 1H), 5.34 (dd, $J = 9.6, 15.2$ Hz, 1H), 7.37 (m, 5H); ^{13}C NMR (125.8 MHz, CDCl_3), δ -3.3, 25.8, 28.4, 29.4, 33.2, 36.3, 62.3, 70.3, 72.8, 123.5, 127.4, 127.6, 128.3, 135.4, 138.6; IR (film) 3395, 1454 cm^{-1} ; MS (FAB+) m/z 321 ($\text{M}^+ + \text{H}$), 136; HRMS calcd for $\text{C}_{19}\text{H}_{36}\text{NO}_2\text{Si}$ ($\text{M}^+ + \text{NH}_4$) 338.2515, found 338.2506.



(±)-(E)-7-(tert-Butyldiphenylsilyloxy)-5-(trimethylsilyl)hept-3-en-1-ol (D2).

Prepared according to the general procedure from disilanyl ether **C3** (2.50 g, 3.46 mmol). Chromatography (10:1 to 5:1 hexanes:EtOAc) gave 0.90 g (59%) of an orange-brown oil. ^1H NMR (400 MHz, CDCl_3) δ -0.01 (s, 9H), 1.08 (s, 9H), 1.52 (m, 1H), 1.63 (td, $J = 11.5, 2.2$ Hz, 1H), 1.76 (m, 1H), 2.22 (q, $J = 6.4$ Hz, 2H), 3.55 (m, 2H), 3.62 (m, 1H), 3.73 (m, 1H), 5.11 (dt, $J = 15.3, 6.9$ Hz, 1H), 5.27 (dd, $J = 15.3, 9.3$ Hz, 1H), 7.42 (m, 6H), 7.69 (m, 4H); ^{13}C NMR (100.6 MHz, CDCl_3) δ -2.8, 19.6, 27.3, 29.6, 31.9, 36.7, 62.7, 63.7, 124.0, 128.0, 129.9, 134.5, 135.2, 136.0; IR (film) 3356, 1427 cm^{-1} ; MS (FAB+) m/z 441 ($\text{M}^+ + \text{H}$), 307, 154; HRMS calcd for $\text{C}_{26}\text{H}_{44}\text{NSi}_2\text{O}_2$ ($\text{M}^+ + \text{NH}_4$) 458.2911, found 458.2914.



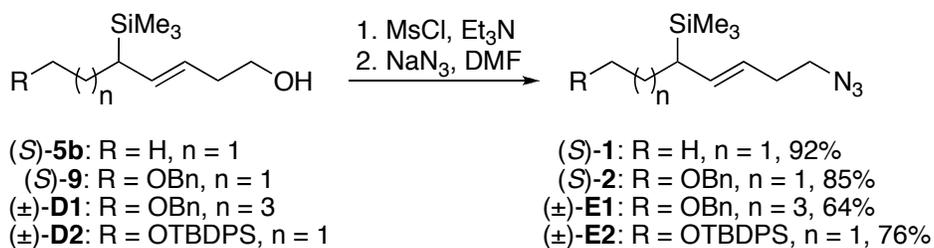
(±)-(E)-7-(4'-Methoxybenzyloxy)-5-(trimethylsilyl)hept-3-en-1-ol (D3).

Prepared according to the general procedure for the conversion of disilanyl ethers to chiral allylsilanes from **C4** (2.0 g, 3.31 mmol). The crude product was purified by chromatography (5:1 hexanes:EtOAc) to give 0.59 g (55%) of an orange-brown oil. ^1H

NMR (400 MHz, CDCl₃) δ -0.103 (s, 9H), 1.61 (m, 2H), 1.80 (m, 1H), 2.27 (q, J = 6.4 Hz, 2H), 3.39 (m, 1H), 3.50 (m, 1H), 3.59 (q, J = 6.0 Hz, 2H), 4.45 (AB q, J = 11.5 Hz, $\Delta\nu$ = 21.8 Hz), 5.20 (dt, J = 15.3, 6.9 Hz, 1H), 5.35 (dd, J = 15.3, 9.0 Hz, 1H), 6.89 (d, J = 8.7 Hz, 2H), 7.28 (d, J = 8.9 Hz, 2H); ¹³C NMR (100.6 MHz, CDCl₃) δ -3.3, 28.8, 30.0, 36.3, 55.3, 62.3, 69.8, 72.6, 113.7, 123.8, 129.3, 130.7, 134.9, 159.1; IR (film) 3400, 1610, 1480 cm⁻¹; MS (ES+) m/z 323 (M⁺ + H), 233, 215; HRMS calcd for C₁₈H₃₁O₃Si (M⁺ + H) 323.2043, found 323.2040.

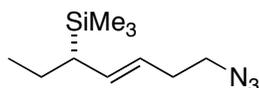
TiCl₄-Promoted Cyclization Reactions

Scheme E. Synthesis of azido allylsilanes.

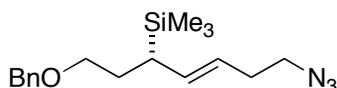


General procedure for the preparation of azido allylsilanes. To a solution of the allylsilyl alcohol (1.0 equiv) and triethylamine (1.5 equiv) in CH₂Cl₂ at 0 °C, was added methanesulfonyl chloride (1.3 equiv). After stirring at 0 °C for 1 h, the reaction was quenched with saturated aqueous NH₄Cl. The layers were separated and the aqueous layer was extracted with CH₂Cl₂ (3 × 20 mL). The combined organic layers were dried (Na₂SO₄), filtered, and concentrated. The crude product was filtered through a pad of silica gel (5:1 hexanes:EtOAc) and the filtrate was concentrated. The mesylate product thus obtained (1.0 equiv) was dissolved in DMF, and sodium azide (4.0 equiv) was added to the resulting solution. The reaction mixture was heated to 110 °C for 2 h. After cooling to room temperature, the reaction mixture was partitioned between water and ether. The layers were separated and the aqueous layer was extracted with ether. The combined

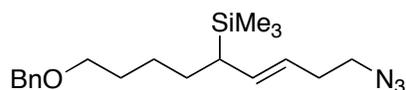
organic layers were washed with brine, dried (Na_2SO_4), filtered, and concentrated. The crude product was purified by chromatography to afford the azido allylsilane.



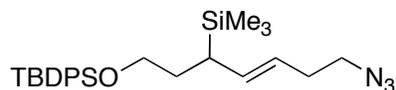
(5*S*, E)-1-Azido-5-trimethylsilylhept-3-ene (1). Prepared according to the general procedure from alcohol **5b** (0.09 g, 0.48 mmol). The crude product was purified by chromatography (20:1 hexanes:EtOAc) to give 0.093 g (92%) of a colorless oil. $[\alpha]_{\text{D}} - 2.64$ (*c* 1.25, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ -0.02 (s, 9H), 0.93 (t, $J = 7.0$ Hz, 3H), 1.35 (m, 2H), 1.55 (m, 1H), 2.34 (q, $J = 6.9$ Hz, 2H), 3.27 (dt, $J = 2.5, 7.0$ Hz, 2H), 5.23 (td, $J = 6.8, 15.2$ Hz, 1H), 5.36 (ddd, $J = 1.1, 8.1, 15.2$ Hz, 1H); ^{13}C (100.6 MHz, CDCl_3) δ -2.8, 14.7, 22.3, 32.8, 35.9, 52.0, 123.7, 135.6; IR (film) 2094, 1726 cm^{-1} ; MS (CI) m/z 184 [$(\text{M}^+ + \text{H}) - \text{N}_2$], 167, 149, 73. It was not possible to obtain a HRMS of the M^+ peak.



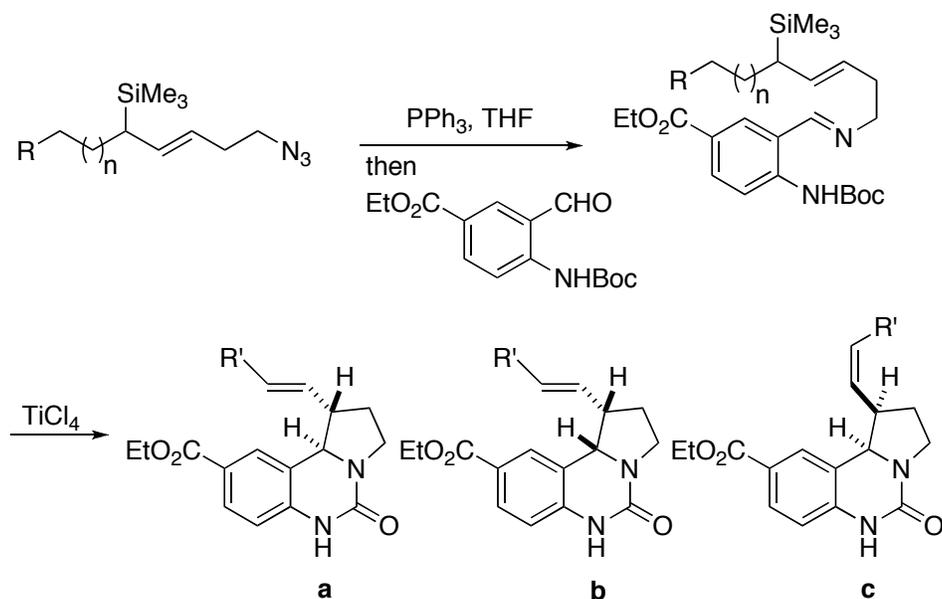
(5*S*, E)-1-Azido-7-benzyloxy-5-trimethylsilylhept-3-ene (2). Prepared according to the general procedure from alcohol **9** (0.58g, 2.0 mmol). The crude product was purified by chromatography (10:1 hexanes:EtOAc) to give 0.54 g (85%) of a colorless oil. $[\alpha]_{\text{D}} + 32.4$ (*c* 0.21, CHCl_3). ^1H NMR (400 MHz, CDCl_3) δ 0.01 (s, 9H), 1.62 (m, 2H), 1.84 (ddd, $J = 12.8, 7.4, 2.3$ Hz, 1H), 2.31 (qd, $J = 6.4, 0.9$ Hz, 2H), 3.24 (t, $J = 7.0$ Hz, 2H), 3.42 (m, 1H), 3.55 (m, 1H), 4.51 (AB q, $J = 11.4$ Hz, $\Delta\nu = 18.5$ Hz), 5.22 (dt, $J = 15.3, 6.8$ Hz, 1H), 5.40 (ddd, $J = 15.2, 8.1, 1.0$ Hz, 1H) 7.4 (m, 5H); ^{13}C NMR (100.6 MHz, CDCl_3) δ -2.9, 29.2, 30.1, 32.7, 51.4, 70.4, 73.4, 124.0, 127.9, 128.0, 128.8, 134.9, 139.1; IR (film) 2100, 1678 cm^{-1} ; MS (CI) m/z 318 ($\text{M}^+ + \text{H}$), 91, 73; HRMS calcd for $\text{C}_{17}\text{H}_{28}\text{SiN}_3\text{O}$ ($\text{M}^+ + \text{H}$) 318.2002, found 318.1984.



(±)-(E)-1-Azido-9-benzyloxy-5-trimethylsilyl-3-nonene (E1). Prepared according to the general procedure from alcohol **D1** (0.23 g, 0.72 mmol). The crude product was purified by chromatography (10:1 hexanes:EtOAc) to give 0.16 g (64%) of a colorless to pale yellow oil. ^1H NMR (500 MHz, CDCl_3) δ -0.01 (s, 9H), 1.27 (m, 1H), 1.35 (m, 1H), 1.45-1.58 (m, 4H), 1.65 (m, 1H), 2.32 (dq, $J = 0.9, 7.0$ Hz, 2H), 3.26 (m, 2H), 3.48 (t, $J = 6.6$ Hz, 2H), 4.52 (s, 2H), 5.23 (td, $J = 6.8, 15.3$ Hz, 1H), 5.36 (dd, $J = 5.9, 15.2$ Hz, 1H), 7.36 (m, 5H); ^{13}C NMR (125.8 MHz, CDCl_3) δ -3.3, 25.9, 28.5, 29.5, 32.3, 33.2, 51.5, 70.4, 72.8, 123.1, 127.4, 127.5, 128.2, 134.3, 135.1, 138.6; IR (film) 2096; MS (FAB+) m/z 318 [$(\text{M}^+ + \text{H}) - \text{N}_2$], 226, 136; HRMS calcd for $\text{C}_{19}\text{H}_{32}\text{N}_3\text{SiO}$ ($\text{M}^+ + \text{H}$) 346.2315, found 346.2315.

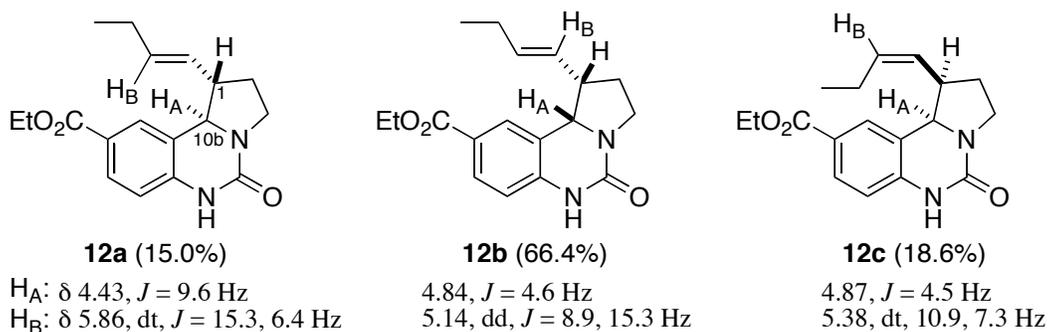


(±)-(E)-1-Azido-7-(tert-butyldiphenylsilyloxy)-5-trimethylsilylhept-3-en-1-ol (E2). Prepared according to the general procedure from alcohol **D2** (0.14 g, 0.32 mmol). Chromatography (10:1 hexanes:EtOAc) gave 0.11 g (76%) of a colorless oil. ^1H NMR (400 MHz, CDCl_3) δ -0.02 (s, 9H), 1.07 (s, 10H), 1.56 (m, 2H), 1.77 (m, 1H), 2.24 (q, $J = 6.7$ Hz, 2H), 3.17 (td, $J = 7.3, 1.6$ Hz, 2H), 3.59 (m, 1H), 3.71 (m, 1H), 5.10 (dt, $J = 15.3, 6.8$ Hz, 1H), 5.27 (dd, $J = 15.2, 9.1$ Hz, 1H), 7.42 (m, 6H), 7.68 (m, 4H); ^{13}C NMR (100.6 MHz, CDCl_3) δ -2.9, 19.6, 27.3, 29.6, 31.9, 32.7, 51.8, 63.8, 123.6, 128.0, 129.9, 134.5, 134.9, 136.0; IR (film) 2095, 1429 cm^{-1} ; MS (FAB+) m/z 466 ($\text{M}^+ + \text{H}$), 438 [$(\text{M}^+ + \text{H}) - \text{N}_2$], 313, 154; HRMS calcd for $\text{C}_{26}\text{H}_{43}\text{N}_4\text{Si}_2\text{O}$ ($\text{M}^+ + \text{NH}_4$) 483.2975, found 483.2970.

Scheme F. $TiCl_4$ -promoted cyclizations of allylsilylimines.

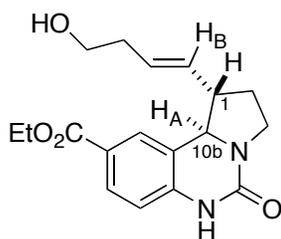
General procedure for $TiCl_4$ -promoted cyclizations of imines. A solution of the azido allylsilane (1.0 equiv) and triphenylphosphine (1.0 equiv) in THF was stirred at room temperature for 3 h. Ethyl 4-(*N*-*tert*-butoxycarbonyl)amino-3-formylbenzoate¹ (1.0 equiv) was then added, and the reaction mixture was heated at reflux for 20 h. After cooling to room temperature, the reaction mixture was concentrated, triturated with ether, filtered through Florisil[®], and the filtrate was concentrated. The crude imine thus obtained was dissolved in CH_2Cl_2 (to form a ca. 0.01 M solution) and the resulting solution was cooled to 0 °C in an ice bath. $TiCl_4$ (5.0 equiv) was then added dropwise to the reaction mixture at 0 °C. Upon complete addition of $TiCl_4$, the ice bath was removed and the reaction mixture was stirred at room temperature for 18 h. The reaction was quenched by the slow addition of a 10% aq NaOH solution, and the mixture was extracted with CH_2Cl_2 . The combined organic layers were washed with brine, dried (Na_2SO_4), filtered, and concentrated. The residue obtained was subjected to chromatography to afford product. 1H NMR integration values of the crude reaction mixtures were used for product ratio determinations (normalized to 100%).

Determination of product stereochemistry and isomer ratios: Chemical shift and coupling constant values for the doublet corresponding to H_A (H10b) of the 1,10b-cis and 1,10b-trans isomers were used for the assignment of relative stereochemistry of the cyclization reaction products. The assignments of doublets corresponding to the cis and trans isomers were made in analogy to previous work and later confirmed through X-ray crystallography (see below).¹ In most cases, the olefin geometry was readily determined by the corresponding coupling constants for the olefinic hydrogens. However, in some examples the corresponding *J* values were obscured in the ¹H NMR for the most minor isomer. In these cases, the olefin geometry was assigned by default.



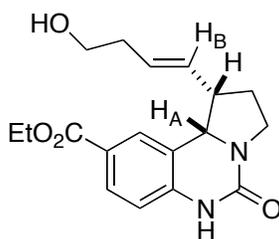
9-Carboethoxy-1-(but-1'-enyl)-2,3,6,10b-hexahydro-1H-pyrrolo[1,2-c]quinazoline-5-one (12a-c). Prepared according to the general procedure from azido allylsilane **1** (0.07 g, 0.34 mmol). Chromatography (1:3 hexanes:EtOAc) gave 0.09 g (85%) of a yellow solid isolated as a mixture of isomers. IR (KBr) 1709, 1673, 1617 cm⁻¹; MS (CI) *m/z* 315 (M⁺ + H), 235, 111; HRMS calcd for C₁₈H₂₃N₂O₃ (M⁺ + H) 315.1709, found 315.1704. Major isomer, **(1S,10bR)-(E)-12b**: ¹H NMR (500 MHz, CDCl₃) δ 0.73 (t, *J* = 7.4 Hz, 3H), 1.39 (t, *J* = 7.1 Hz, 3H), 1.81 (m, 2H), 1.93 (dd, *J* = 7.1, 11.8 Hz, 1H), 2.18 (m, 2H), 3.24 (m, 1H), 3.55 (m, 1H), 3.79 (m, 1H), 4.35 (dq, *J* = 1.6, 7.0, Hz, 2H), 4.84 (d, *J* = 4.6 Hz, 1H), 5.14 (dd, *J* = 8.9, 15.3 Hz, 1H), 5.61 (dt, *J* = 15.3, 6.9 Hz, 1H), 6.84 (d, *J* = 8.4 Hz, 1H), 7.60 (s, 1H), 7.77 (d, *J* = 8.3, 1H), 9.61 (s, 1H); ¹³C NMR (125.8 MHz, CDCl₃) δ 13.7, 14.2, 25.5, 30.1, 43.4, 44.7, 60.5, 61.5, 113.7, 118.1, 123.3, 126.1, 128.7, 129.7, 135.4, 141.8, 153.3, 166.1. **(1R,10bS)-(Z)-12c** (diagnostic peaks only): ¹H NMR (500 MHz, CDCl₃) δ 4.87 (d, *J* = 4.5 Hz, 1H), 5.38 (dt, *J* = 10.9, 7.3 Hz, 1H).

(**1S,10bS**)-(*E*)-**12a** (diagnostic peaks only): ^1H NMR (500 MHz, CDCl_3) δ 4.43 (d, $J = 9.6$ Hz, 1H), 5.86 (dt, $J = 15.3, 6.4$ Hz, 1H).



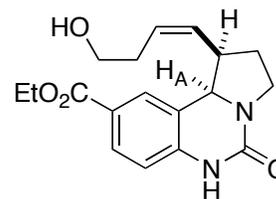
13a (87.3%)

H_A : δ 4.45, $J = 9.7$ Hz
 H_B : δ 5.68, dd, $J = 15.1, 8.9$ Hz



13b (9.8%)

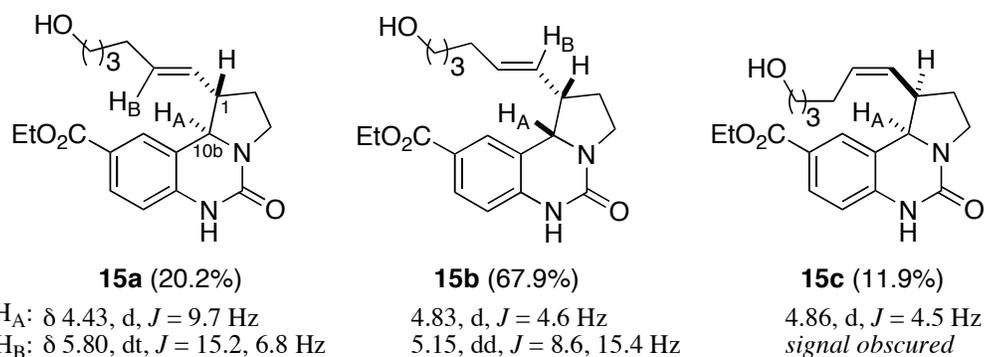
4.81, $J = 4.6$ Hz
 5.26, dd, $J = 9.8, 15.4$ Hz



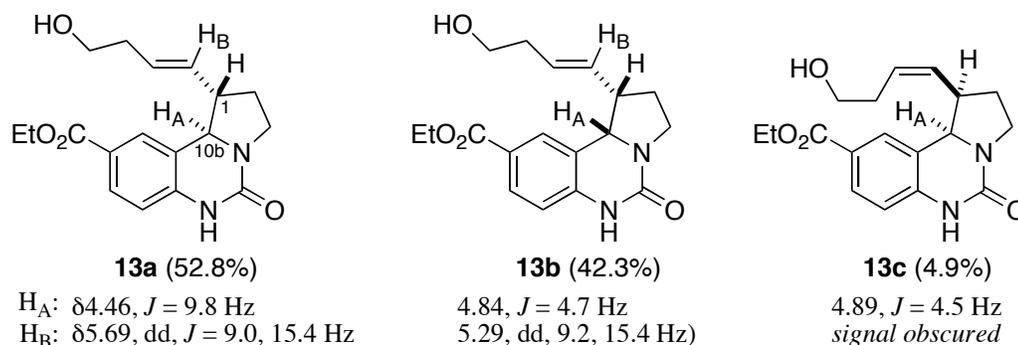
13c (2.9%)

4.85 d, $J = 4.5$ Hz
 signal obscured

9-Carboethoxy-1-(4'-hydroxybut-1'-enyl)-2,3,6,10b-tetrahydro-1H-pyrrolo[1,2-*c*]quinazolin-5-one (13a–c). Prepared according to the general procedure using azido allylsilane **2** (0.10 g, 0.32 mmol). Chromatography (1:3 hexanes:EtOAc) gave 0.085 g (80%) of a yellow solid as a mixture of isomers. The major isomer (**1S,10bS**)-(*E*)-**13a** was isolated by crystallization from the reaction mixture (EtOAc/hexanes, 42%). Major isomer (**1S,10bS**)-(*E*)-**13a**: mp 162–164 °C (EtOAc/hexanes); $[\alpha]_D -22.0$ (c 0.45, CHCl_3). ^1H NMR (500 MHz, CDCl_3) δ 1.38 (t, $J = 7.0$ Hz, 3H), 1.90 (quintet, $J = 11.1$ Hz, 1H), 2.21 (m, 1H), 2.4 (q, $J = 6.1$ Hz, 2H), 2.98 (quintet, $J = 9.5$ Hz, 1H), 3.21 (br, s, 1H), 3.66 (m, 2H), 3.83 (t $J = 5.6$ Hz, 2H), 4.36 (q, $J = 7.1$ Hz, 2H), 4.45 (d, $J = 9.7$ Hz, 1H) 5.68 (dd, $J = 15.1, 8.9$ Hz, 1H), 5.86 (dt, $J = 14.9, 6.7$ Hz, 1H), 6.80 (d, $J = 8.4$ Hz, 1H), 7.83 (d, $J = 8.4$ Hz, 1H), 8.28 (s, 1H), 8.74 (br, s, 1H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 14.3, 30.6, 36.2, 43.5, 50.0, 60.1, 61.0, 61.7, 113.5, 121.1, 123.4, 126.7, 130.0, 131.5, 132.8, 141.3, 152.8, 166.7; IR (KBr) 3216, 1709, 1671, 1615 cm^{-1} ; MS (CI) m/z 331 ($\text{M}^+ + \text{H}$), 232; HRMS calcd for $\text{C}_{18}\text{H}_{22}\text{N}_2\text{O}_4$ ($\text{M}^+ + \text{H}$) 331.1658, found 331.1662. (**1S,10bR**)-(*E*)-**13b** (diagnostic peaks only): ^1H NMR (500 MHz, CDCl_3) δ 4.81 (d, $J = 4.6$ Hz, 1H), 5.26 (dd, $J = 9.8, 15.4$ Hz, 1H). (**1R,10bS**)-(*Z*)-**13c** (diagnostic peaks only): ^1H NMR (500 MHz, CDCl_3) δ 4.85 (d, $J = 4.5$ Hz, 1H).



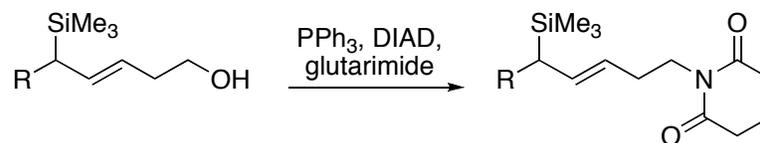
(±)-9-Carboethoxy-1-(6'-hydroxyhex-1'-enyl)-1,2,3,6,10b-tetrahydro-1H-pyrrolo[1,2-c]quinazolin-5-one (15a–c). Prepared according to the general procedure from azido allylsilane **E1** (0.30 g, 0.87 mmol). Chromatography (1:3 hexanes:EtOAc) gave 0.23 g (73%) of a yellow foam as a mixture of isomers. IR (film) 3328, 1675, 1614 cm^{-1} ; MS (CI) m/z 359 ($M^+ + H$), 232; HRMS calcd for $C_{20}H_{27}N_2O_4$ ($M^+ + H$) 359.1971, found 359.1971. Major isomer **(1S,10bR)-(E)-15b**: ^1H NMR (400 MHz, CDCl_3) δ 1.39 (t, $J = 7.1$ Hz, 3H), 1.63 (m, 2H), 1.98 (m, 4H), 2.20 (m, 3H), 3.46 (t, $J = 6.1$ Hz, 2H), 3.67 (m, 2H), 3.81 (m, 1H), 4.35 (m, 2H), 4.83 (d, $J = 4.6$ Hz, 1H), 5.15 (dd, $J = 8.6, 15.4$ Hz, 1H), 5.54 (dt, $J = 15.3, 6.9$ Hz, 1H), 6.78 (d, $J = 8.1$ Hz, 1H), 7.70 (s, 1H), 7.82 (dd, $J = 1.6, 8.4$ Hz, 1H), 9.02 (s, 1H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 14.8, 25.6, 29.6, 32.1, 32.5, 43.7, 45.3, 61.3, 62.2, 63.1, 114.2, 118.6, 124.0, 128.1, 129.5, 130.3, 133.9, 142.2, 153.6, 166.8. **(1S,10bS)-(E)-15a** (diagnostic peaks only): ^1H NMR (400 MHz, CDCl_3) 4.43 (d, $J = 9.7$ Hz, 1H), 5.80 (dt, $J = 15.2, 6.8$ Hz, 1H). **(1R,10bS)-(Z)-15c**: ^1H NMR (400 MHz, CDCl_3) δ 4.86 (d, $J = 4.5$ Hz, 1H).



(±)-9-Carboethoxy-1-(4'-hydroxy-but-1'-enyl)-2,3,6,10b-tetrahydro-1H-pyrrolo[1,2-*c*]quinazolin-5-one (13a–c). Prepared according to the general procedure using azido allylsilane **E2** (0.11 g, 0.24 mmol). Chromatography (1:3 hexanes:EtOAc) gave 0.05 g (64%) of a yellow solid as a mixture of isomers. MS (FAB+) m/z 331 (M^+ + H), 232, 154, 136; HRMS calcd for $C_{18}H_{23}N_2O_4$ 331.1658, found 331.1649. Full product characterization for **13a–c** is given above. (**1S,10bS**)-**13a** (diagnostic peaks only): 1H NMR (500 MHz, $CDCl_3$) δ 4.46 (d, J = 9.8 Hz, 1H), 5.69 (dd, J = 9.0, 15.4 Hz, 1H). (**1S,10bR**)-**13b** (diagnostic peaks only): 1H NMR (500 MHz, $CDCl_3$) δ 4.84 (d, J = 4.7 Hz, 1H), 5.29 (dd, 9.2, 15.4 Hz) 5.63 (m, 1H). (**1R,10bS**)-**13c** (diagnostic peaks only): 1H NMR (500 MHz, $CDCl_3$) δ 4.89 (d, J = 4.5 Hz, 1H).

TFA-Promoted Cyclization Reactions

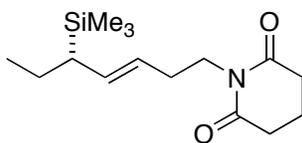
Scheme G. Synthesis of *N*-allylsilyl glutarimides.



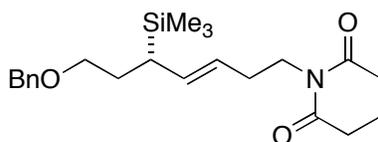
(**S**)-**5b**: R = Et
 (**S**)-**9**: R = $(CH_2)_2OBn$
 (±)-**D1**: R = $(CH_2)_4OBn$
 (±)-**D2**: R = $(CH_2)_2OTBDPS$
D4: R = H
 (±)-**D3**: R = $(CH_2)_2OPMB$

(**S**)-**21**: 82%
 (**S**)-**23**: 80%
 (±)-**25**: 90%
 (±)-**27**: 62%
29: 98%
 (±)-**G1**: 98%

General procedure for the preparation of *N*-allylsilyl glutarimides.¹⁵ To a solution of the allylsilyl alcohol (1.0 equiv), glutarimide (1.5 equiv), and triphenylphosphine (1.5 equiv) in THF at 0°C was added diisopropyl azodicarboxylate (1.5 equiv). The reaction mixture was warmed to room temperature and stirred for 16 h. Water was then added, and the mixture was extracted with ether. The combined organic extracts were washed with brine, dried (Na₂SO₄), filtered, and concentrated. The crude material was purified by chromatography to afford product.

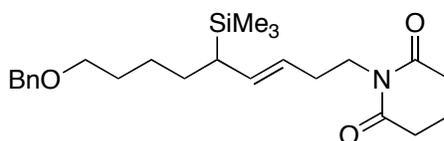


(5'*S*,*E*)-1-[5'-Trimethylsilylhept-3'-enyl]piperidine-2,6-dione (21). Prepared according to the general procedure from alcohol (*S*)-**5b** (0.26 g, 1.40 mmol). Chromatography (5:1 hexanes:EtOAc) gave 0.32 g (82%) of a pale yellow oil. $[\alpha]_D +6.2$ (*c* 0.90, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ -0.05 (s, 9H), 0.88 (t, *J* = 7.1 Hz, 3H), 1.30 (m, 2H), 1.51 (m, 1H), 1.91 (quintet, *J* = 6.7 Hz, 2H), 2.24 (q, *J* = 7.1 Hz, 2H), 2.64 (t, *J* = 6.5 Hz, 4H), 3.82 (m, 2H), 5.21 (m, 2H); ¹³C NMR (100.6 MHz, CDCl₃) δ -2.7, 14.7, 17.6, 22.3, 32.0, 33.3, 35.7, 40.0, 124.5, 134.6, 172.8; IR (film) 1726, 1673 cm⁻¹; MS (CI) *m/z* 282 (M⁺ + H), 90, 73; HRMS calcd for C₁₅H₂₈NsiO₂ (M⁺ + H) 282.1889, found 282.1891.



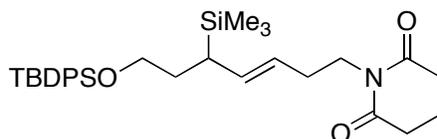
(5'*S*,*E*)-1-[7'-Benzyloxy-5'-trimethylsilylhept-3'-enyl]piperidine-2,6-dione (23). Prepared according to the general procedure from alcohol (*S*)-**9** (0.90 g, 3.0 mmol). Chromatography (6:1 to 3:1 hexanes:EtOAc) gave 0.93 g (80%) of a yellow oil. $[\alpha]_D +17.6$ (*c* 0.68, CHCl₃). ¹H NMR (500 MHz, CDCl₃) δ -0.03 (s, 9H), 1.57 (m, 2H), 1.80

(m, 1H), 1.91 (quintet, $J = 6.7$ Hz, 2H), 2.21 (q, $J = 7.0$, 2H), 2.62 (t, $J = 6.6$ Hz, 4H), 3.39 (m, 1H), 3.51 (ddd, $J = 3.9, 9.0, 17.8$ Hz, 1H), 3.77 (t, $J = 7.6$ Hz, 2H), 4.50 (AB q, $J = 11.4$ Hz, $\Delta\nu = 22.2$ Hz, 2H), 5.20 (dt, $J = 15.2, 6.7$ Hz, 1H), 5.30 (dd, $J = 8.7, 15.3$ Hz, 1H), 7.33 (m, 5H); ^{13}C NMR (125.8 MHz, CDCl_3) δ -3.3, 17.1, 28.7, 29.4, 31.4, 32.8, 39.5, 70.0, 72.8, 124.3, 127.4, 127.5, 128.2, 133.4, 138.6, 172.3; IR (film) 1724, 1675 cm^{-1} ; MS (CI) m/z 388 ($\text{M}^+ + \text{H}$), 298, 91; HRMS calcd for $\text{C}_{22}\text{H}_{34}\text{NO}_3\text{Si}$ 388.2308 ($\text{M}^+ + \text{H}$), found 388.2286.



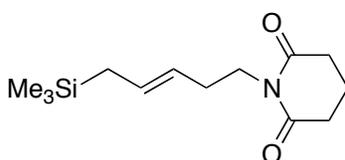
(±)-(E)-1-[9'-Benzyloxy-5'-trimethylsilylnon-3'-enyl]piperidine-2,6-dione (25).

Prepared according to the general procedure from alcohol **D1** (0.24 g, 0.75 mmol). The crude product was purified by chromatography (6:1 to 3:1 hexanes:EtOAc) to give 0.28 g (90%) of a colorless oil. ^1H NMR (500 MHz, CDCl_3) δ -0.05 (s, 9H), 1.28 (m, 2H), 1.43 (m, 4H), 1.55 (m, 1H), 1.64 (m, 1H), 1.89 (m, 2H), 2.25 (q, $J = 6.9$ Hz, 2H), 2.62 (t, $J = 6.4$ Hz, 4H), 3.46 (t, $J = 6.5$ Hz, 2H), 3.80 (m, 2H), 4.51 (s, 2H), 5.18 (dt, $J = 15.2, 6.7$ Hz, 1H), 5.25 (dd, $J = 8.6, 15.3$ Hz, 1H), 7.34 (m, 5H); ^{13}C NMR (100.6 MHz, CDCl_3) δ -2.8, 17.6, 22.1, 26.3, 28.5, 29.0, 30.0, 32.0, 33.3, 40.0, 70.8, 73.2, 124.4, 128.0, 128.7, 134.6, 139.1, 172.8 IR (film) 1727, 1675 cm^{-1} ; MS (CI) m/z 416 ($\text{M}^+ + \text{H}$), 266, 91; HRMS calcd for $\text{C}_{24}\text{H}_{38}\text{NO}_3\text{Si}$ ($\text{M}^+ + \text{H}$) 416.2621, found 416.2622.

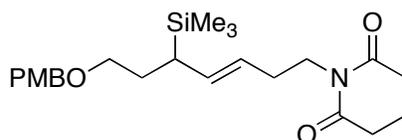


(±)-(E)-1-[7'-(tert-Butyldiphenylsilyloxy)-5'-trimethylsilylhept-3'-enyl]piperidine-2,6-dione (27). Prepared according to the general procedure from alcohol **D2** (0.14 g, 0.36 mmol). Chromatography (3:1 hexanes:EtOAc) afforded 0.12 g (62%) of a

pale yellow oil. ^1H NMR (500 MHz, CDCl_3) δ -0.05 (s, 9H), 1.06 (s, 10H), 1.52 (m, 2H), 1.73 (m, 1H), 1.85 (quintet, $J = 6.4$ Hz, 2H), 2.18 (q, 7.2 Hz, 2H), 2.58 (t, $J = 6.5$ Hz, 4H), 3.59 (m, 1H), 3.73 (m, 3H), 5.12 (dt, $J = 15.3, 6.7$ Hz, 1H), 5.20 (dd, $J = 8.7, 15.3$ Hz, 1H), 7.41 (m, 6H), 7.68 (m, 4H); ^{13}C NMR (125.7 MHz, CDCl_3) δ -3.4, 17.1, 19.1, 21.7, 26.8, 28.8, 31.4, 32.8, 39.4, 63.3, 124.1, 127.5, 129.4, 133.5, 134.3, 135.5, 172.3; IR (film) 1727, 1678 cm^{-1} ; MS (FAB+) m/z 536 ($\text{M}^+ + \text{H}$), 307, 154; HRMS calcd for $\text{C}_{31}\text{H}_{49}\text{N}_2\text{Si}_2\text{O}_3$ ($\text{M}^+ + \text{NH}_4$) 553.3282, found 553.3293.



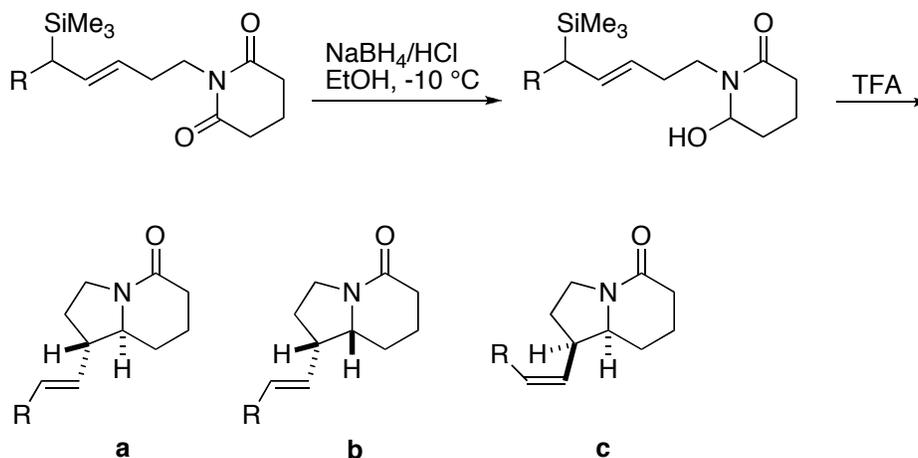
(E)-1-[5'-Trimethylsilylpent-3'-enyl]piperidine-2,6-dione (29). Prepared according to the general procedure using known alcohol **D4** (0.10 g, 0.63 mmol).¹ The crude product was purified by chromatography (5:1 to 1:1 hexanes:EtOAc) to give 0.15 g (94%) of a colorless oil. ^1H NMR (400 MHz, CDCl_3) δ -0.02 (s, 9H), 1.31 (d, $J = 8.0$ Hz, 2H), 1.92 (quintet, $J = 6.5$ Hz, 2H), 2.20 (q, $J = 7.7$ Hz, 2H), 2.64 (t, $J = 6.5$ Hz, 4H), 3.78 (t, $J = 7.5$ Hz, 2H), 5.21 (dt, $J = 15.1, 7.1$ Hz, 1H), 5.44 (td, $J = 8.0, 15.1$ Hz, 1H); ^{13}C NMR (100.6 MHz, CDCl_3) δ -1.6, 17.6, 23.1, 31.8, 33.3, 39.9, 125.0, 129.3, 172.8; IR (film) 1724, 1673 cm^{-1} ; MS (CI) m/z 254 ($\text{M}^+ + \text{H}$) 199, 170, 73; HRMS calcd for $\text{C}_{13}\text{H}_{23}\text{SiNO}_2$ ($\text{M}^+ + \text{H}$) 254.1576, found 254.1573.



(±)-(E)-1-(7'-(4'-Methoxybenzyloxy)-5'-trimethylsilylhept-3'-enyl)piperidine-2,6-dione (G1). Prepared according to the general procedure from **D3** (0.40 g, 1.24 mmol). The crude product was purified by chromatography (3:1 to 2:1 hexanes:EtOAc) to give 0.51 g (98%) of a pale yellow oil. ^1H NMR (400 MHz, CDCl_3) δ

–0.05 (s, 9H), 1.53 (m, 2H), 1.76 (m, 1H), 1.90 (quintet, $J = 6.6$ Hz, 2H), 2.19 (q, $J = 6.7$ Hz, 2H), 2.62 (t, $J = 6.5$ Hz, 4H), 3.35 (m, 1H), 3.46 (m, 1H), 3.75 (t, $J = 7.5$ Hz, 2H), 3.80 (s, 3H), 4.41 (AB q $J = 11.5$ Hz, $\Delta\nu = 17.9$ Hz, 2H), 5.18 (dt, $J = 15.2, 6.5$ Hz, 1H), 5.27 (dd $J = 15.3, 8.6$ Hz, 1H), 6.88 (m, 2H), 7.26 (m, 2H); ^{13}C NMR (100.6 MHz, CDCl_3) δ –3.28, 17.2, 28.7, 29.5, 31.5, 32.9, 39.6, 55.3, 70.0, 72.5, 113.7, 124.4, 129.2, 130.8, 133.5, 159.1, 172.4; IR (film) 1695, 1640, 1475 cm^{-1} ; MS (ES+) m/z 418 ($\text{M}^+ + \text{H}$), 409, 354, 313, 227; HRMS calcd for $\text{C}_{23}\text{H}_{36}\text{NO}_4\text{Si}$ ($\text{M}^+ + \text{H}$) 418.2414, found 418.2431.

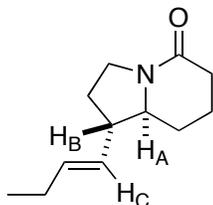
Scheme H. TFA-promoted cyclizations of hydroxy lactams.



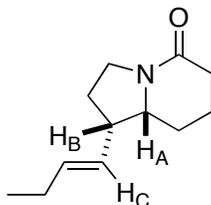
General procedure for TFA-promoted cyclization reactions. According to the procedure of Speckamp.^{15,16} NaBH_4 (4.0 equiv) was added to a solution of the *N*-allylsilyl glutarimide in 100% EtOH at -10°C . At 15 min intervals, 2–3 drops of 2N HCl in 100% EtOH were added while maintaining the temperature at -10°C . After 4 to 6 h (monitored by TLC), the reaction was quenched with water, and the mixture was extracted with CH_2Cl_2 (3×20 mL). The combined organic extracts were dried (Na_2SO_4), filtered, and concentrated. The crude product was passed through a short column of silica gel (EtOAc) to afford a pale yellow oil. The hydroxy lactam thus obtained was dissolved in CH_2Cl_2 (to give a 0.01 to 0.005 M solution), and the resulting solution was cooled to 0°C or -55°C .

CF₃CO₂H (4.0 equiv) was then added dropwise to the reaction mixture and stirring was continued at the indicated temperature for 2h (at 0 °C) or 24 h (at -55 °C). The reaction was quenched with saturated aq NaHCO₃, and the aqueous layer was extracted with CH₂Cl₂. The combined organic layers were washed with brine, dried (Na₂SO₄), filtered, and concentrated. Chromatography afforded lactam product as a mixture of isomers. ¹H NMR integration values of the crude reaction mixtures were used for product ratio determinations (normalized to 100%).

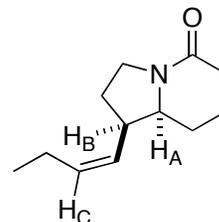
Determination of product stereochemistry and isomer ratios: The assignment of relative configuration for the lactam products was based on the ¹H NMR chemical shifts and coupling patterns of the 1H and 8aH peaks. Chemical shift values for these hydrogens were established by HMQC and ¹H-¹H COSY NMR. The chemical shift of the C8a hydrogen was highly dependent on the relative configuration of the C1 hydrogen in a manner consistent with reports by Speckamp for bicyclic lactams of similar structure.^{15, 17} The C8a hydrogen absorption of the cis diastereomer occurred between 3.54 and 3.57 ppm, while that of the trans isomer occurred between 3.04 and 3.06 ppm. The cis diastereomer also displayed two C1 hydrogen absorptions centered around 3.15 and 2.73 ppm that were established as arising from Z and E olefin isomers, respectively. Diastereomeric and olefin geometric ratios were determined by ¹H NMR integration values.

