## Complete Nucleotide Sequence of the Polymerase 3 Gene of Human Influenza Virus A/WSN/33

JOHN S. KAPTEIN AND DEBI P. NAYAK\*

Department of Microbiology and Immunology, UCLA School of Medicine, Los Angeles, California 90024

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The complete nucleotide sequence of polymerase 3 (P3) gene of a human influenza virus (A/WSN/33) has been determined using cDNA clones except for the last 11 nucleotides which were obtained by direct RNA sequencing. The WSN P3 gene contains 2,341 nucleotides and codes for a protein of 759 amino acids (molecular weight 85,800). The WSN P3 protein, as deduced from the plus-strand DNA sequence, is basic and enriched in positively charged amino acids. In addition, it contains clusters of basic amino acids which may provide sites for the interaction of P3 protein with the capped primer, template, and/or other polymerase proteins during the transcriptive and replicative processes of influenza viral RNA.

Influenza A viruses contain eight RNA segments. After infection these RNA segments are transcribed into mRNA's which are translated into proteins. The proteins encoded by the corresponding individual RNA segments have been identified (11, 22, 29). Seven of these RNAs code for structural proteins which are eventually found in mature virions, and one (RNA8) codes for nonstructural proteins (NS1 and NS2) which are found only in infected cells.

Influenza RNA segments vary considerably in size. The largest ones are over 2,200 nucleotides long and code for the three polymerase (P1, P2, P3) proteins (24). The smaller RNAs range approximately from 1,800 to 900 nucleotides in length and code for hemagglutinin, neuraminidase, nucleoprotein, two or possibly more membrane proteins (17, 23a), and two nonstructural proteins (16). Using recombinant DNA technology and DNA sequencing procedures, the sequences of these five smaller viral RNA segments from one or more influenza A viruses were determined and the primary structures of the corresponding proteins were deduced.

Three polymerase proteins encoded by the three largest RNA segments are required for replication and transcription (15, 29). Both replication and transcription are complex processes which involve the interaction of these polymerase proteins with RNAs, other viral proteins, and host factors (12). Among the three polymerase proteins, P3 has recently been shown to recognize the 5' Cap 1 structure of cellular mRNA which is used as the primer during viral mRNA synthesis (33a). In addition, defective interfering viral RNA segments that have been examined to date also originate only from polymerase genes (20). However, although the three polymerase genes constitute at least 50% of the total RNA mass of influenza virus, virtually no information is available concerning the nucleotide or amino acid sequences of different polymerase genes or their variation among different subtypes or strains of influenza virus.

In an attempt to provide a basis for understanding the structure and function of polymerase proteins and to elucidate their role in the process of viral replication and transcription, we have undertaken DNA cloning and sequence analysis of polymerase genes. In this report we present the complete sequence of P3 gene of A/WSN/33 virus.

### MATERIALS AND METHODS

Virus and cells. Viral RNA used for cloning was obtained from ts52 virus (a group II temperaturesensitive mutant of A/WSN/33) which was grown in MDBK cells at 34°C. Viral RNA was isolated from purified virus preparation and enriched for polymerase genes by fractionating in sucrose velocity gradients (3).

**Recombinant DNA cloning and DNA sequencing of P3** gene. The P3 gene of influenza A/WSN/33 ts52 (21) was cloned as a double-stranded DNA copy in the *PstI* site of pBR322 (3). Accordingly, RNA enriched in polymerase genes was reverse transcribed into a plusstrand DNA copy using the 5'-specific primer 5' dAGCGAAAGCAGG 3'. Approximately full-length plus-strand DNA copies were isolated on 1.4% alkaline agarose gels, and copied into double-stranded DNA using fold-back of the 3' end as the self-primer. After S1 nuclease treatment, double-stranded DNA fragments were size fractionated on neutral agarose gels, and approximately 20 dC residues were added to their 3' ends. Finally the dC-tailed double-stranded



FIG. 1. Restriction map of A/WSN/33 P3 gene. Sites listed across the top of the diagram represent recognition sequences which are found only once in the P3 insert. Those listed along the side represent sequences found more than once. The sites of cleavage are designated by the small vertical bars. The following recognition sequences are not present: AccI, AsuII, AvaIII, AvrII, BamHI, CauII, ClaI, EcoK, GdiII, HgiAI, HgiCI, HgiEII, HpaI, KpnI, MstI, NaeI, NarI, PstI, PvuI, RruI, SaII, SmaI, SnaI, SphI, SstI, SstII, StuI, TthllII, XbaI, XhoI, XmaIII.

