

Supporting Information

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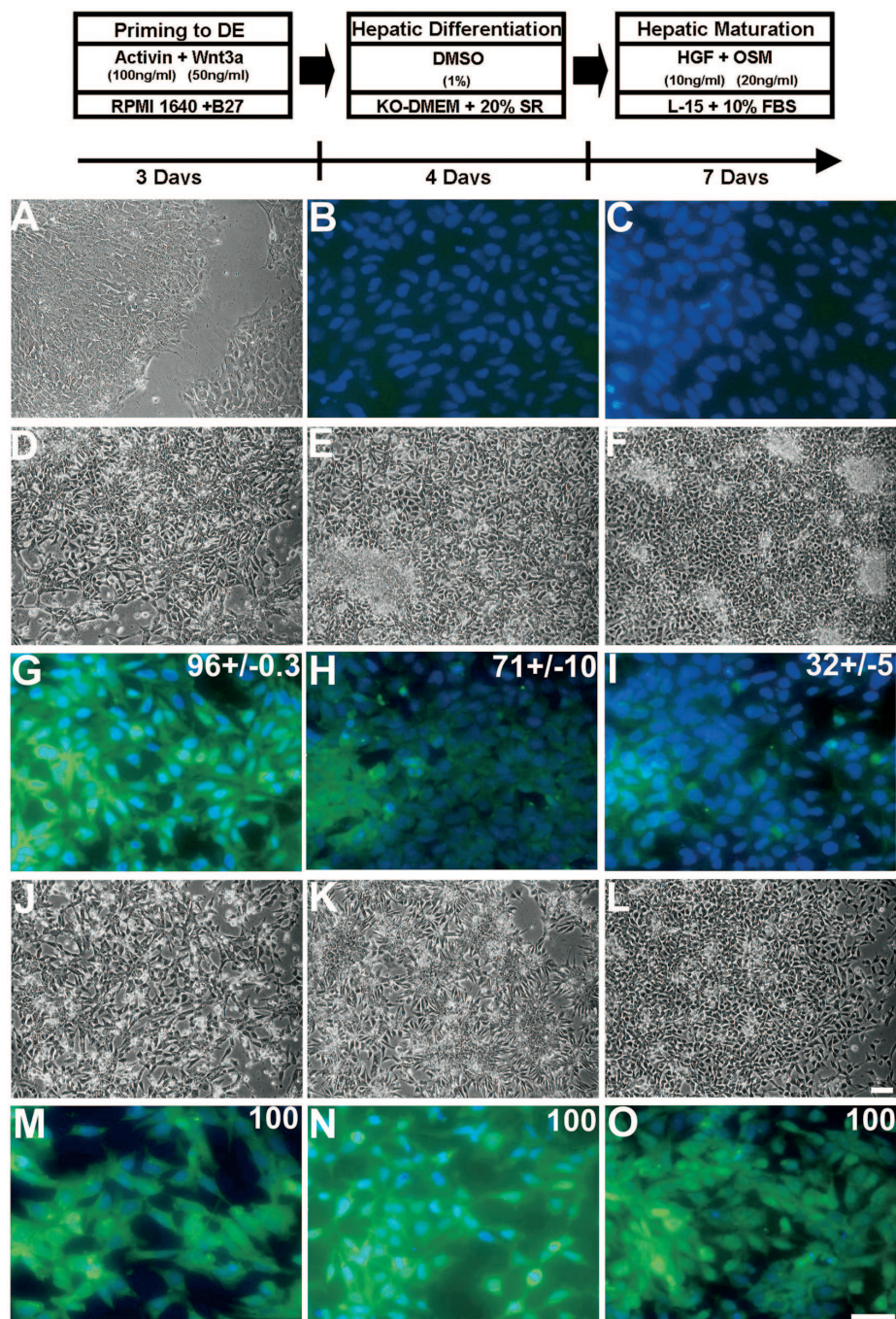


Fig. S1. AW promotes down-regulation of Brachyury in differentiating hESCs. The hESC differentiation protocol is shown at top. hESCs (A) exhibited a drastic change in cellular morphology in response to the different media. AW-differentiated cells rapidly formed a confluent monolayer (D–F) of cells exhibiting similar morphology. In contrast, AB-differentiated cells took longer to reach confluence (J–L). Consistent with different morphology, Brachyury gene expression was differently regulated in both conditions. hESCs did not express Brachyury (B). In contrast, cells differentiated with AW showed high levels of Brachyury at day 1, which decreased by day 3 (G–I). Cells treated with AB exhibited high-level Brachyury expression throughout the 72-h period (M–O). Specificity of staining was assessed using an IgG control (C). Phase images were recorded using a Nikon TE300/U, immunostaining was recorded using a Leica DMIRB. Scale bar = 100 μ M. HGF, hepatocyte growth factor; OSM, Oncostatin M. Results are presented as a mean of 4 fields of view \pm SE.