**22a** (7.9%)

H_A: δ 3.04, td, *J* = 10.6, 3.4 Hz
 H_C: δ 5.27, dd, *J* = 7.9, 15.3 Hz

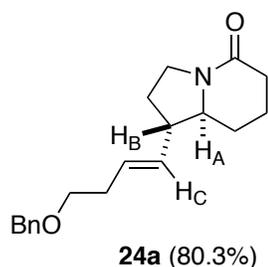
**22b** (28.2%)

H_B: δ 2.75, quint, *J* = 5.5 Hz
 H_C: δ 5.19, dd, *J* = 5.7, 15.3 Hz

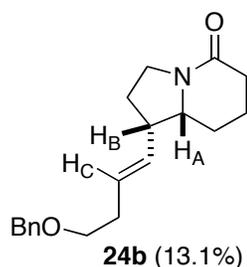
**22c** (63.9%)

H_A: δ 3.54, dt, *J* = 11.5, 4.4 Hz
 H_B: δ 3.11, quint, *J* = 5.5 Hz
 H_C: δ 5.10, t, *J* = 1.5, 10.7 Hz

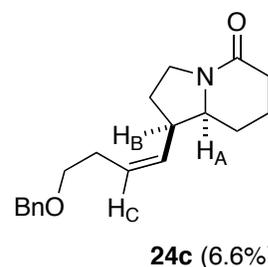
1-(But-1'-enyl)hexahydroindolizin-5-one (22a–c). Prepared according to the general procedure from (*S*)-**21** (0.11 g, 0.39 mmol); cyclization was done at $-55\text{ }^{\circ}\text{C}$. Chromatography (EtOAc) gave 0.070 g (92%) of a colorless oil as a mixture of isomers. IR (film) 1632, 1462 cm^{-1} ; MS (CI) m/z 194 ($\text{M}^+ + \text{H}$), 111, 55; HRMS calcd for $\text{C}_{12}\text{H}_{20}\text{NO}$ ($\text{M}^+ + \text{H}$) 194.1545, found 194.1552. Major isomer (**1R,8aS**)-(*Z*)-**22c**: ^1H NMR (500 MHz, CDCl_3) δ 0.98 (t, *J* = 7.5 Hz, 3H), 1.35 (m, 1H), 1.64–1.81 (m, 3H), 1.92–2.09 (complex, 4H), 2.24 (m, 1H), 2.40 (dd, *J* = 6.1, 17.8 Hz, 1H), 3.11 (quintet, *J* = 5.5 Hz, 1H), 3.49 (m, 1H), 3.54 (dt, *J* = 11.5, 4.4 Hz, 1H), 3.63 (ddd, *J* = 8.5, 8.0, 8.1 Hz, 1H), 5.10 (tt, *J* = 1.5, 10.7 Hz, 1H), 5.51 (td, *J* = 7.6, 10.3 Hz, 1H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 14.2, 20.8, 25.2, 29.2, 31.1, 39.7, 43.2, 61.7, 126.0, 133.4, 169.4. (**1S,8aR**)-(*E*)-**22b** (diagnostic peaks only): ^1H NMR (500 MHz, CDCl_3) δ 2.75 (quintet, *J* = 5.5 Hz, 1H), 5.19 (dd, *J* = 5.7, 15.3 Hz, 1H), 5.53 (dt, *J* = 15.0, 6.3 Hz, 1H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 13.7, 20.7, 25.5, 28.9, 31.1, 43.4, 45.4, 61.8, 126.2, 134.3. (**1S,8aS**)-(*E*)-**22a** (diagnostic peaks only): ^1H NMR (500 MHz, CDCl_3) δ 3.04 (td, *J* = 10.6, 3.4 Hz, 1H), 5.27 (dd, *J* = 7.9, 15.3 Hz, 1H), 5.61 (dt, *J* = 5.8, 15.3 Hz, 1H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 13.6, 20.9, 25.4, 29.4, 44.1, 49.7, 63.4, 127.3, 134.9.



H_A : δ 3.05, td, $J = 10.6, 3.2$ Hz
 H_C : δ 5.39, ddd, $J = 1.1, 8.0, 15.4$ Hz

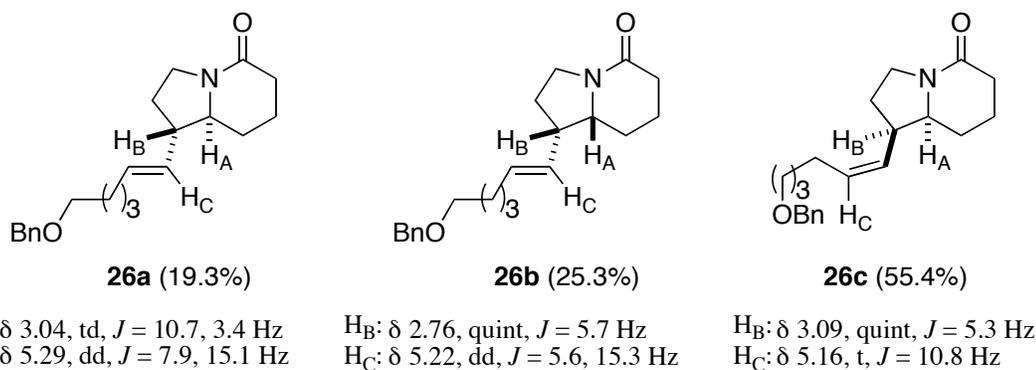


H_B : δ 2.78, quint, $J = 5.5$ Hz
 H_C : δ 5.31, td, $J = 9.5, 15.3$ Hz

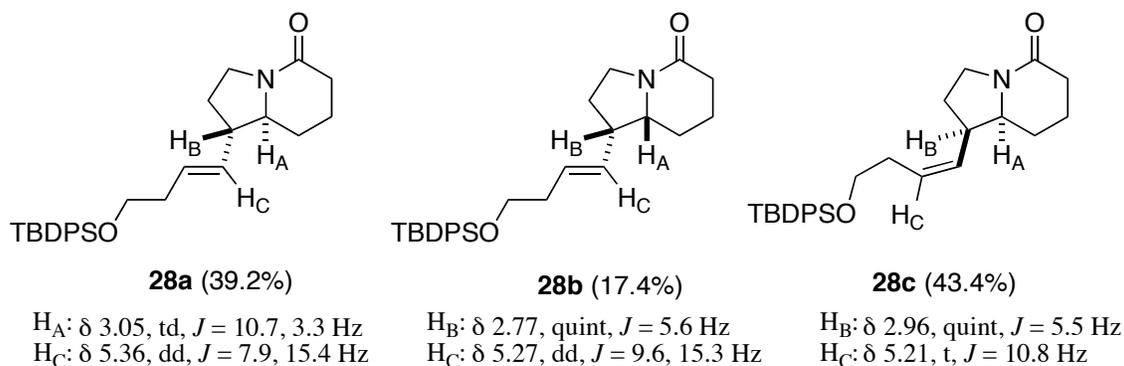


H_B : δ 3.13, quint, $J = 5.5$ Hz
 H_C : δ 5.26, td, $J = 10.6, 1.2$ Hz

1-(4'-Benzyloxy-but-1'-enyl)hexahydroindolizin-5-one (24a–c). Prepared according to the general procedure using **23** (0.25 g, 0.65 mmol); cyclization was done at -55 °C. Chromatography (EtOAc) gave 0.17 g (87%) of a colorless oil as a mixture of isomers. IR (film) 1726, 1644 cm^{-1} ; MS (CI) m/z 300 ($M^+ + H$), 111, 91; HRMS calcd for $C_{19}H_{26}NO_2$ ($M^+ + H$) 300.1964, found 300.1951. Major isomer **(1S,8aS)-(E)-24a**: 1H NMR (500 MHz, $CDCl_3$) δ 1.17 (dq, $J = 1.9, 11.0$ Hz, 1H), 1.64 (m, 2H), 1.95 (m, 1H), 2.06 (m, 2H), 2.26 (m, 2H), 2.38 (q, $J = 6.7$ Hz, 2H), 2.45 (dd, $J = 6.6, 18.0$ Hz, 1H), 3.05 (td, $J = 10.6, 3.2$ Hz, 1H), 3.53 (t, $J = 7.4$ Hz, 4H), 4.53 (s, 2H), 5.39 (ddd, $J = 1.1, 8.0, 15.4$ Hz, 1H), 5.62 (td, $J = 6.8, 15.4$ Hz, 1H), 7.34 (m, 5H); ^{13}C NMR (125.8 MHz, $CDCl_3$) δ 20.9, 27.4, 29.3, 31.0, 33.0, 44.1, 49.8, 63.3, 69.7, 72.8, 127.5, 127.5, 128.3, 129.5, 130.4, 138.3, 169.1. **(1R,8aS)-(Z)-24c** (diagnostic peaks only): 1H NMR (500 MHz, $CDCl_3$) δ 1.35 (m, 1H), 3.13 (quintet, $J = 5.5$ Hz, 1H), 5.26 (td, $J = 10.6, 1.2$ Hz, 1H); **(1S,8aR)-(E)-24b** (diagnostic peaks only): 1H NMR (500 MHz, $CDCl_3$) δ 2.78 (quintet, $J = 5.5$ Hz, 1H), 5.31 (dd, $J = 9.5, 15.3$ Hz, 1H).



(±)-1-(6'-Benzyloxyhex-1'-enyl)hexahydroindolizin-5-one (26a–c). Prepared according to the general procedure from **25** (0.10, 0.24 mmol); cyclization was done at 0 °C. Chromatography (20:1 EtOAc:MeOH) gave 0.070 g (82%) of a colorless oil as a mixture of isomers. IR (film) 1639, 1453 cm^{-1} ; MS (FAB+) m/z 328 ($M^+ + H$), 236, 154; HRMS calcd for $C_{21}H_{30}NO_2$ ($M^+ + H$) 328.2277, found 328.2261. Major isomer (**1R*,8aS***)-**(Z)-26c**: ^1H NMR (400 MHz, CDCl_3) δ 1.32 (m, 2H), 1.45 (quintet, $J = 7.7$ Hz, 2H), 1.65 (m, 4H), 1.76 (m, 1H), 1.94 (m, 2H), 2.03 (m, 3H), 2.24 (m, 1H), 2.41 (m, 1H), 3.09 (quintet, $J = 5.3$ Hz, 1H), 3.47 (t, $J = 6.3$ Hz, 2H), 3.63 (m, 1H), 4.52 (s, 2H), 5.16 (t, $J = 10.8$ Hz, 1H), 5.47 (dt, $J = 10.4, 6.2$ Hz, 1H), 7.33 (m, 5H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 21.3, 25.7, 26.7, 27.7, 29.6, 31.5, 40.2, 43.7, 50.2, 62.2, 70.6, 73.3, 127.4, 127.9, 128.7, 132.0, 132.9, 139.0, 169.9. (**1S*,8aR***)-**(E)-26b** (diagnostic peaks only): ^1H NMR (400 MHz, CDCl_3) δ 2.76 (quintet, $J = 5.7$ Hz, 1H), 5.22 (dd, $J = 5.6, 15.3$ Hz, 1H). (**1S*,8aS***)-**(E)-26a** (diagnostic peaks only): ^1H NMR (400 MHz, CDCl_3) δ 3.04 (td, $J = 10.7, 3.4$ Hz, 1H), 5.29 (dd, $J = 7.9, 15.1$ Hz, 1H), 5.58 (dt, $J = 15.3, 6.7$ Hz, 1H).



(±)-1-[4'-(*tert*-Butyldiphenylsilyloxy)but-1'-enyl]hexahydroindolizin-5-one

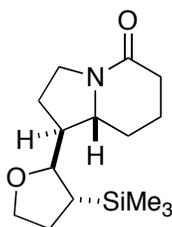
(28a–c). Prepared according to the general procedure from **27** (0.09 g, 0.15 mmol); cyclization was done at 0 °C. Chromatography (EtOAc) gave 0.05 g (74%) of a pale yellow oil as a mixture of isomers. IR (film) 1641, 1111 cm^{-1} ; MS (FAB+) m/z 448 ($M^+ + H$), 390, 154, 136; HRMS calcd for $C_{28}H_{38}N\text{SiO}_2$ ($M^+ + H$) 448.2672, found 448.2653. Major isomer (**1R*,8aS***)-(**Z**)-**28c**: ^1H NMR (500 MHz, CDCl_3) δ 1.06 (s, 9H), 1.28 (m, 1H), 1.63 (m, 4H), 1.87–2.45 (m, 5H), 2.96 (quintet, $J = 5.5$ Hz, 1H), 3.45–3.74 (m, 5H), 5.21 (t, $J = 10.8$ Hz, 1H), 5.54 (dt, 10.2, 7.7 Hz, 1H), 7.41 (m, 6H), 7.69 (m, 4H); ^{13}C NMR (125.7 MHz, CDCl_3) δ 19.1, 20.8, 25.2, 26.7, 27.5, 29.1, 31.0, 35.9, 39.8, 43.2, 49.8, 61.6, 63.4, 127.5, 129.5, 133.8, 135.5, 169.4. (**1S*,8aR***)-(**E**)-**28b** (diagnostic peaks only): ^1H NMR (500MHz, CDCl_3) δ 2.77 (quintet, $J = 5.6$ Hz, 1H), 5.27 (dd, $J = 9.6, 15.3$ Hz, 1H); ^{13}C NMR (127.5 MHz, CDCl_3) δ 19.1, 20.9, 28.9, 43.4, 63.6, 127.7, 135.5. (**1S*,8aS***)-(**E**)-**28a** (diagnostic peaks only): ^1H NMR (500 MHz, CDCl_3) δ 3.05 (td, $J = 10.7, 3.3$ Hz, 1H), 5.36 (dd, $J = 7.9, 15.4$ Hz, 1H), 5.61 (dt, $J = 15.2, 6.9$ Hz, 1H); ^{13}C NMR (127.5 MHz, CDCl_3) δ 19.2, 20.9, 29.3, 44.2, 61.8, 63.4, 127.5, 129.5, 133.7, 135.5, 169.2.



H_A : signal obscured
 H_B : δ 3.08, td, $J = 10.5, 3.2$ Hz

H_A : δ 3.56, dt, $J = 9.0, 4.9$ Hz
 H_B : δ 2.81, quint, $J = 5.7$ Hz

(±)-1-Vinylhexahydroindolizin-5-one (30a,b). Prepared according to the general procedure from **29** (0.23 g, 0.91 mmol); cyclization was done at 0 °C. Chromatography (20:1 EtOAc:MeOH) gave 0.11 g (71%) of a pale yellow oil as a mixture of isomers. IR (film) 1637, 1465, 1447, 1410 cm^{-1} ; MS (CI) m/z 166 ($M^+ + 1$), 150, 111; HRMS calcd for $C_{10}H_{16}NO$ ($M^+ + H$) 166.1232, found 166.1230. Major isomer (**1R*,8aS***)-**30b**: ^1H NMR (500 MHz, CDCl_3) δ 1.35 (m, 1H), 1.69 (m, 1H), 1.83 (m, 2H), 1.93-2.03 (complex, 2H), 2.27 (m, 1H), 2.42 (dd, $J = 4.8, 14.3$ Hz, 1H), 2.81 (quintet, $J = 5.7$, 1H), 3.50 (t, $J = 8.3$ Hz, 1H), 3.56 (dt, $J = 9.0, 4.9$ Hz, 1H), 3.65 (q, $J = 8.3$ Hz, 1H), 5.10 (dd, $J = 1.5, 11.5$ Hz, 2H), 5.63 (m, 1H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 20.9, 25.5, 28.5, 31.3, 43.3, 46.6, 61.6, 116.5, 135.9, 169.4. (**1R*,8aR***)-**30a** (diagnostic peak only): ^1H NMR (500 MHz, CDCl_3) δ 3.08 (td, $J = 10.5, 3.2$ Hz, 1H).

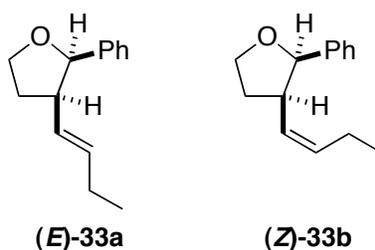


(1S*,8aR*2'S*,3'R*)-1-(3'-Trimethylsilyltetrahydrofuran-2'-yl)hexahydroindolizin-5(1H)-one (37). Prepared according to the general procedure from **G1** (0.28 g, 0.68 mmol); cyclization was done at 0 °C. The crude product was purified by chromatography (EtOAc) to give 0.03 g (20%) of a colorless oil. ^1H NMR (400 MHz, CDCl_3) δ 0.05 (s, 9H), 1.19 (m, 1H), 1.26 (m, 1H), 1.54 (m, 1H), 1.64–1.78 (complex, 2H), 1.90 (m, 3H), 2.08 (m, 1H), 2.33 (m, 2H), 2.42 (m, 2H), 3.35 (m, 1H), 3.46 (m, 1H),

3.61 (m, 1H), 3.70 (q, $J = 7.3$ Hz, 1H), 3.76 (dd, $J = 7.7, 4.8$ Hz, 1H), 3.81 (m, 1H); ^{13}C NMR (100.6 MHz, CDCl_3) δ -2.4, 21.1, 26.8, 28.9, 29.8, 30.2, 30.9, 44.1, 50.7, 62.4, 67.4, 83.0, 169.4; IR (film) 1640, 1460 cm^{-1} ; MS (ES+) m/z 282 ($\text{M}^+ + \text{H}$); HRMS calcd for $\text{C}_{15}\text{H}_{28}\text{NO}_2\text{Si}$ 282.1889 ($\text{M}^+ + \text{H}$), found 282.1914.

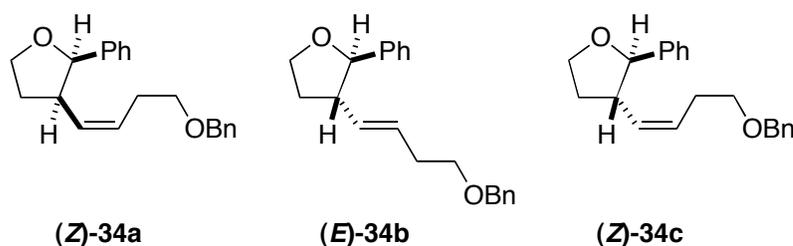
TMSOTf-Promoted Oxenium Cyclization Reactions

General procedure for oxenium ion cyclization reactions. According to the procedure of Ito,¹⁸ TMSOTf (0.83 mmol) was added to a solution of the allylsilyl alcohol (0.75 mmol) and aldehyde (0.83 mmol) in CH_2Cl_2 at -78 °C. After stirring at -78 °C for 2.5 h, a 10% aq NaOH solution (10 mL) was added to the reaction mixture, which was subsequently warmed to room temperature. The layers were separated and the aqueous layer was extracted with CH_2Cl_2 (3×20 mL). The combined organic layers were washed with brine, dried (Na_2SO_4), filtered, and concentrated. The crude reaction mixture was purified by chromatography to afford the corresponding products.



(±)-3-But-1-enyl-2-phenyl tetrahydrofuran (33a,b). Prepared according to the general procedure using allylsilane (±)-**1** (0.080 g, 0.44 mmol) and benzaldehyde (0.050 g, 0.49 mmol). The crude product was purified by chromatography (20:1 hexanes:ether) to give 0.061 g (67%) of a colorless oil as a 2:1 mixture of olefin isomers based on ^1H NMR integration. IR (film) 1604, 1493, 1451 cm^{-1} ; MS (CI) m/z 203 ($\text{M}^+ + \text{H}$), 159, 96, 81; HRMS calcd for $\text{C}_{14}\text{H}_{19}\text{O}$ ($\text{M}^+ + \text{H}$) 203.1436, found 203.1426. Major isomer (**2R*,3R***)-**(E)-33a**: ^1H NMR (500 MHz, CDCl_3) δ 0.80 (t, $J = 7.5$ Hz, 3H), 1.85 (quintet, $J = 6.7$ Hz, 2H), 1.93 (m, 1H), 2.16 (m, 1H), 3.08 (quintet, $J = 7.5$ Hz, 1 H), 4.00 (q, $J =$

8.1 Hz, 1H), 4.30 (m, 1H), 4.83 (dd, $J = 9.0, 15.5$ Hz, 1H), 5.04 (d, $J = 7.2$ Hz, 1H), 5.42 (td, $J = 6.4, 15.3$ Hz, 1H), 7.27 (m, 5H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 13.5, 25.3, 32.2, 47.2, 68.0, 83.5, 126.5, 126.7, 127.6, 128.1, 133.2, 140.6. **(2R*,3R*)-(Z)-1.70b** (diagnostic peaks only): ^1H NMR (400 MHz, CDCl_3) δ 3.43 (m, 1H), 5.25 (dt, $J = 10.8, 7.3$ Hz, 1H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 14.7, 21.2, 33.4, 42.0, 68.4, 83.7, 126.9, 127.2, 128.1, 128.4, 132.8, 141.0.



(±)-3-(4'-Benzyloxybut-1'-enyl)-2-phenyl tetrahydrofuran (34a–c). Prepared according to the general procedure from allylsilane (±)-**2** (0.22 g, 0.75 mmol) and benzaldehyde (0.09 g, 0.83 mmol). The crude product was purified by chromatography (20:1 to 5:1 hexanes:ether) to give 0.13 g (56%) of a colorless oil as a 3:1 (E/Z) mixture of olefin isomers, and 1:12 overall cis:trans (i.e., **a**:(**b+c**) diastereomeric ratio based on ^1H NMR integration. IR (film) 1603, 1494, 1453 cm^{-1} ; MS (CI) m/z 309 ($\text{M}^+ + \text{H}$), 217, 104; HRMS calcd for $\text{C}_{21}\text{H}_{25}\text{O}_2$ ($\text{M}^+ + \text{H}$) 309.1855, found 309.1847. Major isomer **(2R*,3S*)-(E)-34b**: ^1H NMR (500 MHz, CDCl_3) δ 1.97 (m, 1H), 2.28 (m, 1H), 2.36 (q, $J = 6.8$ Hz, 2H), 2.68 (quintet, $J = 8.2$ Hz, 1H), 3.51 (t, $J = 6.8$ Hz, 2H), 4.10 (td, $J = 8.4, 4.3$ Hz, 1H), 4.14 (q, $J = 8.3$ Hz, 1H), 4.51 (d, $J = 8.4$ Hz, 1H), 4.54 (s, 2H), 5.46 (td, $J = 6.7, 15.4$ Hz, 1H), 5.56 (dd, $J = 7.9, 15.4$ Hz, 1H), 7.33 (m, 10 H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 33.0, 33.7, 47.3, 51.7, 68.1, 69.9, 72.8, 125.9, 127.2, 127.4, 127.6, 127.6, 128.1, 128.3, 131.2, 138.4, 141.7. **(2R*,3S*)-(Z)-34c** (diagnostic peaks only): ^1H NMR (500 MHz, CDCl_3) δ 3.11 (quintet, $J = 8.1$ Hz, 1H), 3.30 (t, $J = 6.9$ Hz, 2H), 4.00 (q, $J = 8.1$ Hz, 1H), 4.30 (td, $J = 7.0, 2.7$ Hz, 1H), 4.46 (s, 2H), 5.04 (d, $J = 7.2$ Hz, 1H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 32.1, 32.7, 68.0, 69.9, 72.7, 126.4, 126.7, 128.3, 128.6,

131.3138.4. (**2R***,**3R***)-(**Z**)-**34a** (diagnostic peaks only): ^1H NMR (500 MHz, CDCl_3) δ 5.31 (dt, $J = 11.2, 3.4$ Hz, 1H).

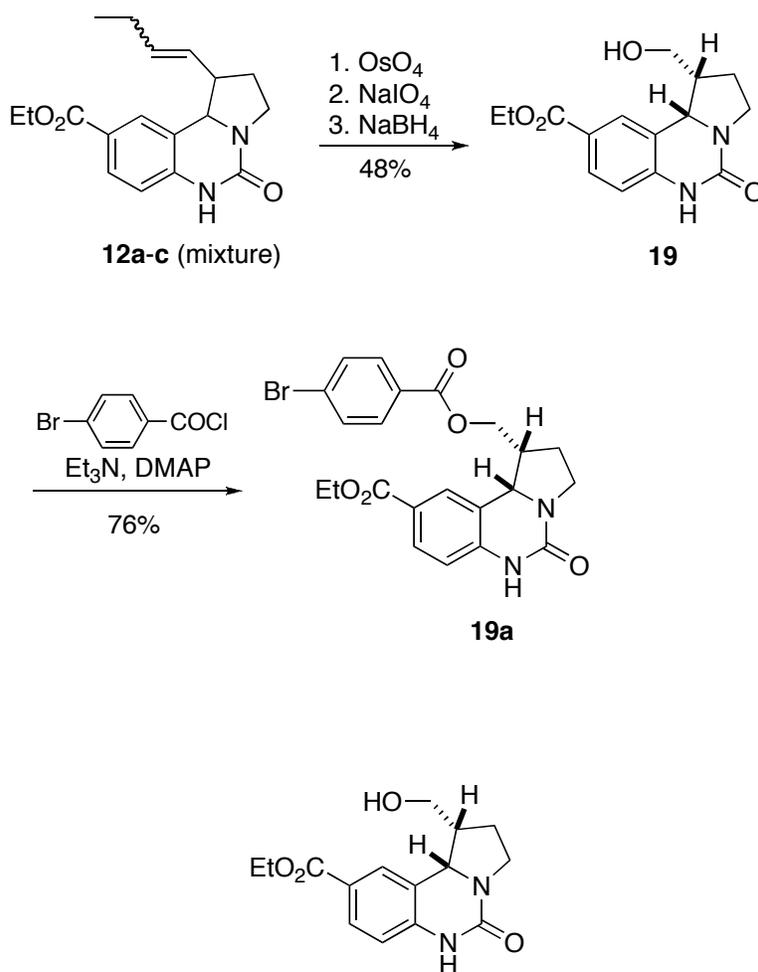
Determination of Absolute Configurations

The absolute configuration of the major isomer obtained in the cyclization reactions employing the enantioenriched azido allylsilanes (*S*)-**1.10** and (*S*)-**1.11** were determined by heavy atom anomalous dispersion X-ray analysis of the corresponding *p*-bromobenzoyl derivatives. In general, preparation of the latter was accomplished by oxidative cleavage of the side chain olefin followed by reduction to the corresponding alcohol and esterification. X-Ray data is given in the Appendix.

General procedure for oxidative cleavage/reduction reactions. OsO_4 (5 mg/mL solution in water, 0.05 equiv) was added to a solution of the olefin (1.0 equiv) and NMO (50% w/v solution in water, 1.5 equiv) in acetone/water/*t*-BuOH (5:5:1, 11 mL total volume), and the resulting mixture was stirred at room temperature for 24 h. Saturated aq NaHSO_3 was then added, and the mixture was stirred for 20 min at room temperature. The reaction mixture was extracted several times (EtOAc), and the combined organic extracts were dried (Na_2SO_4), filtered, and concentrated. The resulting residue was passed through a pad of silica gel (10:1 CH_2Cl_2 :MeOH) to give the corresponding diol. The diol thus obtained (1.0 equiv) was dissolved in 10% aq THF (ca. 10 mL) and cooled to 0 °C. NaIO_4 (1.3 equiv) was then added and the mixture was stirred at 0 °C for 1 h. The reaction mixture was diluted with water (ca. 5 mL) and extracted with EtOAc. The combined organic extracts were dried (Na_2SO_4), filtered through Celite, and concentrated. The resulting residue was dissolved in MeOH (ca. 10 mL) and cooled to 0 °C. NaBH_4 (5.0 equiv) was then added, the ice bath was removed, and stirring at room temperature was continued for 2 h. The reaction was quenched with water and extracted with CH_2Cl_2 . The combined organic extracts were dried (Na_2SO_4), filtered, and concentrated. Chromatography afforded the corresponding alcohol.

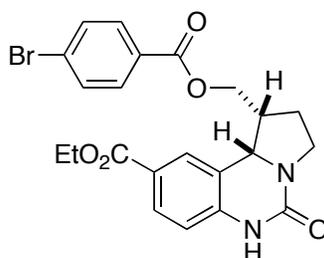
General procedure for the preparation of *p*-bromobenzoyl esters. To a solution of the alcohol (1.0 equiv), triethylamine (3.0 equiv), and a spatula tip amount of DMAP in CH₂Cl₂ (ca. 5.0 mL) at 0 °C, was added 4-bromobenzoyl chloride (1.2 equiv). The reaction mixture was subsequently allowed to warm to room temperature and stirred for 16 h. A saturated aqueous solution of NH₄Cl (10 mL) was then added to the reaction mixture and the aqueous layer was extracted with CH₂Cl₂ (3 × 20 mL). The combined organic extracts were washed with brine (1 × 20 mL), dried (Na₂SO₄), filtered, and concentrated. Chromatography gave the corresponding *p*-bromobenzoyl ester.

Scheme H. Preparation of *p*-bromobenzoyl ester **19a**.

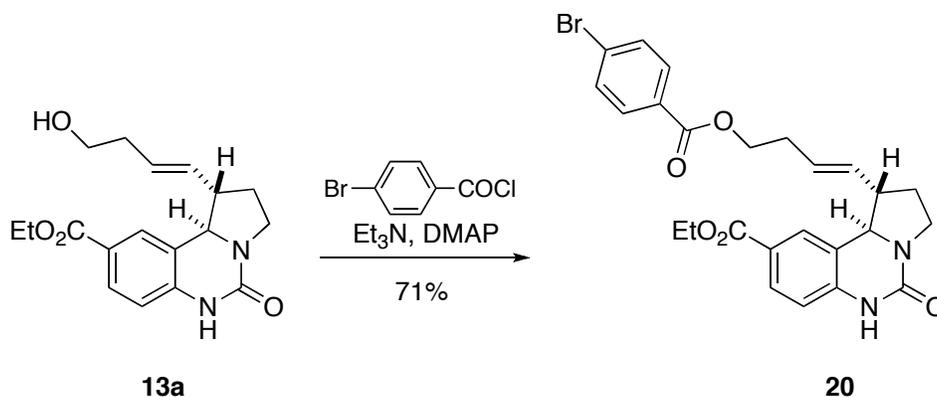


(1*R*,10*aS*)-9-Carboethoxy-1-hydroxymethyl-2,3,6,10*b*-tetrahydro-1*H*-pyrrolo[1,2-*c*]quinazolin-5-one (19**).** Prepared according to the general procedure for

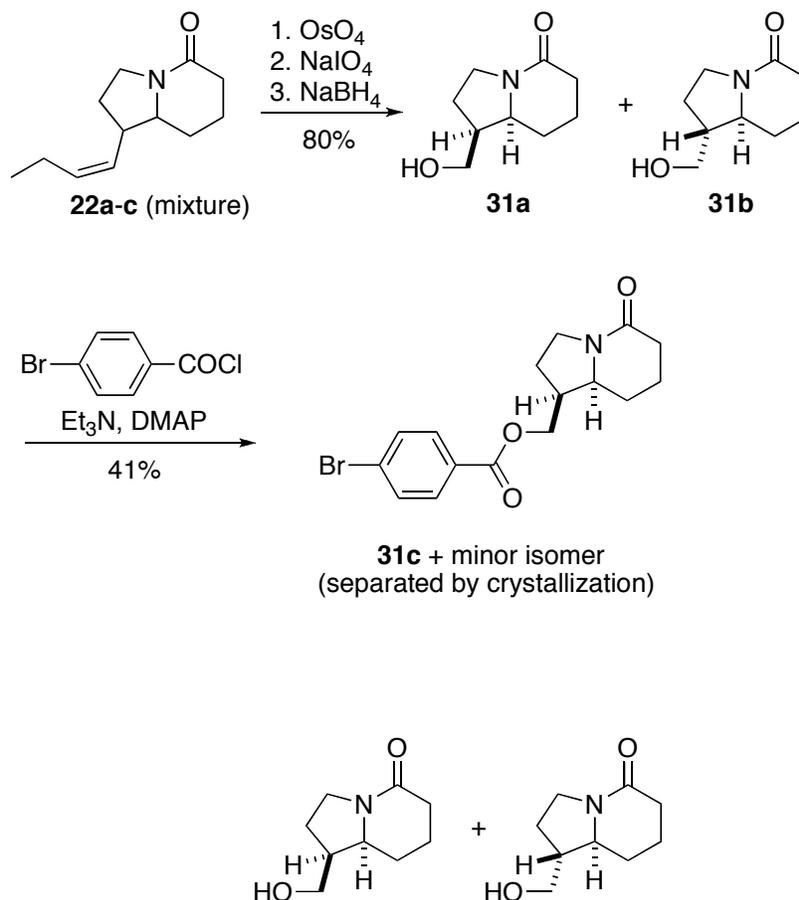
the oxidative cleavage/reduction reaction sequence using the purified product mixture containing **12a–c** (0.18 g, 0.58 mmol). Chromatography (20:1 EtOAc:MeOH) gave 0.081 g (48%) of **19** (isolated as a single isomer) as a white foam. $[\alpha]_D -54.0$ (*c* 0.05, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.40 (t, *J* = 7.1 Hz, 3H), 2.06 (m, 1H), 2.27 (ddd, *J* = 2.2, 8.5, 12.8 Hz, 1H), 2.81 (m, 1H), 3.33 (m, 1H), 3.50 (dt, *J* = 1.2, 10.6 Hz, 1H), 3.69 (m, 1H), 3.87 (m, 1H), 4.36 (t, *J* = 7.6 Hz, 2H), 4.93 (d, *J* = 4.9 Hz, 1H), 6.81 (d, 8.4 Hz, 1H), 7.78 (s, 1H), 7.84 (d, *J* = 8.4 Hz, 1H), 8.62 (br, s, 1H); ¹³C NMR (125.8 MHz, CDCl₃) δ 14.1, 14.3, 24.7, 43.2, 43.3, 60.5, 60.9, 113.9, 117.0, 123.7, 128.0, 130.2, 141.2, 152.4, 166.0; IR (film) 3320, 1669, 1614 cm⁻¹; MS (CI) *m/z* 291 (M⁺ + H), 122, 105; HRMS calcd for C₁₅H₁₉N₂O₄ (M⁺ + H) 291.1345, found 291.1326.



(1R,10aS)-1-(4'-Bromobenzoyloxymethyl)-9-carboethoxy-2,3,6,10b-tetrahydro-1H-pyrrolo[1,2-c]quinazolin-5-one (19a). Prepared according to the general procedure for preparation of *p*-bromobenzoyl esters using **19** (0.06 g, 0.21 mmol). Chromatography (1:2 hexanes:EtOAc) gave 0.070 g (76%) of a white solid that was crystallized from EtOAc/hexanes. Mp 186–188 °C; $[\alpha]_D -95.6$ (*c* 0.25, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.37 (t, *J* = 7.1 Hz, 3H), 2.20 (m, 2H), 3.15 (m, 1H), 3.55 (dt, *J* = 2.7, 10.3 Hz, 1H), 3.96 (q, *J* = 8.9 Hz, 1H), 4.13 (m, 1H), 4.32 (m, 3H), 5.05 (d, *J* = 4.8 Hz, 1H), 6.83 (d, *J* = 8.2 Hz, 1H), 7.47 (d, *J* = 8.5 Hz, 2H), 7.74 (d, *J* = 8.5 Hz, 2H), 7.83 (d, *J* = 8.1 Hz, 2H) 9.11 (s, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 14.8, 25.9, 41.2, 43.6, 61.1, 61.3, 64.1, 114.6, 116.8, 124.5, 128.6, 128.7, 128.8, 130.9, 131.5, 132.0, 141.6, 152.8, 166.0, 166.3; IR (film) 1715, 1670, 1618, 1457; MS (CI) *m/z* 473 (M⁺ + H), 475 (M⁺ + 3), 272, 185; HRMS calcd for C₂₂H₂₂N₂O₅Br (M⁺ + H) 473.0712, found 473.0718. A single crystal was subjected to X-ray analysis.

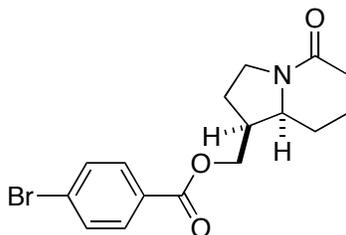


(1*S*,10*bR*)-(E)-9-Carboethoxy-1-[4'-(4''-bromobenzoyloxy)but-1'-enyl]-2,3,6,10*b*-tetrahydro-1*H*-pyrrolo[1,2-*c*]quinazolin-5-one (20). Prepared according to the general procedure for preparation of *p*-bromobenzoyl esters from **13a** (0.070 g, 0.20 mmol). Chromatography (1:1 hexanes:EtOAc to EtOAc) gave 0.069 g (71%) of a pale yellow solid that was crystallized from EtOAc/hexanes. Mp 166-168 °C; $[\alpha]_D -27.2$ (*c* 0.25, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.36 (t, *J* = 7.0 Hz, 3H), 1.88 (quintet, *J* = 10.2 Hz, 1H), 2.20 (m, 1H), 2.65 (q, *J* = 6.7 Hz, 2H), 3.01 (quintet, *J* = 8.9 Hz, 1H), 3.67 (m, 2H), 4.31 (q, *J* = 7.1 Hz, 2H), 4.46 (m, 3H), 5.75 (dd, *J* = 8.5, 15.5 Hz, 1H), 5.89 (td, *J* = 6.8, 15.4 Hz, 1H), 6.80 (d, *J* = 8.3 Hz, 1H), 7.54 (d, *J* = 8.4 Hz, 2H), 7.88 (t, *J* = 6.8 Hz, 3H), 8.11 (s, 1H), 8.42 (br, s, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 14.8, 31.3, 32.5, 44.0, 49.6, 60.9, 61.2, 64.5, 114.0, 121.5, 124.4, 126.7, 128.5, 129.3, 129.5, 130.7, 131.5, 132.1, 133.5, 141.8, 153.3, 166.2, 166.4; IR (KBr) 1714, 1673, 1619, 1591 cm⁻¹; MS (CI) 513 (M⁺ + H), 515 (M⁺ + 3), 232; HRMS calcd for C₂₅H₂₅N₂O₅Br (M⁺) 512.0947, found 512.0938. A single crystal was subjected to X-ray analysis.

Scheme I. Preparation of *p*-bromobenzoyl ester **31c**.

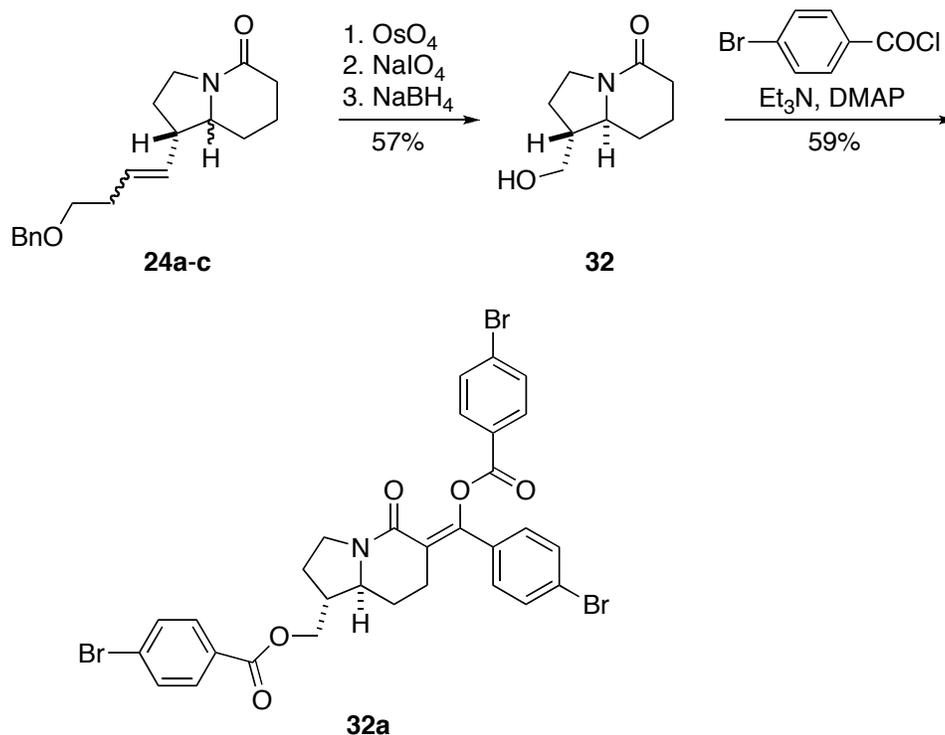
(1*S*,8*aS*)- and (1*R*,8*aS*)-1-Hydroxymethylhexahydroindolizin-5-one (31a,b).
 Prepared according to the general procedure for the oxidative cleavage/reduction reaction sequence using the purified product mixture containing **22a-c** (0.08 g, 0.41 mmol). Chromatography (20:1 CH_2Cl_2 :MeOH) gave 0.061 g (80% over 3 steps) of a colorless oil as a mixture of isomers. IR (film) 3380, 1608, 1470 cm^{-1} ; MS (CI) m/z 170 ($\text{M}^+ + \text{H}$), 111, 82; HRMS calcd for $\text{C}_9\text{H}_{16}\text{NO}_2$ ($\text{M}^+ + \text{H}$) 170.1181, found 170.1188. Major isomer **(1*S*,8*aS*)-31a**: ^1H NMR (400 MHz, CDCl_3) δ 1.48 (dq, $J = 3.1, 11.9$ Hz, 1H), 1.69 (m, 1H), 1.91 (m, 1H), 1.97 (m, 3H), 2.27 (m, 1H), 2.42 (m, 2H), 2.49 (br, s, 1H), 3.45 (m, 2H), 3.55-3.74 (m, 3H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 21.3, 24.8, 28.5, 31.0, 43.4, 43.6, 60.8, 61.0, 169.4. Minor isomer **(1*R*,8*aS*)-31b** (diagnostic peaks only): ^1H NMR

(400 MHz, CDCl_3) δ 1.28 (m, 1H), 3.24 (dt, $J = 3.5, 10.3$ Hz, 1H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 20.9, 25.4, 44.0, 48.2, 61.5, 63.1.



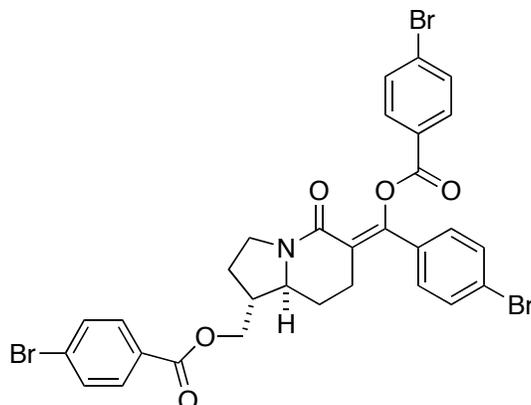
(1S,8aS)-1-(4-Bromobenzoyloxymethyl)hexahydroindolizin-5-one (31c).

Prepared according to the general procedure for preparation of *p*-bromobenzoyl esters using a mixture of **31a** and **31b** (0.060 g, 0.37 mmol). Chromatography (20:1 CH_2Cl_2 : MeOH) gave 0.12 g (92%) of a white solid that was crystallized (EtOAc/hexanes) to afford 0.070 g (56%) of **31c** as a single isomer. Mp 85-87 °C; $[\alpha]_{\text{D}} -2.7$ (c 0.47, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 1.47 (dq, $J = 2.9, 12.8$ Hz, 1H), 1.69 (m, 1H), 2.02 (m, 4H), 2.26 (m, 1H), 2.46 (dd, $J = 6.1, 17.9$ Hz, 1H), 2.68 (quintet, $J = 6.3$ Hz, 1H), 3.52 (dt, $J = 1.8, 9.1$ Hz, 1H), 3.70 (m, 2H), 4.18 (dd, $J = 6.6, 11.3$ Hz, 1H), 4.35 (dd, $J = 6.5, 11.3$ Hz, 1H), 7.60 (d, $J = 8.5$ Hz, 2H), 7.86 (d, $J = 8.5$ Hz, 2H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 21.3, 25.0, 25.4, 31.1, 40.6, 43.4, 60.5, 63.9, 128.3, 128.6, 131.0, 131.8, 165.6, 169.1; IR (film) 1719, 1621, 1590 cm^{-1} ; MS (CI) m/z 352 ($\text{M}^+ + 1$), 354 ($\text{M}^+ + 3$), 151, 82; HRMS calcd for $\text{C}_{16}\text{H}_{19}\text{NO}_3\text{Br}$ ($\text{M}^+ + \text{H}$) 352.0548, found 352.0548. A single crystal was subjected to X-ray analysis.

Scheme J. Preparation of *p*-bromobenzoyl ester **32a.**

(1*R*,8*aS*)-1-Hydroxymethylhexahydroindolizin-5-one (32). Prepared according to the general procedure for the oxidative/reduction reaction sequence using the purified product mixture containing **24a-c** (0.22 g, 0.72 mmol). Chromatography (20:1 CH₂Cl₂:MeOH) gave 0.11 g (86%) of a white solid that was crystallized (EtOAc/hexanes) to afford 0.068 g (57%) of **32** as a single isomer. Mp 114-116 °C; [α]_D -26.4 (*c* 0.55, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.32, (m, 1H), 1.67 (m, 3H), 1.98 (m, 2H), 2.04 (m, 1H), 2.30 (m, 2H), 2.47 (dd, *J* = 6.6, 18.0 Hz, 1H), 3.25 (dt, *J* = 3.4, 10.5 Hz, 1H), 3.51 (td, *J* = 1.5, 11.7 Hz, 1H), 3.66 (m, 1H), 3.77 (dd, *J* = 1.4, 5.4 Hz, 2H); ¹³C NMR (125.8 MHz, CDCl₃) δ 21.0, 25.4, 28.5, 31.0, 44.0, 48.2, 61.5, 63.2, 169.1; IR (film)

3396, 1614 cm^{-1} ; MS (FAB+) m/z 170 ($\text{M}^+ + \text{H}$) 110; HRMS calcd for $\text{C}_9\text{H}_{16}\text{NO}_2$ ($\text{M}^+ + \text{H}$) 170.1181, found 170.1181.