DNA was inserted into the *PstI* site of pBR322 to which approximately 20 dG residues had been added. *Escherichia coli*  $\chi$ 1776 cells were transformed, screened for tetracycline resistance, and characterized for insert size. Clones with inserts of approximately 2.2 to 2.4 kilobases (kb) were tentatively designated as clones of polymerase genes and analyzed to identify them as either of P1, P2, or P3 origin.

DNA sequencing of the P3 inserts was carried out by the methods of Maxam and Gilbert (18). In all cases, asymmetric cleavage by a second restriction enzyme was used for isolating DNA fragments uniquely labeled at one 5' end and, thereby, allowing sequence analysis of a fragment from its labeled ends.

### RESULTS

Identification of DNA clones of the P3 gene. Before extensive sequence analysis, a number of approaches were used to identify clones containing an insert of P3 origin. (i) As mentioned before, inserts of all clones in this group were approximately 2.2 to 2.4 kb and were thus larger than the expected size of any influenza gene except for the polymerase genes. (ii) All of these clones hybridized to combined polymerase gene RNAs isolated from gels, but not to other viral RNA segments. (iii) Restriction analyses classified these clones into three groups as expected for three polymerase genes. (iv) Clones of specific polymerase genes were identified by hybridization to specific defective interfering RNAs originating from known polymerase genes. For example, defective interfering RNA L1 and L2a of P3 origin hybridized only to the DNA from 1-14b and 1-26b clones. These defective interfering RNAs are easily separable by gel electrophoresis and have been extensively characterized (2). (v) Finally, similarity of the sequences at the 5' and 3' ends of the plus strands of these clones to the previously reported end sequences of P3 gene confirmed the identity of these clones (6a, 26).

Nucleotide sequence analysis. A preliminary restriction map was constructed using several of the enzymes which have six-nucleotide recognition sequences. The orientation of the P3 gene with respect to the pBR322 DNA was determined and found to be the same for both clones with the 3' end of the plus-strand DNA in close proximity to the PvuI site of pBR322. Initially, a cleavage map of the insert DNA was determined with a number of restriction enzymes. Appropriate cleavages were then employed to obtain the fragments used for sequencing. A detailed restriction map obtained from the complete sequence information and also partly confirmed by actual restriction enzyme analyses is shown in Fig. 1.

The series of fragments and restriction sites which were used in sequencing is shown in Fig. 2. All sites used as either the site of labeling or the site of second cleavage were also read through from another site to verify the continuity of overlaps. The sequence through EcoRII(*BstNI*) sites was verified by sequencing through the site from both strands, mapping of *BstNI* sites, and kinasing and sequencing from *BstNI* sites. Thus all gaps in the sequencing ladder, due to the presence of methylcytosine (22), were resolved.

The sequence presented was obtained from the clone 1-14b except for the nucleotides from 2322 to 2341 (Fig. 3). At the 5' end of the plus strand, clone 1-14b contains the complete sequence of the oligonucleotide primer used for priming DNA synthesis on the viral RNA (vRNA) template and is linked to pBR322 through 15 dG residues. At the 3' end, clone 1-14b ends at nucleotide 2321 followed by a tail of 11 dC residues. Clone 1-26b contains an additional viral sequence of 9 nucleotides, extending to position 2330 followed by a tail of 19 dC residues. However, it is not possible to ascertain whether the two cytosines at position 2329 and 2330 originated from reverse transcription of the viral RNA or from the addition of G:C residues. Finally, the last 11 nucleotides (position 2331-2341) were obtained by direct RNA sequencing of the vRNA (2). Confirmation of the correct overlap was also obtained by comparison of this sequence to the partial sequence of P3 gene of fowl plague virus (26).

The sequence at the 3' end of full length cDNA,

# J'UCAUCUUUGUUC, 5'---AAUAGUUUAAAAACGAC),

shows the likely loop structure involved in priming the double-stranded DNA synthesis. Arrows show the position of possible S1 nuclease cleavage sites which could generate the insert of the 1-14b and 1-26b clones. This fortuitous selfcomplementarity at the 3' end of the cDNA may have enabled us to obtain nearly full-length clones of this gene. The other genes of influenza virus differ in the nucleotide sequence beyond the last 13 nucleotides and therefore would not be expected to generate clones with the same degree of completeness by this procedure.

Characterization of the A/WSN/33 P3 (WSN P3) gene. The entire nucleotide sequence of the WSN P3 gene and the amino acid sequence of



FIG. 2. Sequencing strategy of cloned P3 DNA. Listed along the side are the restriction endonuclease sites which were kinased as shown by vertical bars. The site of cleavage by a second restriction enzyme is not shown. The solid lines represent the sum of the nucleotide sequence as determined from a series of overlapping gels. The dashed lines represent nucleotides which were run off the bottom of the gel and were not determined from those sites.