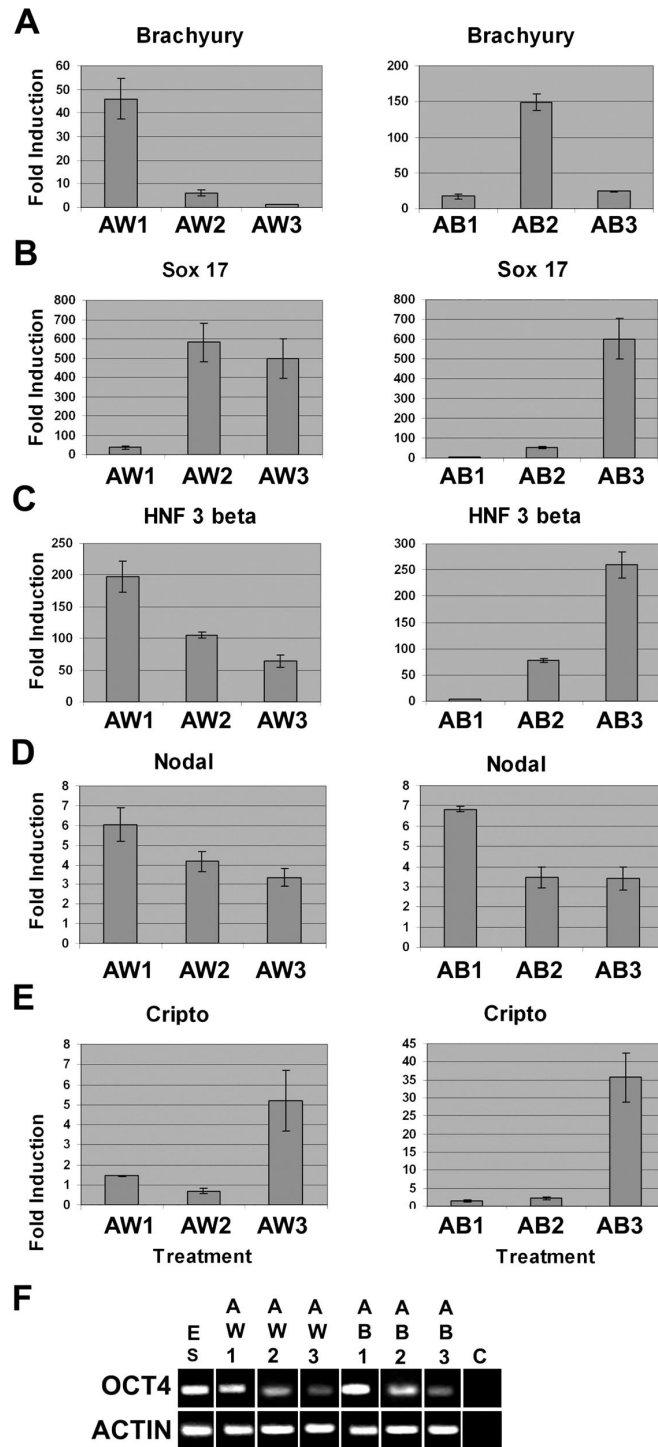


Fig. S2. AW treatment promotes accelerated progression of cells through the PS. hESC-directed mesoderm and endoderm differentiation was monitored by qPCR to *Brachyury*, *SOX17*, and *HNF3 β* . Cells differentiated with AW proceeded through mesoendoderm differentiation more rapidly than cells differentiated with AB. AW treatment exhibited rapid down-regulation of the mesoderm marker *Brachyury* (A, AW1–3 and AB1–3) and more rapid progression through the PS (*SOX17*, *HNF3 β*) than cells exposed to AB (B and C, AW1–3 and AB1–3). (D) *Nodal* levels decreased over the 3-day time course in both models (AW1–3 and AB1–3). (E) *CRIPTO* levels were maximally expressed at day 3 in both protocols, indicating cellular transition to the endoderm. (F) Down-regulation of the hESC marker *OCT4* was observed in both protocols over the 3-day period and absent in the –RT control. C, qPCR amplicons were measured using an ABI Prism 7900.