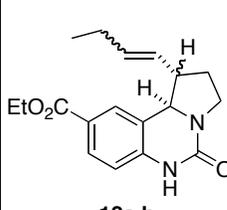
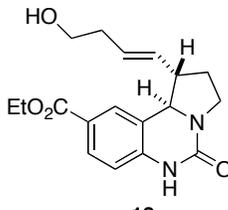
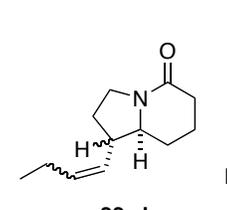
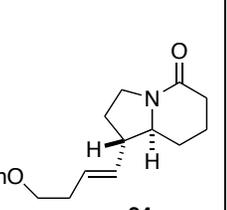
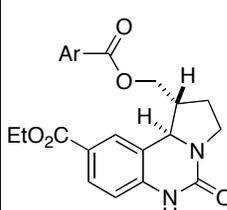
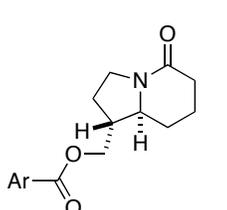
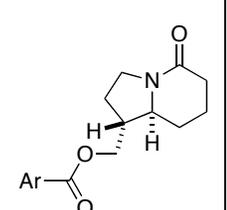


(1R,8aS)-(Z)-6-[4'-Bromobenzoyloxy(4''-bromophenyl)methylene]-1-(4'-bromobenzoyloxymethyl)octahydroindolizin-5-one (32a). Prepared according to the general procedure for preparation of *p*-bromobenzoyl esters from **32** (0.05 g, 0.30 mmol) with a modification of the workup: upon completion of the reaction, the reaction mixture was diluted with EtOAc (10 mL) and washed sequentially with cold 1M HCl (2 × 10 mL), followed by saturated aqueous NaHCO_3 (2 × 10 mL) and brine (1 × 10 mL). The organic layer was dried (Na_2SO_4), filtered, and concentrated. Chromatography (10:1 to 1:1 hexanes:EtOAc) gave 0.14 g (64%) of a white solid that was crystallized (EtOAc/hexanes) to afford 0.080 g (40%) of **32a**. Mp 186-187 °C; $[\alpha]_D -49.4$ (*c* 0.35, CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 1.55 (dq, $J = 4.2, 13.3$ Hz, 1H), 1.74 (quintet, $J = 9.9$ Hz, 1H), 2.16 (quintet, $J = 6.4$ Hz, 1H), 2.26 (m, 2H), 2.60 (dt, $J = 4.3, 13.6$ Hz, 1H), 2.79 (td, $J = 3.5, 15.7$ Hz, 1H), 3.45 (dt, $J = 3.1, 10.8$ Hz, 1H), 3.58 (m, 1H), 3.67 (m, 1H), 4.36 (dd, $J = 6.7, 11.4$ Hz, 1H), 4.46 (dd, $J = 5.1, 11.4$ Hz, 1H), 7.37 (d, $J = 8.3$ Hz, 2H), 7.55 (d, $J = 8.3$ Hz, 2H), 7.62 (dd, $J = 4.1, 8.3$ Hz, 4H), 7.88 (d, $J = 8.3$ Hz, 2H), 8.00 (d, $J = 8.3$ Hz, 2H); ^{13}C NMR (125.8 MHz, CDCl_3) δ 26.0, 27.4, 28.5, 44.5, 45.2, 61.7, 65.0, 118.4, 123.7, 128.4, 128.4, 128.5, 128.6, 130.3, 131.0, 131.6, 131.7, 131.7, 131.8, 134.1, 149.6, 161.4, 164.1, 165.5; IR (film) 1721, 1650, 1590 cm^{-1} ; MS (FAB+) m/z 716 ($\text{M}^+ + \text{H}$), 718 ($\text{M}^+ + 3$), 720 ($\text{M}^+ + 5$), 722 ($\text{M}^+ + 7$), 307, 154; HRMS calcd for $\text{C}_{30}\text{H}_{25}\text{NO}_5\text{Br}_3$ ($\text{M}^+ + \text{H}$) 715.9283, found 715.9296. A single crystal was subjected to X-ray analysis.

Determination of Enantiomeric Purities

The enantiomeric ratios of the major products obtained in the cyclization reactions employing the enantiomerically enriched allylsilanes (*S*)-**1** and (*S*)-**2** were determined by chiral HPLC. In all cases, with the exception of **13a** (which was amenable to analysis without derivatization), the *p*-bromobenzoyl esters were required to facilitate separation of the component enantiomers in the racemic standards. All compound derivatizations for the purpose of HPLC analysis were carried out as described above with the exception that crystallization was avoided throughout to prevent alteration of the enantiomeric purity obtained in the original products.

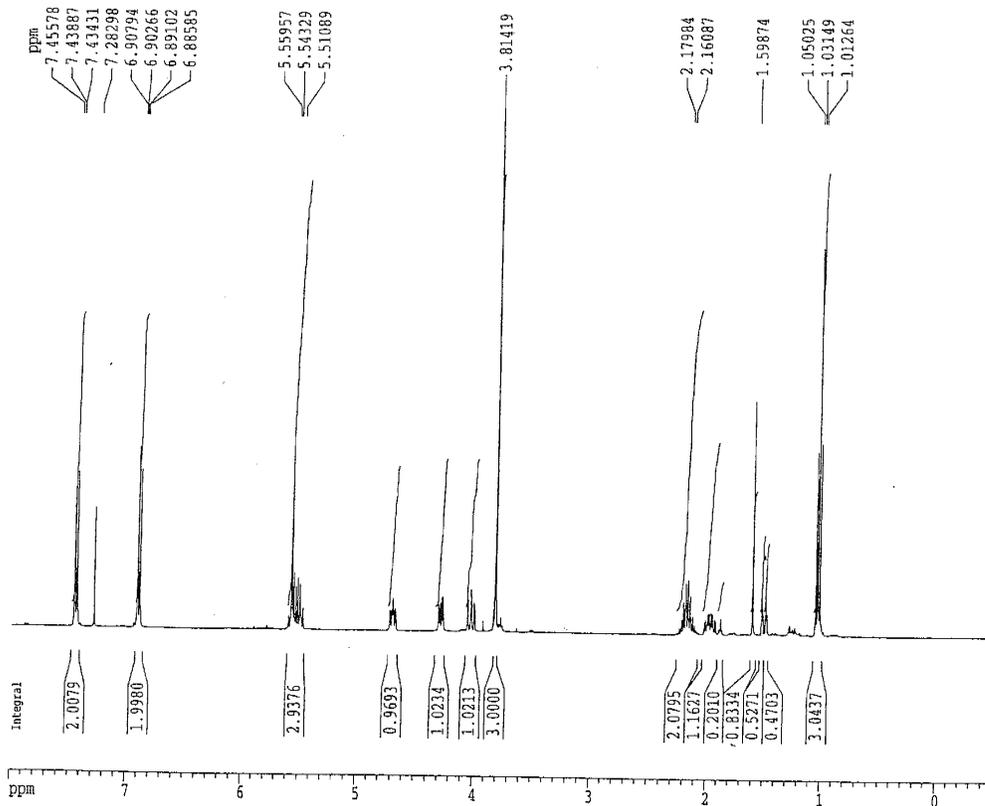
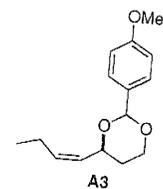
Table A. Summary of er determination.

Parent Compound	 12a,b	 13a	 22a,b	 24a
Derivative Analyzed ^a		N/A		
chiral column ^b	Chiralcel OD-H	Chirobiotic T	Chiralcel OD-H	Chiralcel OD-H
eluent ^c	5-10% IPA/hexanes gradient over 60 min	30% EtOH/hexanes	5-10% IPA/hexanes gradient over 60 min	5-10% IPA/hexanes gradient over 60 min
RT ^d (major isomer)	33.4 min	18.2 min	37.9 min	37.5 min
RT (minor isomer)	40.4 min	20.9 min	34.7 min	35.4 min
er	97.1:2.9 (minor diastereomer)	99:1	93.3:6.7 (minor diastereomer)	97.0:3.0

^aAr = *p*-bromophenyl; ^bDiacel Chemical; ^cIPA = 2-propanol, EtOH = ethanol; ^dRT = retention time

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F2 - Acquisition Parameters

Date_ 20011115
 Time 12.13
 INSTRUM drx400
 PROBHD 5 mm Multinu
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 1.4210291 sec
 RG 181
 DW 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec

=====**CHANNEL f1**=====

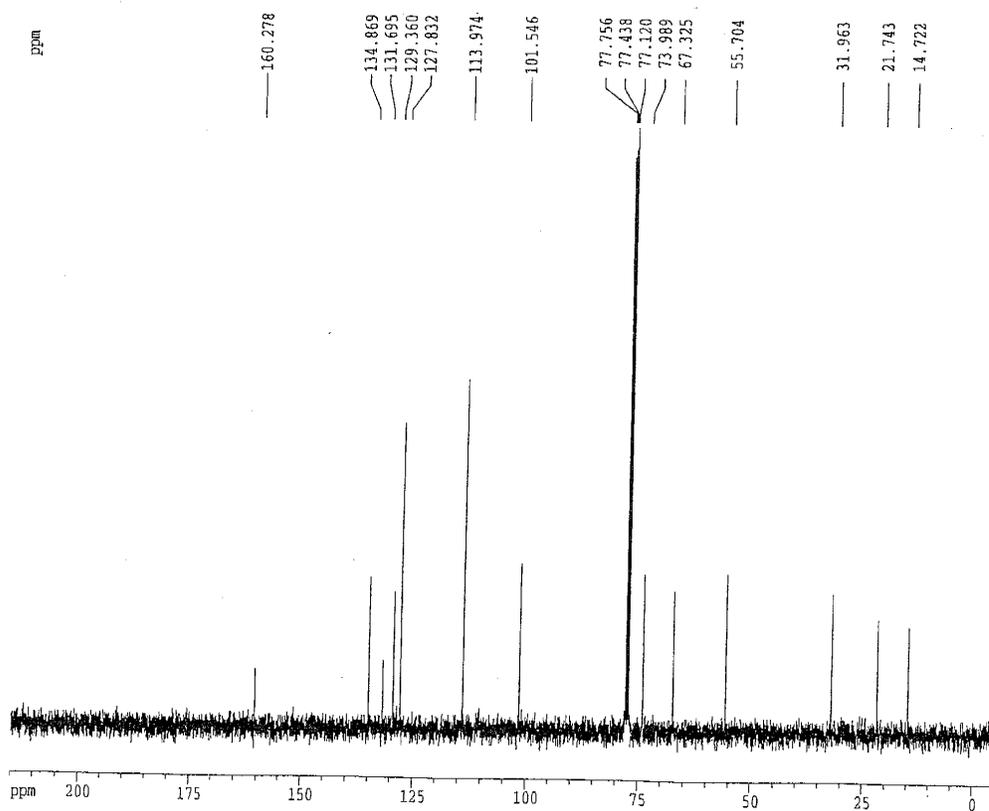
NUC1 1H
 P1 7.70 usec
 PL1 -6.00 dB
 SFO1 400.1320007 MHz

F2 - Processing parameters

SI 16384
 SF 400.1300000 MHz
 HW 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters

CX 20.00 cm
 F1P 8.000 ppm
 F1 3201.04 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.42500 ppm/cm
 HZCM 170.05525 Hz/cm



Current Data Parameters

NAME wj2-128.2
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20011115
 Time 12.20
 INSTRUM drx400
 PROBHD 5 mm Multinu
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 223
 DS 2
 SWH 23140.148 Hz
 FIDRES 0.353213 Hz
 AQ 1.4156276 sec
 RG 4096
 DW 21.600 usec
 DE 4.50 usec
 TE 300.0 K
 D1 0.05000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

=====**CHANNEL f1**=====

NUC1 13C
 P1 12.30 usec
 PL1 2.00 dB
 SFO1 100.6232933 MHz

=====**CHANNEL f2**=====

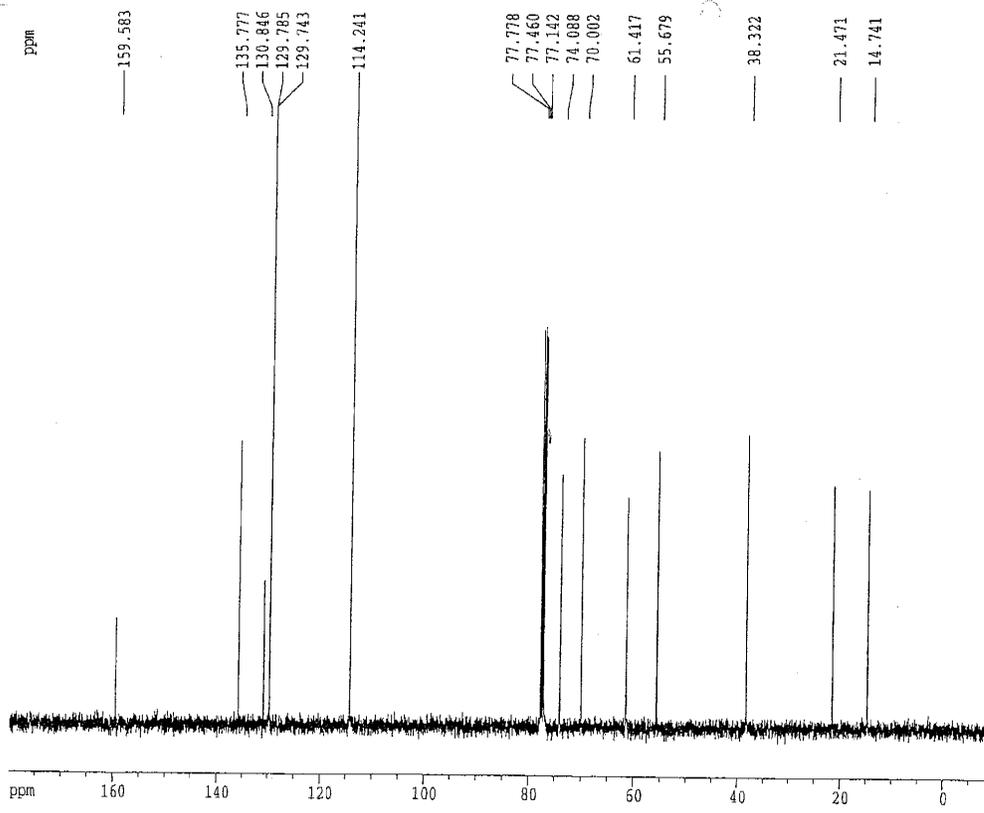
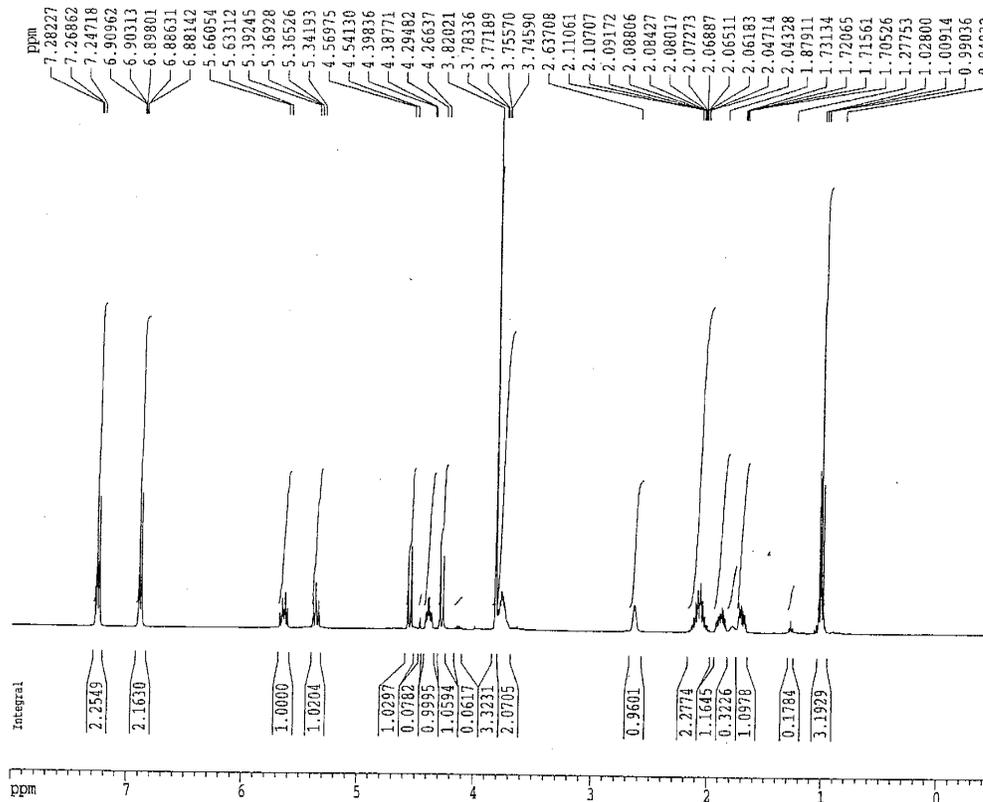
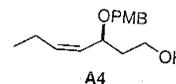
CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316005 MHz

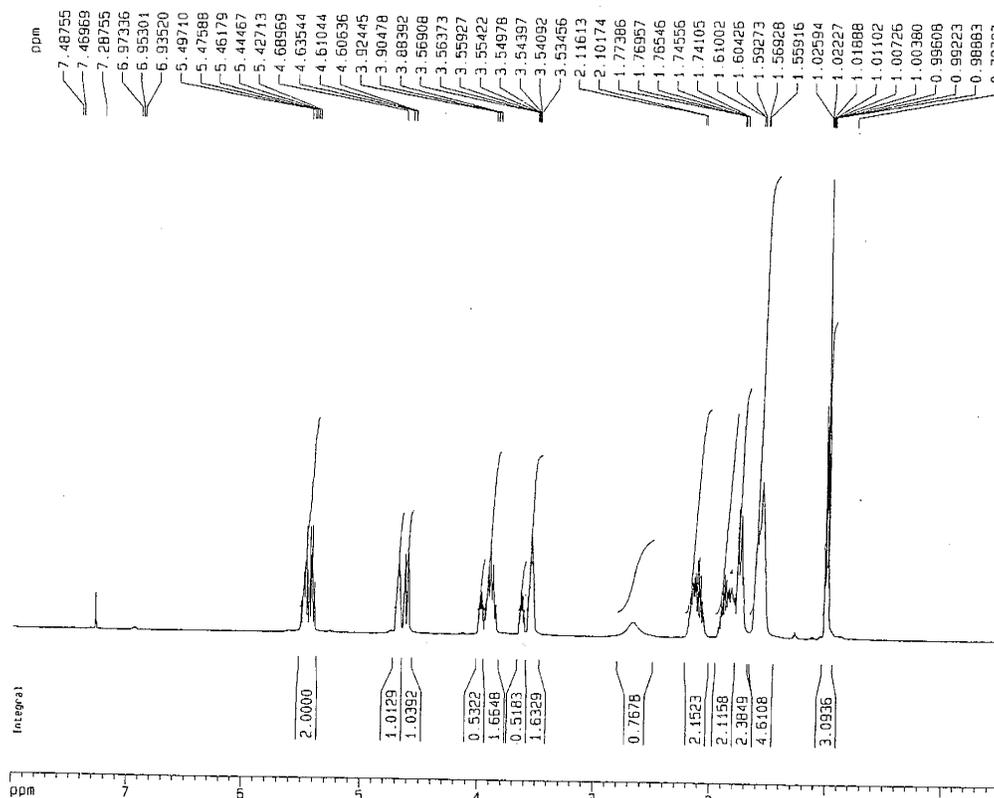
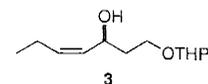
F2 - Processing parameters

SI 32768
 SF 100.6127290 MHz
 HW 0
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters

CX 20.00 cm
 F1P 215.000 ppm
 F1 21631.74 Hz
 F2P -5.000 ppm
 F2 -503.06 Hz
 PPMCM 11.00000 ppm/cm
 HZCM 1106.73999 Hz/cm





Current Data Parameters
 NAME wj2-251
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20020417
 Time 20.56
 INSTRUM spect
 PROBRD 5 mm BBO 50-1H
 PULPROG zg30
 TO 32788
 SOLVENT CDCl3
 NS 15
 DS 2
 SWH 6009.618 Hz
 FIDRES 0.193398 Hz
 AQ 2.7864308 sec
 RG 203.2
 DM 83.200 usec
 DE 5.00 usec
 TE 300.0 K
 D1 0.0300000 sec

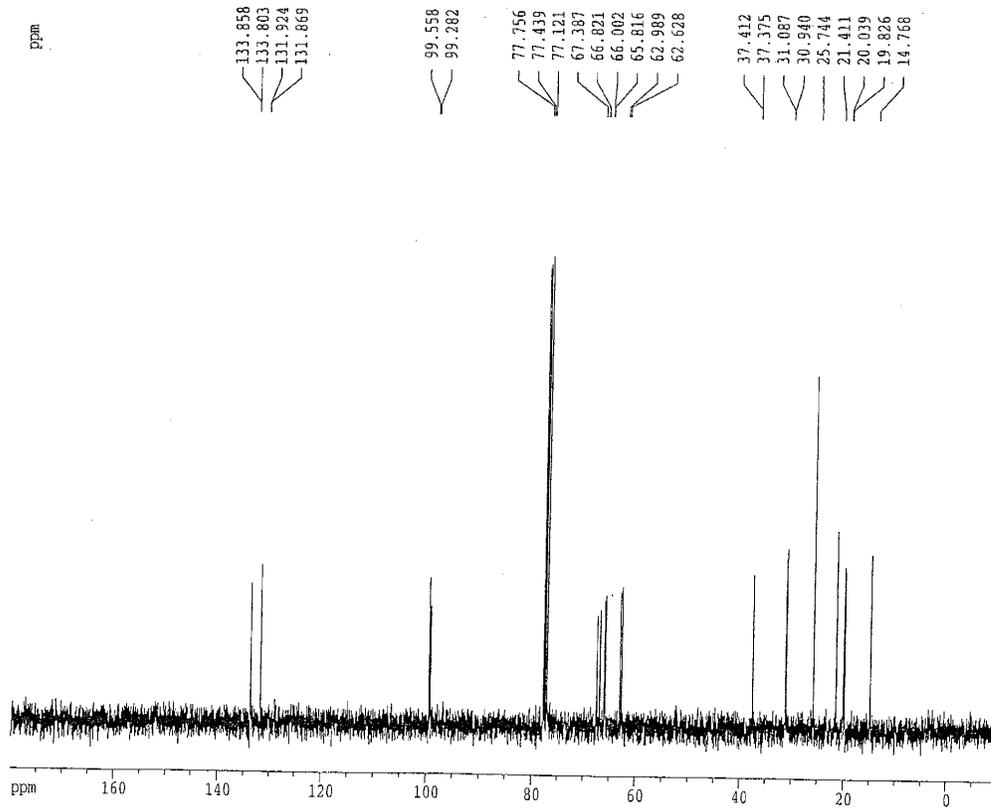
***** CHANNEL f1 *****
 NUC1 1H
 P1 7.70 usec
 PL1 -4.00 dB
 SFO1 500.130005 MHz

F1 - Acquisition parameters
 ND0 2
 TO 256
 SFO1 500.1305 MHz
 FIDRES 23.475050 Hz
 SW 12.016 ppm

F2 - Processing parameters
 SI 32768
 SF 500.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

F1 - Processing parameters
 SI 1024
 MC2 OF
 SF 500.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 8.000 ppm
 F1 4001.04 Hz
 F2P -0.500 ppm
 F2 -950.07 Hz
 PPMCH 9.42500 ppm/cm
 HZCH 212.55825 Hz/cm



Current Data Parameters
 NAME wj2-214
 EXPNO 2
 PROCNO 1

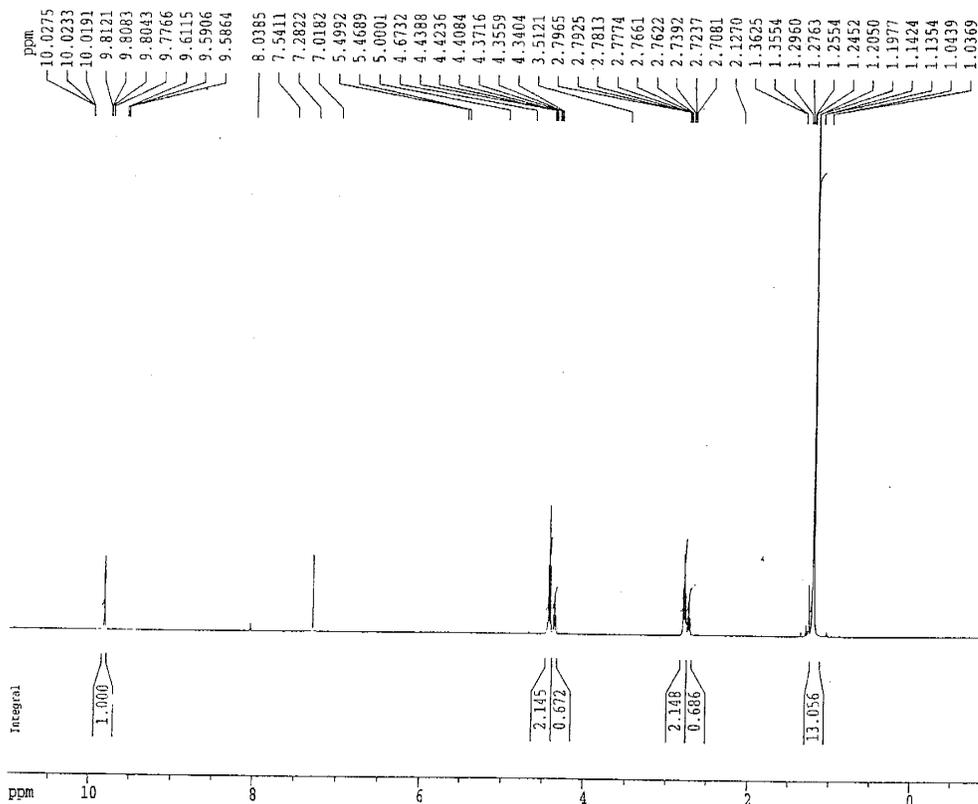
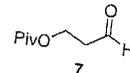
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 Date_ 20020304
 Time 13.03
 INSTRUM dxt400
 PROBRD 5 mm Multinu
 PULPROG zgpg30
 TO 65336
 SOLVENT CDCl3
 NS 100
 DS 2
 SWH 23148.148 Hz
 FIDRES 0.353213 Hz
 AQ 1.4156276 sec
 RG 3649.1
 DM 21.600 usec
 DE 4.50 usec
 TE 300.0 K
 D1 0.0500000 sec
 d11 0.0100000 sec
 d12 0.0000200 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 12.30 usec
 PL1 2.00 dB
 SFO1 100.6232933 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127290 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 CY 180.000 ppm
 F1 18110.29 Hz
 F2P -1009.74 Hz
 F2 -1009.74 Hz
 PPMCH 9.50179 ppm/cm
 HZCH 956.00128 Hz/cm



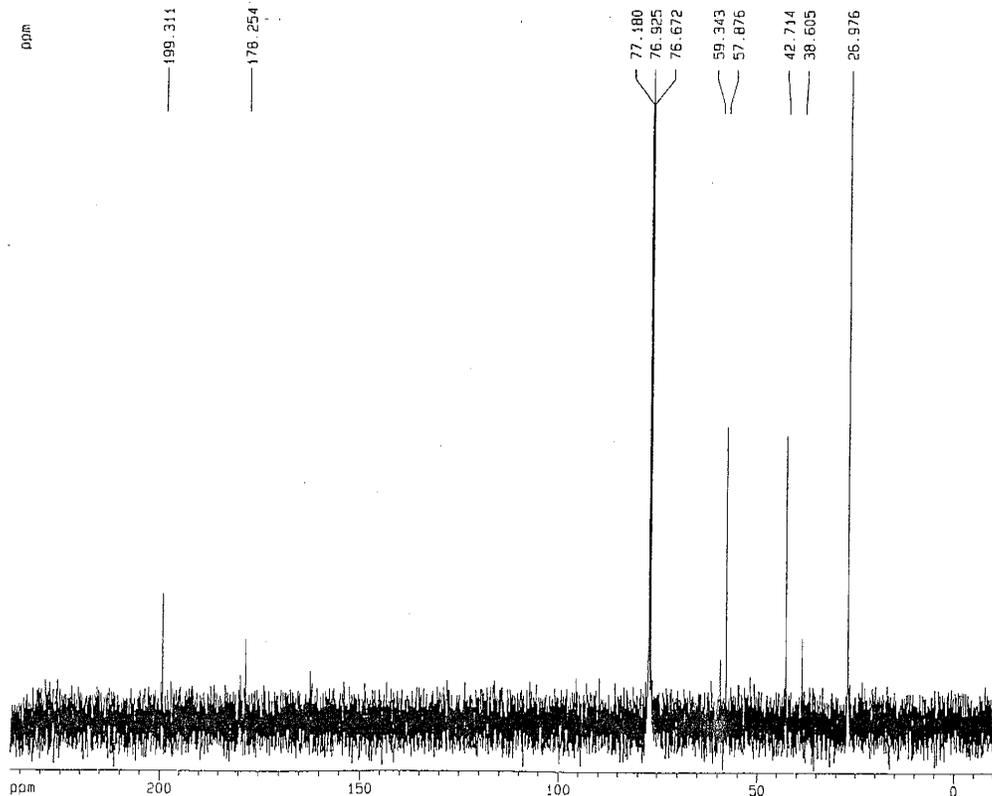
Current Data Parameters
 NAME wj2-145
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20011206
 Time 20.44
 INSTRUM dirx400
 PROBHD 5 mm Multinu
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 161.3
 EW 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.70 usec
 PL1 -6.00 dB
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

ID NMR plot parameters
 CX 20.00 cm
 FIP 10.985 ppm
 F1 4395.28 Hz
 F2P -0.985 ppm
 F2 -391.99 Hz
 PPMCM 0.59846 ppm/cm
 HZCM 239.46359 Hz/cm



Current Data Parameters
 NAME wj2-145
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20020227
 Time 13:30
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 4
 DS 4
 SWH 31446.541 Hz
 FIDRES 0.478206 Hz
 AQ 1.0400083 sec
 RG 7296.2
 DW 15.500 usec
 DC 6.00 usec
 TE 300.0 K
 D1 0.10000000 sec
 D11 0.03000000 sec
 D12 0.00020000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 6.50 usec
 PL1 5.00 dB
 SFO1 125.7719472 MHz

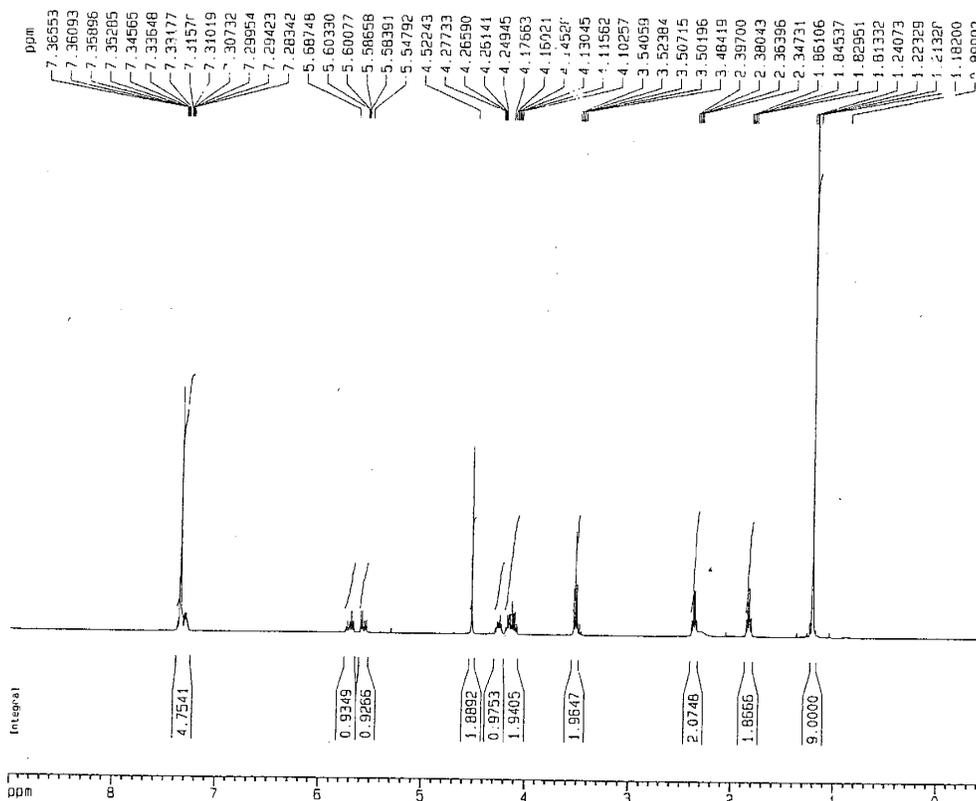
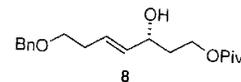
===== CHANNEL f2 =====
 NUC2 1H
 P2 95.00 usec
 PL2 -4.00 dB
 PL12 19.00 dB
 PL13 30.00 dB
 SFO2 500.1325000 MHz

F1 - Acquisition Parameters
 NQD 2
 TD 256
 SFO1 500.1325 MHz
 FIDRES 23.475080 Hz
 SM 12.016 ppm

F2 - Processing parameters
 SI 65536
 SF 125.7578000 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

F1 - Processing parameters
 SI 1024
 MCF 0F
 SF 500.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0

ID NMR plot parameters
 CX 20.00 cm
 FIP 0.00 ppm
 F1 237.517 ppm
 F1 29869.63 Hz
 F2P -12.338 ppm
 F2 -1576.91 Hz
 PPMCM 12.50282 ppm/cm
 HZCM 1572.32703 Hz/cm

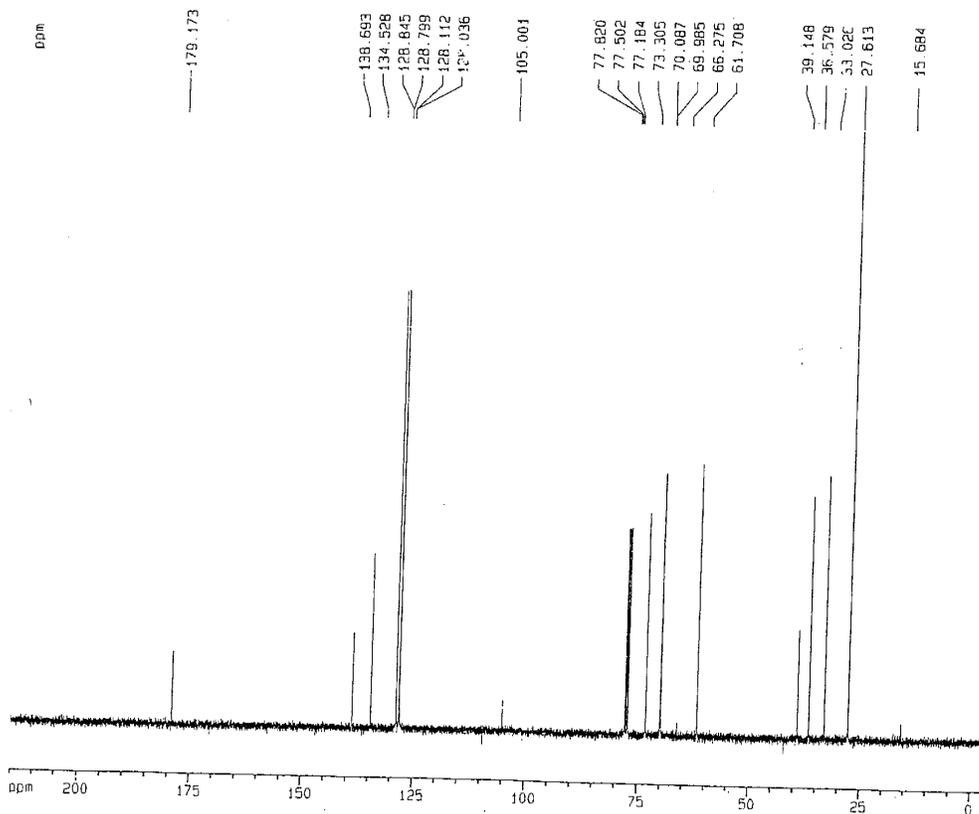


Current Data Parameters
 NAME wj1-145
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 1000515
 Time 12:23
 INSTRUM grx400
 PROBHD 5 mm Multinu
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 64
 DM 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 P1 7.70 usec
 DE 4.50 usec
 SFO1 400.1320007 MHz
 NUC1 1H
 PL1 -6.00 dB

F2 - Processing parameters
 SI 16384
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 9.000 ppm
 F1 3601.17 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PRACH 0.47500 ppm/cm
 HZCN 190.06175 Hz/cm

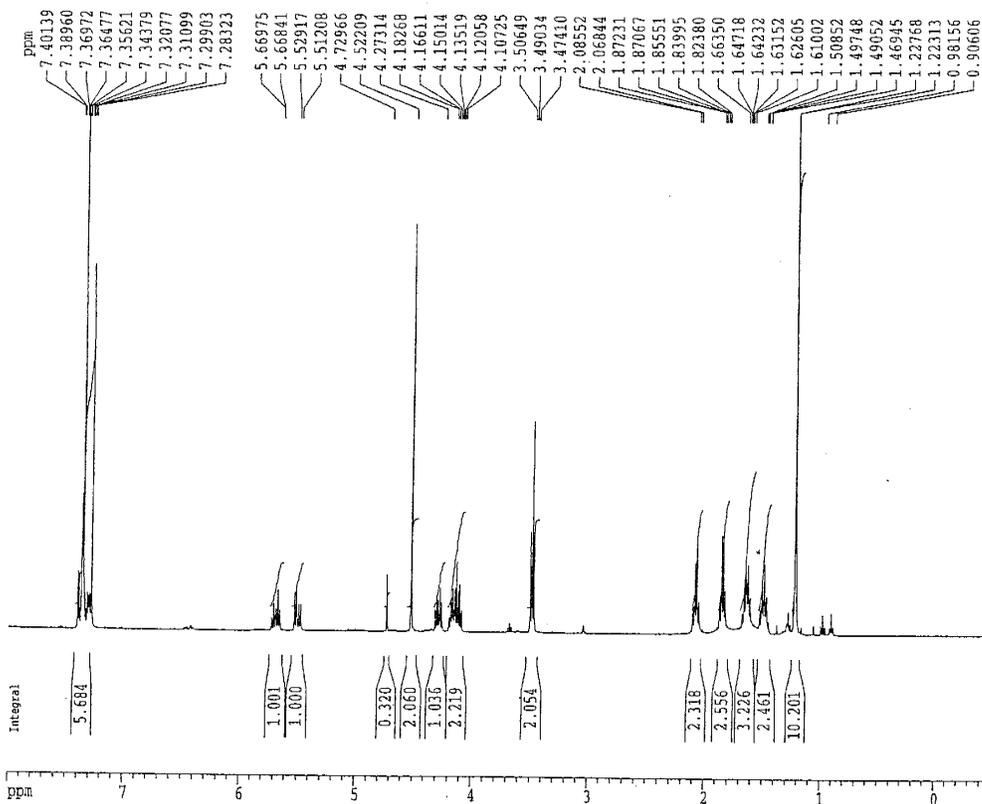
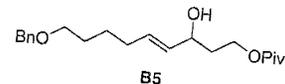


Current Data Parameters
 NAME wj1-145c
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 1000515
 Time 12:12
 INSTRUM grx400
 PROBHD 5 mm Multinu
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 239
 DS 2
 SWH 23148.146 Hz
 FIDRES 0.353213 Hz
 AQ 1.4156276 sec
 RG 2048
 DM 21.600 usec
 DE 4.50 usec
 TE 300.0 K
 d11 0.03000000 sec
 d12 0.00000000 sec
 PL13 1.00 dB
 D1 0.05000000 sec
 CPDPRG2 waitz16
 PCDP2 100.00 usec
 SFO2 400.1315005 MHz
 NUC2 13C
 PL2 -6.00 dB
 PL12 18.00 dB
 P1 6.90 usec
 DE 4.50 usec
 SFO1 100.6252933 MHz
 NUC1 13C
 PL1 -6.00 dB

F2 - Processing parameters
 SI 32768
 SF 100.6127290 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 215.000 ppm
 F1 21631.74 Hz
 F2P -5.000 ppm
 F2 -503.06 Hz
 PRACH 11.00000 ppm/cm
 HZCN 1106.73999 Hz/cm



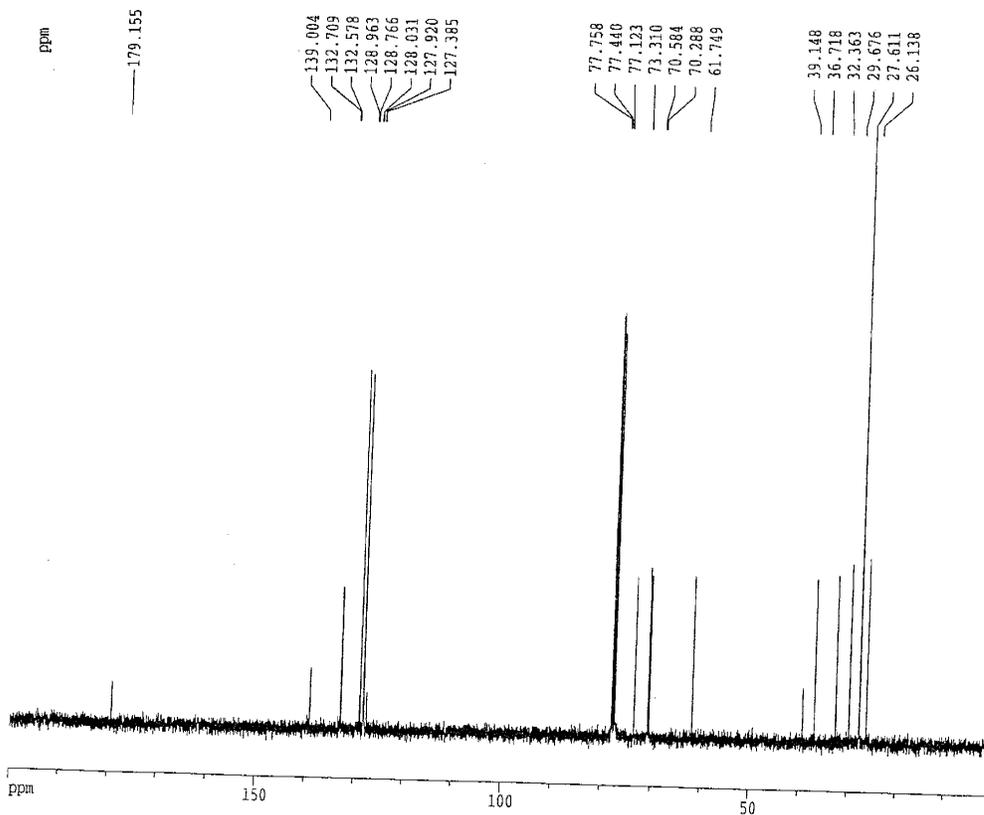
Current Data Parameters
 NAME wj4-41
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20030709
 Time 12.22
 INSTRUM drx400
 PROBHD 5 mm Multinucl
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 256
 DW 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.70 usec
 PL1 -5.00 dB
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 8.000 ppm
 F1 3201.04 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.42500 ppm/cm
 HZCM 170.35525 Hz/cm