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VRNA CRNA	3'	UCB( AGC(		CGUCC 3CAGG	CAGUL BUCA	MUAN MUAN	NUAAG JAUUG	auua Caau	30 UAC AUG	CUU GAA GLU	UCU AGA ARG	UAU AUA ILE	UUU AAA Lys	CUU GAA GLU	GAU CUA LEU	UCC AGG ARG	UUA AAU ASN	GAU CUA 10 LEU	60 UAC AUG MET	AGC UCG SER	GUC CAG GLN	AGA UCU SER	GCG CGC ARG	UGA ACU THR	GCG CGC ARG	CUC GAG GLU	UAU AUA ILE	BAG CUC 20 LEU	90 UBU ACA THR
UUU	UGB ACC	UBG ACC	CAC GUG	CUG GAC	GUA CAU	UAC AUG	C66 6CC	UAU	120 UAG AUC	UUC	UUC	AUG	UGU ACA	AGU UCA	CCU GGA	UCU AGA	GUC CAG	CUC GAG	150 UUC AAG		GGU CCA	CGU GCA	GAA CUU	UCC AGG	UAC AUG	UUU	ACC UGB	UAC	180 UAC AUG
LYS	THR	THR	VAL	ASP	HIS	MET	ALA	30 ILE	ILE	LYS	LYS	TYR	THR	SER	GLY	ARG	GLN	40 GLU	LYS	ASN	PRO	ALA	LEU	ARG	MET	LYS	TRP	50 MET	HET
CBU GCA	UAC AUB	UUU AAA	AUA UAU	GGU CCA	UAA AUU	UGU ACA	CGU GCA	CUG GAC 60	210 UUC AAG	UCC AGG	UAU AUA	UGC ACG	CUU GAA	UAC AUG	uaa Auu	GGA CCU	CUC GAG	UCU AGA 70	240 UUA AAU	CUC GAG	GUC CAG	CCU GGA	guu Caa	UGA ACU	AAU UUA	ACC UGG	UCA AGU	UUU AAA B0	270 UAC AUG
ALA	HET	LYS	TYR	PRO	ILE	THR	ALA	ASP	LYS	ARG	ILE	THR	GLU	MET	ILE	PRO	GLU	ARG	ASN	GLU	GLN	GLY	GLN	THR	LEU	TRP	SER	LYS	HET
UUA AAU	CUB BAC	C86 8CC	CCU BGA	AGU UCA	CUG GAC	gcu Cga	CAC GUG	UAC AUG 90	300 Cau Gua	AGU UCA	GGA CCU	GAC CUG	CGA GCU	CAC GUG	UGU ACA	ACC UGG	ACC UGG	UUA AAU 100	330 UCC AGG	UUA AAU	CCU GGA	GGU CCA	CAC GUG	UGU ACA	UCA AGU	UGU ACA	CAA GUU	GUA CAU 110	360 AUA UAU
ABN	ABP	ALA	GLY	SER	ASP	ARG	VAL	MET	VAL	SER	PRO	LEU	ALA	VAL	THR	TRP	TRP	ASN	ARG	ASN	GLY	PRO	VAL	THR	SER	THR	VAL	MIS	TYR
96U CCA		UAB AUC	AUG		UGA ACU	AUA UAU		CUU GAA 120	390 UUU AAA	CAG GUC	CUU GAA	UCC AGG	AAU		gua Cau	CCU GGA	UGG ACC	AAA UUU 130	420 CCG GGC	GGA CCU	CAG GUC	GUA CAU		UCU AGA	UUG AAC		CAB BUC	UUU AAA 140	450 UAU AUA
PRO	LYS	ILE	TYR	LYS	THR	TYR	PHE	GLU	LYS	VAL	GLU	ARG	LEU	LYS	HIS	GLY	THR	PHE	GLY	PRU	VAL	MIS	PHE	AKU	ASA	ULN	VAL	LTB	ILE
BCA CGU	GCU CGA		CAA GUU VAL				GGA CCU PRO	CCA BGU 150 BLY	480 GUA CAU HIS	CGU GCA	CUA GAU ASP	GAG CUC	UCA AGU SER	CGG GCC		CUC GAG GLU	CGU GCA	GUC CAG 160 GLN	GAU ASP	CAU GUA VAL	UAG AUC ILE	UAC AUG HET	CUU GAA GLU	CAA GUU VAL	CAA BUU VAL	AAG UUC PHE	GGA CCU PRO	UUB AAC 170 ASN	CUU GAA BLU
									570										600										630
