

## ***Supporting Information for***

### **Preferential DNA Cleavage under Anaerobic Conditions by a DNA Binding Ruthenium Dimer**

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### Plasmid DNA Cleavage Assay

Cleavage reactions were carried out in a total volume of 20  $\mu\text{L}$  in 0.5 mL Eppendorf tubes in a 7 mM  $\text{Na}_3\text{PO}_4$  buffer medium (pH 7) containing 2  $\mu\text{L}$  of supercoiled pUC18 DNA (1  $\mu\text{g}/1 \mu\text{L}$ ). Samples were prepared by first dissolving the DNA in 13  $\mu\text{L}$  buffer and then adding the GSH in 3  $\mu\text{L}$  and finally by adding  $[\text{P}]\text{Cl}_4$  in 2  $\mu\text{L}$ . Final concentrations are indicated in the figure legends. In case of Figure 2, the same concentration of  $\text{P}^{4+}$  (0.0128 mM) and Fe-BIm (0.0128 mM) were used with respect to the [DNA]. After incubation (times given in the figure captions), the DNA was precipitated by adding 2  $\mu\text{L}$  sodium acetate (pH 5.2) and 80  $\mu\text{L}$  ethanol followed by cooling overnight at  $-20 \text{ }^\circ\text{C}$ . The precipitated DNA was then dried for about 30 minutes and resuspended in 40  $\mu\text{L}$  of a storage buffer (e.g., 40 mM Tris-Cl, 1 mM EDTA at pH 8.0), 65  $\mu\text{L}$  deionized water and 12  $\mu\text{L}$  of a loading buffer (e.g., 30% glycerol in water with 0.1% w/v bromophenol blue). Samples were loaded on 1% agarose gel containing ethidium bromide (0.2  $\mu\text{L}/1 \text{ mL}$ ) and electrophoresed at 80 V for 90 minutes using TAE buffer (40 mM Tris-acetate, 1 mM EDTA, pH 8).

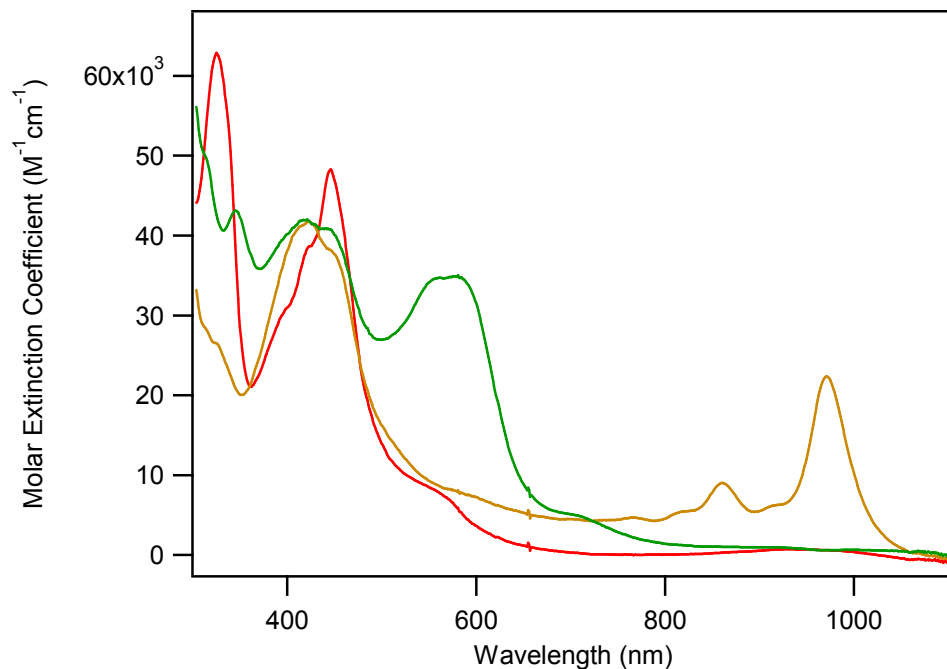
Anaerobic experiments were conducted in a glove box under a nitrogen atmosphere. All reagents and solutions, including pUC18 DNA, were subject to five freeze-pump-thaw cycles under  $\text{N}_2$  prior to introduction to the glove box. Incubations were stopped by precipitating the DNA using 2  $\mu\text{L}$  of degassed sodium acetate at pH 5.2 and 80  $\mu\text{L}$  degassed ethanol under  $\text{N}_2$  after which the samples were removed from the glove box and treated as the aerobic samples for further work-up. Control experiments established that no aerobic cleavage reactions are observed after the ethanol precipitation step.