Current Data Parameters
 NAME wj4-41
 EXPNO 2
 PROCNO 1

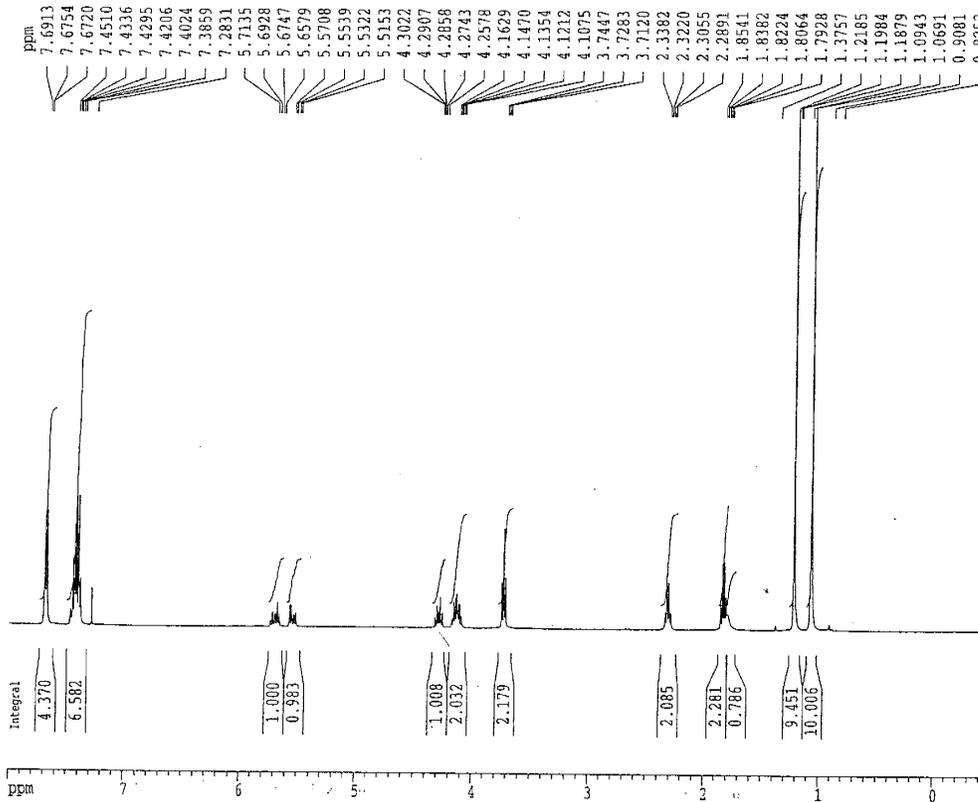
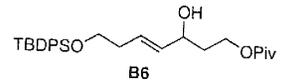
F2 - Acquisition Parameters
 Date_ 20030709
 Time 12.29
 INSTRUM drx400
 PROBHD 5 mm Multinucl
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 375
 DS 2
 SWH 23148.148 Hz
 FIDRES 0.353213 Hz
 AQ 1.4156276 sec
 RG 32768
 DW 21.600 usec
 DE 4.50 usec
 TE 300.0 K
 D1 0.05000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 12.30 usec
 PL1 2.00 dB
 SFO1 100.6232933 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127290 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 200.000 ppm
 F1 20122.55 Hz
 F2P -0.500 ppm
 F2 -50.31 Hz
 PPMCM 10.02500 ppm/cm
 HZCM 1008.64264 Hz/cm



Current Data Parameters

NAME wj4-191.13
EXNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20040404
Time 15.17
INSTRUM drx400
PROBHD 5 mm Multinucl
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 128
DM 104.400 usec
DE 5.50 usec
TE 1809.2 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

***** CHANNEL f1 *****

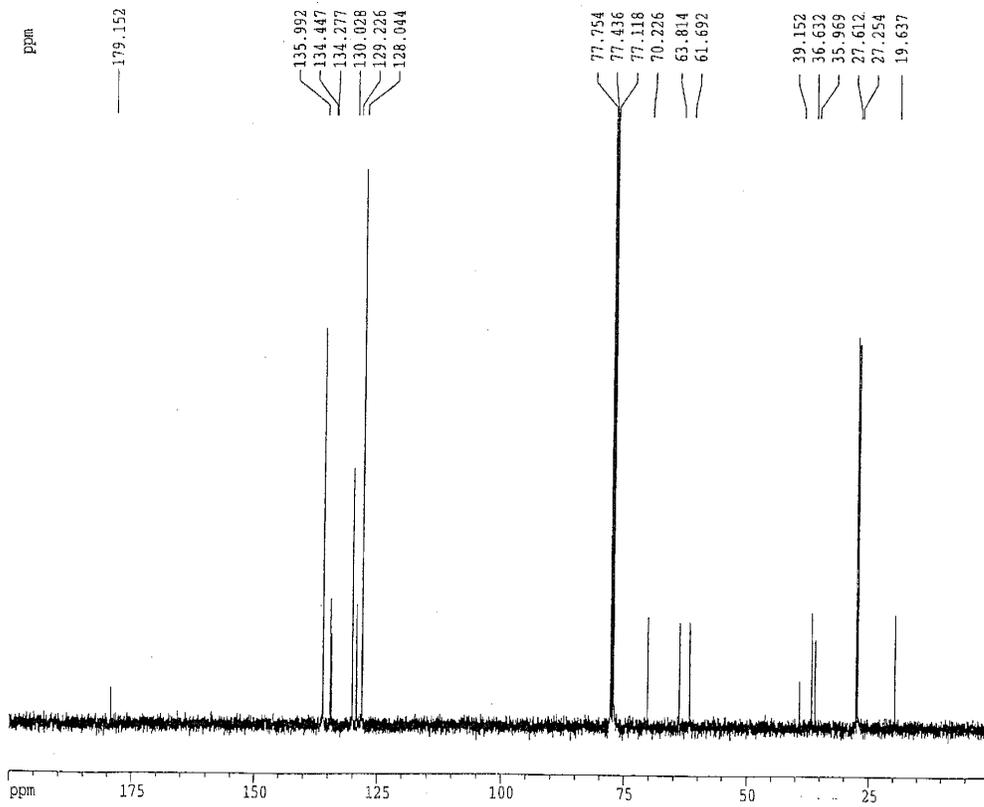
NUC1 1H
P1 7.70 usec
PL1 -6.00 dB
SFO1 400.1320007 MHz

F2 - Processing parameters

SI 16384
SF 400.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters

CX 20.00 cm
CY 0.00 cm
F1P 8.000 ppm
F2P 3201.04 Hz
F2P -0.500 ppm
F2P -200.07 Hz
PPMCM 0.42500 ppm/cm
HZCM 170.05525 Hz/cm



Current Data Parameters

NAME wj4-191.13
EXNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20040404
Time 15.22
INSTRUM drx400
PROBHD 5 mm Multinucl
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 548
DS 4
SWH 23148.148 Hz
FIDRES 0.153213 Hz
AQ 1.4156276 sec
RG 32768
DM 21.600 usec
DE 5.50 usec
TE 1809.2 K
D1 0.15000001 sec
d11 0.03000000 sec
DELTA 0.05000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

***** CHANNEL f1 *****

NUC1 13C
P1 12.30 usec
PL1 2.00 dB
SFO1 100.6232933 MHz

***** CHANNEL f2 *****

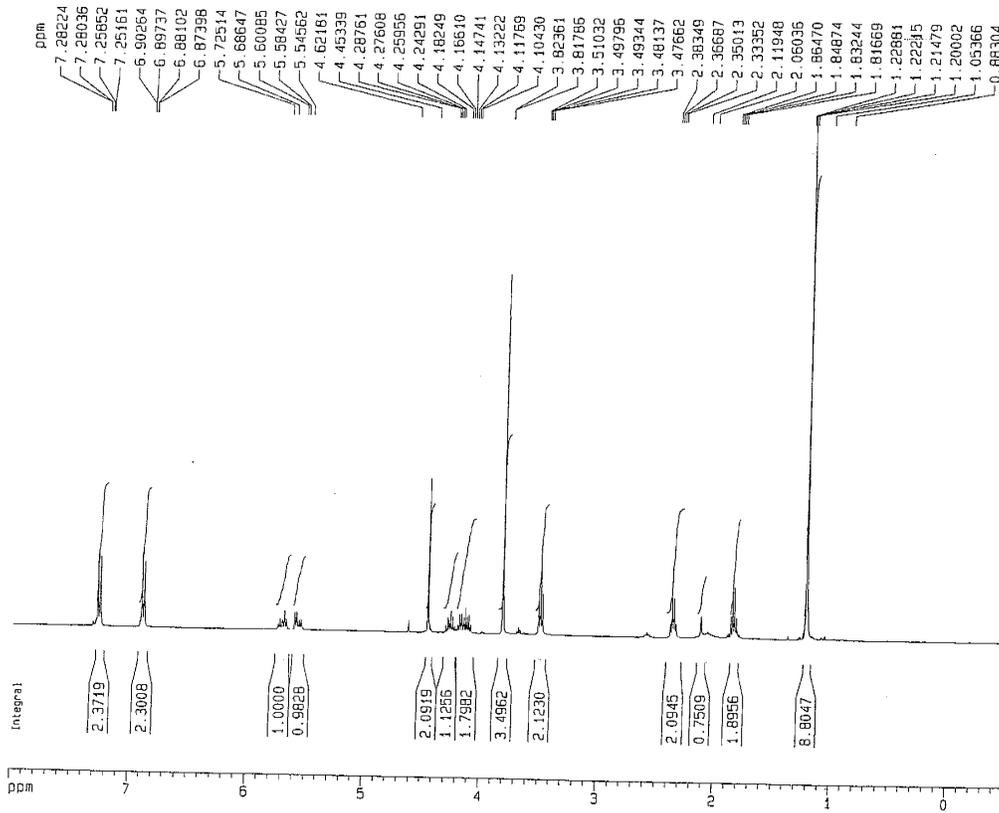
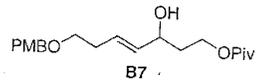
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1316005 MHz

F2 - Processing parameters

SI 65536
SF 100.6127290 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters

CX 20.00 cm
CY 0.00 cm
F1P 200.300 ppm
F2P 20122.55 Hz
F2P -0.500 ppm
F2P -50.31 Hz
PPMCM 10.02500 ppm/cm
HZCM 1008.64264 Hz/cm



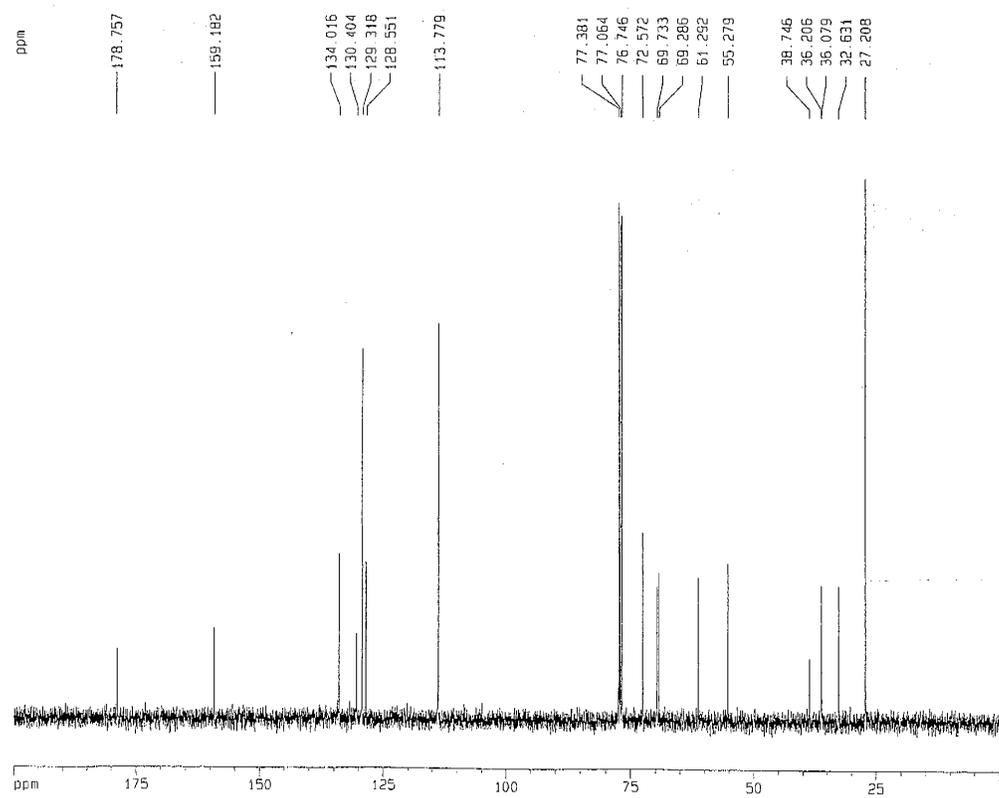
Current Data Parameters
 NAME wj5-146
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20050320
 Time 15.12
 INSTRUM spect
 PROBHD 5 mm QNP 1H/15
 PULPROG zg30
 TD 32768
 SOLVENT CDC13
 NS 15
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 114
 DW 104.400 usec
 DE 6.00 usec
 TE 295.6 K
 O1 1.00000000 sec
 MCREST 0.00000000 sec
 MCHRX 0.01500000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.05 usec
 PL1 -3.00 dB
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 8.000 ppm
 F2P 3201.04 Hz
 F2 -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.42500 ppm/cm
 HZCM 170.05525 Hz/cm



Current Data Parameters
 NAME wj5-168
 EXPNO 2
 PROCNO 1

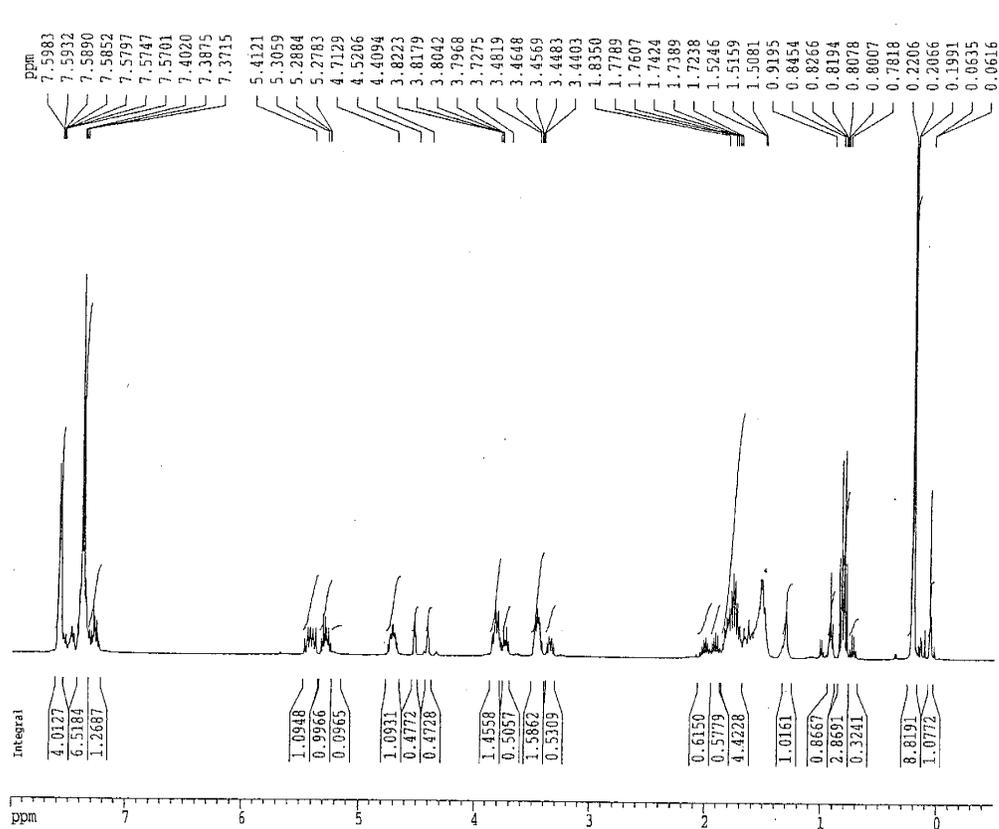
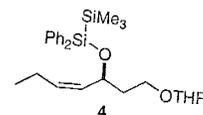
F2 - Acquisition Parameters
 Date_ 20050421
 Time 21.03
 INSTRUM spect
 PROBHD 5 mm QNP 1H/15
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 108
 DS 4
 SWH 23880.814 Hz
 FIDRES 0.385918 Hz
 AQ 1.9864706 sec
 RG 2048
 DW 20.850 usec
 DE 6.00 usec
 TE 295.6 K
 O1 0.15000001 sec
 d11 0.03000000 sec
 DELTA 0.05000000 sec
 MCREST 0.00000000 sec
 MCHRX 0.01500000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 7.20 usec
 PL1 -2.00 dB
 SFO1 100.628268 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 98.40 usec
 PL2 -3.00 dB
 PL12 19.85 dB
 PL13 120.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127680 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 200.000 ppm
 F2P 20122.55 Hz
 F2 -0.500 ppm
 F2 -50.21 Hz
 PPMCM 10.02500 ppm/cm
 HZCM 1008.64307 Hz/cm



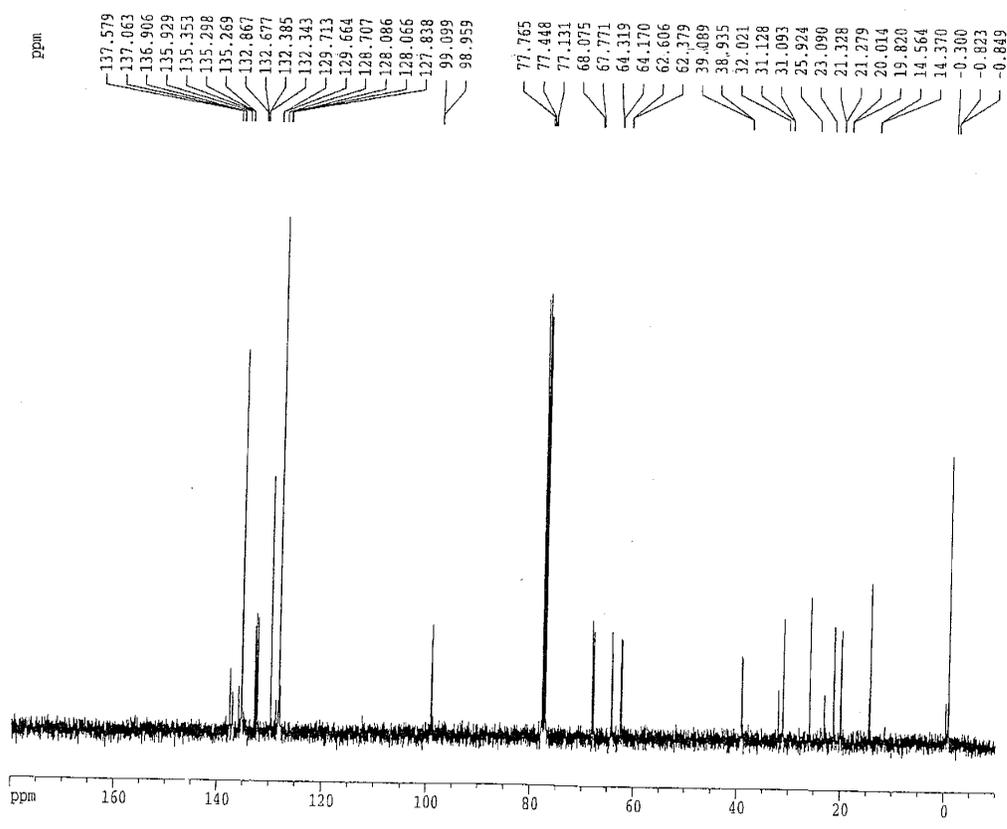
Current Data Parameters
 NAME wj2-220
 EXPRNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20020313
 Time 14.28
 INSTRUM drx400
 PROBRD 5 mm Multinu
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 64
 DW 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.0000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.70 usec
 PL1 -6.00 dB
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 8.000 ppm
 F1 3201.04 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.42500 ppm/cm
 HZCM 170.05525 Hz/cm



Current Data Parameters
 NAME wj2-220
 EXPRNO 2
 PROCNO 1

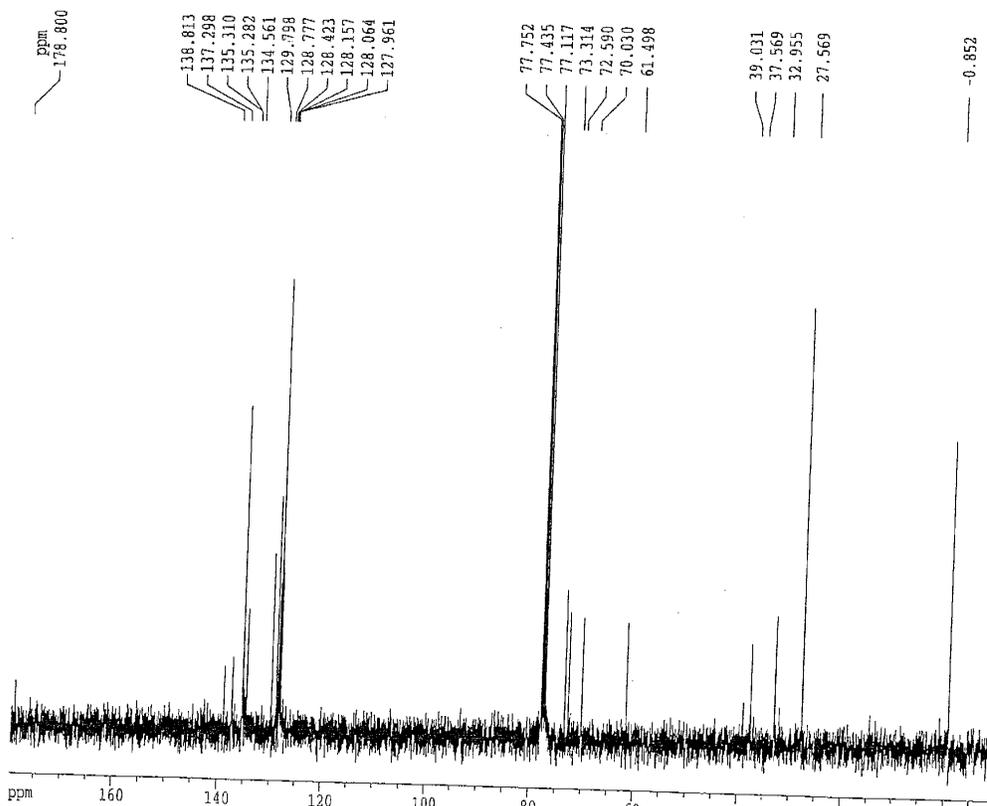
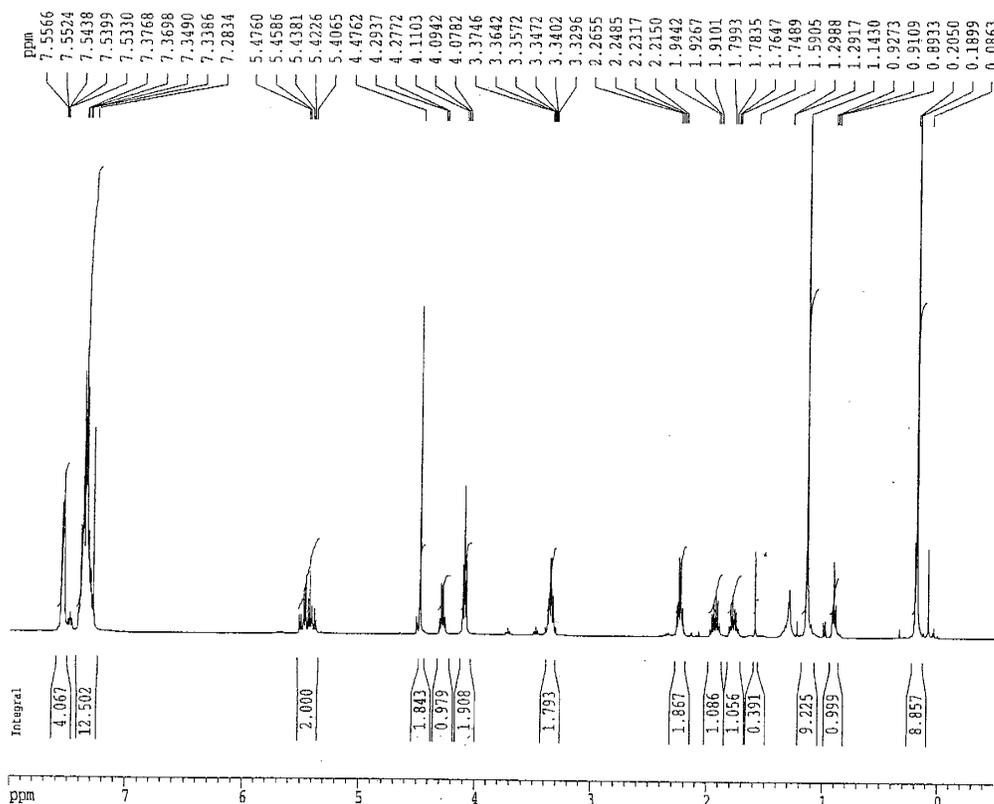
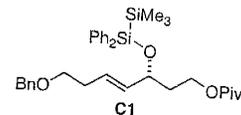
F2 - Acquisition Parameters
 Date_ 20020313
 Time 14.32
 INSTRUM drx400
 PROBRD 5 mm Multinu
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 180
 DS 2
 SWH 23148.148 Hz
 FIDRES 0.353213 Hz
 AQ 1.4156276 sec
 RG 8192
 DW 21.600 usec
 DE 4.50 usec
 TE 300.0 K
 D1 0.0500000 sec
 d11 0.0300000 sec
 d12 0.0000200 sec

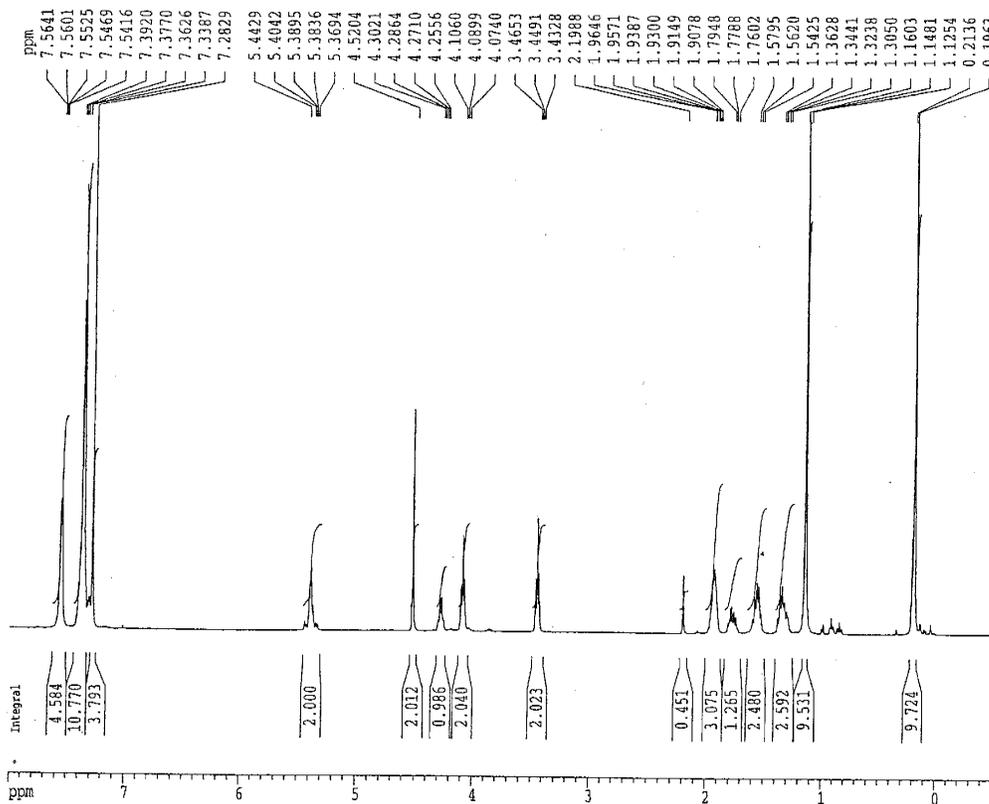
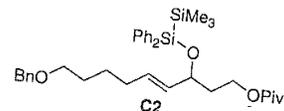
===== CHANNEL f1 =====
 NUC1 13C
 P1 12.10 usec
 PL1 2.00 dB
 SFO1 100.6232933 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127290 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 180.000 ppm
 F1 18110.29 Hz
 F2P -10.036 ppm
 F2 -1009.74 Hz
 PPMCM 9.50179 ppm/cm
 HZCM 956.00140 Hz/cm





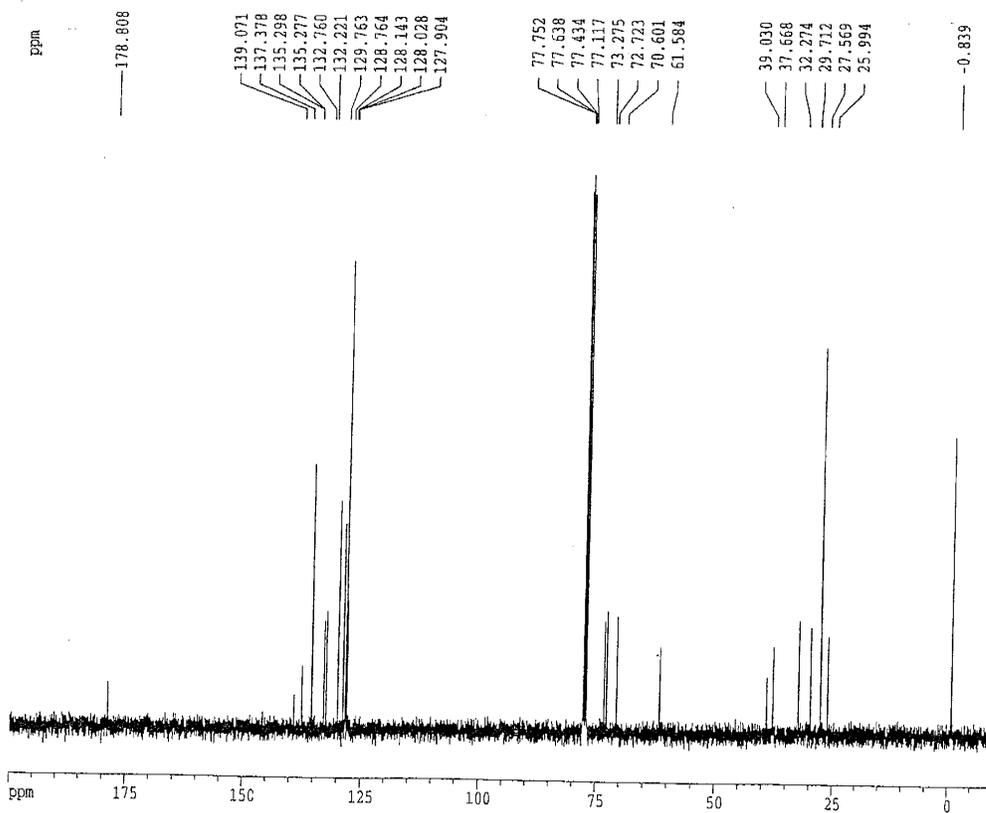
Current Data Parameters
 NAME wj3-266
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20030416
 Time 11.29
 INSTRUM drx400
 PROBHD 5 mm Multinucl
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 90.5
 DW 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.70 usec
 PL1 -6.00 dB
 SFO1 400.132007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 FC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 8.000 ppm
 F1 3201.04 Hz
 F2P -0.560 ppm
 F2 -200.07 Hz
 PPMCM 0.42500 ppm/cm
 HZCM 170.05525 Hz/cm



Current Data Parameters
 NAME wj3-266
 EXPNO 2
 PROCNO 1

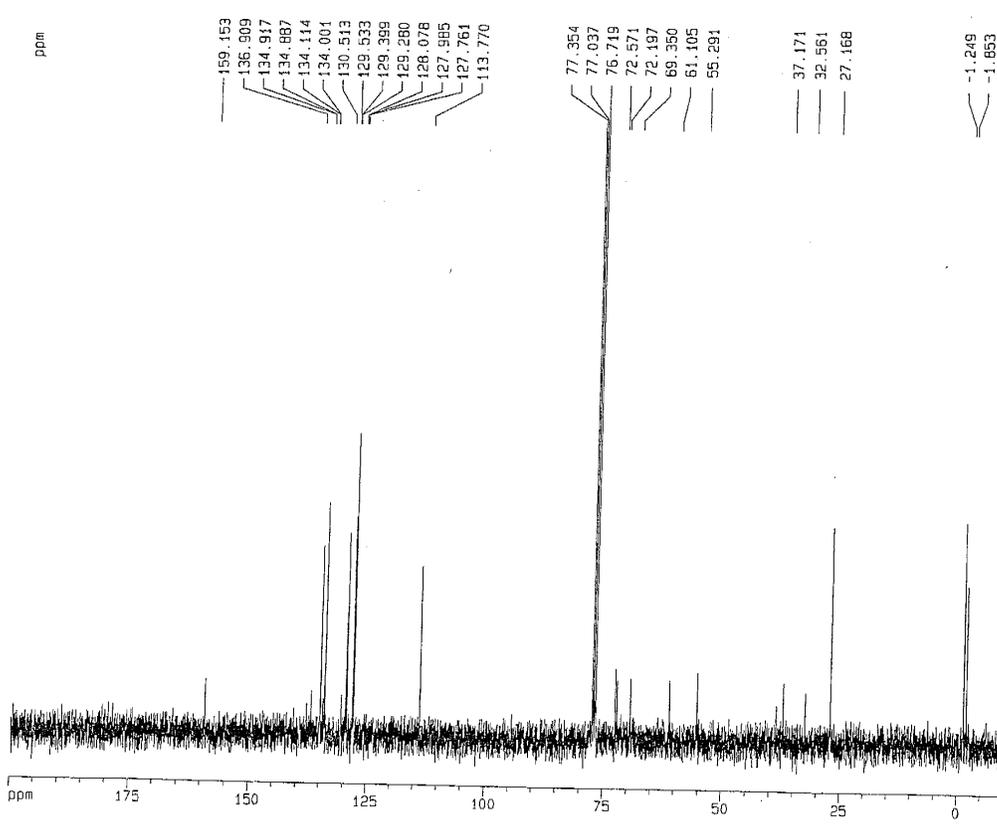
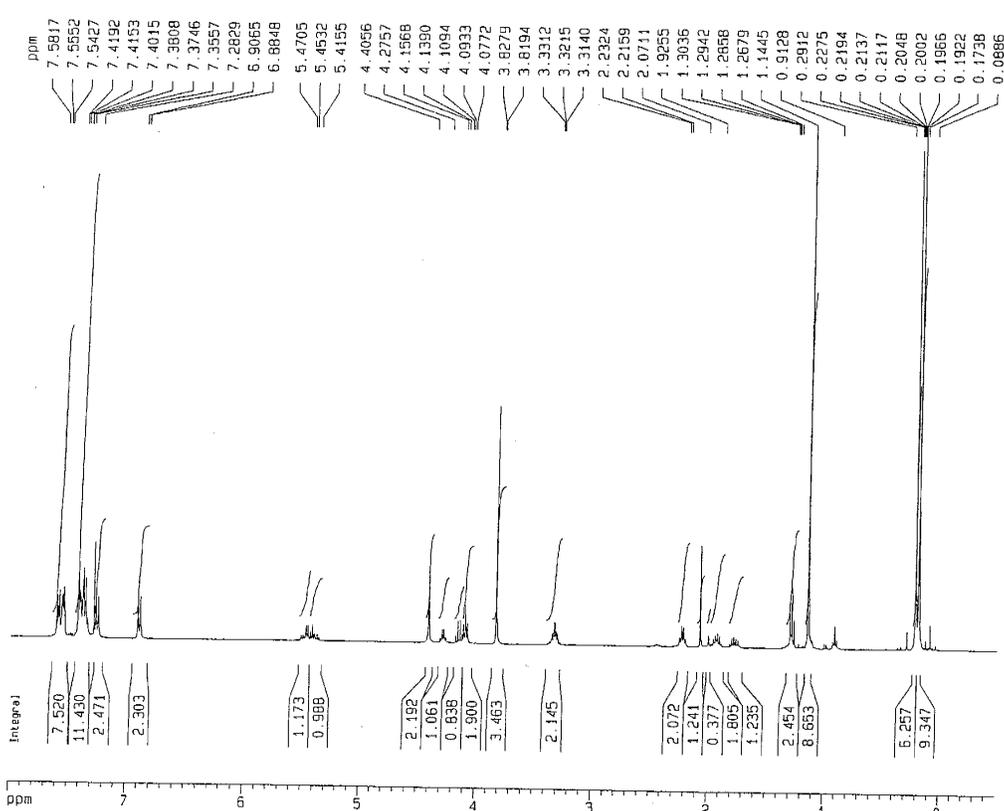
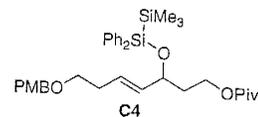
F2 - Acquisition Parameters
 Date_ 20030416
 Time 11.33
 INSTRUM drx400
 PROBHD 5 mm Multinucl
 PULPROG zpg30
 TD 65536
 SOLVENT CDCl3
 NS 338
 DS 2
 SWH 23148.148 Hz
 FIDRES 0.353213 Hz
 AQ 1.4156276 sec
 RG 12768
 DW 21.600 usec
 DE 4.50 usec
 TE 300.0 K
 D1 0.05000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

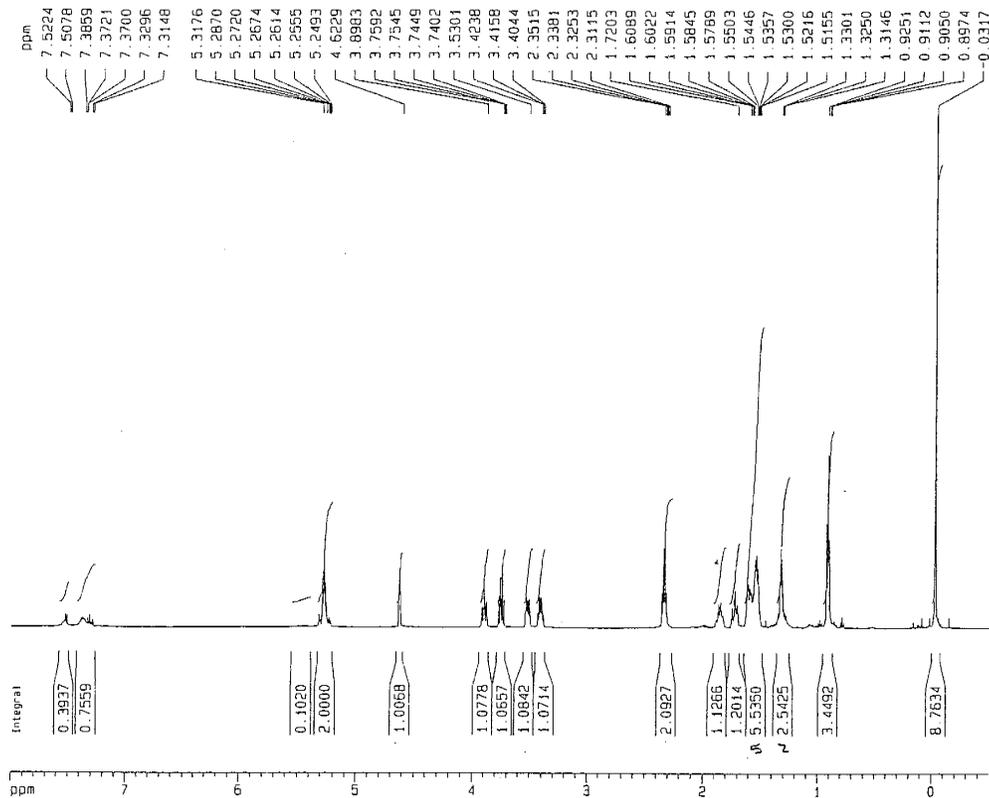
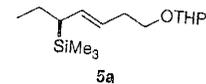
===== CHANNEL f1 =====
 NUC1 13C
 P1 12.30 usec
 PL1 2.00 dB
 SFO1 100.6232933 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127290 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 260.000 ppm
 F1 20122.55 Hz
 F2P -10.000 ppm
 F2 -1006.13 Hz
 PPMCM 10.50000 ppm/cm
 HZCM 1056.43372 Hz/cm





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Current Data Parameters
NAME w12-229
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20020317
Time 14.18
INSTRUM spect
PROBHD 5 mm BBO 90-1H
PULPROG zg30
TD 32768
SOLVENT CCl3
NS 16
DS 2
SWH 6009.815 Hz
FIDRES 0.183359 Hz
AQ 2.7264309 sec
RG 32
DK 83.200 usec
DE 5.00 usec
TE 300.0 K
D1 0.0300000 sec

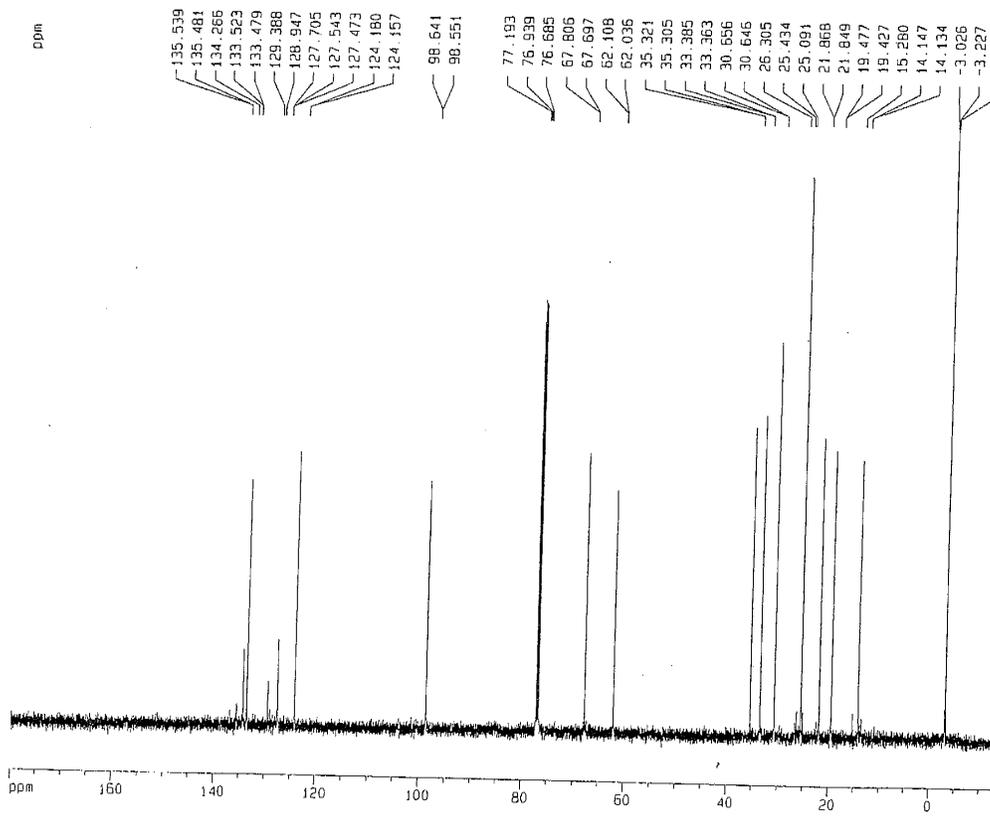
***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -4.00 dB
SFO1 500.1320005 MHz

F1 - Acquisition parameters
ND0 2
TD 256
SFO1 500.1325 MHz
FIDRES 23.478560 Hz
SW 12.016 ppm

F2 - Processing parameters
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

F3 - Processing parameters
SI 1024
MC2 GF
SF 500.1300000 MHz
WDW OBTUNE
SSB 0
LB 0.30 Hz
GB 0

ID NMR plot parameters
CX 20.00 cm
CY 0.00 cm
FIP 8.000 ppm
F1 4001.04 Hz
FAP -0.500 ppm
F2 -250.07 Hz
PPHCH 0.42500 ppm/cm
MECH 212.95525 Hz/cm
  
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Current Data Parameters
NAME w12-229
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20020317
Time 14.22
INSTRUM spect
PROBHD 5 mm BBO 90-1H
PULPROG zgpg30
TD 65536
SOLVENT CCl3
NS 192
DS 4
SWH 31446.541 Hz
FIDRES 0.478936 Hz
AQ 1.0420883 sec
RG 5180.6
WDW EM
SSB 0
LB 15.800 usec
DE 6.00 usec
TE 300.0 K
D1 0.1000000 sec
d11 0.0300000 sec
d12 0.0002000 sec

***** CHANNEL f1 *****
NUC1 13C
P1 6.50 usec
PL1 5.00 dB
SFO1 125.7718472 MHz

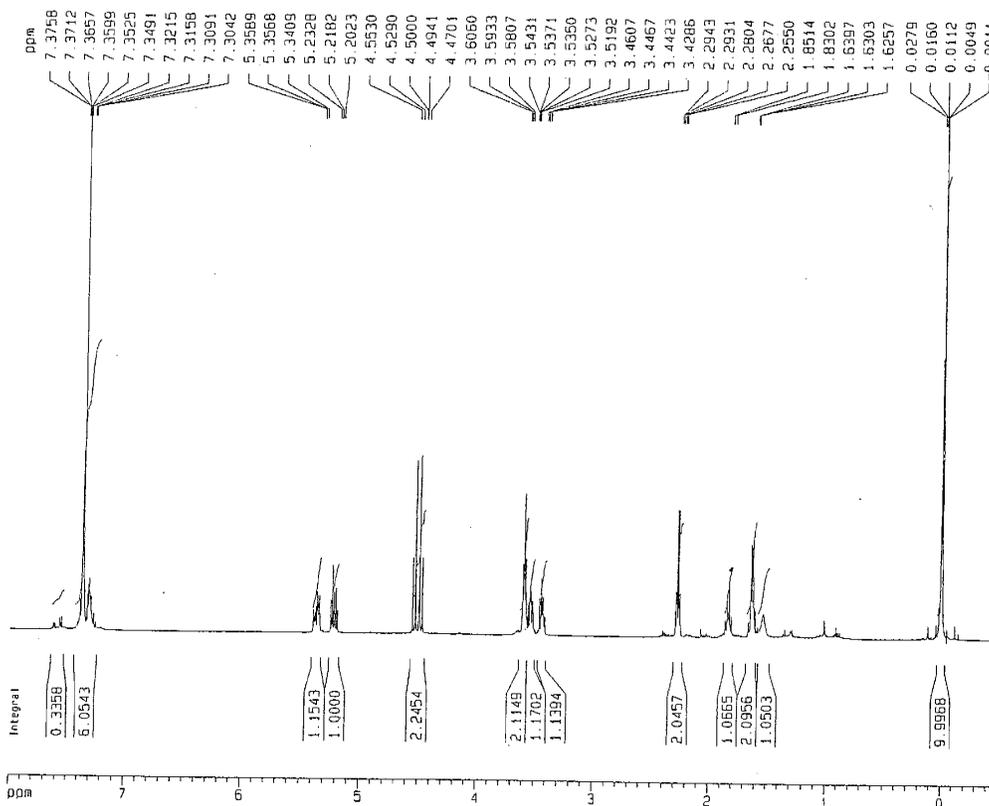
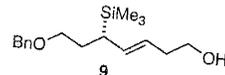
***** CHANNEL f2 *****
CPDPRG2 meltz16
NUC2 1H
PROBHD 5 mm BBO 90-1H
PULPROG zgpg30
PL2 -4.00 dB
PL12 19.00 dB
PL13 30.00 dB
SFO2 500.1325000 MHz