CAC GUG VAL	CCU GGA GLY	CBB BCC ALA	UCC AGG ARG	UAU AUA ILE	GAU CUA LEU	UGU ACA THR	AGC UCG SER	CUU GAA 180 GLU	AGC UCG SER	GUU CAA GLN	GAU CUA LEU	UGC ACG THR	UGU ACA THR	UGG ACC THR	UUU AAA Lys	CUC GAG GLU	UUC AAG LYS	UUU AAA 190 LYS	CUU GAA GLU	CUU GAA GLU	GAG CUC LEU	GUC CAG GLN	GGU GGU GLY	ACG UGC CYS	UUU AAA LYS	UAA AUU ILE	AGA UCU SER	GGA CCU 200 PRO	GAC CUG LEU
									660										690						~~~			~	720
NET	GUG VAL	COU BCA ALA	AUG UAC TYR	NET	LEU	GLU	AGA ARG	GAA 210 GLU	CUG LEU	GUC VAL	GCG CGC ARG	LYS	ACG	AGA	UUC PHE	CUC	CCA PRD	GUG 220 VAL	GCU ALA	GGU GLY	GGA GLY	ACA	AGC	AGU	GUG VAL	UAC	AUU	6AA 230 6LU	OUG VAL
									750	-									780		C114	<b>C114</b>		C114	0111				810
UUG	CAU	UUG	ACC	CAA	GGA GGA	ACA	UGC	UGG 240	GAA	CAG	AUG	UAC	ACU	CCA	GGA	6666 61 Y	GAG	8C6 250	AGG	AAU	GAU	GAU	GUU	GAU	CAA	ABC	UUA	AUU 260	AUU
LEU							0.0		840	02.0									870										700
CGA BCU	CGA GCU	UCU AGA	UUG AAC	UAU AUA	CAU GUA	UCU AGA	UCU AGA	CGG GCC 270	UGU ACA	CAC GUG	AGU UCA	CGU GCA	CUA GAU	GGU CCA	GAU CUA	CGU GCA	AGA UCU	AAU UUA 280	UUG	GAG	AUG	ACG UGC	GUG CAC	UCG <b>AGC</b>	ACG	GUC		CCA 660 290	CCU 86A
ALA	ALA	ARG	ASN	ILE	VAL	ARG	ARG	ALA	THR	VAL	SER	ALA	ASP	PRO	LEU	ALA	SER	LEU	LEU	GLU	MET	CYS	HIS	SER	THR	GLN	ILE	GLY	GLY
UAU AUA	UCC AGG	UAC AUG	CAU GUA	UUG AAC	UAG AUC	GAA CUU	UCC AGG	GUC CAG 300	930 UUG AAC	GGU CCA	UGU ACA	CUU GAA	CUC GAG	GUU CAA	CGG 6CC	CAC GUG	CUA GAU	UAA AUU 310	960 ACG UGC	UUC	CGA GCU	CGU GCA	uac Aug	CCU 664	GAC	UCU		UC8 ABC 320	990 ABU UCA
ILE	ARG	MET	VAL	ASN	ILE	LEU	ARG	GLN	ASN	PRO	THR	GLU	GLU	GLN	ALA	VAL	ASP	ILE	CYS	LYS	ALA	ALA	MET	GLY	LEU	ARG	IL	SEA	SER
AGG UCC	AAG	UCA AGU		CCA GGU	CCU GGA		UGU ACA	AAA UUU 330	1020 UUC AAG	UCU AGA	UGU ACA	UCG AGC	CCU GGA	AGU UCA	AGU UCA	CAG GUC		UCU AGA 340	1050 CUC GAG	CUU GAA	GAG	GUG	GAA	UGC	: CC6 660	UUA AAL	GA CU	900 J CAE 350	1080 UBU ACA
SER	PHE	SER	PHE	ULT	gr 1	PHE	t Pak	r nE	L15	AKG	1 1986	9EK	ULT	9FK	9EK	VAL	L18					VHL	LEU			- R81		J ULI	THR
			UCU AGA	CAC GUG	BUA CAU	CUC GAG		AUA UAU 360	GAA	CUC GAG		UGU ACA	UAC AUG	CAA GUU	CCC 666			GCA 370									G AG	C UCI 3 AG	1170 J AAC A UUG
F	LYB IG.	л.е 3.	P3	gen	e of	οιυ [ Α/	WS	N/3	ыси 3. 1	The	nuc	leo	⊓⊾⊺ tide	sec	uen	HRG	oft	, ⊪⊾≉ hoth	the	- m	, 1LE	 . (v]		) LTE	trar	n Mi nda	к АR nd	⊎AR® the	e LEU nius