Table S1. Primer sequences and RT-PCR conditions

Gene	Primer Sequence	Annealing temperature, °C	Cycles	Size, bp
<i>AFP</i>	For-AGAACCTGTCACAAGCTGTG Rev-GACAGCAAGCTGAGGATGTC	55	30	675
<i>ALB</i>	For-CCTTTGGCACAATGAAGTGGGTAACC Rev-CAGCAGTCAGCCATTTACCATAGG	55	30	354
<i>AAT</i>	For-AGACCCTTTGAAGTCAAGCGACC Rev-CCATTGCTGAAGACCTTAGTGATGC	55	30	358
<i>TO</i>	For-GGCAGCGAAGAAGACAAATC Rev-TCGAACAGAATCCAATCCC	55	30	218
<i>HNF4α</i>	For-CTGCTCGGAGCCACCAAGAGATCCATG Rev-ATCATCTGCCACGTGATGCTCTGCA	55	30	370
<i>OCT4</i>	For-GACAACAATGAAAATCTTCAGGAGA Rev-TTCTGGCGCCGGTTACAGAACCA	55	30	218
<i>CYP3A4</i>	For-CCTTACATATACACACCCCTTG Rev-GGTTGAAGAAGTCCTCCTAAGCT	50	35	169
<i>TAT</i>	For-ACTGTGTTTGGAAACCTGCC Rev-GCAGCCACTTGTGAGAATGA	55	30	188
<i>APOF</i>	For-GGAAGCGATCAAACCTACCA Rev-ATCAGCCTGACAACCAGCTT	58	35	347
<i>CYP7A1</i>	For- CTGCCAATCCTCTTGAGTTCC Rev- ACTCGGTAGCAGAAAGAATACATC	57	35	387
β - <i>ACTIN</i>	For-TCACCACCACGGCCGAGCG Rev-TCTCCTTCTGCATCCTGTCC	58	25	350

[A]**AFP*, α fetoprotein; *ALB*, albumin; *TO*, tryptophan dioxygenase; *HNF4 α* , hepatocyte nuclear factor 4 α ; *OCT4*, Octamer 3/4; *CYP3A4*, cytochrome p450 3A4; *TAT*, tyrosine amino transferase; *APOF*, apolipoprotein F; *CYP7A1*, cytochrome p450 7A1; β -*ACTIN*, β actin.

Table S2. qPCR primer sequences and conditions

Gene	Primer/probe sequence
<i>BRACHYURY</i>	Applied Biosystems - hs00610079-ml
<i>HNF3β</i>	Applied Biosystems - hs00936490-ml
<i>SOX17</i>	Applied Biosystems - hs00751752-sl
<i>NODAL</i>	Applied Biosystems - hs01080749-ml
<i>CRIPTO</i>	Applied Biosystems - hs00414425-ml
<i>ALBUMIN</i>	Applied Biosystems - hs 00910225-m1
	For-TCCTCGCTGGCTACCCAAT
<i>WNT3</i>	Rev-CGGCAGAAGCGCAGTTG
	Probe-CCAGGGCCAGGGACCACCAA
	For-GCCCGCTCAGCCATGA
<i>WNT3α</i>	Rev-CCGTGGCACTTGCACTTGA
	Probe-CAGGCCATGCCAGCCACATG
	For-ACTTCTGCATGAAGAATGAGAAGGT
<i>WNT11</i>	Rev-TCGCTTCCGTTGGATGTCTT
	Probe-CCCACGGGACACAAGACAGGCAG
	For-GAGTATGCCTGCCGTGTG
<i>β2M</i>	Rev-AATCCAAATGCGGCATCT
	Probe-CCTCCATGATGCTGCTTACATGTCTC

WNT, wingless; *HNF3 β* , hepatocyte nuclear factor 3 β ; *SOX17*, SRY (sex-determining region Y)-box 17; *β -2M*, β 2 microglobulin.

Table S3. Immunostaining: primary and secondary antibodies

Antigen	Type	Source	Dilution
Wnt3a	Mo mono	R&D Systems	1/20
DPPIV	Go poly	R&D Systems	1/1000
Brachyury	Go poly	R&D Systems	1/100
Albumin	Mo mono	Sigma	1/500
Albumin	Rb poly	AbCam	1/600
CK18	Mo mono	DAKO	1/50
CK19	Mo mono	DAKO	1/50
hKi67	Mo mono	DAKO	1/400
Anti-Mo 488	Rb anti-Mo	Molecular Probes	1/500
Anti-Go488	Do anti-Go	Molecular Probes	1/500
Anti-Mo HRP	Rb anti-Mo	DAKO	1/100
Anti-Go HRP	Rb anti-Go	DAKO	1/100

Mo mono, mouse monoclonal antibody; Go poly, goat polyclonal antibody; Rb anti-Mo, rabbit antimouse; Rb-anti-Go, rabbit antigoat; Do anti-Go, donkey antigoat.