F1 - Acquisition parameters
ND0 2
TD 256
SFO1 500.1325 MHz
FIDRES 23.478560 Hz
SW 12.016 ppm

F2 - Processing parameters
SI 65536
SF 125.7570000 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

F3 - Processing parameters
SI 1024
MC2 GF
SF 500.1300000 MHz
WDW OBTUNE
SSB 0
LB 0.30 Hz
GB 0

ID NMR plot parameters
CX 20.00 cm
CY 0.00 cm
FIP 180.000 ppm
F1 22836.40 Hz
FAP -12.529 ppm
F2 -1578.31 Hz
PPHCH 9.66286 ppm/cm
MECH 1210.56337 Hz/cm
  
```

Current Data Parameters
 NAME w12-227
 EXPNO 2
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20020314
 Time 17.39
 INSTRUM spect
 PROBHD 5 mm BBO BB-H
 PULPROG zgpg30
 TO 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.103398 Hz
 AQ 2.7264309 sec
 RG 57
 DW 83.000 usec
 DE 5.00 usec
 TE 300.0 K
 D1 0.03000000 sec

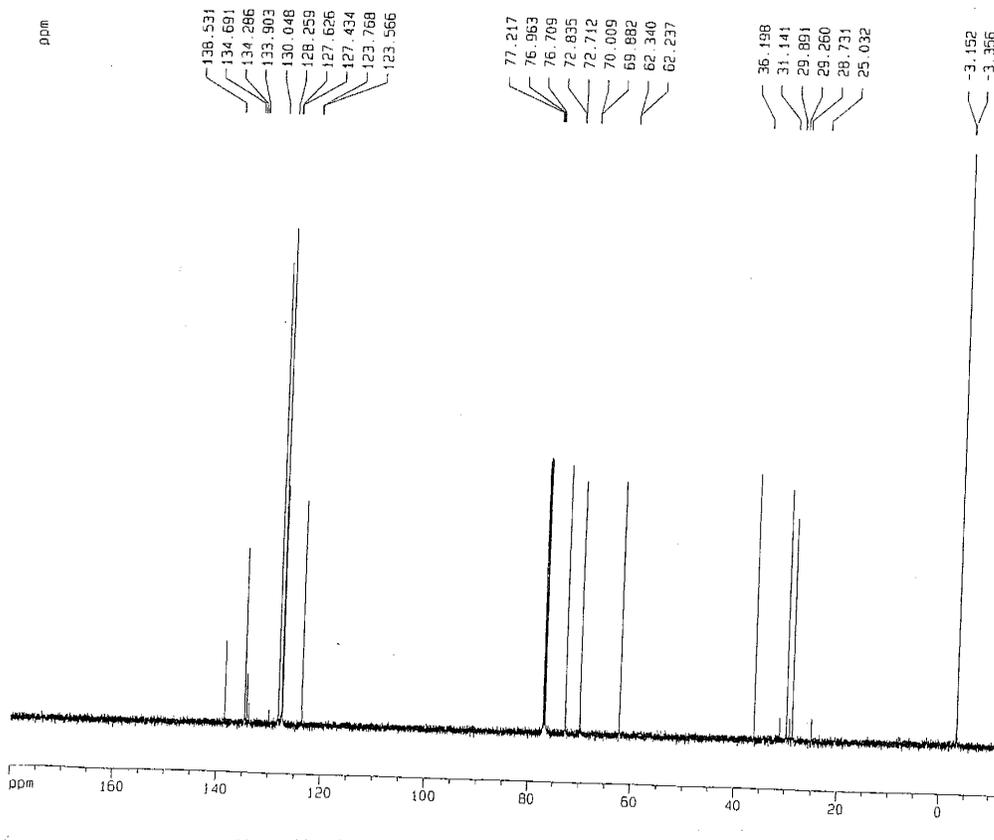
***** CHANNEL f1 *****
 NUC1 1H
 P1 7.70 usec
 PL1 -4.00 dB
 SFO1 500.130005 MHz

F1 - Acquisition parameters
 HQD 2
 TO 256
 SFO1 500.1325 MHz
 FIDRES 23.475060 Hz
 SW 12.016 ppm

F2 - Processing parameters
 SI 32768
 SF 500.130000 MHz
 MDW 0
 SSB 0
 LB 0.20 Hz
 GB 0
 PC 1.00

F1 - Processing parameters
 SI 1024
 MC2 0
 SF 500.130000 MHz
 MDW 0
 SSB 0
 LB 0.30 Hz
 GB 0

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 FIP 8.000 ppm
 F1 4001.04 Hz
 F2 -0.500 ppm
 FZ -250.07 Hz
 GPCCH 0.42500 ppm/cm
 HZCH 212.55525 Hz/cm



Current Data Parameters
 NAME w12-227
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20020314
 Time 17.43
 INSTRUM spect
 PROBHD 5 mm BBO BB-H
 PULPROG zgpg30
 TO 6536
 SOLVENT CDCl3
 NS 32
 DS 2
 SWH 31446.541 Hz
 FIDRES 0.475026 Hz
 AQ 1.0426803 sec
 RG 2048
 DW 15.500 usec
 DE 5.00 usec
 TE 300.0 K
 D1 0.10000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 5.50 usec
 PL1 5.00 dB
 SFO1 125.7719472 MHz

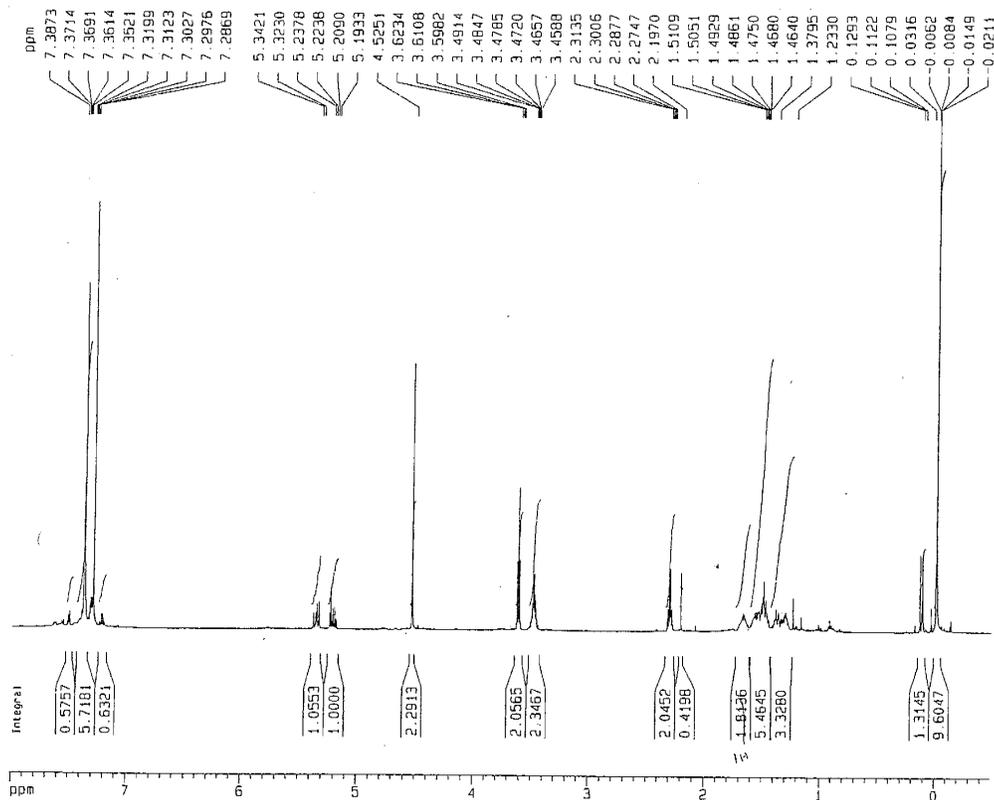
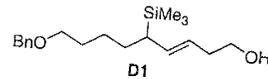
***** CHANNEL f2 *****
 CHPROG2 w12-110
 MIC2 1H
 PPR02 55.00 usec
 PL2 +4.00 dB
 PL12 18.00 dB
 PL13 30.00 dB
 SFO2 500.130500 MHz

F1 - Acquisition parameters
 HQD 2
 TO 256
 SFO1 500.1325 MHz
 FIDRES 23.475060 Hz
 SW 12.016 ppm

F2 - Processing parameters
 SI 6536
 SF 125.757800 MHz
 MDW 0
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

F1 - Processing parameters
 SI 1024
 MC2 0
 SF 500.130000 MHz
 MDW 0
 SSB 0
 LB 0.30 Hz
 GB 0

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 FIP 8.000 ppm
 F1 226.764 Hz
 F2 -12.939 ppm
 FZ -1576.91 Hz
 GPCCH 8.83036 ppm/cm
 HZCH 1210.65577 Hz/cm



Current Data Parameters
 NAME w13-268
 EXPNO 1
 PROCNO 1

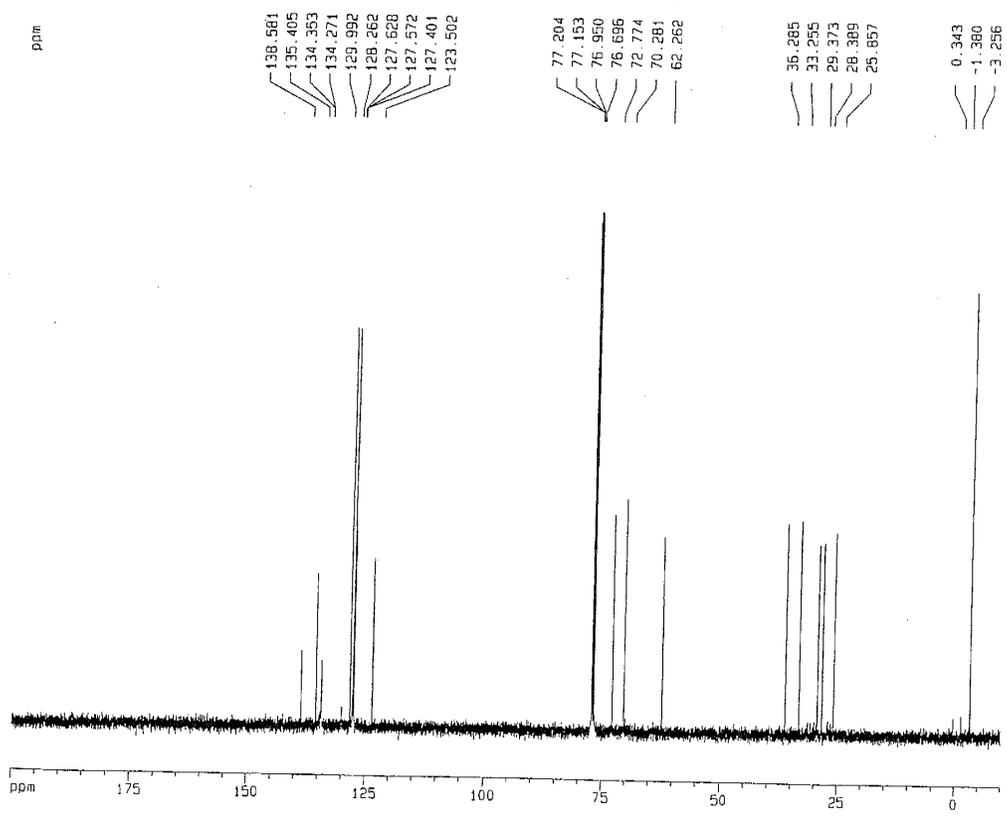
F2 - Acquisition Parameters
 Date_ 20030418
 Time 19.06
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 5009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7264309 sec
 RG 101.6
 OW 83.200 usec
 DE 6.00 usec
 TE 300.0 K
 D1 0.0300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.70 usec
 PL1 -4.00 dB
 SFO1 500.1320000 MHz

F2 - Processing parameters
 SI 32768
 SF 500.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

F1 - Processing parameters
 SI 1024
 HC2 OF
 SF 500.1300000 MHz
 WDW no
 SSB 0
 LB 0.30 Hz
 GB 0

ID NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 FIP 8.000 ppm
 F1 4001.04 Hz
 F2P -0.500 ppm
 F2 -250.07 Hz
 PPMCH 0.42500 ppm/cm
 HZCN 212.55825 Hz/cm



Current Data Parameters
 NAME w13-268
 EXPNO 1
 PROCNO 2

F2 - Acquisition Parameters
 Date_ 20030418
 Time 19.11
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 320
 DS 4
 SWH 31446.541 Hz
 FIDRES 0.479856 Hz
 AQ 1.0420083 sec
 RG 2048
 OW 15.900 usec
 DE 6.00 usec
 TE 300.0 K
 D1 0.1000000 sec
 d11 0.0300000 sec
 d12 0.0000200 sec

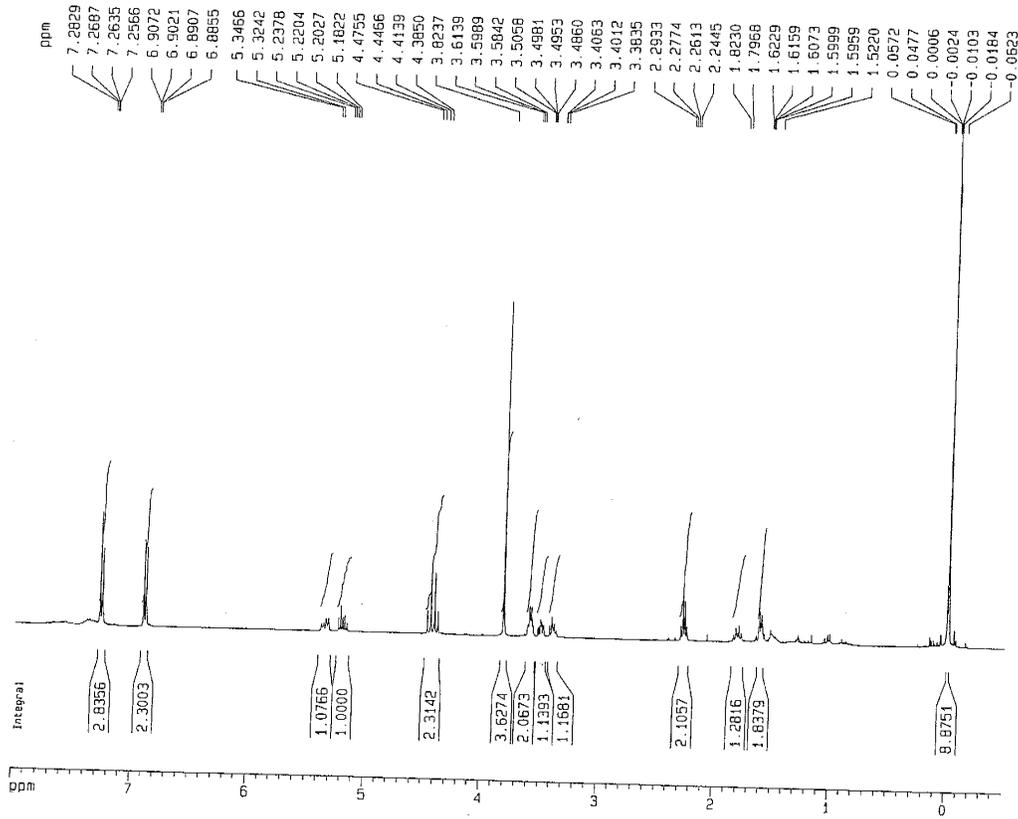
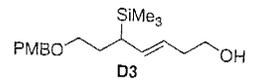
***** CHANNEL f1 *****
 NUC1 13C
 P1 6.20 usec
 PL1 3.00 dB
 SFO1 125.7718472 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCR02 95.00 usec
 PL2 -4.00 dB
 PL12 19.00 dB
 PL13 30.00 dB
 SFO2 500.1325000 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7578008 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

F1 - Processing parameters
 SI 1024
 HC2 OF
 SF 500.1300000 MHz
 WDW no
 SSB 0
 LB 0.30 Hz
 GB 0

ID NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 FIP 260.000 ppm
 F1 25151.56 Hz
 F2P -10.000 ppm
 F2 -1257.58 Hz
 PPMCH 10.30000 ppm/cm
 HZCN 1320.45931 Hz/cm



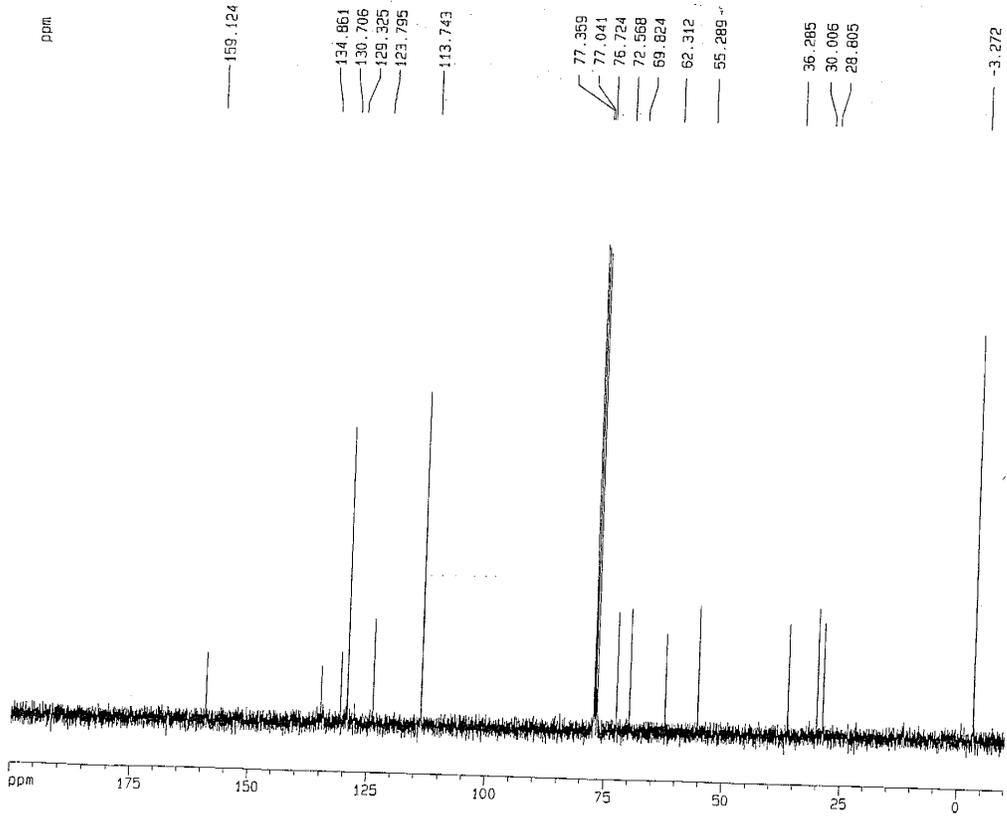
Current Data Parameters
 NAME wj5-148
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20050322
 Time 18.30
 INSTRUM spect
 PROBHD 5 mm QNP 1H/15
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 161.3
 DW 104.400 usec
 DE 6.00 usec
 TE 295.1 K
 D1 1.00000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.05 usec
 PL1 -3.00 dB
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 8.000 ppm
 F1 3201.04 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPHCM 0.42500 ppm/cm
 HZCM 170.05525 Hz/cm



Current Data Parameters
 NAME wj5-148
 EXPNO 2
 PROCNO 1

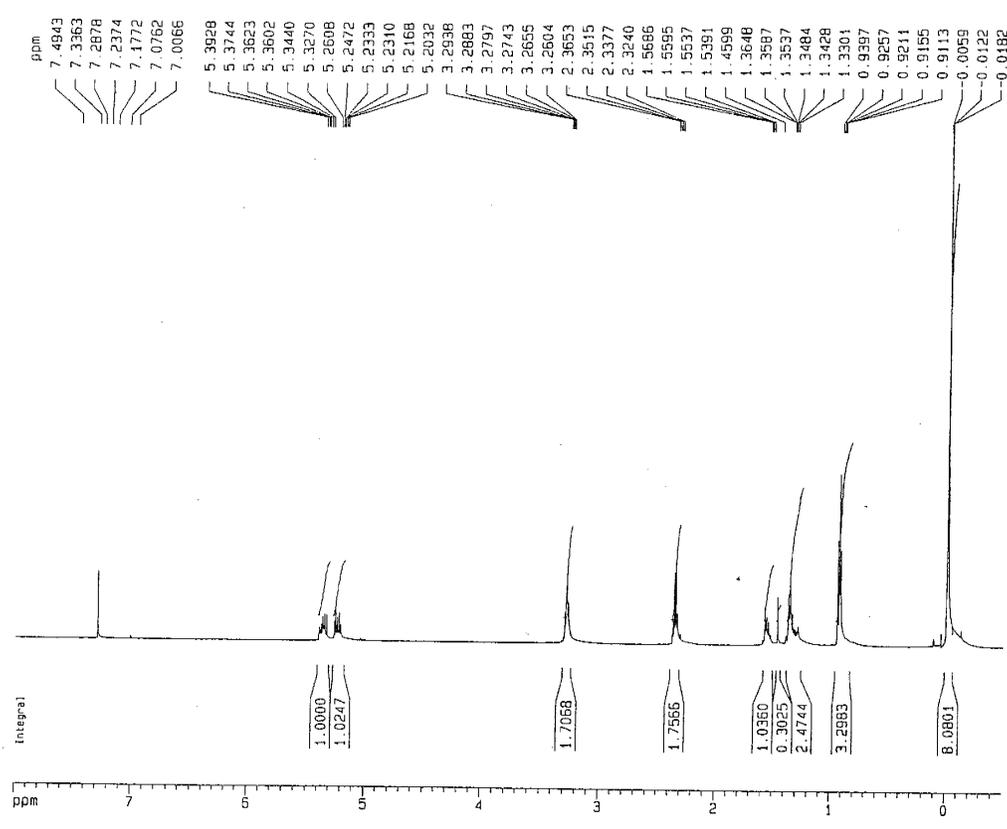
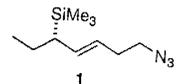
F2 - Acquisition Parameters
 Date_ 20050322
 Time 18.34
 INSTRUM spect
 PROBHD 5 mm QNP 1H/15
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 263
 DS 4
 SWH 25980.814 Hz
 FIDRES 0.365818 Hz
 AQ 1.3564756 sec
 RG 2048
 DW 20.050 usec
 DE 6.02 usec
 TE 296.2 K
 D1 0.15000001 sec
 D11 0.03000000 sec
 DELTA 0.05000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 7.20 usec
 PL1 -2.00 dB
 SFO1 100.6282898 MHz

***** CHANNEL f2 *****
 CHSPRG2 waltz16
 NUC2 1H
 PCPD2 88.40 usec
 PL2 -3.00 dB
 PL12 19.96 dB
 PL13 120.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127650 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 200.000 ppm
 F1 20122.55 Hz
 F2P -10.000 ppm
 F2 -1006.13 Hz
 PPHCM 10.50000 ppm/cm
 HZCM 1056.43408 Hz/cm



Current Data Parameters
NAME wj3-153
EXNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20011213
Time 16.44
INSTRUM spect
PROBHD 5 mm Txi 1H-13
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 6005.615 Hz
FIDRES 0.193358 Hz
AQ 2.7264309 sec
RG 203.2
DW 83.200 usec
DE 5.00 usec
TE 300.0 K
D1 0.03000000 sec

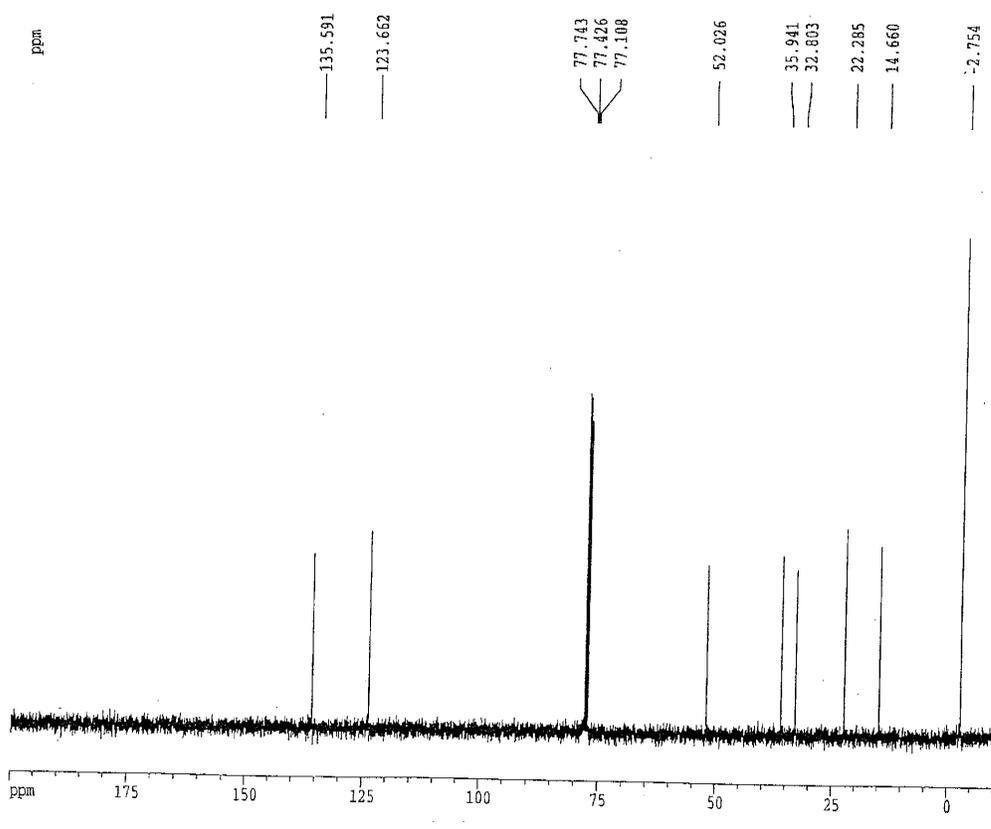
==== CHANNEL f1 =====
NUC1 1H
P1 7.70 usec
PL1 -4.00 dB
SFO1 500.1320005 MHz

F1 - Acquisition Parameters
NUC2 13C
TD 256
SFO1 500.1325 MHz
FIDRES 23.475050 Hz
SW 12.015 ppm

F2 - Processing parameters
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0.0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 DF
SF 500.1300000 MHz
WDW OSINE
SSB 0
LB 0.30 Hz
GB 0

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
FIP 8.900 ppm
F1 4001.04 Hz
F2 -0.500 ppm
FZ -250.07 Hz
PPHCK
HZCM 212.5525 Hz/cm



Current Data Parameters
NAME wj3-24.c
EXNO 2
PROCNO 1

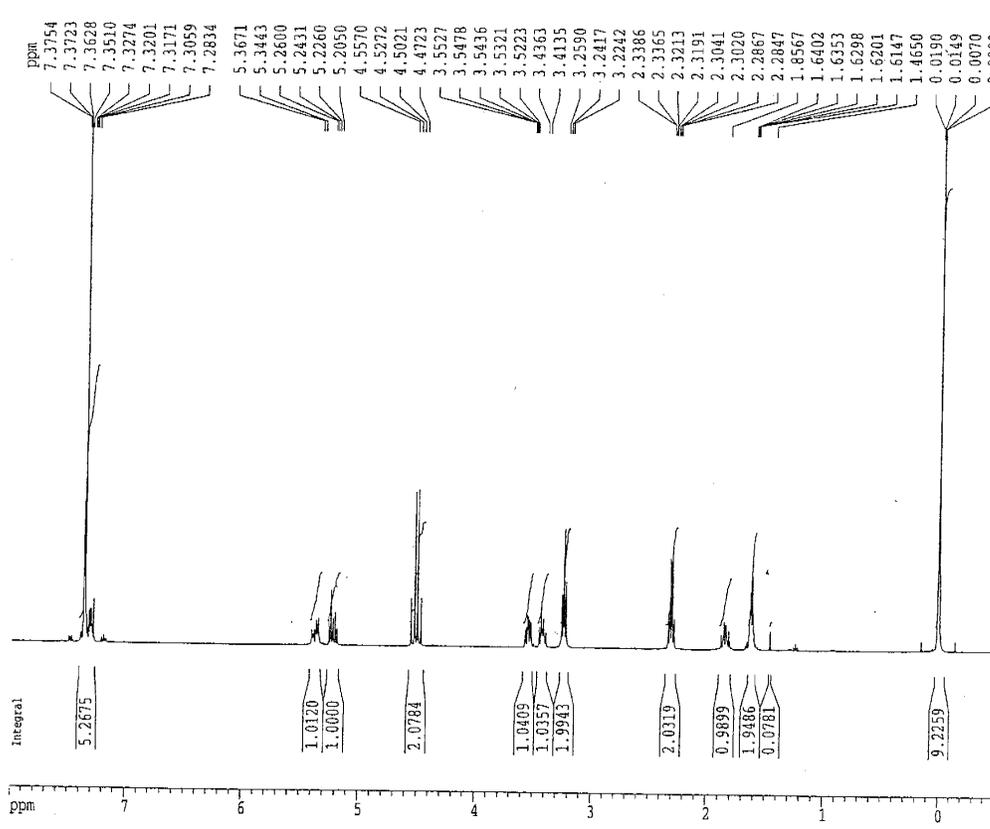
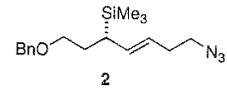
F2 - Acquisition Parameters
Date_ 20030506
Time 15.08
INSTRUM dxt400
PROBHD 5 mm Multinuc1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 120
DS 2
SWH 23148.148 Hz
FIDRES 0.351213 Hz
AQ 1.4156276 sec
RG 32768
DW 21.600 usec
DE 4.50 usec
TE 300.0 K
D1 0.05000000 sec
d11 0.03000000 sec
d12 0.00020000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 12.30 usec
PL1 2.00 dB
SFO1 100.6232933 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127230 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
FIP 200.000 ppm
F1 20122.55 Hz
F2P -10.000 ppm
FZ -1006.13 Hz
PPHCK
HZCM 1056.43372 Hz/cm



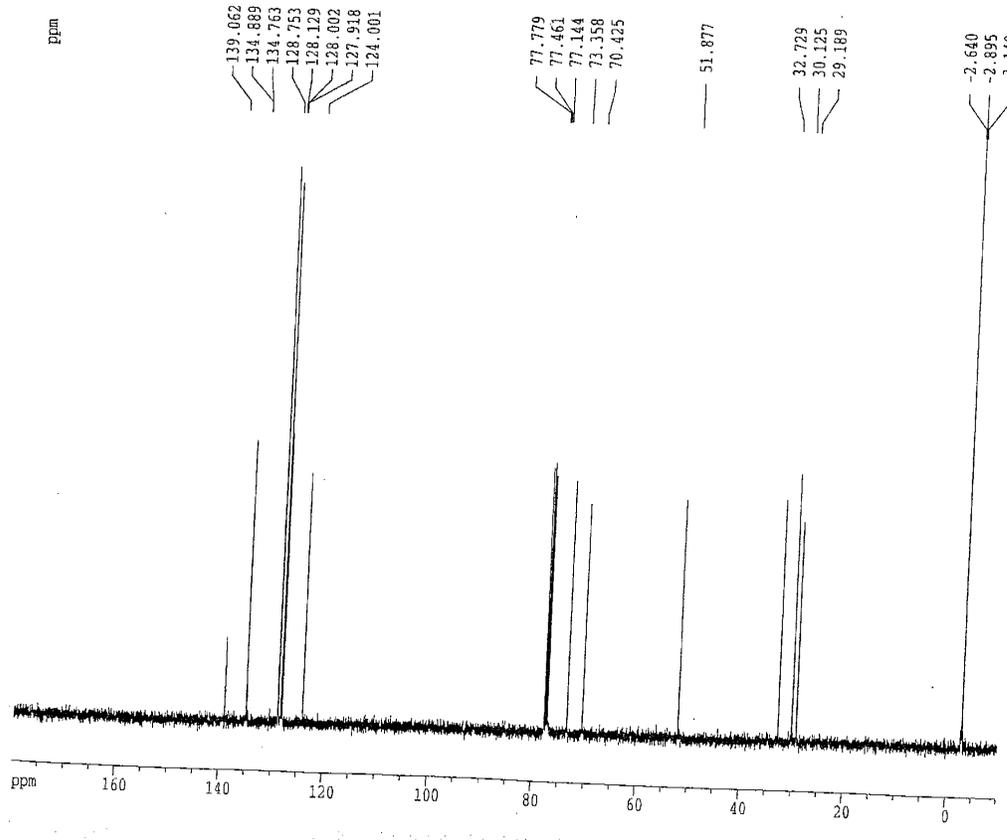
Current Data Parameters
 NAME wj2-200
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20020218
 Time 11.08
 INSTRUM drx400
 PROBHD 5 mm Multino
 PULPROG zgpg30
 TD 32768
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 64
 DW 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 DI 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.70 usec
 PL1 -6.00 dB
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 8.000 ppm
 F1 3201.04 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.42500 ppm/cm
 HZCM 170.05525 Hz/cm



Current Data Parameters
 NAME wj2-200
 EXPNO 2
 PROCNO 1

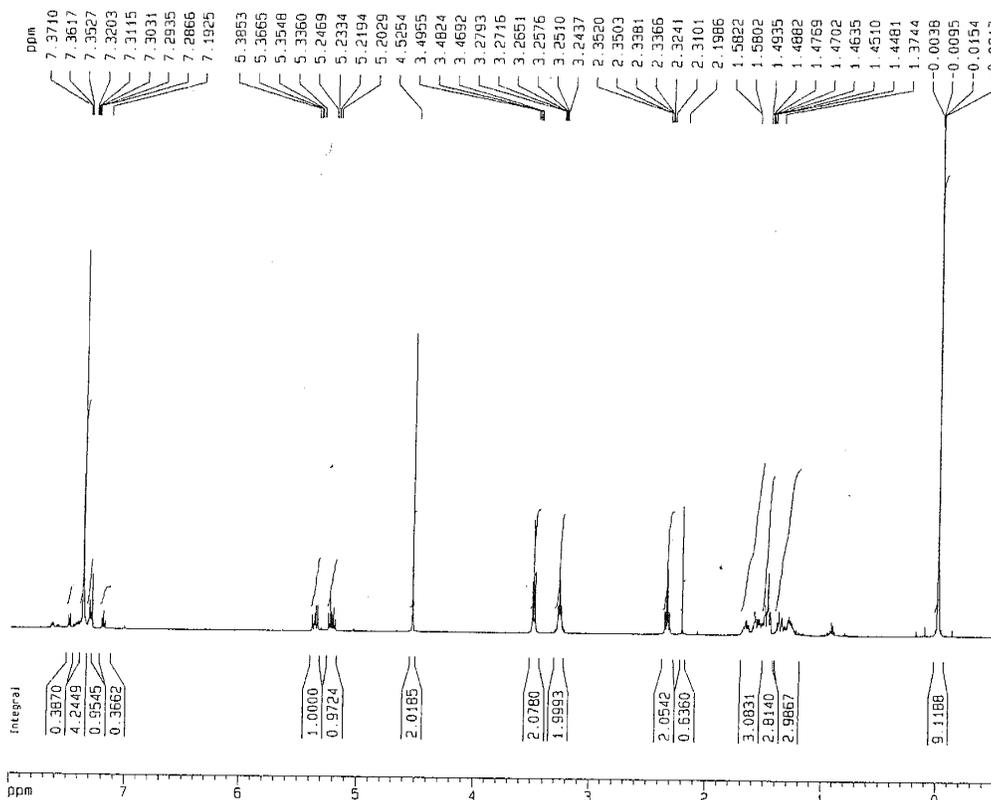
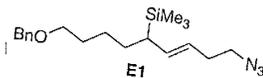
F2 - Acquisition Parameters
 Date_ 20020218
 Time 11.12
 INSTRUM drx400
 PROBHD 5 mm Multino
 PULPROG zgpg30
 TD 65536
 FIDRES 0.351213 Hz
 AQ 1.4156276 sec
 RG 1024
 DW 21.600 usec
 DE 4.50 usec
 TE 300.0 K
 DI 0.05000000 sec
 DI1 0.03000000 sec
 DI2 0.00002000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 12.30 usec
 PL1 2.00 dB
 SFO1 100.6232933 MHz

===== CHANNEL f2 =====
 NUC2 1H
 P2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127290 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 180.000 ppm
 F1 18110.29 Hz
 F2P -10.036 ppm
 F2 -1008.73 Hz
 PPMCM 9.50179 ppm/cm
 HZCM 956.00128 Hz/cm



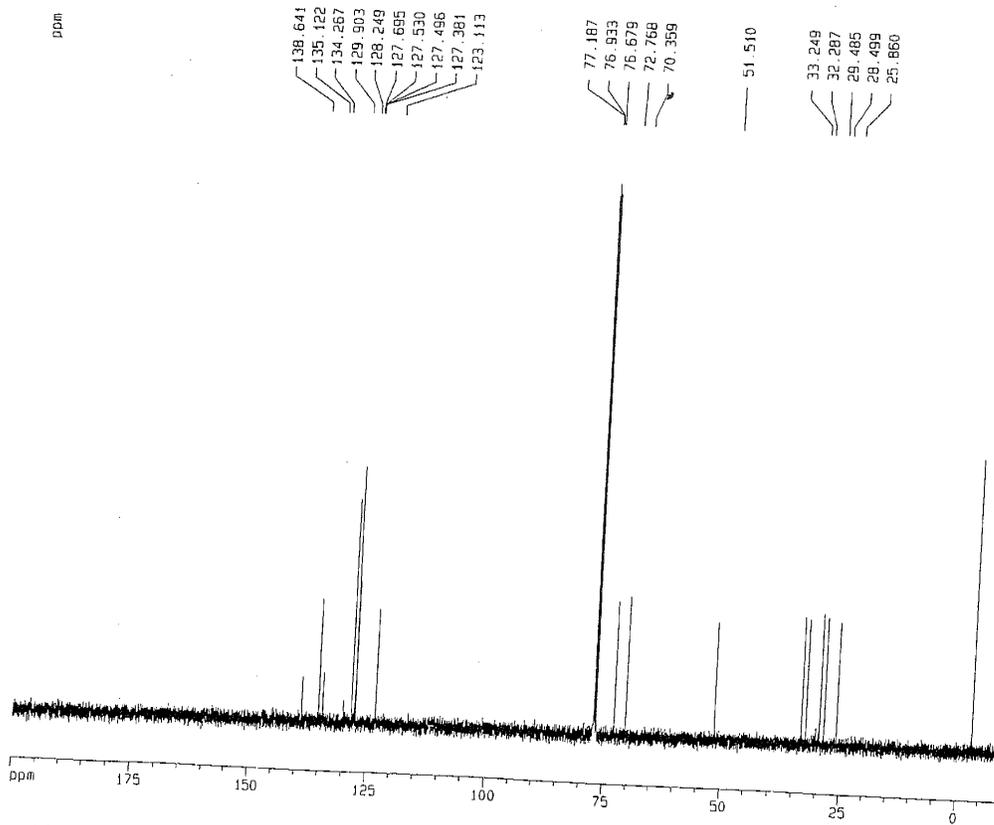
Current Data Parameters
 NAME w13-278
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20030427
 Time 15.38
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SMH 609.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7284309 Sec
 RG 181
 DM 83.200 usec
 DE 5.00 usec
 TE 300.0 K
 D1 0.0300000 Sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.70 usec
 PL1 -4.00 dB
 SFO1 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 500.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

F1 - Processing parameters
 SI 1024
 MC2 OF
 SF 500.1300000 MHz
 WDW 9514C
 SSB 0
 LB 0.30 Hz
 GB 0

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 8.000 ppm
 F1 4001.04 Hz
 F2 -9.500 ppm
 F2 -250.07 Hz
 PRMCM 0.42500 ppm/cm
 HZCM 212.55525 Hz/cm



Current Data Parameters
 NAME w13-278
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20030427
 Time 16.42
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 216
 DS 4
 SMH 31446.641 Hz
 FIDRES 0.479836 Hz
 AQ 1.0420693 Sec
 RG 2048
 DM 19.500 usec
 DE 5.00 usec
 TE 300.0 K
 D1 0.1000000 Sec
 D11 0.0200000 Sec
 D12 0.0000000 Sec

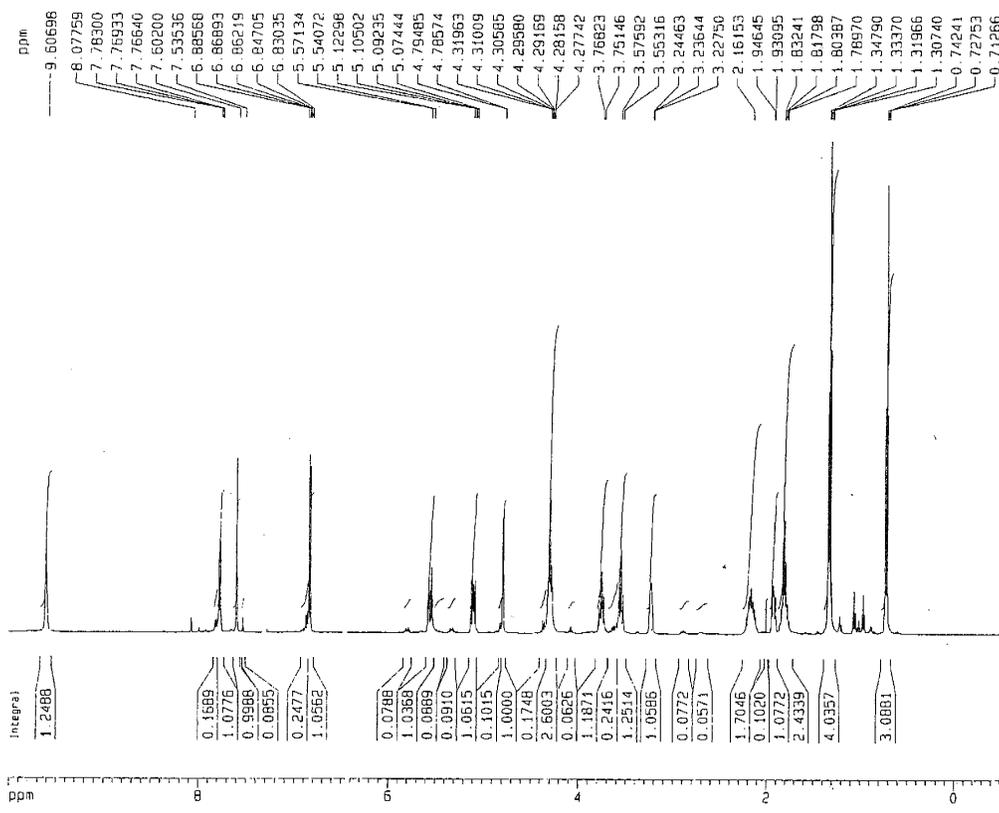
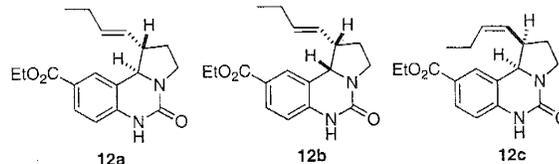
***** CHANNEL f1 *****
 NUC1 13C
 P1 6.50 usec
 PL1 5.00 dB
 SFO1 125.7718472 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 95.00 usec
 PL2 -4.00 dB
 PL12 19.00 dB
 PL13 30.00 dB
 SFO2 500.1325000 MHz

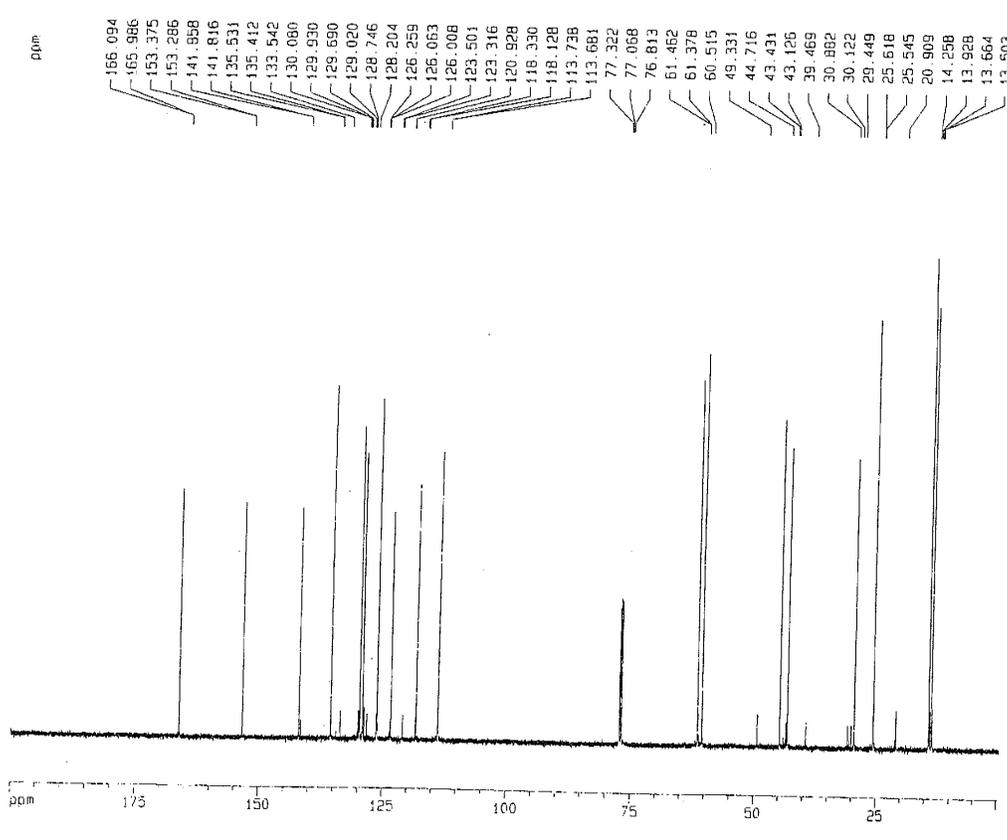
F2 - Processing parameters
 SI 65536
 SF 125.7578008 MHz
 WDW EN
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

F1 - Processing parameters
 SI 1024
 MC2 OF
 SF 500.1300000 MHz
 WDW 9514C
 SSB 0
 LB 0.30 Hz
 GB 0

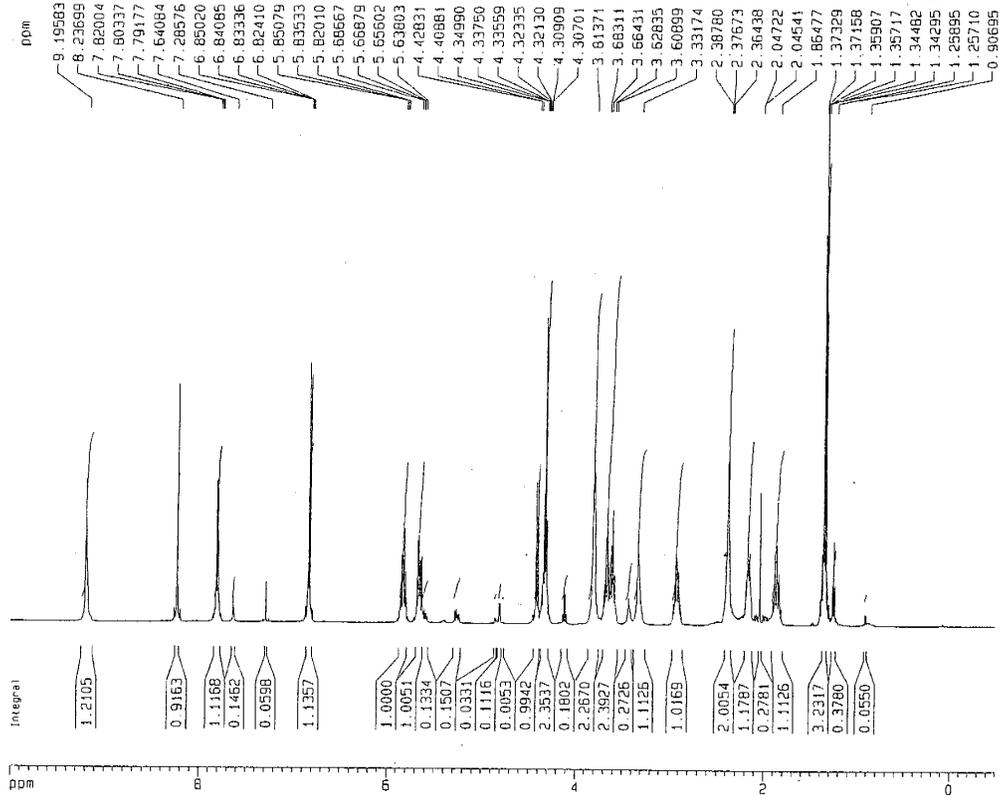
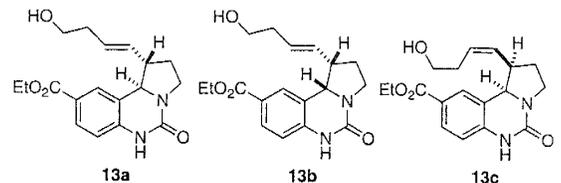
1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 200.000 ppm
 F1 25151.56 Hz
 F2 -1257.58 Hz
 PRMCM 10.30000 ppm/cm
 HZCM 1320.45891 Hz/cm



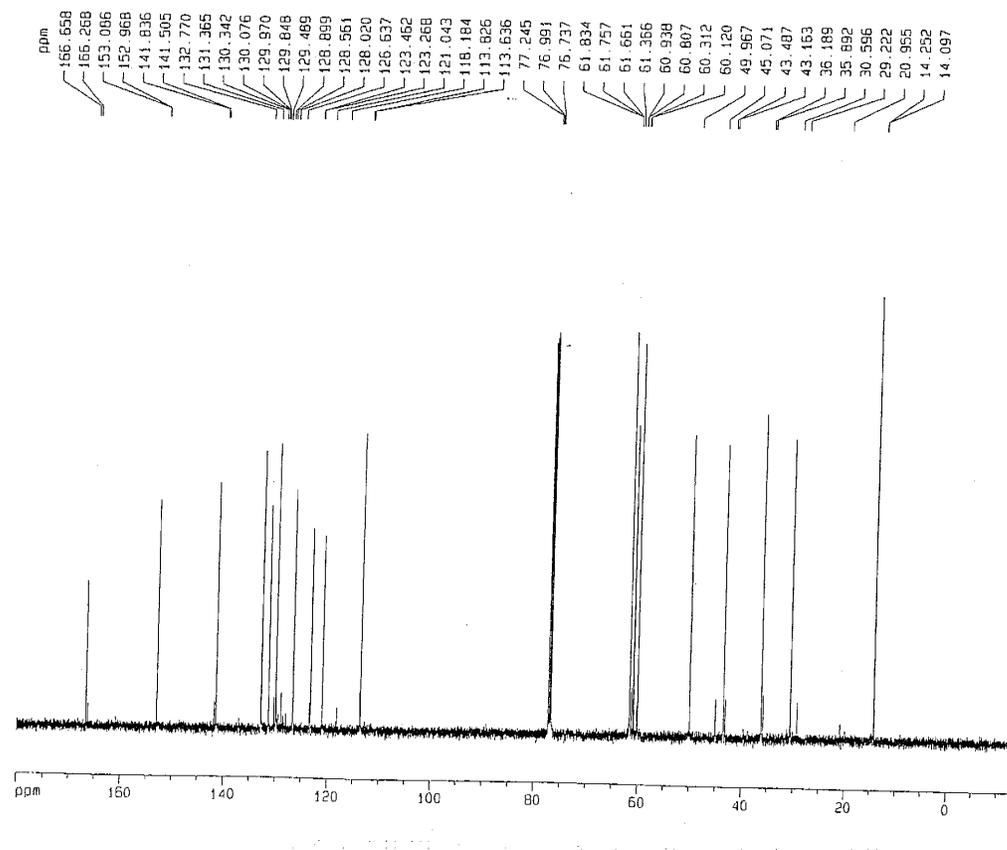
Current Data Parameters
 NAME wj3-197
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20030115
 Time 13.24
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183389 Hz
 AQ 2.7284309 sec
 RG 32
 OW 83.200 usec
 DE 6.00 usec
 TE 300.0 K
 D1 0.0300000 sec
 ***** CHANNEL f1 *****
 NUC1 1H
 P1 7.70 usec
 PL1 -4.00 dB
 SFO1 500.1300005 MHz
 F2 - Processing parameters
 SI 32768