(cRNA) strand is shown. Numbering of the nucleotides is from the 5' end of the plus strand. Also shown is the amino acid sequence of the P3 protein as deduced from translation of the nucleotide sequence starting from the first AUG of the plus strand.

the P3 protein, as deduced from the plus-strand sequence starting from the first AUG (12, 13), are presented in Fig. 3. The WSN P3 gene is 2,341 nucleotides long. It is initiated and terminated by the known 13 nucleotide conserved sequences at the 5' and 3' ends.

The plus strand at the 5' region contains 27 untranslated nucleotides prior to the first AUG. This reading frame is then open for almost the entire gene and ends with a termination codon (UAG) at nucleotide position 2305 followed by a second in-phase termination codon (UAA) at

58

UAA	GUC	BAC	UNU	CAC	UCA	CCC	UCC	CUG	1200 CUU	OUC	AGC	UAA	CGG	CUU	CGU	UAU	UAA	CAC	1230 COO	UAC	CAU	-	ABU	<b>8U</b> U	CUC	CUA	ACA	UNC	260 UAU
ILE	GLN	LEU	ILE	VAL	SER	GEG	AGG	GAC 390 ASP	GAA	GLN	SER	AUU	ALA	GLU	ALA	ILE	AUU	400 400 VAL	ALA	NUG	gua Val	PHE	UCA SER	GLN	GAG GLU	GAU	CYS	AUB 410 MET	ILE
									1 290										1 3 20										150
UUU AAA	CGU GCA	CAA GUU	UCU AGA	CCA GGU	CUG GAC	GAC CUG	UUA AAU	AAG UUC	CAG	UUA AAU	UCC AGG	CGC GCG	UUA AAU	GUC CAG	GCU CGA	AAC UUG	UUG AAC	600 CCC	UAC AUB	GUG CAC	BUU CAA	GAA CUU	AAC UUS	UCU AGA	GUA CAU		OUC CAG		CUA
LYS	ALA	VAL	ARG	GLY	ASP	LEU	ASN	420 PHE	VAL	ASN	ARG	ALA	ASN	GLN	ARG	LEU	ASN	430 PR0	HET	HIS	<b>OL</b> N	LEU	LEU	ARG	HIS	PHE	OLN	440 LYS	ASP
COU	UUC	COC	GAG		GULL	ULA	ACC	ccu	1380	CUN	<b>6</b> 66	UAG	CUB		CAC	UAC	CCU		1410	000			886	0.18				1	440
GCA	AAG	000	CUC	UUU	CAA	AAU	UGG	66A 450	AUU	GAA	ÜCC	AUC	GAC	AAU	GUG	AUS	OGA	AUB 460	AUC	666	AUA	UUG	CCC	BAC	AUG	ACU	CCA	ABC 470	ACC
nLn	LTB	ALA	LEU	PHE	GLN	ASN	TRP	GLY	ILE	GLU	SER	ILE	ASP	ASN	VAL	MET	GL Y	MET	ILE	OL Y	ILE	LEU	PRO	ASP	HET	THIR	PHO	SER	THE
CUC GAG	UAC	AGU UCA	UAC		CCU GGA	CAC	UCU	UAG	1470 UCG AGC		UAC	CCC 666	CAU	CUA		AUA	AGG		1500 COC		UUC	UAU	CAC	CAC	UCS			OCA COU	530 AAA
<b>BL</b> U	HET	SER	NET	ARG	GLY	VAL	ARG	480 ILE	SER	LYS	HET	GLY	VAL	ASP	GLU	TYR	SER	490 SER	ALA	BLU	LYS	ILE	VIL	WAL	SER	ILE	ARP	500 ARE	PHE
									1560										1590									1	420
MAC UUS	UCU AGA	GUU	AGG	CUG GAC	GUU CAA	GCA CGU	CCC 666	UUA AAU 510	CAU GUA	gau Gua	GAC CUG	AGA UCU	666 CCC	CUC GAG	CUC GAG	CAG GUC	UÇA AGU	CUU GAA 520	UGU ACA	OUC CAB	CCU OGA	UQU ACA	CUC BAB		GAC CUB	UBU ACA	NNU	UBA ACU 530	AUB UAC
LEU	ARG	VAL	ARG	ASP	GLN	ARG	GLY	ASN	VAL	LEU	LEU	SER	PRO	GLU	GLU	VAL	SER	OLU	THR	<b>BL</b> N	<b>BL</b> Y	THR	<b>OL</b> U	LYS	LEU	THR	ILE	THR	TYR
ABU	AGC	AGU	UAC	UAC	ACC	CUC	UAA	UUA	1650 CCA	GGA	CUU	AGU	CAC	MAC	CAG	UUA	U86	AUA	1480 BUC	ACC	UAG	UAG	UCU	UUB	ACC	CUU	UBA	CAA	710 UVU