### Synthesis of $[(\text{phen})_2\text{Ru}(\text{tatpp}^-)\text{Ru}(\text{phen})_2][\text{Cl}]_3$ ( $[\text{P}]\text{Cl}_3$ )

Complex  $[\text{P}](\text{PF}_6)_4^1$  (200 mg, 0.1 mmol) was dissolved in 100 mL of degassed acetonitrile under nitrogen inside the glove box. 19 mg (0.1 mmol) of cobaltocene (Alfa) dissolved in 3 mL of degassed acetonitrile was added and the solution stirred for 30 minutes. Slow addition of  $\sim 200\text{mL}$  of diethylether precipitated the product. The solid was filtered, washed with ether and dried under nitrogen. Yield 110 mg (60 %). The complex,  $[\text{P}](\text{PF}_6)_3$ , was converted to the chloride salt by dissolving in a minimum amount of acetone and then slow addition of a saturated solution of tetrabutylammonium chloride in acetone. The chloride salt cleanly precipitates and is filtered, washed with acetone and dried. The complex  $[\text{P}](\text{PF}_6)_3$  was characterized by preparing a dilute solution (26  $\mu\text{M}$ ) in degassed acetonitrile in an airtight 1 cm glass cuvette. The absorption spectrum was identical to that reported previously.<sup>2</sup>(See Supporting figure S1)

### Synthesis of $[(\text{phen})_2\text{Ru}(\text{H}_2\text{tatpp})\text{Ru}(\text{phen})_2][\text{Cl}]_4$ ( $[\text{H}_2\text{P}]\text{Cl}_4$ )

Complex  $[\text{P}][\text{PF}_6]_4^1$  (100 mg, 0.05 mmol) was dissolved in 5 mL of degassed MeCN inside the glove box. Ascorbic acid (0.5 mmol) was dissolved in a minimum amount of water and added to the MeCN solution and the resulting mixture stirred for 1 h at RT. Addition of a aqueous solution of  $\text{NH}_4\text{PF}_6$  (10 mg/mL) caused the product to precipitate. The solution was filtered and the solid washed with water ( $\sim 2 \text{ mL} - 4$  times) and then ether and dried in vacuo. Yield 80 mg (80%). <sup>1</sup>H NMR ( $\delta$ , 500 MHz, MeCN- $d_6$ ). The sample required that a small amount of ascorbic acid be included in the NMR solution or the spectrum noticeably broadened presumably due to the formation of paramagnetic  $\text{P}^{3+}$ : 9.39 (d,  $J = 7.0 \text{ Hz}$ , 2H), 8.58-8.63 (m,  $J = 9.0 \text{ Hz}$ , 5.9 Hz, 8H), 8.45 (d,  $J = 8.7 \text{ Hz}$ , 2H), 8.26 (d,  $J = 4.1\text{Hz}$ , 8H), 8.17 (d,  $J = 5.1 \text{ Hz}$ , 2H), 8.06 (d,  $J = 4.6 \text{ Hz}$ , 2H), 8.02 (d,  $J = 5.0 \text{ Hz}$ , 2H), 7.99 (d,  $J = 4.1 \text{ Hz}$ , 4H), 7.85 (d,  $J = 5.1 \text{ Hz}$ , 2H), 7.69-7.60 (m, 10H), 7.56 (dd,  $J = 8.7\text{Hz}$ , 5.5Hz, 2H), 6.94 (s, 2H). The identity of the complex was further established by obtaining the absorption spectrum of  $[\text{H}_2\text{P}](\text{PF}_6)_4$  (26  $\mu\text{M}$ ) in degassed acetonitrile. The spectra shown in Figure S1 is the same as previously reported for  $[\text{H}_2\text{P}](\text{PF}_6)_4$  generated in situ.<sup>2</sup> The complex was converted to chloride salt with tetrabutylammonium chloride in acetone as described for  $[\text{P}]\text{Cl}_3$ .



**Figure S1:** Absorption Spectra of complex  $\text{P}^{4+}$ ,  $\text{P}^{3+}$  and  $\text{H}_2\text{P}^{4+}$  in MeCN ( $\text{PF}_6^-$  salts). Concentration for all species is 26  $\mu\text{M}$ .

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