 SF 500.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00
 F1 - Processing parameters
 SI 1024
 MC2 OF
 SF 500.1300000 MHz
 WDW QSINE
 SSB 0
 LB 0.30 Hz
 GB 0
 ID NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 FIP 10.000 ppm
 F1 5001.30 Hz
 F2P -0.500 ppm
 F2 -250.07 Hz
 PPMCH 0.02500 ppm/cm
 HZCM 262.56824 Hz/cm

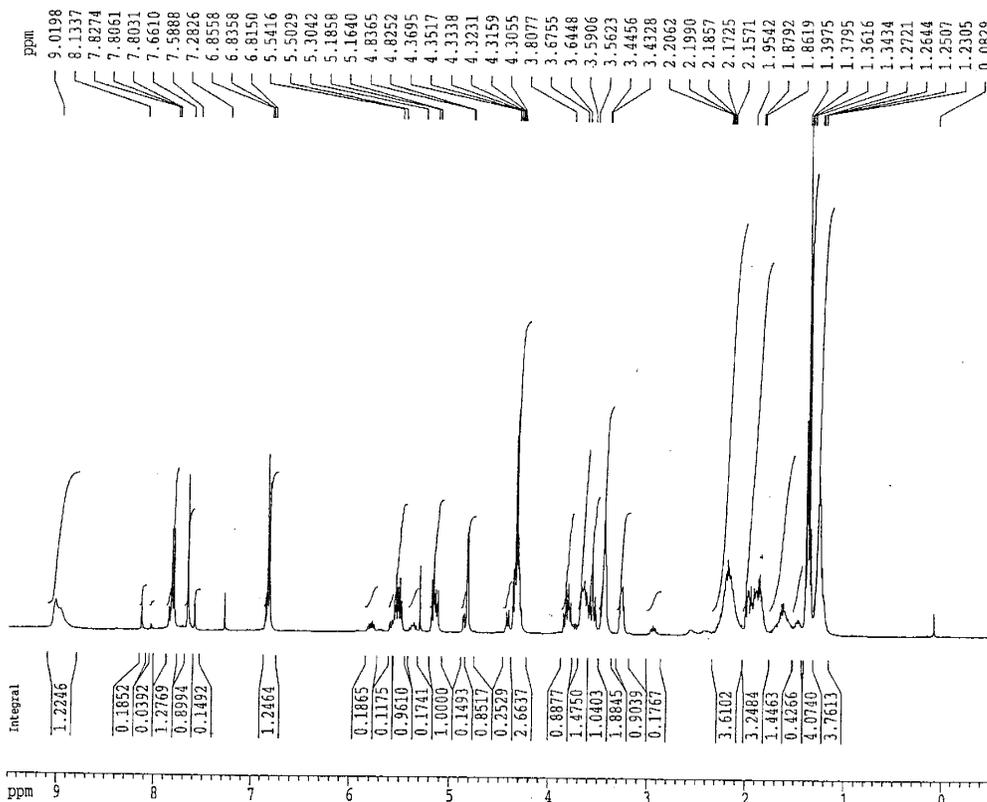
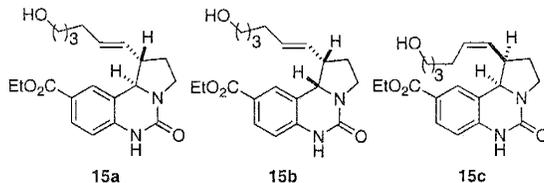


Current Data Parameters
 NAME wj3-197
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20030115
 Time 13.30
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 252
 DS 4
 SWH 31448.541 Hz
 FIDRES 0.478838 Hz
 AQ 1.0428888 sec
 RG 14588.5
 OW 15.900 usec
 DE 6.00 usec
 TE 300.0 K
 D1 0.1000000 sec
 D11 0.0300000 sec
 D12 0.0002000 sec
 ***** CHANNEL f1 *****
 NUC1 13C
 P1 6.50 usec
 PL1 5.00 dB
 SFO1 125.7718472 MHz
 ***** CHANNEL f2 *****
 NUC2 1H
 P2 7.70 usec
 PL2 -4.00 dB
 SFO2 500.1300000 MHz
 F2 - Processing parameters
 SI 65536
 SF 125.7578008 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40
 F1 - Processing parameters
 SI 1024
 MC2 OF
 SF 500.1300000 MHz
 WDW NO
 SSB 0
 LB 0.30 Hz
 GB 0
 ID NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 FIP 100.000 ppm
 F1 2510.56 Hz
 F2P -0.500 ppm
 F2 -62.88 Hz
 PPMCH 0.02500 ppm/cm
 HZCM 1260.78192 Hz/cm



EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20010821
 Time 10.08
 INSTRUM spect
 PROBMG 5 mm BBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183398 Hz
 AQ 2.7263477 sec
 RG 57
 DW 83.200 usec
 DE 5.00 usec
 TE 300.2 K
 D1 0.0300000 sec
 ----- CHANNEL f1 -----
 NUC1 1H
 P1 7.70 usec
 PL1 -4.00 dB
 SFO1 500.1320005 MHz
 F1 - Acquisition parameters
 MD 2
 TD 256
 SFO1 500.1325 MHz
 FIDRES 23.475050 Hz
 SW 12.016 ppm
 F2 - Processing parameters
 SI 32768
 SF 500.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00
 F1 - Processing parameters
 SI 1024
 HC 0F
 SF 500.1300000 MHz
 WDW no
 SSB 0
 LB 0.30 Hz
 GB 0
 ID NMR plot parameters
 CX 20.00 cm
 FIP 10.005 ppm
 FL 5005.33 Hz
 FFP -0.500 ppm
 FZ -250.07 Hz
 PWDW 0.52540 ppm/cm
 HZCM 282.76935 Hz/cm
 Current Data Parameters
 NAME 13-68
 EXPNO 7
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20010821
 Time 16.14
 INSTRUM spect
 PROBMG 5 mm BBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 387
 DS 4
 SWH 31446.541 Hz
 FIDRES 0.479838 Hz
 AQ 1.0480724 sec
 RG 2860.3
 DW 15.900 usec
 DE 6.00 usec
 TE 300.2 K
 D1 0.1000000 sec
 D11 0.0300000 sec
 D12 0.00002000 sec
 ----- CHANNEL f1 -----
 NUC1 13C
 P1 6.50 usec
 PL1 5.00 dB
 SFO1 125.7719472 MHz
 ----- CHANNEL f2 -----
 CPDPRG2 waltz16
 NUC2 1H
 PWDW 95.00 usec
 PL2 17.50 dB
 PL12 19.00 dB
 PL13 30.00 dB
 SFO2 500.1325000 MHz
 F1 - Acquisition parameters
 MD 2
 TD 256
 SFO1 500.1325 MHz
 FIDRES 23.475050 Hz
 SW 12.016 ppm
 F2 - Processing parameters
 SI 65536
 SF 125.7576000 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40
 F1 - Processing parameters
 SI 1024
 HC 0F
 SF 500.1300000 MHz
 WDW no
 SSB 0
 LB 0.30 Hz
 GB 0
 ID NMR plot parameters
 CX 20.00 cm
 FIP 100.000 ppm
 FL 22836.00 Hz
 FFP -12.539 ppm
 FZ 1576.91 Hz
 PWDW 3.62686 ppm/cm
 HZCM 1210.66527 Hz/cm





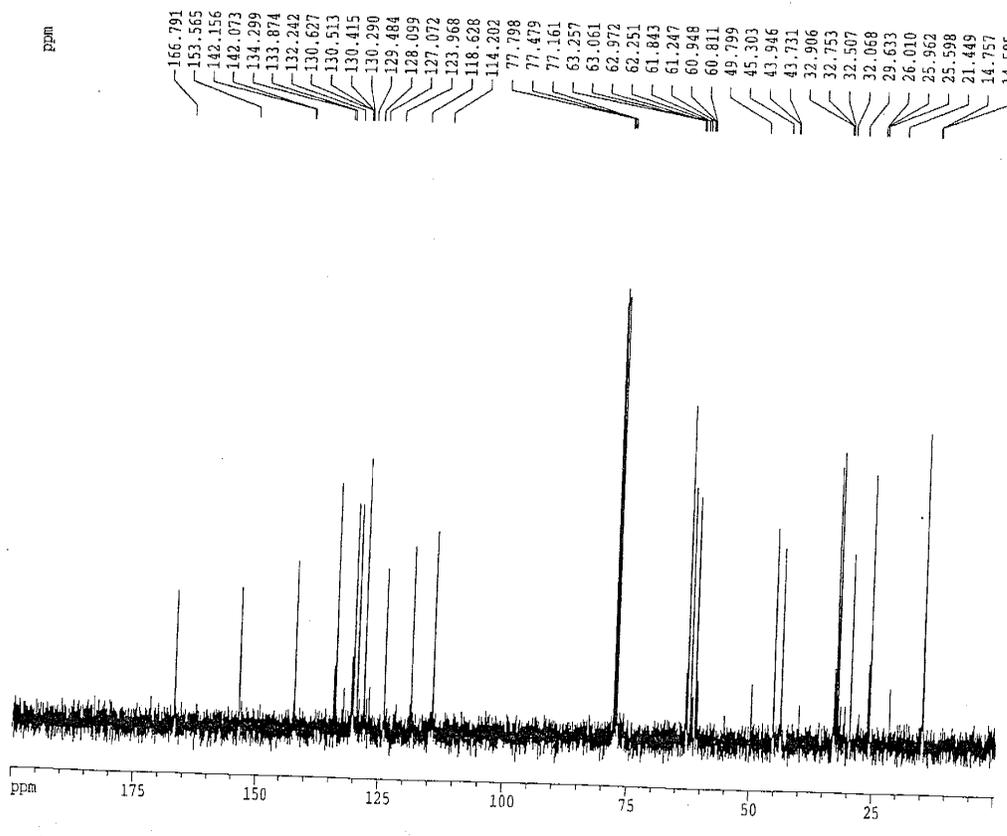
NAME wj4-56
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20030729
Time 14.39
INSTRUM drx400
PROBHD 5 mm Multinucl
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 64
DW 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 7.70 usec
PL1 -6.00 dB
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ID NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 9.500 ppm
F1 3801.24 Hz
F2P -0.500 ppm
F2 -200.07 Hz
FPMCH 0.50000 ppm/cm
HZCM 200.06500 Hz/cm



Current Data Parameters
NAME wj4-56, 24
EXPNO 2
PROCNO 1

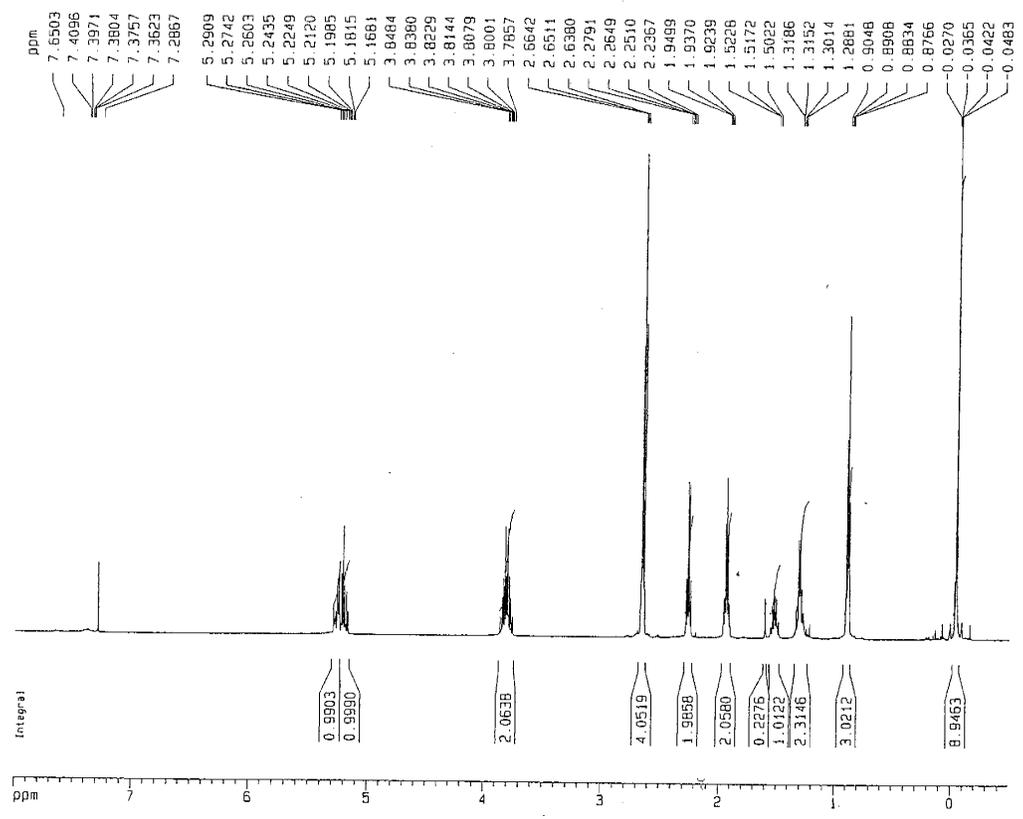
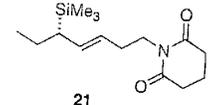
F2 - Acquisition Parameters
Date_ 20030729
Time 16.27
INSTRUM drx400
PROBHD 5 mm Multinucl
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 182
DS 2
SWH 23148.148 Hz
FIDRES 0.353213 Hz
AQ 1.4156276 sec
RG 32768
DW 21.600 usec
DE 4.50 usec
TE 300.0 K
D1 0.05000000 sec
d11 0.03000000 sec
d12 0.00002000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 12.30 usec
PL1 2.00 dB
SFO1 100.6232933 MHz

===== CHANNEL f2 =====
CFPRPG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127290 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

ID NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 200.000 ppm
F1 20122.55 Hz
F2P -50.31 Hz
F2 -10.02500 ppm/cm
FPMCH 1008.64264 Hz/cm



Current Data Parameters
NAME w13-185
EXPNO 1
PROCNO 1

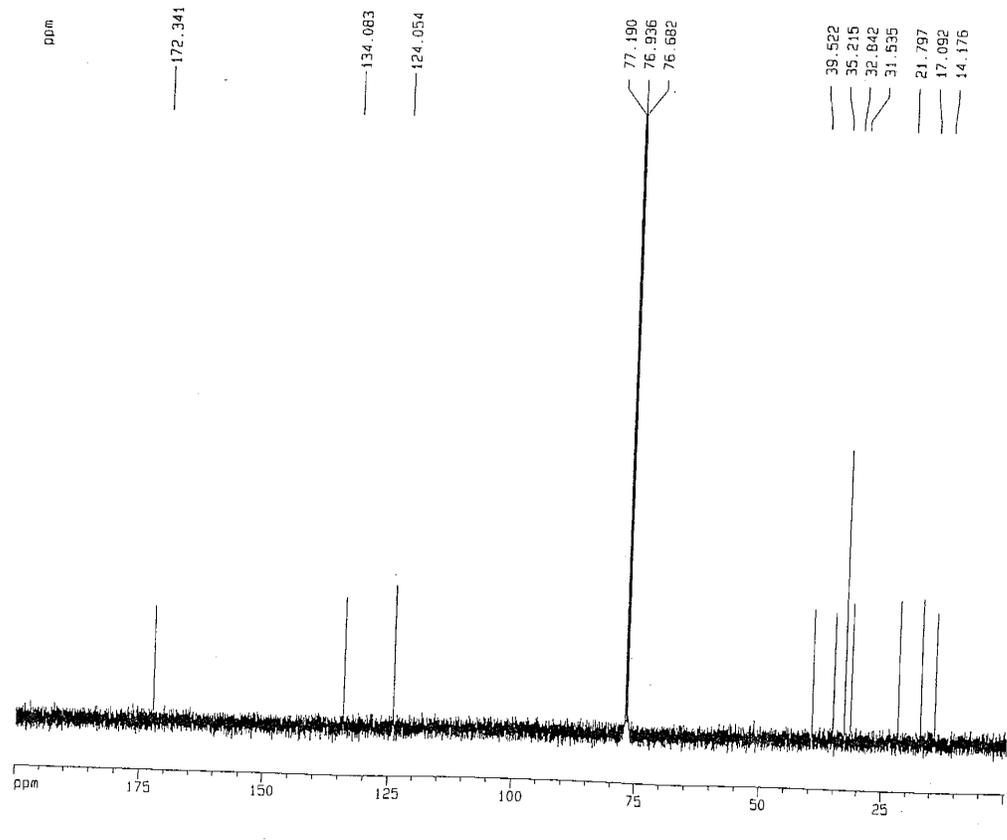
F2 - Acquisition Parameters
Date_ 20030120
Time 12.43
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 6009.615 Hz
FIDRES 0.183399 Hz
AQ 2.7264309 sec
RG 256
OW 83.200 usec
DE 6.00 usec
TE 300.0 K
D1 0.03000000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -4.00 dB
SF01 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 OF
SF 500.1300000 MHz
WDW OSINE
SSB 0
LB 0.30 Hz
GB 0

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 8.000 ppm
F1 4001.04 Hz
F2P -0.500 ppm
F2 -250.07 Hz
PACH 0.42500 ppm/cm
HZCM 212.55525 Hz/cm



Current Data Parameters
NAME w13-185
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20030120
Time 12.48
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 204
DS 4
SWH 31446.541 Hz
FIDRES 0.479836 Hz
AQ 1.0420003 sec
RG 18384
DW 15.900 usec
DE 5.00 usec
TE 300.0 K
D1 0.10000000 sec
d11 0.03000000 sec
d12 0.00002000 sec

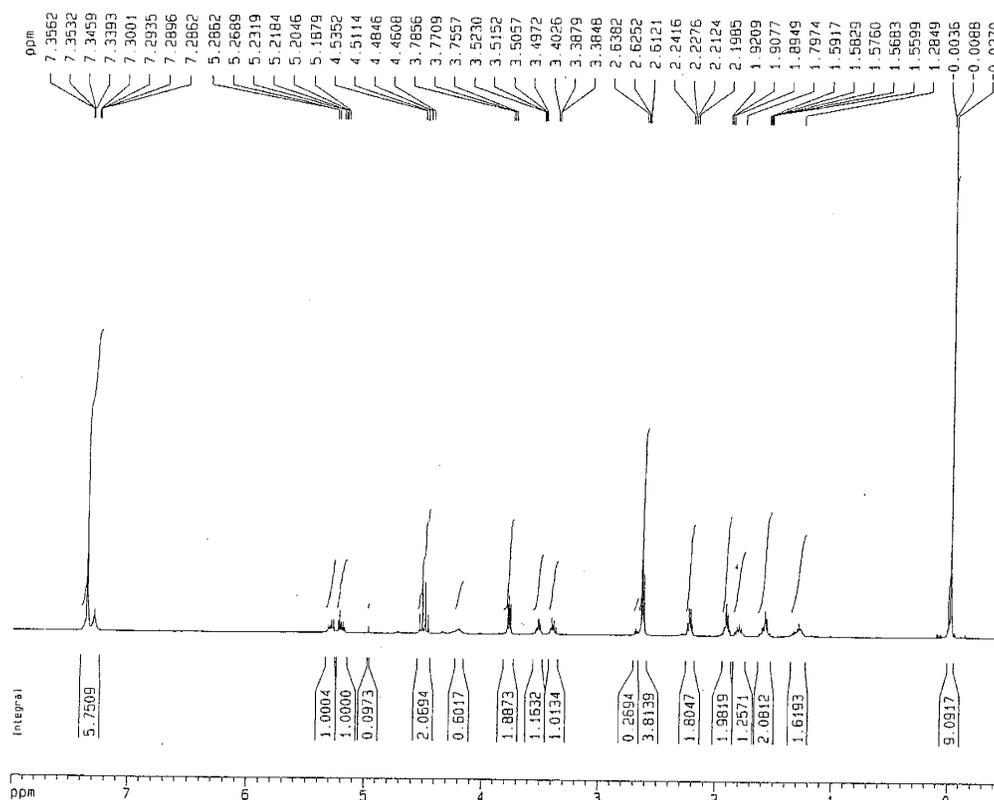
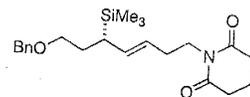
***** CHANNEL f1 *****
NUC1 13C
P1 6.50 usec
PL1 5.00 dB
SF01 125.7719472 MHz

***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 95.00 usec
PL2 -4.00 dB
PL12 19.00 dB
PL13 30.00 dB
SF02 500.1325000 MHz

F2 - Processing parameters
SI 65536
SF 125.7578008 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 OF
SF 500.1300000 MHz
WDW no
SSB 0
LB 0.30 Hz
GB 0

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 200.000 ppm
F1 25151.55 Hz
F2P -0.300 ppm
F2 -62.08 Hz
PACH 10.82500 ppm/cm
HZCM 1260.72192 Hz/cm



Current Data Parameters

NAME w12-74

EXPNO 1

PROCNO 1

F2 - Acquisition Parameters

Date_ 20010902

Time 14.47

INSTRUM spect

PROBHD 5 mm BBO BB-

PULPROG zg30

TD 32768

SOLVENT CDCl3

NS 16

DS 2

SWH 6009.615 Hz

FIDRES 0.133399 Hz

AQ 2.7263477 sec

RG 57

DM 83.200 usec

DE 6.00 usec

TE 300.0 K

D1 0.03000000 sec

----- CHANNEL f1 -----

NUC1 1H

P1 7.70 usec

PL1 -4.00 dB

SFO1 500.1320005 MHz

F1 - Acquisition parameters

TD 2

SI 256

SF 500.1325000 MHz

FIDRES 23.475060 Hz

SW 12.016 ppm

F2 - Processing parameters

SI 32768

SF 500.1300000 MHz

WDW EM

SSB 0

LB 0.30 Hz

GB 0

PC 1.00

F1 - Processing parameters

SI 1024

MC2 OF

SF 500.1300000 MHz

WDW no

SSB 0

LB 0.30 Hz

GB 0

10 NMR plot parameters

CX 20.00 cm

F1P 6.000 ppm

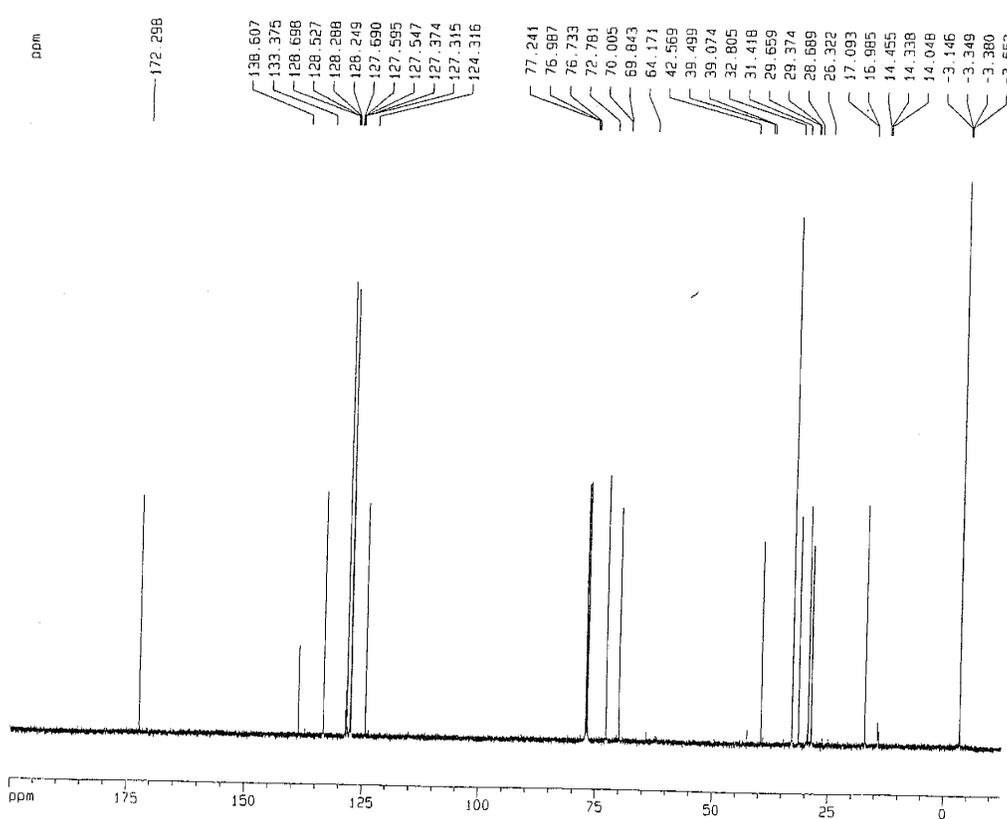
F1 4001.04 Hz

F2P -0.500 ppm

F2 -250.07 Hz

PPMCH 0.42500 ppm/cm

HzCH 212.59525 Hz/cm



Current Data Parameters

NAME w12-74

EXPNO 2

PROCNO 1

F2 - Acquisition Parameters

Date_ 20010902

Time 14.58

INSTRUM spect

PROBHD 5 mm BBO BB-

PULPROG zgpg30

TD 65536

SOLVENT CDCl3

NS 16

DS 4

SWH 31446.541 Hz

FIDRES 0.478205 Hz

AQ 1.0420724 sec

RG 8192

DM 15.900 usec

DE 6.50 usec

TE 300.0 K

D1 0.10000000 sec

D11 0.03000000 sec

D12 0.00020000 sec

----- CHANNEL f1 -----

NUC1 13C

P1 6.50 usec

PL1 5.00 dB

SFO1 125.7719472 MHz

----- CHANNEL f2 -----

CPDPRG2 waltz16

NUC2 1H

PCPD2 95.00 usec

PL2 17.50 dB

PL12 19.00 dB

PL13 30.00 dB

SFO2 500.1325000 MHz

F1 - Acquisition parameters

TD 2

SI 65536

SF 125.7578968 MHz

WDW EM

SSB 0

LB 1.00 Hz

GB 0

PC 1.40

F1 - Processing parameters

SI 1024

MC2 OF

SF 500.1300000 MHz

WDW no

SSB 0

LB 0.30 Hz

GB 0

10 NMR plot parameters

CX 20.00 cm

F1P 200.000 ppm

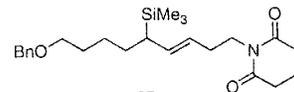
F1 25151.58 Hz

F2P -12.438 ppm

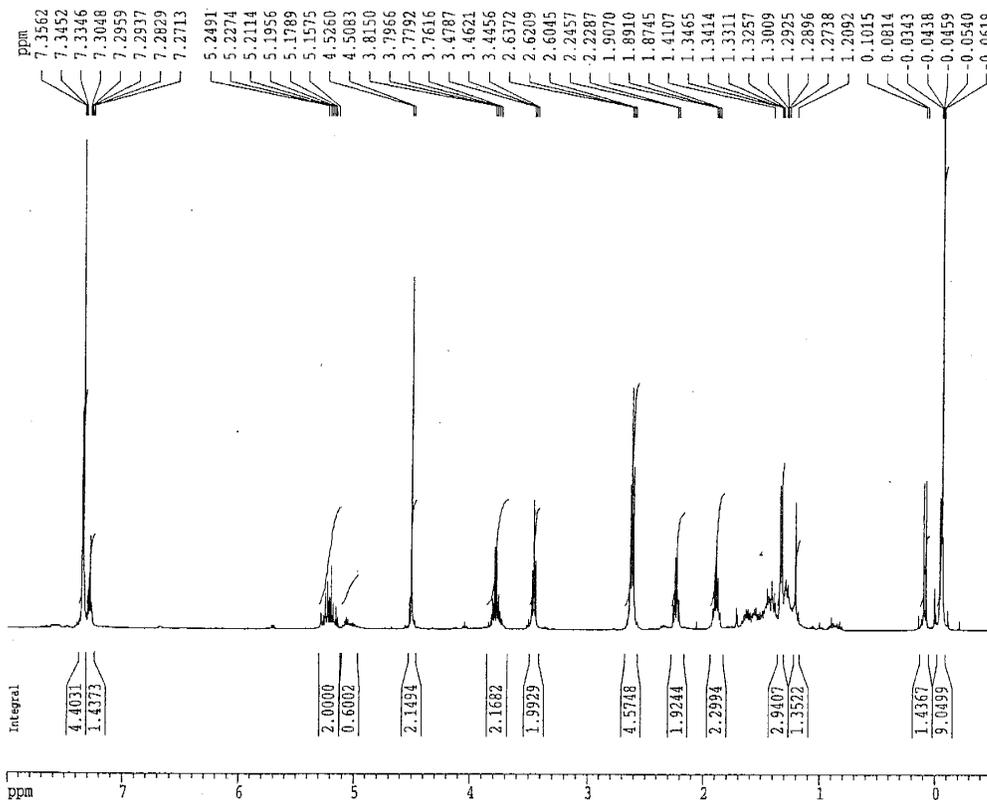
F2 -1976.91 Hz

PPMCH 10.60588 ppm/cm

HzCH 1336.42358 Hz/cm



25



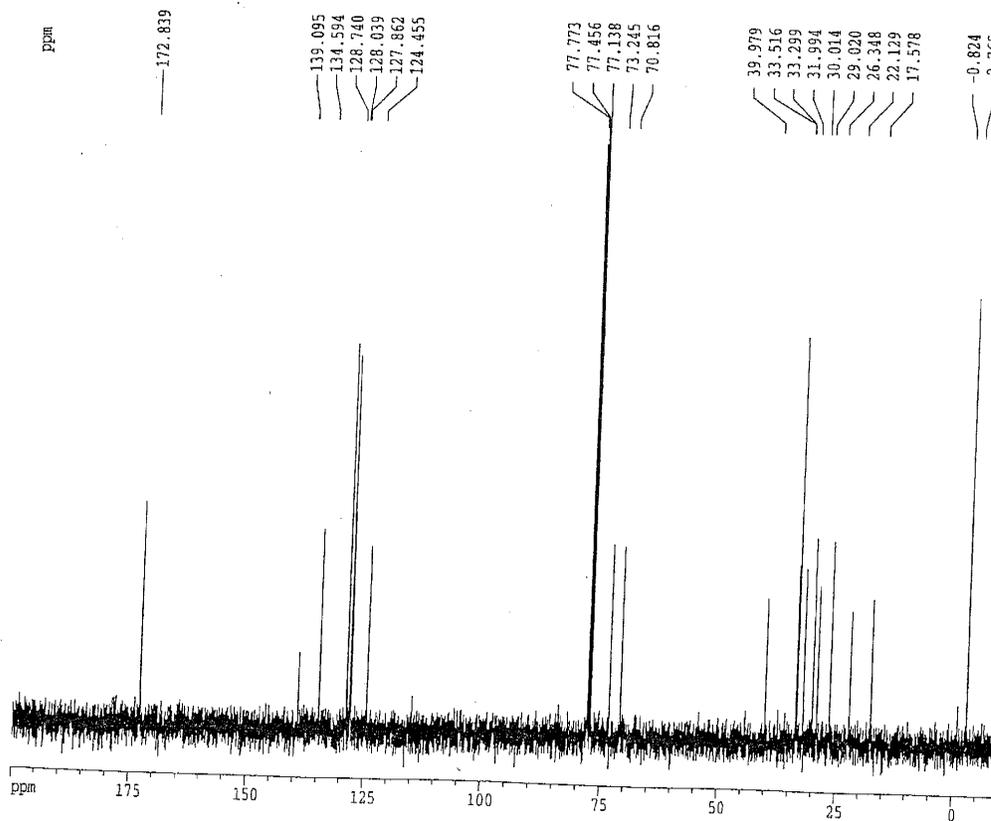
Current Data Parameters
NAME wj3-286
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20030505
Time 13.56
INSTRUM drx400
PROBHD 5 mm Multinucl
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 64
DW 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 7.70 usec
PL1 -6.00 dB
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 8.000 ppm
F1 3201.04 Hz
F2P -200.07 Hz
F2 -200.07 Hz
PPMCM 0.42500 ppm/cm
HZCM 170.05525 Hz/cm



Current Data Parameters
NAME wj3-286
EXPNO 2
PROCNO 1

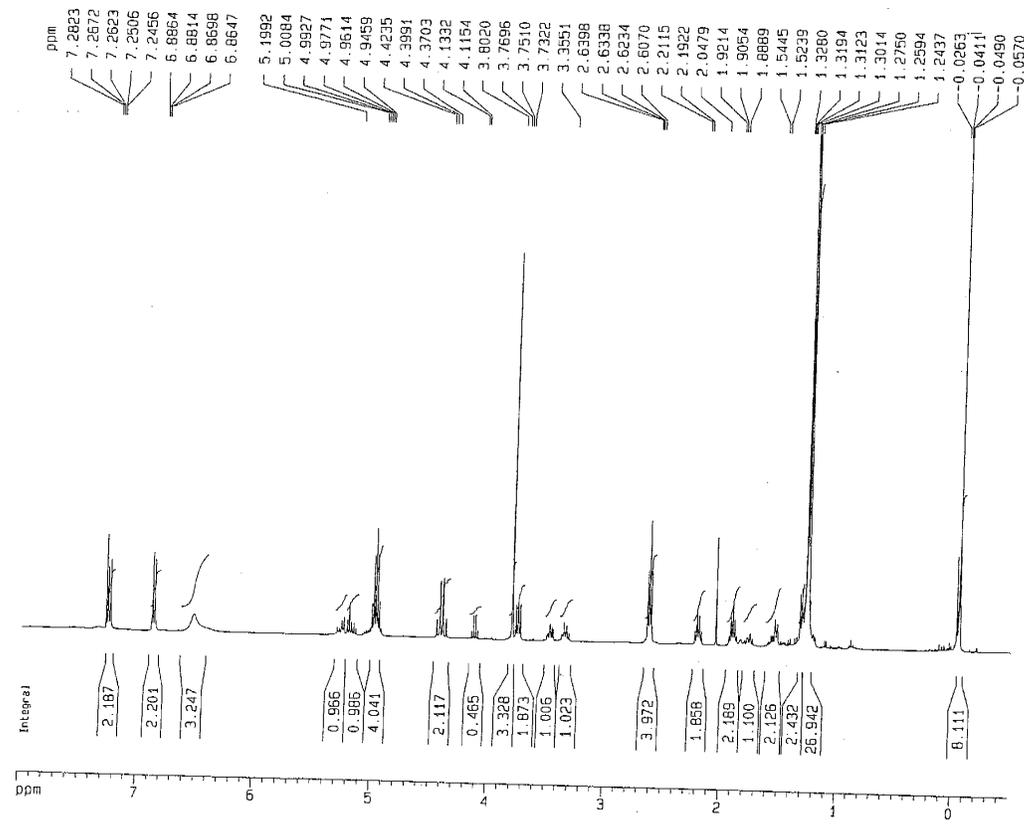
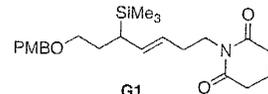
F2 - Acquisition Parameters
Date_ 20030505
Time 13.59
INSTRUM drx400
PROBHD 5 mm Multinucl
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 90
DS 2
SWH 23148.148 Hz
FIDRES 0.353213 Hz
AQ 1.4156276 sec
RG 32768
DW 21.600 usec
DE 4.50 usec
TE 300.0 K
D1 0.05000000 sec
d11 0.03000000 sec
d12 0.00602000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 12.30 usec
PL1 2.00 dB
SFO1 100.6232933 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127290 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 200.000 ppm
F1 20122.55 Hz
F2P -10.000 ppm
F2 -1006.13 Hz
PPMCM 16.50000 ppm/cm
HZCM 1056.43372 Hz/cm



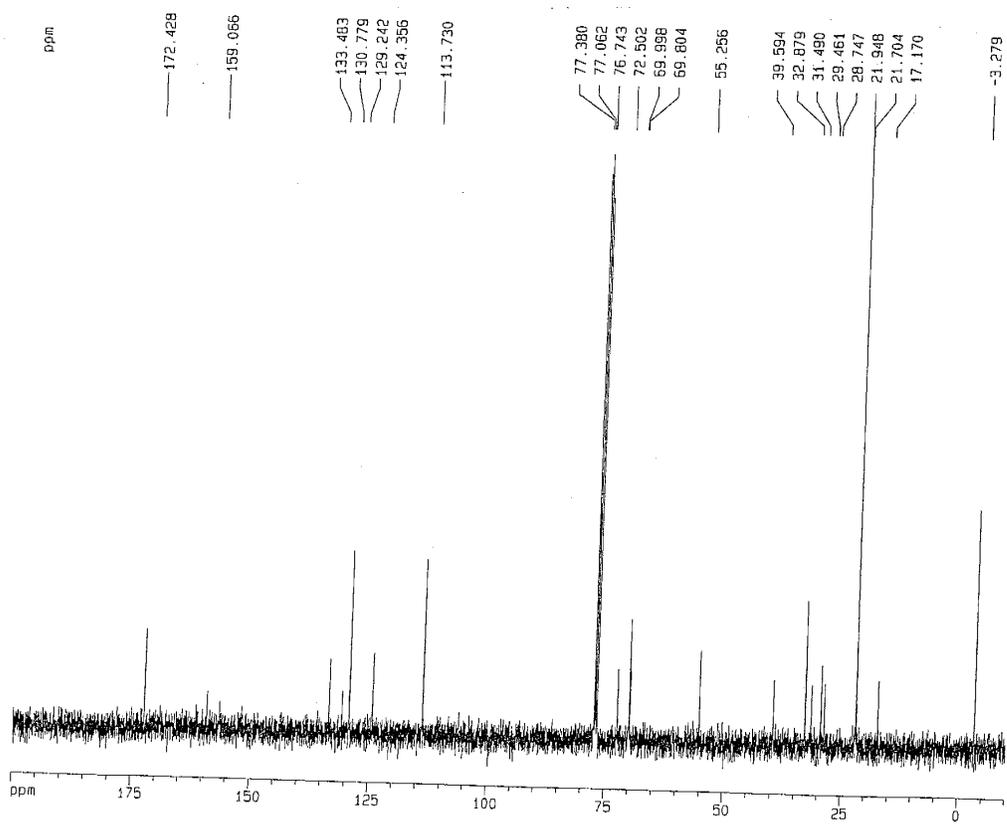
Current Data Parameters
 NAME w15-147-16-21
 EXPNO 149
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20050323
 Time 17.18
 INSTRUM spect
 PROBHD 5 mm QNP 1H/15
 PULPROG zg30
 TO 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 90.5
 DW 104.400 usec
 DE 6.00 usec
 TE 297.1 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWAK 0.0150000 sec

CHANNEL f1
 NUC1 1H
 P1 7.05 usec
 PL1 -3.00 dB
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1320000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 8.000 ppm
 F1 3201.04 Hz
 F2P -0.500 ppm
 F2 -200.67 Hz
 PPMCM 0.42500 ppm/cm
 HZCM 170.05525 Hz/cm



NAME w15-147-16-21
 EXPNO 149
 PROCNO 2

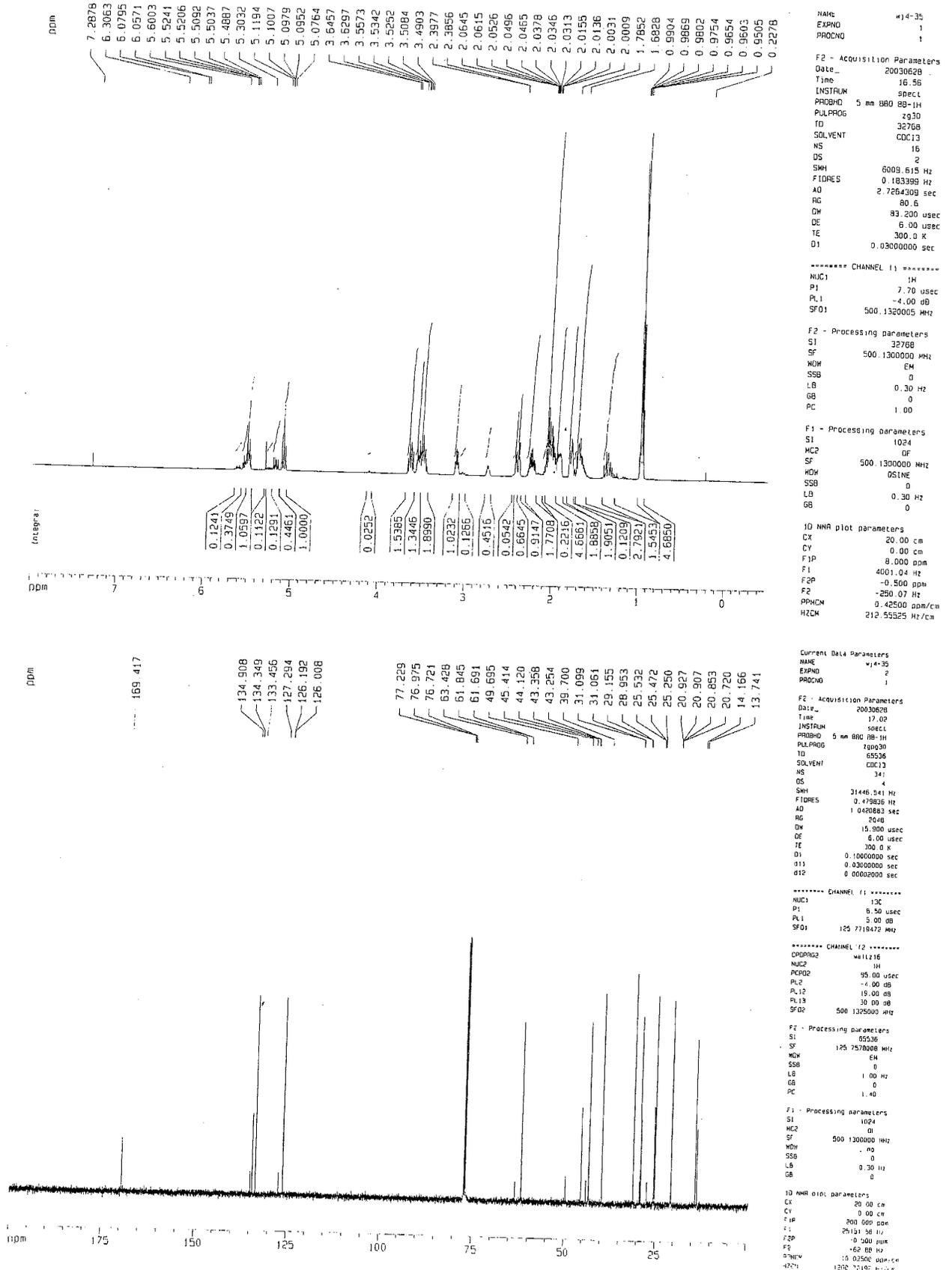
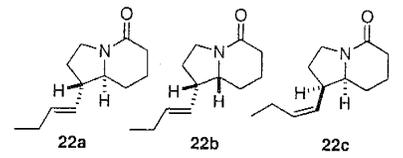
F2 - Acquisition Parameters
 Date_ 20050323
 Time 17.24
 INSTRUM spect
 PROBHD 5 mm QNP 1H/15
 PULPROG zgpg30
 TO 65335
 SOLVENT CDCl3
 NS 382
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.369918 Hz
 AQ 1.3664756 sec
 RG 5048
 DW 20.850 usec
 DE 6.00 usec
 TE 297.2 K
 D1 0.1500001 sec
 D11 0.0300000 sec
 DELTA 0.0500000 sec
 MCREST 0.0000000 sec
 MCWAK 0.0150000 sec

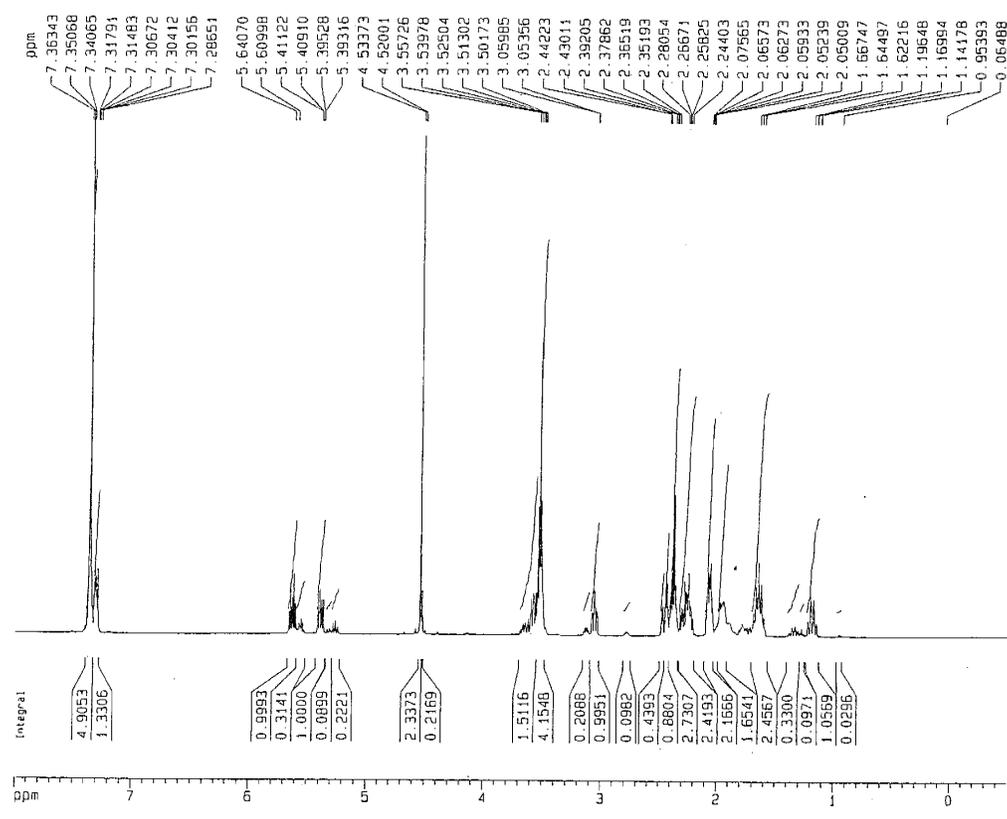
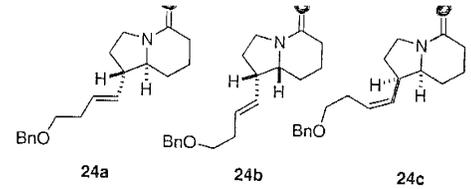
CHANNEL f1
 NUC1 13C
 P1 7.20 usec
 PL1 -2.00 dB
 SFO1 100.6228258 MHz

CHANNEL f2
 CAPPROG2 waltz16
 NUC2 1H
 PCPD2 98.40 usec
 PL2 -3.00 dB
 PL12 19.98 dB
 PL13 120.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127650 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 200.000 ppm
 F1 20122.55 Hz
 F2P -10.000 ppm
 F2 -1008.13 Hz
 PPMCM 10.50000 ppm/cm
 HZCM 1096.43408 Hz/cm





Current Data Parameters
NAME wj3-126
EXPNO 1
PROCNO 1

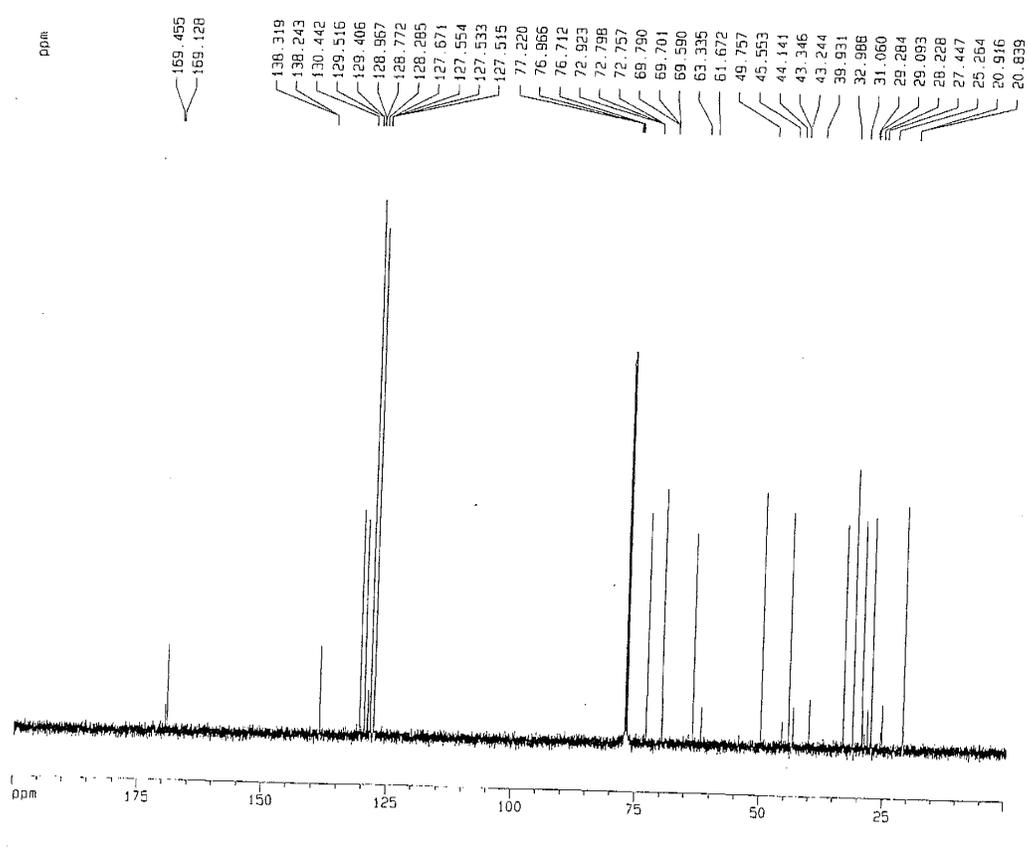
F2 - Acquisition Parameters
Date_ 20020920
Time 11.00
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 32768