	UCG	UCA	AUG	AUG	UGG	GAG	AUU	540	GGU	CCU	GAA		GUG	UUG	GUC	AAU	ACC	UAU 550	CAG	U96	AUC	AUC	A6A	AAC	U96	GAA GAA	ACU	6UU 540	
																									· · ·			••••	
	OUC CAG	ACC UGG	AGG UCC	GUC CAG	UUA AAU	GGA CCU	UGU ACA	UAC AUG	GAC CUG	AUG UAC	UUA AAU		UAC AUG	CUU GAA		CUC GAG	66U CCA	-	BUC CAG	AGA UCU	AAU UUA	CAA BUU	00U CCA	UUC AAG	C96 9CC	CAA GUU	UCU AGA		
ILE	GLN	TRP	SER	GLN	ASN	PRO	THR	570 MET	LEU	TYR	ASN	LYS	HET	GLU	PHE	GLU	PRO	580 Phe	GLN	SER	LEU	VAL	PRO	LYS	ALA	VAL	ARG	570 BL Y	QL.H
A110						1104	RAC		1830				C114						1860			~~~						1	
UAC	AGU	666	ΰΰΰ	GUG	AGA	ACU	CUG	UUC 600	CAA	CAA	AUG	AGG	GAU	GUG	CUU	606	ACA	UUU 610	GAU	ACC	ecu	CAB	AUA	AUA	***	CUU	CUU	CCC 620	UUC
TYR	SER	GLY	PHE	VAL	ARG	THR	LEU	PHE	GLN	GLN	MET	ARG	ASP	VAL	LEU	GLY	THR	PHE	ASP	THR	ALA	GLN	ILE	ILE	LYS	LEU	LEU	PRO	PHE
COU	C96	CGA	GGU	86U	UUC	GUU		CCU	UAC	GUC	AAG	AGG	AGU	AAC	UGA	UAU	UUA	CAC	1950 UCC	CCU	AGU	CCU	UAC	UCU	UNU	BAA	CAU	UCC	780 CC8
ALA	ALA	ALA	PRO	PRO	LYS	GLN	SER	630 GL Y	MET	GLN	PHE	SER	SER	LEU	THR	ILE	ASN	640 VAL	ARG	GLY	SER	GLY	MET	ARG	ILE	LEU	VAL	450 AR6	BEC BLY
								:	2010										2040										070
	AGA UCU	66U CCA	UAU AUA	AAG UUC	UUG AAC	AUG UAC	UUG AAC	UUC	UGG ACC	UGA ACU	UUU AAA	UCU AGA	GAG CUC	UGU ACA	CAA GUU	GAG CUC	CCU GGA		CUA GAU	CGA GCU	CC6 06C	GGA CCU	AAU UUA	UGA ACU	CUU BAA	CUB GAC	66U CCA	CUA	CUU
ASN	SER	PRO	ILE	PHE	ASN	TYR	ASN	LYS	THR	THR	LYS	ARG	LEU	THR	VAL	LEU	GL Y	670 LYS	ASP	ALA	GLY	PRO	LEU	THR	BLU	ASP	PRO	480 ASP	9LU
668	UGU	CBA	ccu	CAA	CUC	AGG	CGU	CAA	2100 GAC	исо	CCU	846	GAG	-	BAC	000	111.91	CIRI	2130			<b>A</b> 11A		0011	-				140
99C	ACA	OCU	GGA	GUU	GAG	UCC	BCA	GUU 690	CUG	AGA	GGA	UUC	CUC	AUU	CUG	GGC	AAA	6AA 700	GAC	AGG	AGA	UAU	GGA	CCA	GCA	UUA	ABC	AUA 710	AAU
ULT	INK	MLA		VINL	GLU	SER	ALA	VAL	LEU	AKU	ULT	PHE	LEU	ILE	LEU	GLY	LTS	GLU	ASP	ARG	ARG	TYR	GLY	PRO	ALA	LEU	SER	ILE	ASN
CUU	GAC		UUG	GAA	CGC 6C6				2190 UUC	CGA	UUA	CAC	GAU		000	GUU	CCU	CUG	2220 CAC	CAC	AAC	CAU	UAC	UUU	OCC	UUU	900	006	250 ABA
OLU	LEU	SER	ASN	LEU	ALA	LYS	GLY	720 GLU	LYS	ALA	ASN	VAL	LEU	ILE	GLY	GLN	GLY	730 ASP	VAL	VAL	LEU	VAL	NET	LYS	ARG	LYS	ARB	AAC 740 ASH	UCU
								-	280										2310										
UC8 AGC	UAU AUA	GAA CUU	UGA ACU	CUG GAC	UCG AGC	GUC CAG	UGU ACA	CGC	UGG	UUU AAA	UCU AGA	uaa Auj	8CC C66	UAC AUG	CGG GCC	uag AUC	UUA AAU	AUC UAG	ACA	BCUU CGAAI	NUCAI	JUUAA	JUUGO	CUGG/ BACCI	MACA JUGU		JGA S	5' 3'	
SER	ILE	LEU	THR	ASP	SER	GLN	THR	ALA	THR	LYS	ARG	ILE	ARG	HET	ALA	ILE	759 ASN												
												FI	G. 3	3—0	Con	tinu	ed												

position 2320 (Fig. 3 and 4). The 37 nucleotides at the 3' end which are not translated contain the proposed polyadenylation site (positions 2321 to 2325) of the mRNA (27).