SOLVENT COC13
NS 16
DS 2
SWH 6009.615 Hz
FIDRES 0.183399 Hz
AQ 2.7264308 sec
RG 128
DW 83.200 usec
DE 5.00 usec
TE 300.0 K
D1 0.0300000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -4.00 dB
SFO1 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 OF
SF 500.1300000 MHz
WDW GSINE
SSB 0
LB 0.30 Hz
GB 0

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 8.000 ppm
F1 4001.04 Hz
F2P -0.500 ppm
F2 -250.07 Hz
PFMCM 0.42500 ppm/cm
HZCM 212.55525 Hz/cm



Current Data Parameters
NAME wj3-126
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20020920
Time 11.08
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 85336
SOLVENT COC13
NS 422
DS 4
SWH 31446.541 Hz
FIDRES 0.478836 Hz
AQ 1.0420803 sec
RG 15
DW 14596.5
DE 5.00 usec
TE 300.0 K
D1 0.1000000 sec
D11 0.0300000 sec
D12 0.0000200 sec

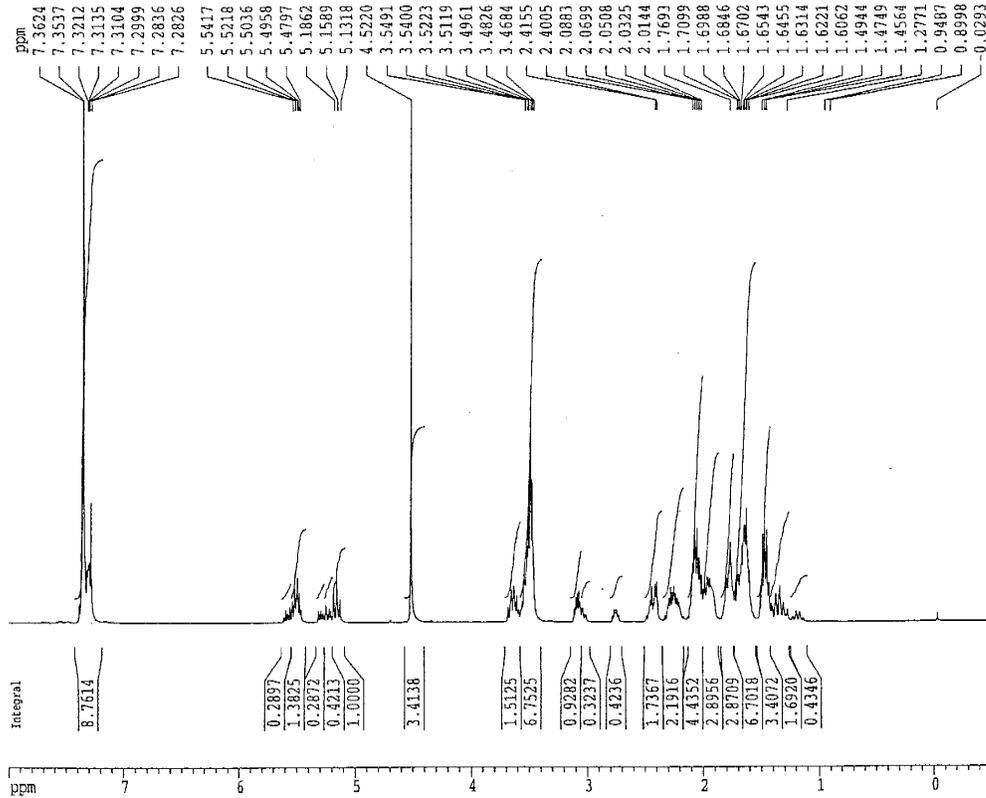
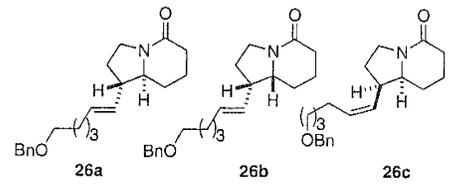
***** CHANNEL f1 *****
NUC1 13C
P1 6.50 usec
PL1 5.00 dB
SFO1 125.7719472 MHz

***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 95.00 usec
PL2 +4.00 dB
PL12 19.00 dB
PL13 30.00 dB
SFO2 500.1325000 MHz

F2 - Processing parameters
SI 85336
SF 125.7570000 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 OF
SF 500.1300000 MHz
WDW - no
SSB 0
LB 0.30 Hz
GB 0

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 200.000 ppm
F1 25151.56 Hz
F2P -0.500 ppm
F2 -42.88 Hz
PFMCM 10.02508 ppm/cm
HZCM 1800.72190 Hz/cm



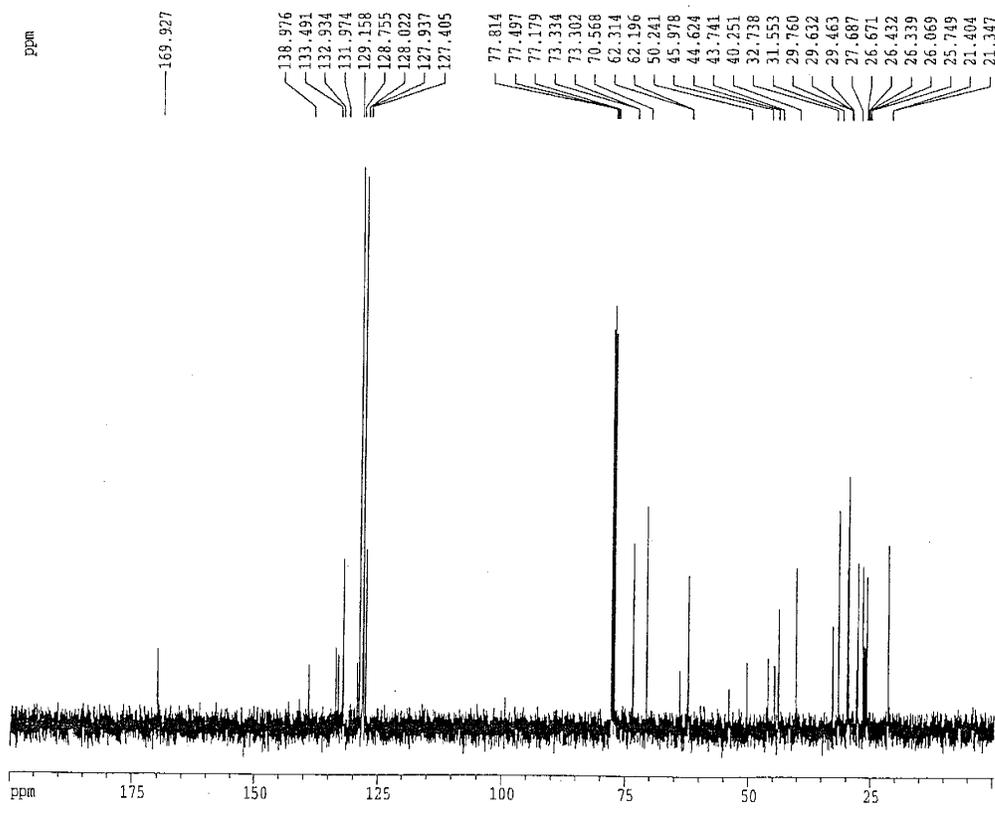
Current Data Parameters
NAME wj4-53
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20030729
Time 14.29
INSTRUM drx400
PROBHD 5 mm Multinuc1
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210391 sec
RG 143.7
DW 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 7.70 usec
PL1 -6.00 dB
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300000 MHz
MEW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 8.000 ppm
F1 3201.04 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PPMCH 0.42500 ppm/cm
HZCM 170.05525 Hz/cm



Current Data Parameters
NAME wj4-53
EXPNO 2
PROCNO 1

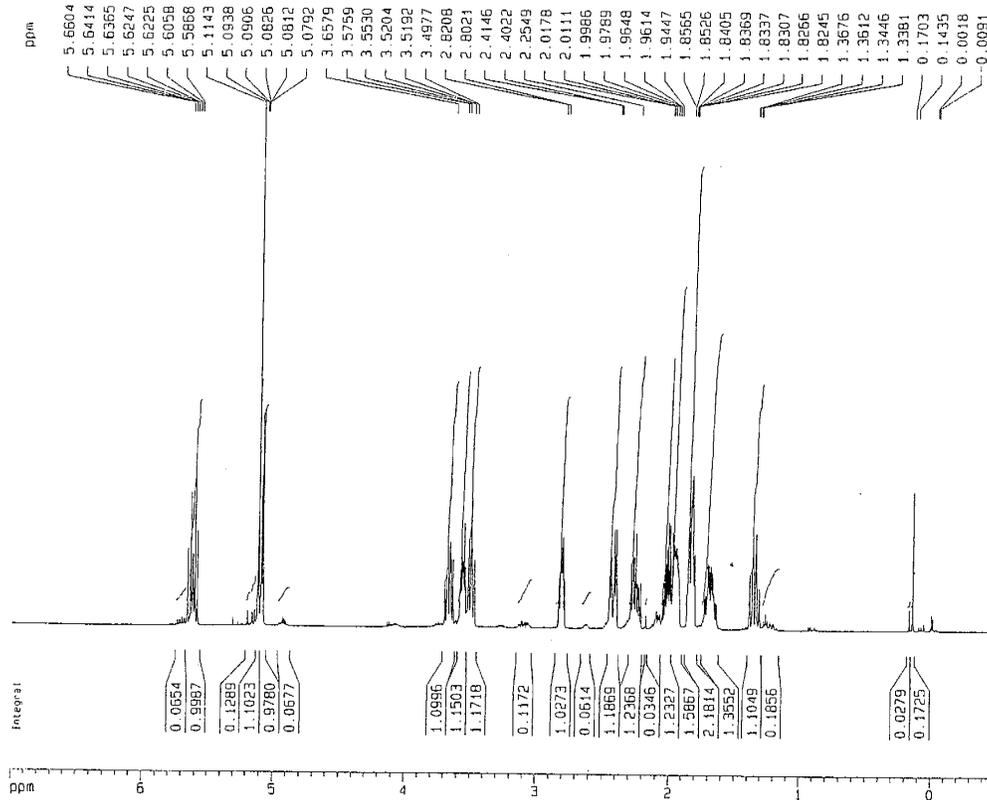
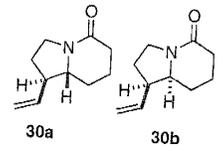
F2 - Acquisition Parameters
Date_ 20030729
Time 16.20
INSTRUM drx400
PROBHD 5 mm Multinuc1
PULPROG zgpg30
TD 65336
SOLVENT CDCl3
NS 105
DS 2
SWH 23148.148 Hz
FIDRES 0.353213 Hz
AQ 1.4156276 sec
RG 32768
DW 21.600 usec
DE 4.50 usec
TE 300.0 K
D1 0.05000000 sec
d11 0.03000000 sec
d12 0.00002000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 12.30 usec
PL1 2.00 dB
SFO1 100.6232933 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 180.00 usec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127290 MHz
MEW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 200.000 ppm
F1 20122.55 Hz
F2P -0.500 ppm
F2 -50.31 Hz
PPMCH 10.02500 ppm/cm
HZCM 1008.64264 Hz/cm



Current Data Parameters
NAME wj3-242
EXPNO 2
PROCNO 1

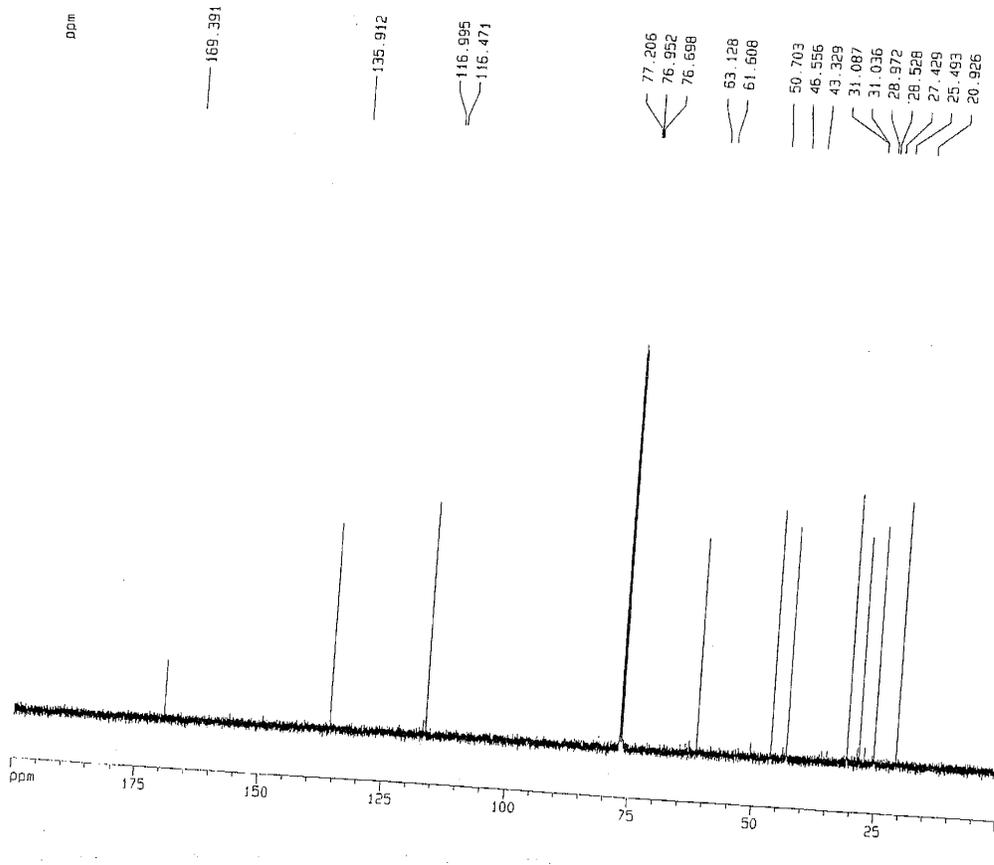
F2 - Acquisition Parameters
Date_ 20030305
Time 18.14
INSTALM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 5009.615 Hz
FIDRES 0.183398 Hz
AQ 2.7264309 sec
RG 161.3
DM 83.200 usec
DE 6.00 usec
TE 300.0 K
D1 0.03000000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -4.00 dB
SF01 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 OF
SF 500.1300000 MHz
WDW no
SSB 0
LB 0.30 Hz
GB 0

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 7.000 ppm
F1 3500.91 Hz
F2P -0.500 ppm
F2 -250.07 Hz
PPMCH 0.37500 ppm/cm
HZCX 187.54875 Hz/cm



Current Data Parameters
NAME wj3-242
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20030305
Time 18.20
INSTALM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 288
DS 4
SWH 31446.541 Hz
FIDRES 0.479836 Hz
AQ 1.0420803 sec
RG 2048
DM 15.900 usec
DE 6.00 usec
TE 300.0 K
D1 0.10000000 sec
d11 0.03000000 sec
d12 0.00002000 sec

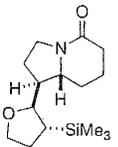
***** CHANNEL f1 *****
NUC1 13C
P1 6.50 usec
PL1 5.00 dB
SF01 125.7719472 MHz

***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 95.00 usec
PL2 -4.00 dB
PL12 19.00 dB
PL13 30.00 dB
SF02 500.1325000 MHz

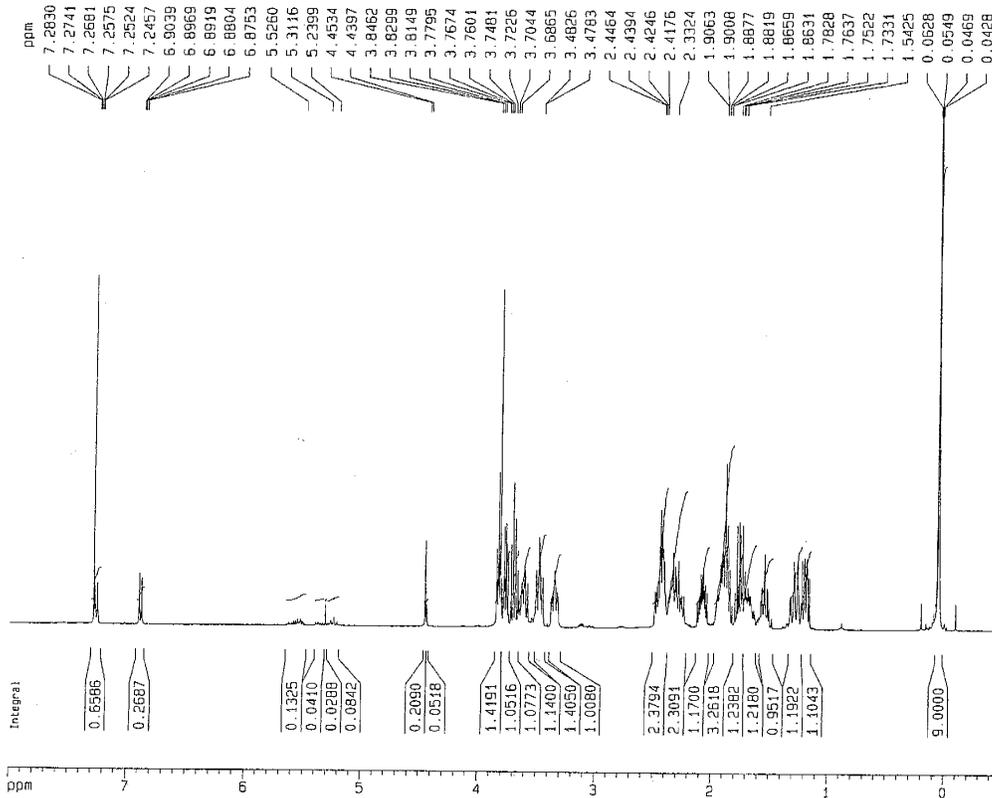
F2 - Processing parameters
SI 65536
SF 125.7578000 MHz
WDW EV
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 OF
SF 500.1300000 MHz
WDW no
SSB 0
LB 0.30 Hz
GB 0

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 200.000 ppm
F1 25151.56 Hz
F2P -0.500 ppm
F2 -62.88 Hz
PPMCH 10.02500 ppm/cm
HZCX 1260.72132 Hz/cm



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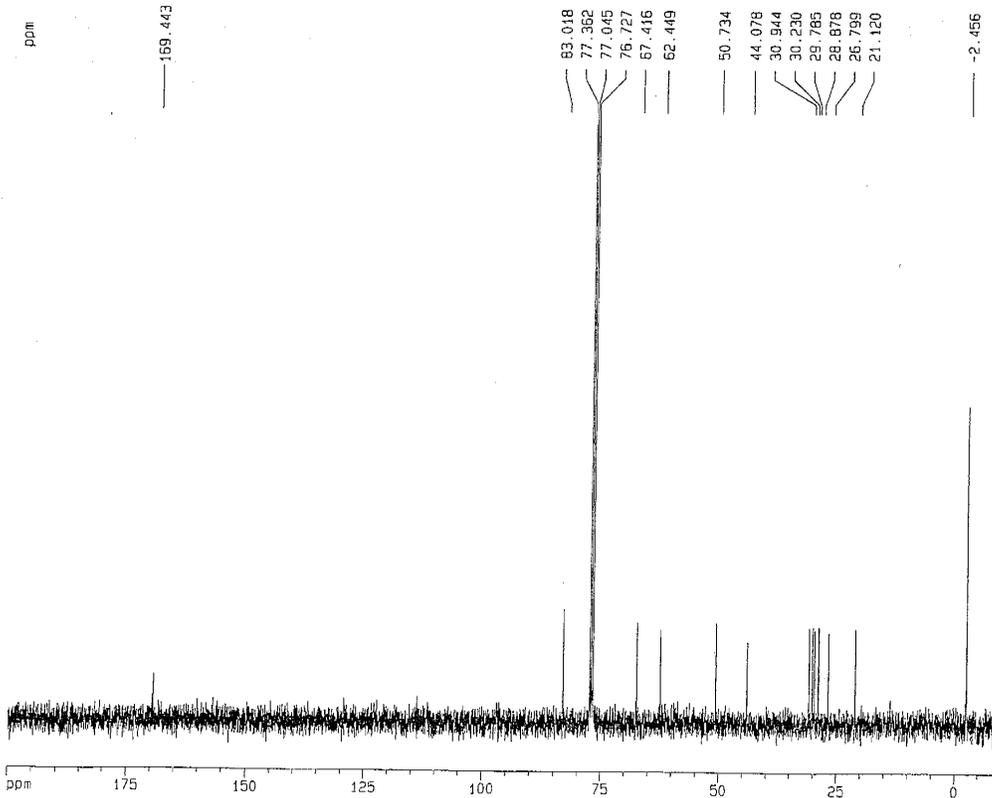


F2 - Acquisition Parameters
Date_ 20051015
Time 14.48
INSTRUM spect
PROBHD 5 mm QNP 1H/15
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4769.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 203.2
DW 104.400 usec
DE 6.00 usec
TE 295.9 K
D1 1.0000000 sec
MCREST 0.0000000 sec
MCHWK 0.01500000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.05 usec
PL1 -3.00 dB
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 32768
SF 400.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 8.000 ppm
F1 3201.04 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PPHM 0.42500 ppm/cm
HZCM 170.05525 Hz/cm



Current Data Parameters
NAME w16-22,51-59
EXPNO 2
PROCNO 1

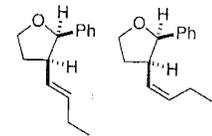
F2 - Acquisition Parameters
Date_ 20051015
Time 14.56
INSTRUM spect
PROBHD 5 mm QNP 1H/15
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 632
DS 4
SWH 23390.814 Hz
FIDRES 0.365918 Hz
AQ 1.3684756 sec
RG 2048
DW 20.850 usec
DE 6.00 usec
TE 298.1 K
D1 0.1500001 sec
D11 0.0200000 sec
DELTA 0.0500000 sec
MCREST 0.0000000 sec
MCHWK 0.01500000 sec

***** CHANNEL f1 *****
NUC1 13C
P1 7.20 usec
PL1 -2.00 dB
SFO1 100.6228298 MHz

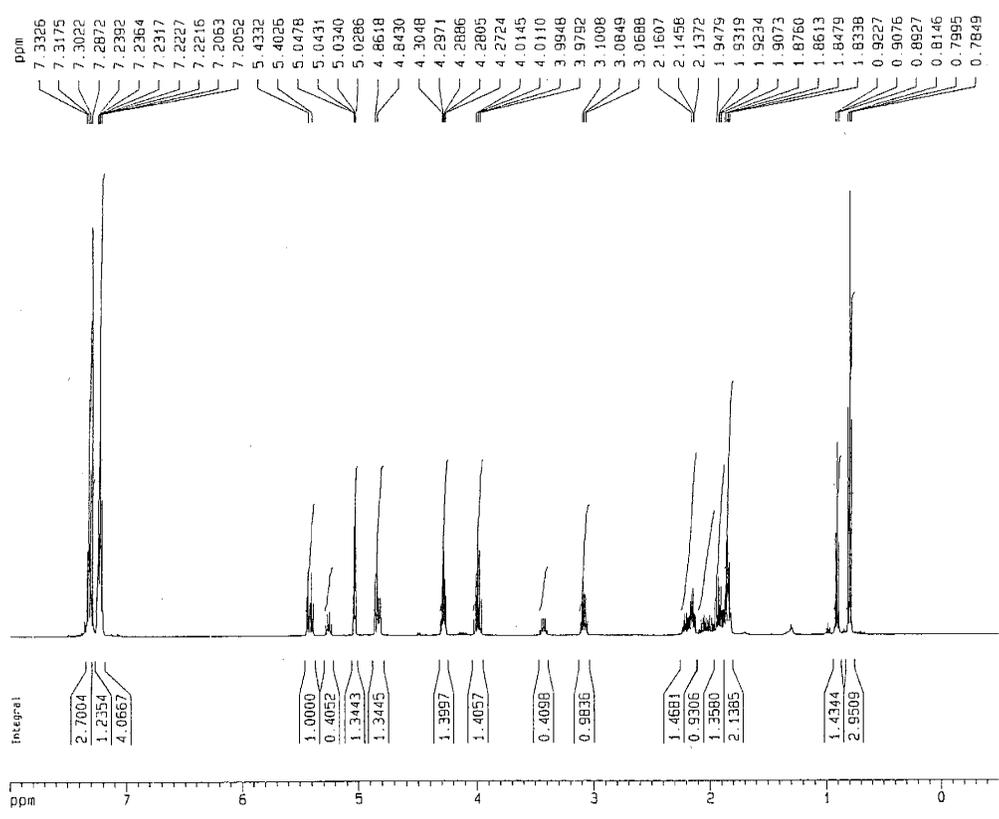
***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 98.40 usec
PL2 -3.00 dB
PL12 19.96 dB
PL13 120.00 dB
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127650 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 300.000 ppm
F1 20132.55 Hz
F2P -10.000 ppm
F2 -1006.13 Hz
PPHM 10.50000 ppm/cm
HZCM 1056.43408 Hz/cm



(E)-33a (Z)-33b



Current Data Parameters
NAME wj3-274
EXPNO 1
PROCNO 1

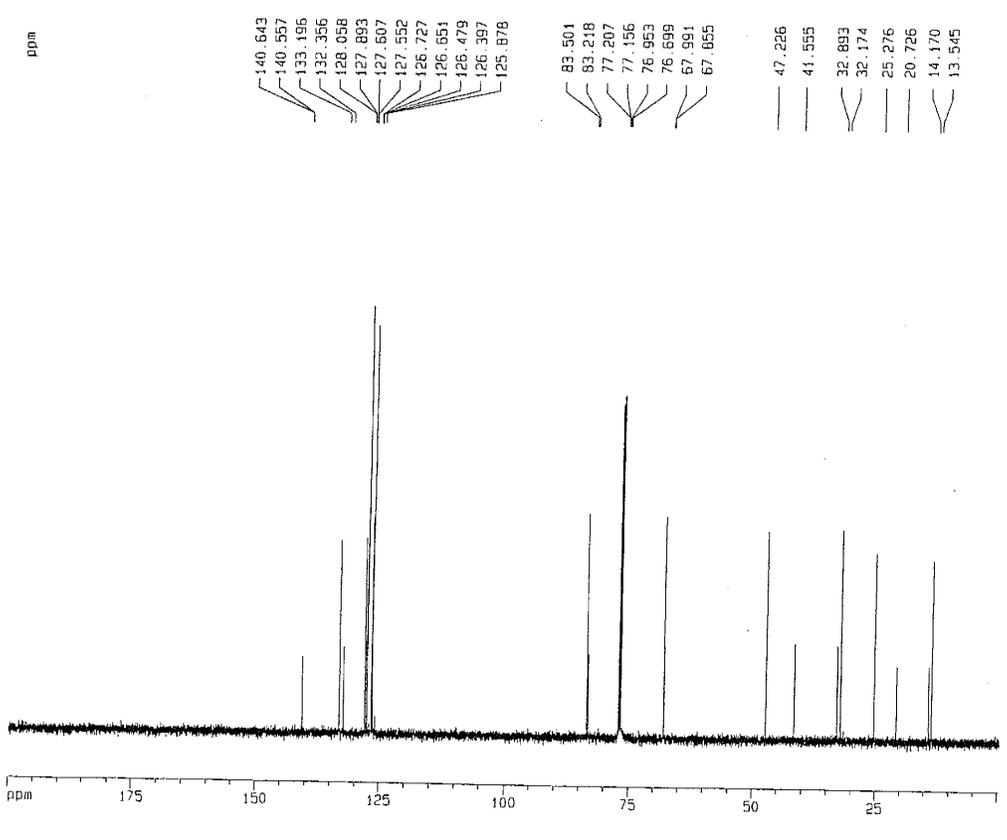
F2 - Acquisition Parameters
Date_ 20030422
Time 23.39
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 6009.615 Hz
FIDRES 0.18339 Hz
AQ 2.7264308 sec
RG 90.5
DM 83.200 usec
DE 6.00 usec
TE 300.0 K
D1 0.0300000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -4.00 dB
SFO1 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
HC2 OF
SF 500.1300000 MHz
WDW QSINE
SSB 0
LB 0.30 Hz
GB 0

ID NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 8.000 ppm
FJ 4001.04 Hz
F2P -0.500 ppm
F2 -250.07 Hz
PPMCH 0.42500 ppm/cm
HZCH 212.55525 Hz/cm



Current Data Parameters
NAME wj3-274
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20030422
Time 23.44
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 256
DS 4
SWH 31446.541 Hz
FIDRES 0.479836 Hz
AQ 1.0420983 sec
RG 76
DM 2040
DE 15.300 usec
TE 300.0 K
D1 0.1600000 sec
d11 0.0300000 sec
d12 0.00002000 sec

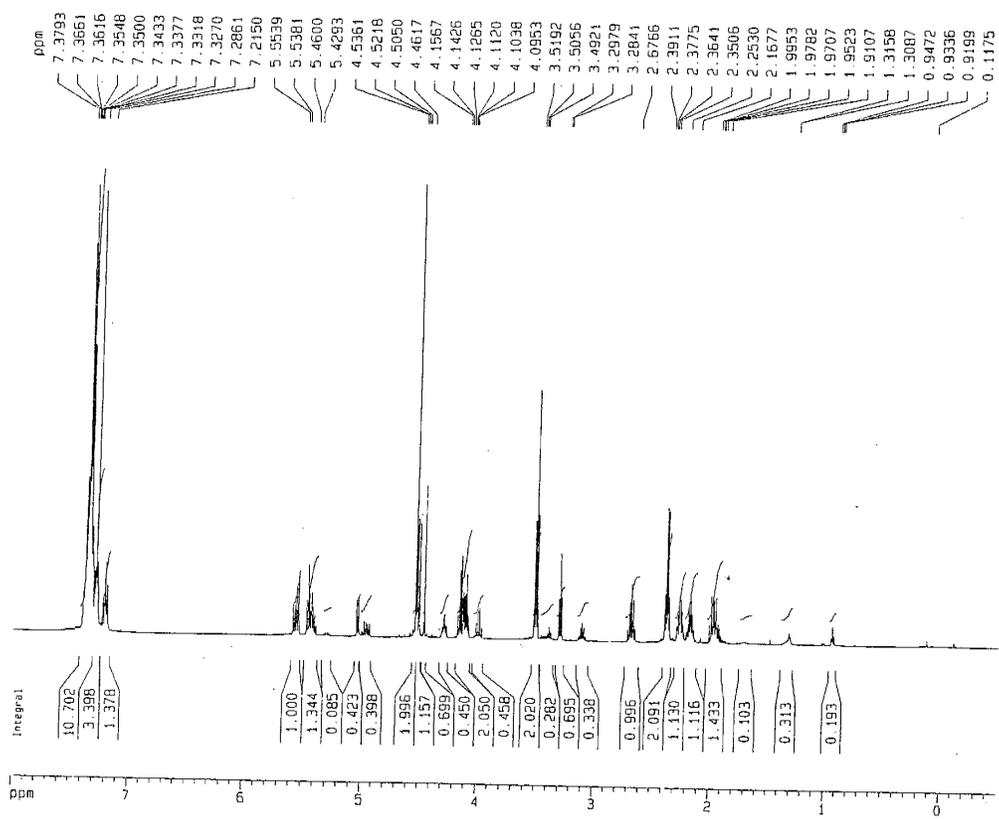
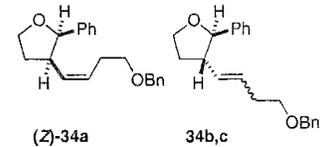
***** CHANNEL f1 *****
NUC1 13C
P1 6.50 usec
PL1 5.00 dB
SFO1 125.7719472 MHz

***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 95.00 usec
PL2 -4.00 dB
PL12 19.00 dB
PL13 30.00 dB
SFO2 500.1325000 MHz

F2 - Processing parameters
SI 65536
SF 125.7578008 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
HC2 OF
SF 500.1300000 MHz
WDW NO
SSB 0
LB 0.30 Hz
GB 0

ID NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 200.500 ppm
F1 25151.56 Hz
F2P -62.88 Hz
F2 10.02500 ppm/cm
PPMCH 1260.72192 Hz/cm



Current Data Parameters
NAME w13-270
EXPNO 1
PROCNO 1

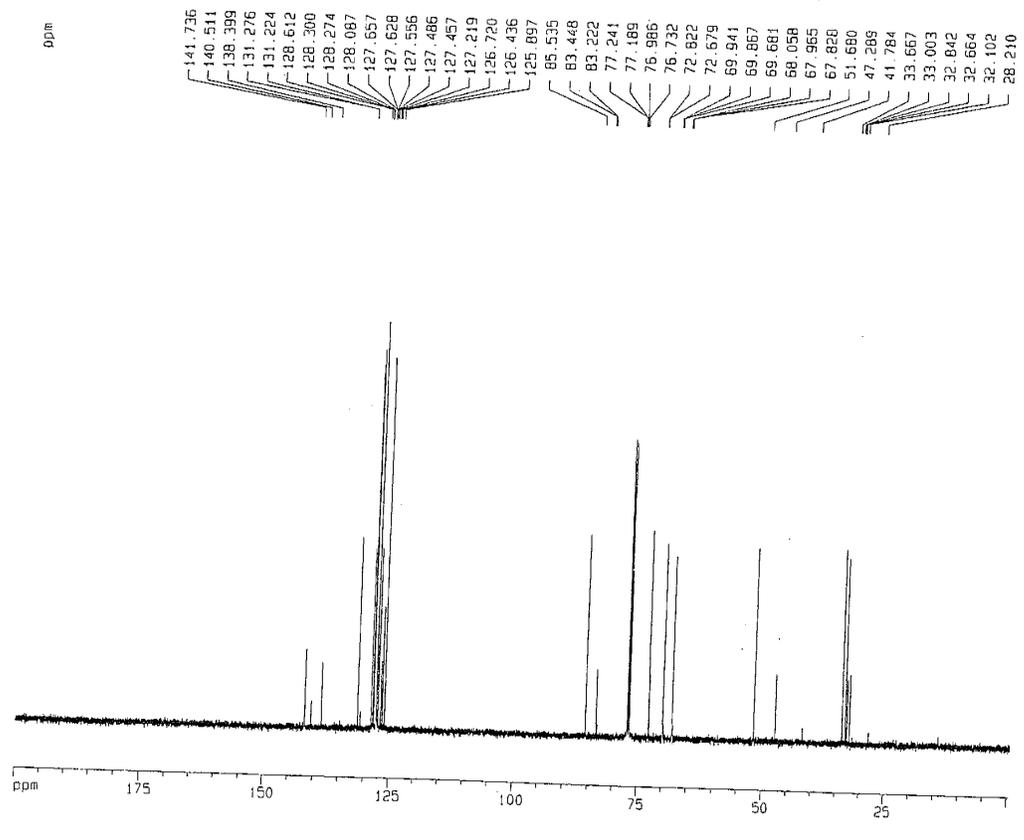
F2 - Acquisition Parameters
Date_ 20030422
Time 23.30
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SMH 6009.615 Hz
FIDRES 0.183389 Hz
AQ 2.7264309 sec
RG 71.8
DW 83.200 usec
DE 6.00 usec
TE 300.0 K
D1 0.0300000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -4.00 dB
SFO1 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 500.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 GF
SF 500.1300000 MHz
WDW DSINE
SSB 0
LB 0.30 Hz
GB 0

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 8.000 ppm
F1 4001.04 Hz
F2P -0.500 ppm
F2 -250.07 Hz
PPHMC 0.42500 ppm/cm
HZCN 212.55525 Hz/cm



Current Data Parameters
NAME w13-270
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20030422
Time 23.36
INSTRUM spect
PROBHD 5 mm BBO BB-1H
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 4
DS 4
SMH 31446.541 Hz
FIDRES 0.478038 Hz
AQ 1.0426883 sec
RG 20.48
DW 15.800 usec
DE 6.00 usec
TE 300.0 K
D1 0.1000000 sec
S11 0.0350000 sec
S12 0.0002000 sec

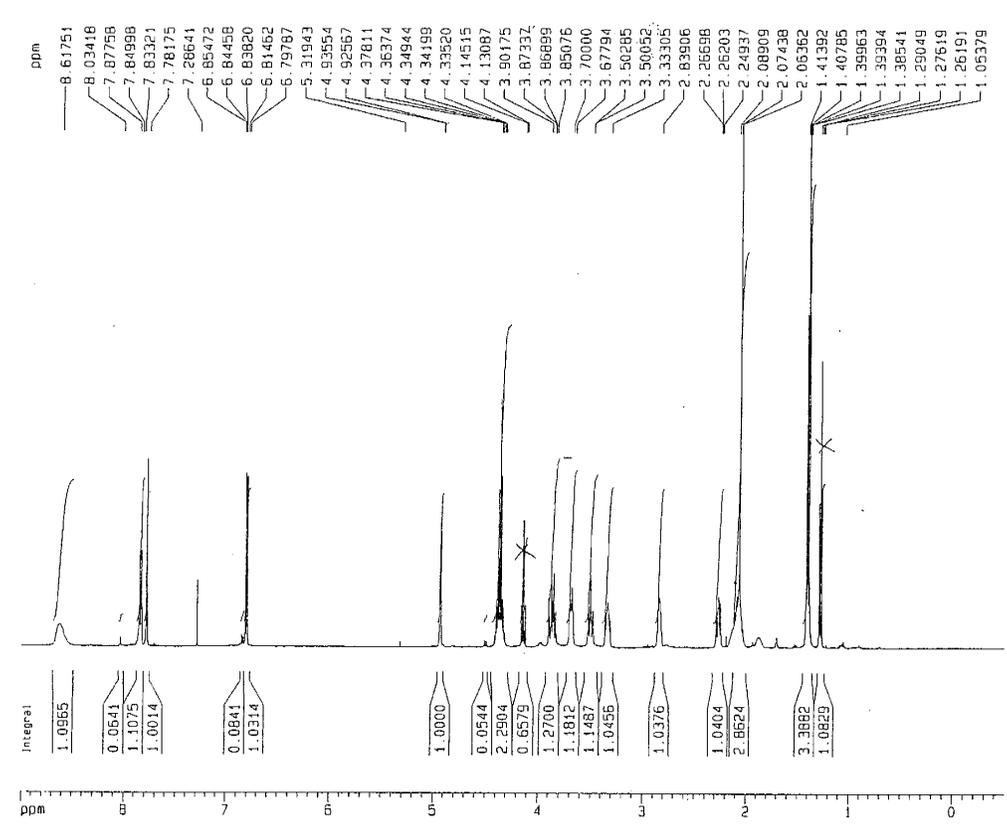
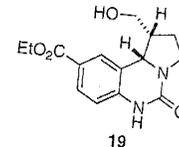
***** CHANNEL f1 *****
NUC1 13C
P1 6.50 usec
PL1 5.00 dB
SFO1 125.7719472 MHz

***** CHANNEL f2 *****
CPDPRG2 w13z16
NUC2 1H
PCPD2 95.00 usec
PL2 -4.00 dB
PL12 19.00 dB
PL13 30.00 dB
SFO2 500.1325000 MHz

F2 - Processing parameters
SI 65536
SF 125.7570008 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 GF
SF 500.1300000 MHz
WDW no
SSB 0
LB 0.30 Hz
GB 0

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 200.000 ppm
F1 25151.56 Hz
F2P -62.88 Hz
F2 -32.88 Hz
PPHMC 10.02500 ppm/cm
HZCN 1280.72192 Hz/cm



Current Data Parameters
 NAME wj3-196
 EXPNO 1
 PROCNO 1

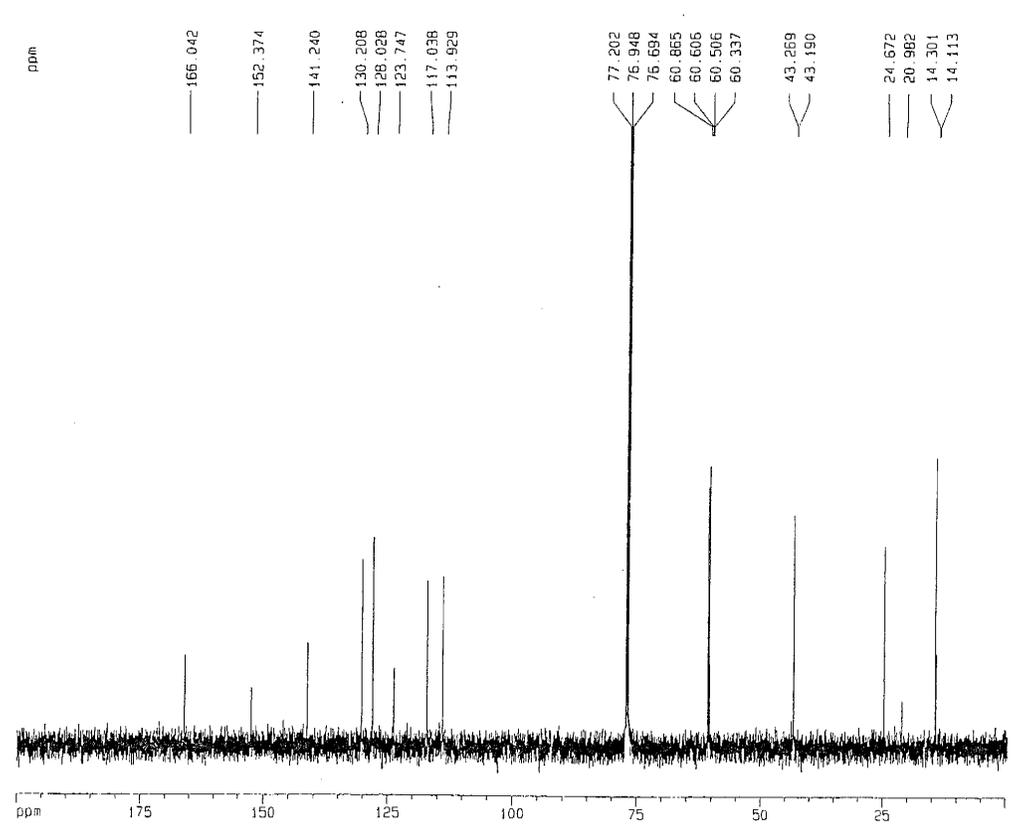
F2 - Acquisition Parameters
 Date_ 20030115
 Time 13.13
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.284399 sec
 RG 328.1
 DW 83.200 usec
 DE 6.00 usec
 TE 300.0 K
 O1 0.0300000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.70 usec
 PL1 -4.00 dB
 SFO1 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 500.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

F1 - Processing parameters
 SI 1024
 MC2 GF
 SF 500.1300000 MHz
 WDW QSTINE
 SSB 0
 LB 0.30 Hz
 GB 0

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 9.000 ppm
 F1 4501.17 Hz
 F2P -0.500 ppm
 F2 -250.07 Hz
 PRMCM 0.47500 ppm/cm
 HZCM 237.56175 Hz/cm



Current Data Parameters
 NAME wj3-196
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20030115
 Time 13.19
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 362
 DS 4
 SWH 31446.541 Hz
 FIDRES 0.470836 Hz
 AQ 1.040583 sec
 RG 8192
 DW 15.900 usec
 DE 8.00 usec
 TE 300.0 K
 O1 0.1000000 sec
 O11 0.0300000 sec
 O12 0.0000200 sec

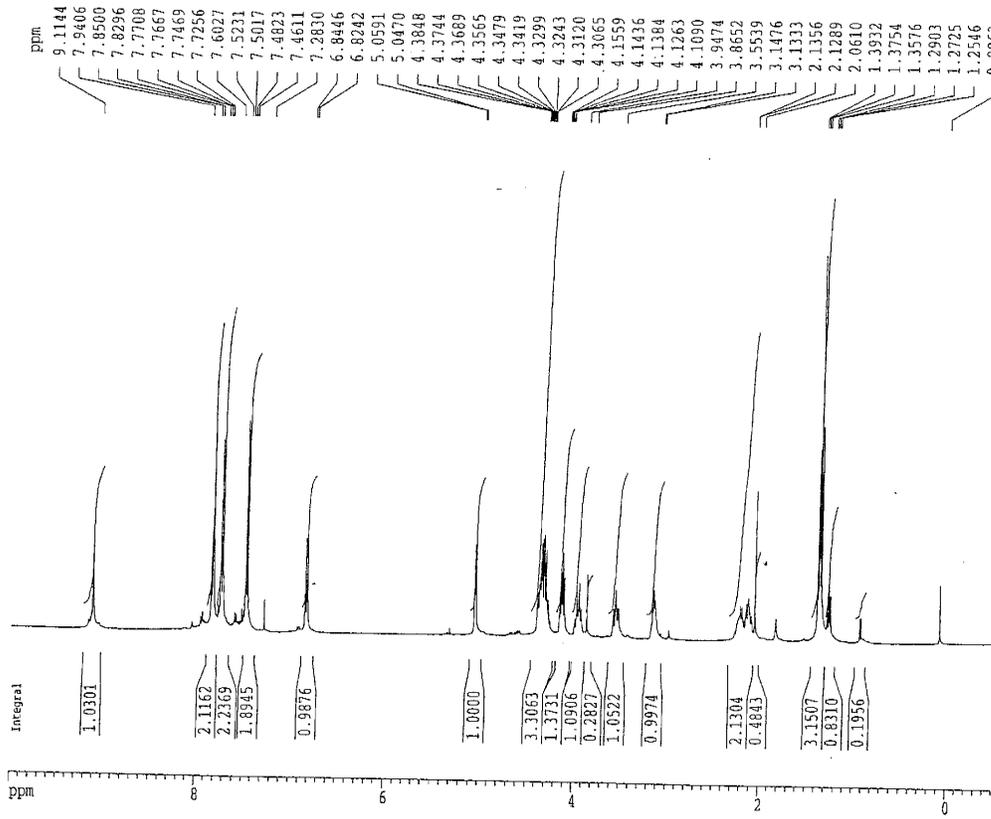
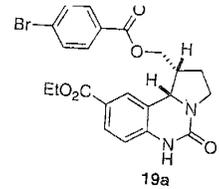
***** CHANNEL f1 *****
 NUC1 13C
 P1 6.50 usec
 PL1 5.00 dB
 SFO1 125.7719472 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 95.00 usec
 PL2 -4.00 dB
 PL12 19.00 dB
 PL13 30.00 dB
 SFO2 500.1325000 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7578208 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

F1 - Processing parameters
 SI 1024
 MC2 GF
 SF 500.1300000 MHz
 WDW ac
 SSB 0
 LB 0.30 Hz
 GB 0

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 200.000 ppm
 F1 25151.56 Hz
 F1P -0.500 ppm
 F2 -62.60 Hz
 PRMCM 10.02500 ppm/cm
 HZCM 1260.72152 Hz/cm