The frequency of codon usage is shown in Table 1. As found for other eucaryotic genes, CG-containing codons are relatively few. On a random basis considering the base composition of P3 plus-strand RNA, one would expect 83 CG-containing codons, but only 28 occur in the P3 gene. This bias against CG is particularly evident in the usage of CCG and CGN coding for proline and arginine, respectively. Furthermore, the occurrence of CG dinucleotide irrespective of its presence in codons is also low (2.4% compared to expected 4.7%). A similar deficien-

cy in CG dinucleotide whether present in codons or between adjacent codons has been reported for other genes of influenza virus and for other viruses as well. Thus the bias against CG dinucleotide appears to be operating at the level of both DNA (33) as well as RNA genomes.

The portion of the gene which can be translated extends from the nucleotide position 28 to 2304 and codes for 759 amino acids. The other two reading frames of the plus strand and all three reading frames of the minus strand are blocked repeatedly by termination codons and are unlikely to be used in synthesizing functional proteins (Fig. 4). However, sequences over 300 nucleotides without any termination codons are present in these reading frames. Of interest in this respect is the first AUG of the minus strand (phase 3) at nucleotide position 40, which is present approximately in the same relative position as the first AUG of the plus strand. This AUG is followed by an open reading frame extending for 300 nucleotides. However, the significance of an open reading frame in the minus strand is unknown.

Sites which closely resemble consensus donor or acceptor sites for splicing

reference 30) can be found in both the plus and minus strands (in the plus strand, donor sites at positions 12, 399, 1057, 1274, and 1579, and acceptor sites at positions 1344 and 1894; and in the minus strand, donor sites at positions 413, 1278, 1673, 2172, and 2329, and acceptor sites at positions 230 and 965). Since neither altered mRNA's nor altered P3 proteins have been demonstrated, the significance of these potential splicing sites is unknown. Furthermore, splicing sites do not appear to be involved in generating influenza defective interfering RNAs (D. P. Nayak, N. Sivasubramanian, A. R. Davis, R. Cortini, and J. Sung, Proc. Natl. Acad. Sci. U.S.A., in press).

The P3 protein predicted from our sequence data contains 759 amino acids and has a molecular weight of 85,800. This compares favorably with previous estimates of the size of the P3 protein, the smallest of the three polymerase proteins with molecular weights ranging from  $80 \times 10^3$  to  $100 \times 10^3$  (24, 29). Our data therefore suggest that the primary translation product is probably the functional protein. However, in the absence of either the amino- or carboxyterminal amino acid sequences, any minor proteolytic cleavage or additional modification cannot be ruled out.

In polyacrylamide gel electrophoresis, P3 RNA runs as the largest RNA segment whereas P3 protein migrates as the smallest of three polymerase proteins (24). Our sequence data, however, do not support a reduced coding capacity of P3 mRNA or a major cleavage of the primary translation product to generate the P3 protein. It is therefore likely that this represents an anomalous migratory behavior of either polymerase RNAs or polymerase proteins, or both, in polyacrylamide gel electrophoresis or a posttranslational modification of P1 and P2 proteins or a combination of these factors.

Table 2 shows the amino acid composition of the predicted P3 protein. Clearly it is a basic protein. Horisberger (10) has also reported that P1 and P3 are basic proteins and that P2 is acidic. In addition we find that the P3 protein contains a large excess of methionine (36 residues) and fewer cysteine residues (5 residues) when compared to the average composition of proteins (5). P3 protein has a very hydrophilic amino end and does not contain any large clusters of hydrophobic or nonpolar amino acids at either amino- or carboxy-terminus. Therefore, this protein is unlikely to be attached to membranes during its biosynthesis, transport, or assembly into virions.

## DISCUSSION

The P3 gene reported here contains 2341 nucleotides and is the largest of the influenza viral genes sequenced to date. The influenza WSN P3 gene contains sequences which are similar to the known partial sequences of this gene from other influenza viruses. For example, in the plus strand, a comparison with A/PR/8 P3 gene (6a) shows only two changes in the first 110 nucleotides (position 51,  $G \rightarrow A$ , and position 75,  $U\rightarrow C$ ). Similarly, at the 5' end, the partial sequence of fowl plague virus segment one RNA (26) differs at two positions out of 49 (position 17, U $\rightarrow$ A, and position 33, A $\rightarrow$ G), and at the 3' end two positions out of 63 vary (position 2307,  $G \rightarrow A$ , and position 2316,  $A \rightarrow U$ ). These changes occur in either the noncoding region or in the last position of a codon without altering any of the amino acid residues. However, additional sequence studies of other A viruses will be required before assessing the diversity and lineage of the P3 genes among subtype A viruses.