```

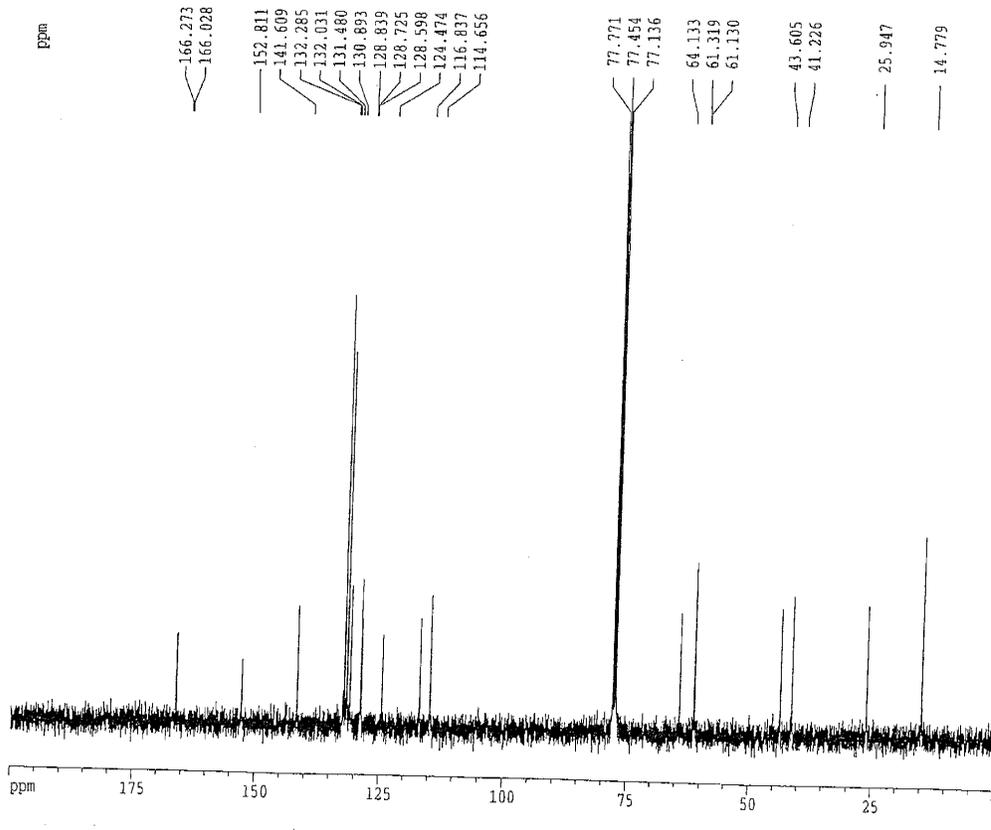
NAME          wj3-225
EXPNO         1
PROCNO        1

F2 - Acquisition Parameters
Date_         20030215
Time          18.23
INSTRUM       dx400
PROBHD        5 mm Multinucl
PULPROG       zg30
TD             32768
SOLVENT       CDCl3
NS             16
DS             2
SWH            4789.272 Hz
FIDRES        0.146157 Hz
AQ             3.4210291 sec
RG             114
DW            104.400 usec
DE             4.50 usec
TE             300.0 K
D1             1.00000000 sec

===== CHANNEL f1 =====
NUC1           1H
P1              7.70 usec
PL1             -6.00 dB
SFO1           400.1320007 MHz

F2 - Processing parameters
SI             16384
SF             400.1300000 MHz
WDW            EM
SSB            0
LB             0.30 Hz
GB             0
PC             1.00

ID NMR plot parameters
CX             20.00 cm
CY             0.00 cm
F1P           10.000 ppm
F1             4001.30 Hz
F2P           -0.500 ppm
F2            -200.07 Hz
PPMCH         0.52500 ppm/cm
HZCH          210.06825 Hz/cm
  
```



```

Current Data Parameters
NAME          wj3-225
EXPNO         2
PROCNO        1

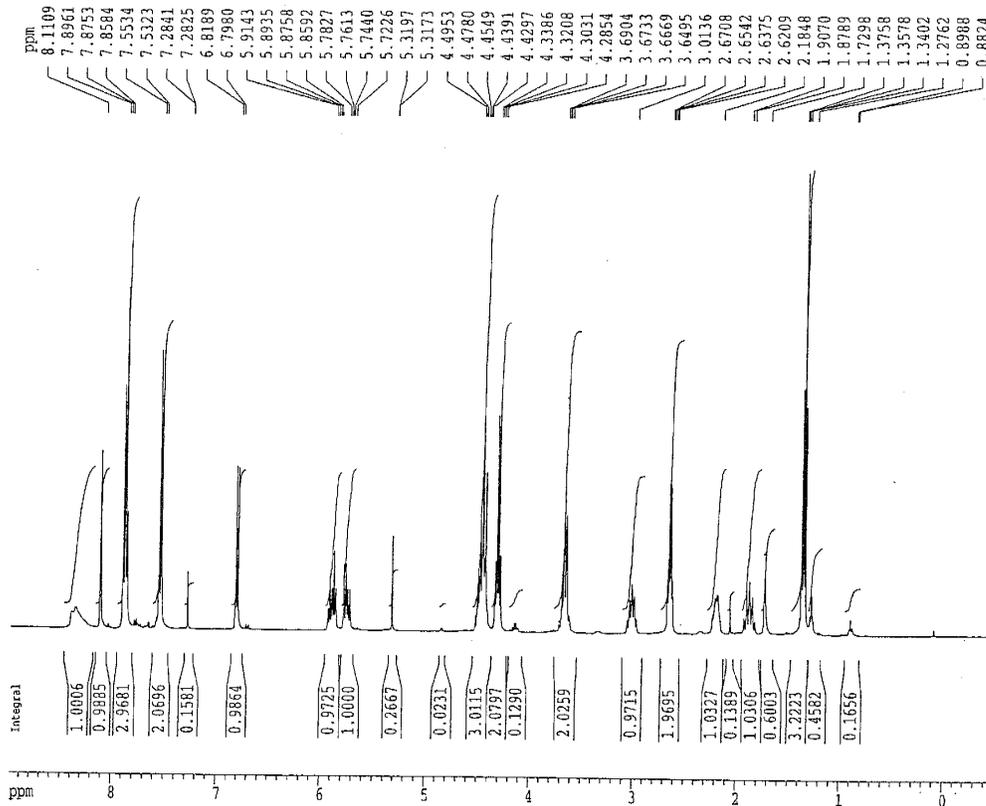
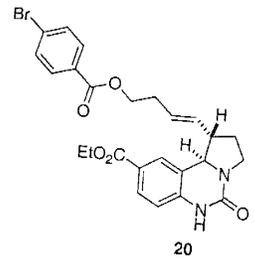
F2 - Acquisition Parameters
Date_         20030215
Time          18.27
INSTRUM       dx400
PROBHD        5 mm Multinucl
PULPROG       zgpg30
TD             65536
SOLVENT       CDCl3
NS             435
DS             2
SWH            23148.148 Hz
FIDRES        0.353213 Hz
AQ             1.4156276 sec
RG             32768
DW            21.600 usec
DE             4.50 usec
TE             300.0 K
D1             0.05000000 sec
d11            0.03000000 sec
d12            0.00002000 sec

===== CHANNEL f1 =====
NUC1           13C
P1             12.30 usec
PL1             2.00 dB
SFO1           100.6232933 MHz

===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2           1H
PCPD2         100.00 usec
PL2            0.00 dB
PL12           18.00 dB
PL13           18.00 dB
SFO2           400.1316005 MHz

F2 - Processing parameters
SI             32768
SF             100.6127290 MHz
WDW            EM
SSB            0
LB             1.00 Hz
GB             0
PC             1.40

ID NMR plot parameters
CX             20.00 cm
CY             0.00 cm
F1P           200.000 ppm
F1             20122.55 Hz
F2P           -50.30 Hz
F2            -50.30 Hz
PPMCH         10.02500 ppm/cm
HZCH          1008.64258 Hz/cm
  
```

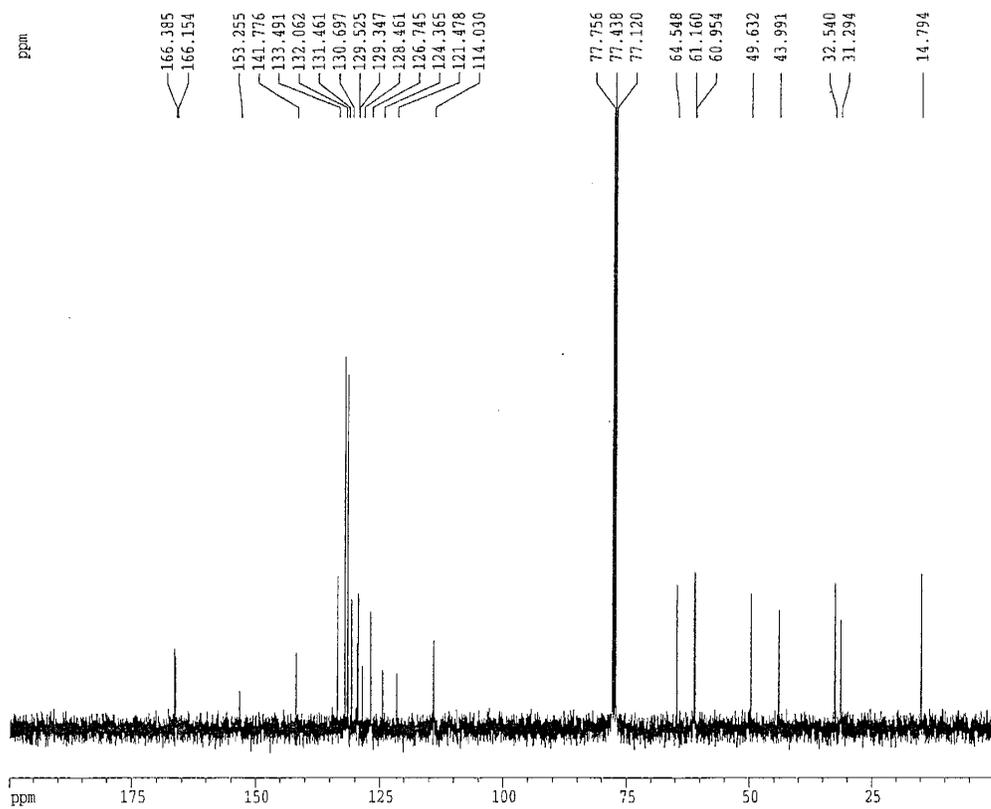


F2 - Acquisition Parameters
Date_ 20021127
Time 17.01
INSTRUM drx400
PROBHD 5 mm Multinucl
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 128
DW 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -6.00 dB
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 9.000 ppm
F1 3601.17 Hz
F2P -0.500 ppm
F2 -200.07 Hz
PRCM 0.47500 ppm/cm
HZCM 190.06175 Hz/cm



Current Data Parameters
NMR wj3-165
EXPNO 2
PROCNO 1

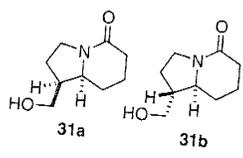
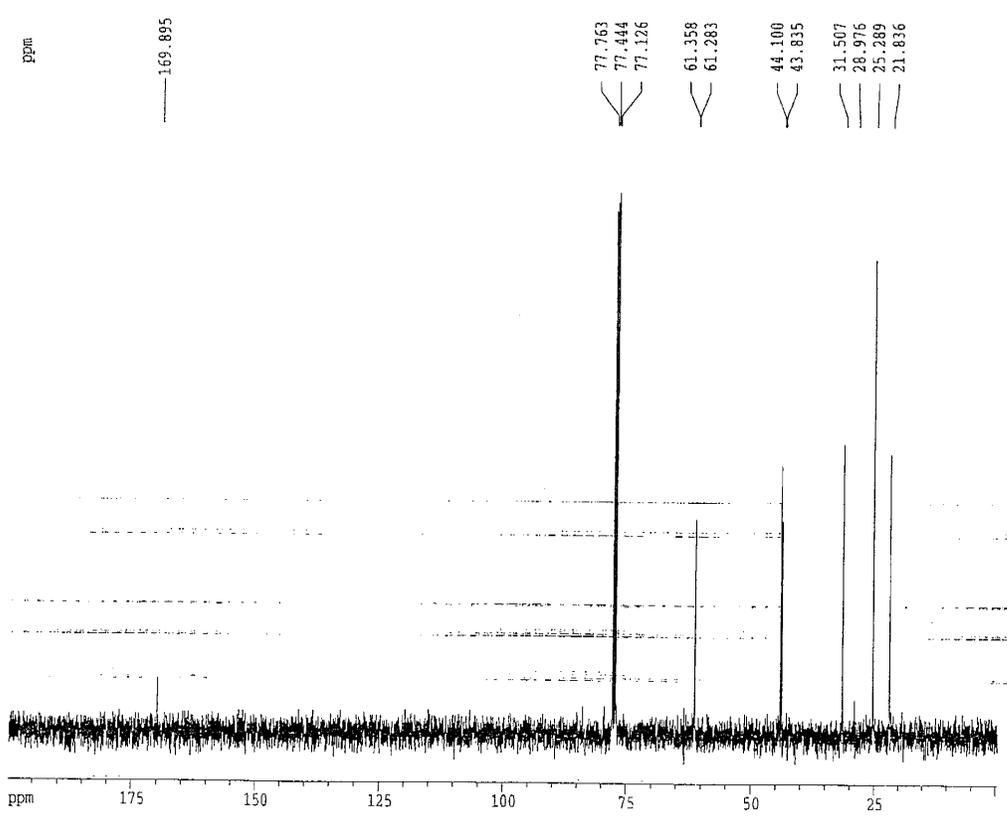
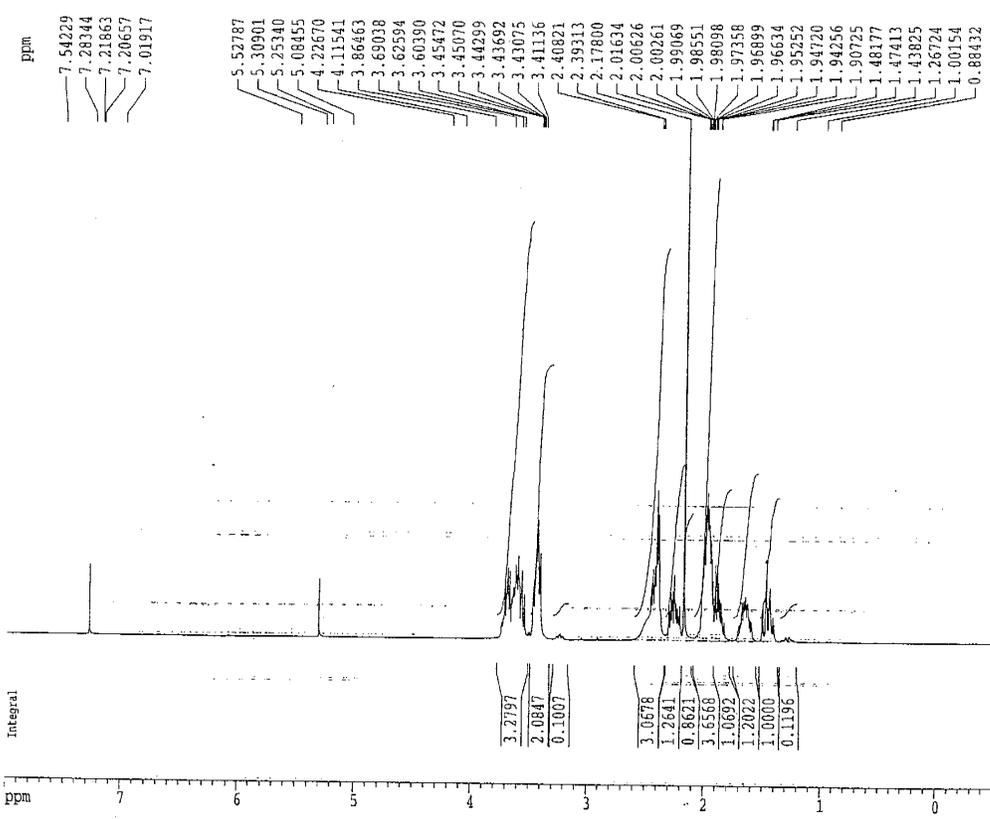
F2 - Acquisition Parameters
Date_ 20021127
Time 17.07
INSTRUM drx400
PROBHD 5 mm Multinucl
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 425
DS 2
SWH 23148.148 Hz
FIDRES 0.351213 Hz
AQ 1.4156276 sec
RG 3648.3
DW 21.600 usec
DE 4.50 usec
TE 300.0 K
D1 0.05000000 sec
d11 0.03000000 sec
d12 0.00002000 sec

***** CHANNEL f1 *****
NUC1 13C
P1 12.30 usec
PL1 2.00 dB
SFO1 100.6232933 MHz

***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127290 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
CY 0.00 cm
F1P 200.000 ppm
F1 20322.55 Hz
F2P -50.31 Hz
F2 10.02500 ppm/cm
PRCM 1008.64270 Hz/cm
HZCM 1008.64270 Hz/cm



Current Data Parameters
 NAME wj4-38
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20030702
 Time 16.08
 INSTRUM drx400
 PROBHD 5 mm Multinucl
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 143.7
 DW 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 DI 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.70 usec
 PL1 -6.00 dB
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

ID NMR plot parameters
 CX 20.00 cm
 CY 20.00 cm
 F1P 8.000 ppm
 F1 3201.04 Hz
 F2P -0.500 ppm
 F2 -200.07 Hz
 PPMCM 0.42500 ppm/cm
 HZCM 170.05525 Hz/cm

Current Data Parameters
 NAME wj4-38
 EXPNO 2
 PROCNO 1

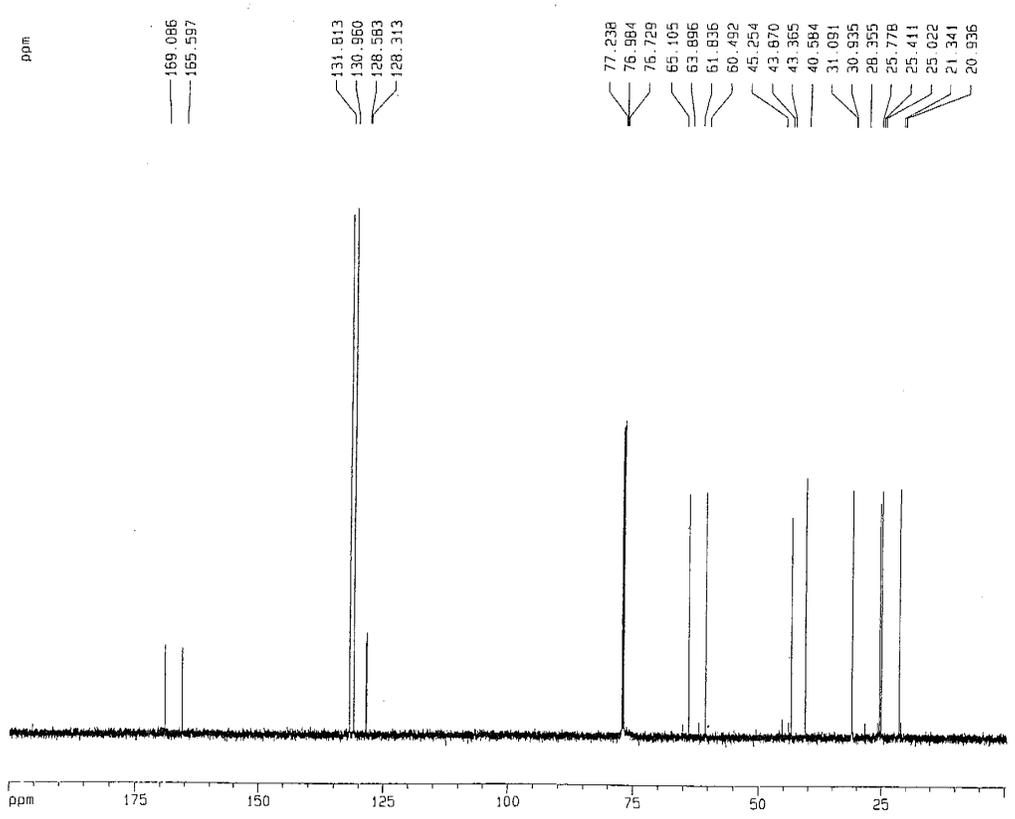
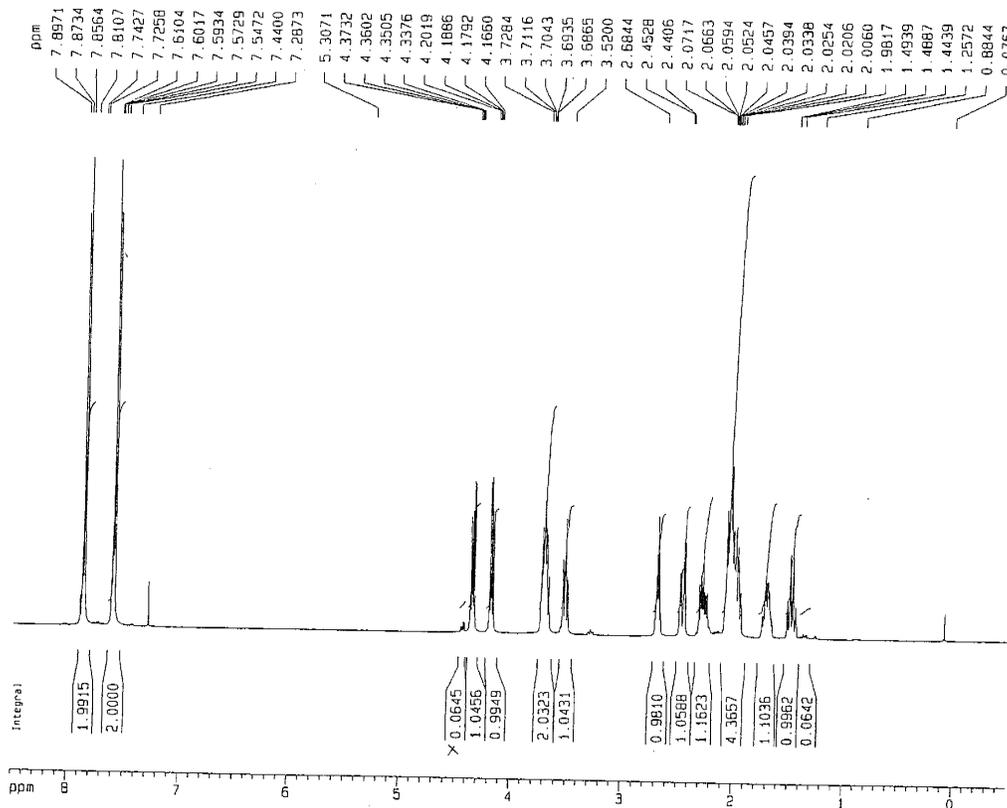
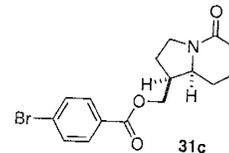
F2 - Acquisition Parameters
 Date_ 20030702
 Time 16.11
 INSTRUM drx400
 PROBHD 5 mm Multinucl
 PULPROG zgpg30
 TD 85536
 SOLVENT CDCl3
 NS 175
 DS 2
 SWH 23148.148 Hz
 FIDRES 0.353213 Hz
 AQ 1.4156276 sec
 RG 32768
 EM 4.50 usec
 DE 21.600 usec
 TE 300.0 K
 DI 0.05000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

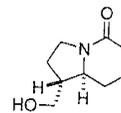
===== CHANNEL f1 =====
 NUC1 13C
 P1 12.30 usec
 PL1 2.00 dB
 SFO1 100.6213933 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316005 MHz

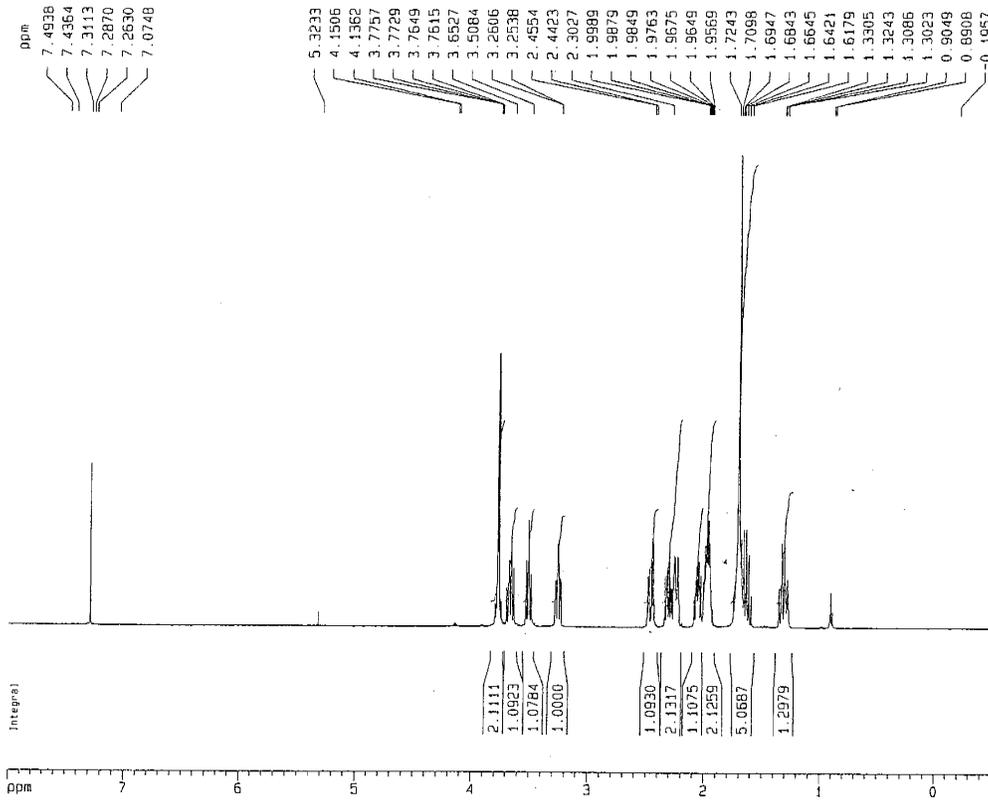
F2 - Processing parameters
 SI 32768
 SF 100.5127290 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

ID NMR plot parameters
 CX 20.00 cm
 CY 8.00 cm
 F1P 200.000 ppm
 F1 20122.55 Hz
 F2P -0.500 ppm
 F2 -50.31 Hz
 PPMCM 10.02500 ppm/cm
 HZCM 1008.64264 Hz/cm





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Current Data Parameters
 NAME wj3-42.crv1
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20020706
 Time 11:18
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6089.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7264309 sec
 RG 574.7
 DM 83.200 usec
 DE 6.00 usec
 TE 300.0 K
 D1 0.0300000 sec

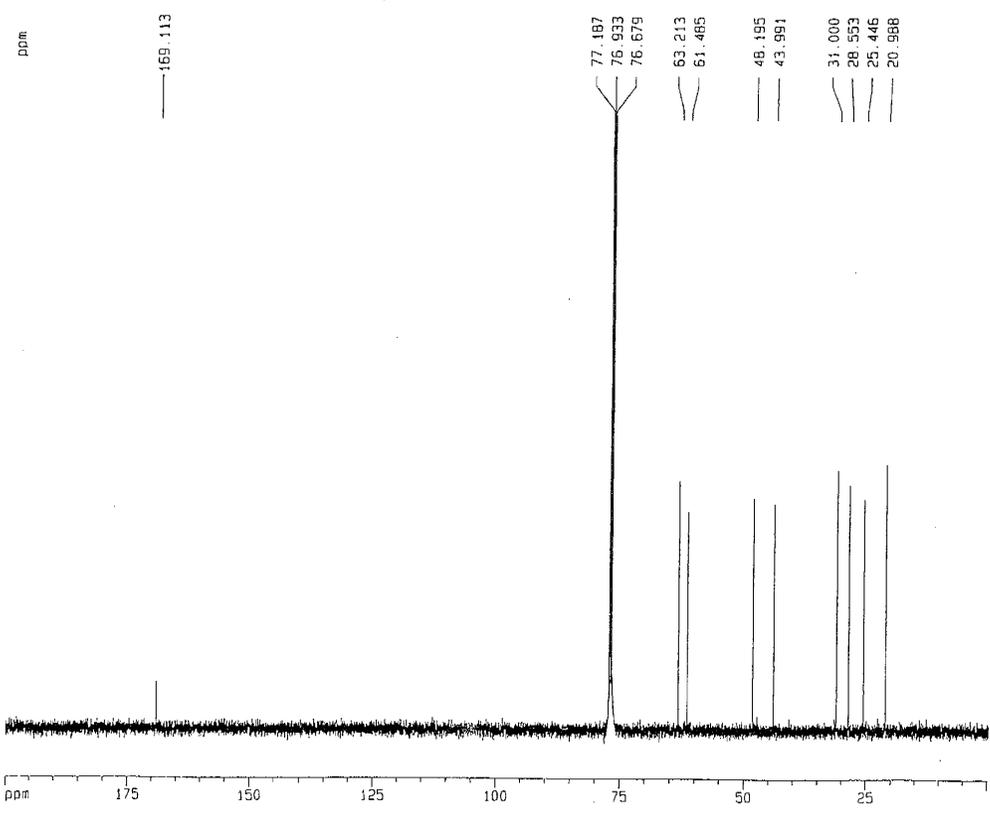
***** CHANNEL f1 *****
 NUC1 1H
 P1 7.70 usec
 PL1 -4.00 dB
 SFO1 500.1320005 MHz

F1 - Acquisition Parameters
 ND 2
 TD 256
 SFO1 500.1325 MHz
 FIDRES 23.475000 Hz
 SW 12.016 ppm

F2 - Processing parameters
 SI 32768
 SF 500.1300000 MHz
 KW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

F1 - Processing parameters
 SI 1024
 MC2 GF
 SF 500.1300000 MHz
 KW no
 SSB 0
 LB 0.30 Hz
 GB 0

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 8.000 ppm
 F1 4001.04 Hz
 F2P -0.500 ppm
 F2 -250.07 Hz
 PPRCM 0.42500 ppm/cm
 HZCM 212.55925 Hz/cm



Current Data Parameters
 NAME wj3-42.crv1
 EXPNO 2
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20020706
 Time 15:29
 INSTRUM spect
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 3712
 DS 4
 SWH 31446.541 Hz
 FIDRES 0.479836 Hz
 AQ 1.0420093 sec
 RG 1024.3
 DM 15.300 usec
 DE 6.00 usec
 TE 300.0 K
 D1 0.1000000 sec
 d11 0.0300000 sec
 d12 0.0002000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 6.50 usec
 PL1 5.00 dB
 SFO1 125.7719472 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 P2P2P 95.00 usec
 PL2 -4.00 dB
 PL12 19.00 dB
 PL13 30.00 dB
 SFO2 500.132500 MHz

F1 - Acquisition Parameters
 ND 2
 TD 256
 SFO1 500.1325 MHz
 FIDRES 23.475000 Hz
 SW 12.016 ppm

F2 - Processing parameters
 SI 65536
 SF 125.7570000 MHz
 KW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

F1 - Processing parameters
 SI 1024
 MC2 GF
 SF 500.1300000 MHz
 KW no
 SSB 0
 LB 0.30 Hz
 GB 0

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1P 200.000 ppm
 F1 25151.56 Hz
 F2P -0.500 ppm
 F2 -453.07 Hz
 PPRCM 10.02500 ppm/cm
 HZCM 1280.72192 Hz/cm