Studies of temperature-sensitive mutants have shown that all three polymerase genes are involved in viral RNA (both plus and minus strand) synthesis (15, 29). Moreover, Krug and his colleagues (33a) have recently shown that the P3 protein recognizes the 5' terminal Cap I structure of host mRNA which is used as a primer in viral transcription. They have also shown that the P3 protein remains associated



FIG. 4. Termination codons in the sequence of the P3 gene of A/WSN/33. The vertical bars represent termination codons found in the plus strand (top) and minus strand (bottom). 1, 2, and 3 represent the reading frames phased from the first, second, and third nucleotides, respectively. The arrows represent the position of the first AUG in the plus and minus strands.

with this cap structure throughout the transcription process.

As mentioned earlier, P3 is a basic protein and contains 115 basic amino acid residues (60 Arg, 45 Lys, 10 His) compared to 81 acidic amino acid residues (29 Asp, 52 Glu). Charge calculations indicate that P3 is more basic than nucleoprotein and membrane proteins, the other two basic influenza proteins for which the primary structure has been deduced (35, 36a). At pH 6.5, each molecule of WSN P3 protein has a net charge of +29 compared to +14 for PR/8 nucleoprotein (36) and +9.5 for PR/8 membrane proteins (36).

When a secondary structure analysis was per

TABLE 1. Frequency of codon usage in A/WSN/33 P3 gene

				F	requency			
		U		ċ		Α		G
U	Phe	13	Ser	7	Tyr	7	Cys	1 U
		11		6		9		4 C
	Leu	7		14	Term	0	Term	0 A
		13		5		1	Тгр	10 G
С	Leu	11	Pro	9	His	8	Arg	3 U
		9		4		2		3 C
		7		16	Gln	15		3 A
		13		0		21		3 G
Α	Ile	19	Thr	12	Asn	23	Ser	9 U
		10		11		12		13 C
		24		25	Lys	28	Arg	31 A
	Met	36		5		17		17 G
G	Val	14	Ala	11	Asp	14	Gly	7 U
		8		11		15		7 C
		9		17	Glu	27		27 A
		26		6		25		9 G

formed using Chou-Fasman (1) analysis supplemented by the helix wheel plot (28), P3 protein was found to contain several clusters of basic amino acids in non- $\alpha$ -helical regions (e.g., amino acid residues 140, 213, 375, 736, and 752). These clusters contain three to four arginine and lysine residues in close proximity without being interrupted by acidic residues. These clusters of basic amino acids are much more pronounced than those reported for PR/8 nucleoprotein (36a) and PR/8 membrane (35, 36) proteins. In addition, the P3 protein also contains an  $\alpha$ -helical region where basic amino acids are spaced three

TABLE 2. Amino acid composition of P3 protein (A/WSN/33) as deduced from the nucleic acid sequence

Amino acid	No. of residues
Ala	 45
Arg	 60
Asn	 35
Asp	 29
Cys	 5
Gln	 36
Glu	 52
Gly	 50
His	 10
Ile	 53
Leu	 60
Lys	 45
Met	 36
Phe	 24
Pro	 29
Ser	 54
Thr	 53
Trp	 10
Tyr	 16
Val	 57

to four amino acids apart (amino acid residues 431 to 448). In this region one side of the  $\alpha$ -helix presents a cluster of basic groups. Since the P3 protein interacts with the capped primer RNA (33a), and probably also with the template viral RNA and other polymerase proteins as well (e.g., P2), the clusters of basic amino acid residues may provide areas of interaction. Similar RNA-protein interaction via clusters of basic amino acids has been proposed for influenza nucleoprotein (36a), Semliki forest virus nucleocapsid (7), VP1 of simian virus 40 (34) and polyoma virus (31), and the core antigen of hepatitis B virus (25).

One of the major difficulties in studying the polymerase proteins has been the small amount of P proteins present either in infected cells or in virions (probably one to two molecules per RNA segment). Therefore, although a peptide mapping analysis of fowl plague P3 protein (11) has been performed, neither the amino acid composition nor a direct amino acid sequencing of polymerase proteins of any influenza virus has been possible. However, since cloned influenza genes can now be expressed in eucaryotic (8, 8a, 31) as well as procaryotic (4, 5, 9) systems, it should be possible to express P3 clones and to produce relatively large amounts of functional protein. This would then help in defining the role of the P3 protein in the transcription/replication process of influenza viruses.

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