

Characterization of the tautomycin biosynthetic gene cluster from *Streptomyces spiroverticillatus* unveiling new insights into dialkylmaleic anhydride and polyketide biosynthesis*

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Running title: Tautomycin biosynthetic gene cluster from *S. spiroverticillatus*

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TABLE S1. Targeted gene inactivation by the REDIRECT technology in *S. spiroverticillatus* (1)

gene	primers ^a	cosmid ^b	strain
<i>ΔtmJ</i>	<i>tmJF1</i> : 5' <u>GCCGGT</u> CACGAACCGGACCGGCTCACGTCCGCCACCGAGATTCCGGGGATCCGTCGACC-3' <i>tmJR1</i> : 5'- <u>CACCGCAACGGAATCGGACGGCGCCTCGGGCGT</u> CGCGTCTGTAGGCTGGAGCTGCTTC-3'	pBS6018	SB6003
<i>ΔtmK</i>	<i>tmKF1</i> : 5'- <u>GTGGTGGAGACGTCGTCCGGCTCGATT</u> CGGGGCTTCTGATTCCGGGGATCCGTCGACC-3' <i>tmKR1</i> : 5'- <u>TCAGTCCGCCGGTACGCCGATCCGGTGC</u> GGGGCCGGTGTGTAGGCTGGAGCTGCTTC-3'	pBS6019	SB6004
<i>ΔtmP</i>	<i>tmPF1</i> : 5'- <u>ATGGCGATTCCGCGCGGCAGAACCGAATGTGGCAGGCCGA</u> -TTCCGGGGATCCGTCGACC-3' <i>tmPR1</i> : 5'- <u>TCATGCCCGGCTCCCCGGCCGTCGAGCGGGCGGGCGCTGTAGGCTGGAGCTGCTTC</u> -3'	pBS6021	SB6006
<i>ΔtmR</i>	<i>tmRF1</i> : 5'- <u>GTGACCCCGGCCGAACCGGCTGGGCGCGCTGGCCGA</u> ACTCATTCCGGGGATCCGTCGACC-3' <i>tmRR1</i> : 5'- <u>TCAGGTCGCCGAACGCAGCAGGCAGGTGATCGCCCGGGTGTAGGCTGGAGCTGCTTC</u> -3'	pBS6022	SB6007
<i>ΔtmS</i>	<i>tmSF1</i> : 5'- <u>ATGACGG-ACGCGCGAACCGGTGCGCTGCTGGCCGTGCTG</u> ATTCCGGGGATCCGTCGACC-3' <i>tmSR1</i> : 5'- <u>TCACGCGCCCACCACCGCATCGGCCGGATCGGGGA</u> ACTGTAGGCTGGAGCTGCTTC-3'	pBS6023	SB6008

^aUnderlined letters represent the 39 nt homologous to the DNA regions internal to targeted genes.

^bpBS6018 and pBS6019 are based on pBS6016, and pBS6021, pBS6022 and pBS6023 are based on pBS6017.

TABLE S2. Southern Analysis Confirming the Genotypes of Mutant Strains^a

strains	gene targeted	probe		fragment replaced (bp)	Restriction digestion	Signal Size (kp)	
		primers used to amplify the probe	Size (bp)			WT	mutant
SB6003	<i>ΔtmJ</i>	<i>tmJFP2</i> : 5'-GGACGCCGAATACTGGTGC-3' <i>tmJRP2</i> : 5'-GGGCGAGATGCCGAAGAA-3'	918	288	<i>Bam</i> HI	4.0	3.5
SB6004	<i>ΔtmK</i>	<i>tmKFP2</i> : 5'-CGGTCAGGGCCGCCACGAGCTCTG-3' <i>tmKRP2</i> : 5'-CCCGAGTGGAGGTGTTTCG-3'	853	1292	<i>Bam</i> HI	3.7	1.3
SB6006	<i>ΔtmP</i>	<i>tmPFP2</i> : 5'-GGC-TGGTGTCCATGGTCGGC-3' <i>tmPRP2</i> : 5'-GAACTTCAGCCGGGCACCC-3'	1405	1156	<i>Pvu</i> I	3.3	1.7 1.4
SB6007	<i>ΔtmR</i>	<i>tmRFP2</i> : 5'-GAACCGGTCGCCGAGAACG-3' <i>tmRRP2</i> : 5'-GGCATGCCCTGCTCCCCG-3'	1490	1324	<i>Sma</i> I	1.6	2.3
SB6008	<i>ΔtmS</i>	<i>tmSFP2</i> : 5'-ACGCGTCGGTCTGTTCCGCG-3' <i>tmSRP2</i> : 5'-TACCTGGGGCACGCGACC-3'	659	747	<i>Apa</i> I	1.6	2.2

^aSee Figures S1, S2, S3, S4, S5 for details.

FIGURE S1. Inactivation of *ttmJ* by gene replacement. (A) Construction of the Δ *ttmJ* gene replacement mutant and restriction maps of *S. spiroverticillatus* wild-type and SB6003 mutant strains showing predicted fragment sizes upon *Bam*HI digestion. (B) Southern analysis of the wild-type (lane 5) and SB6003 (lanes 2, 3 and 4 are three individual isolates) genomic DNAs digested with *Bam*HI using the 918-bp amplified DNA fragment as a probe. Lane 1, molecular weight standard.

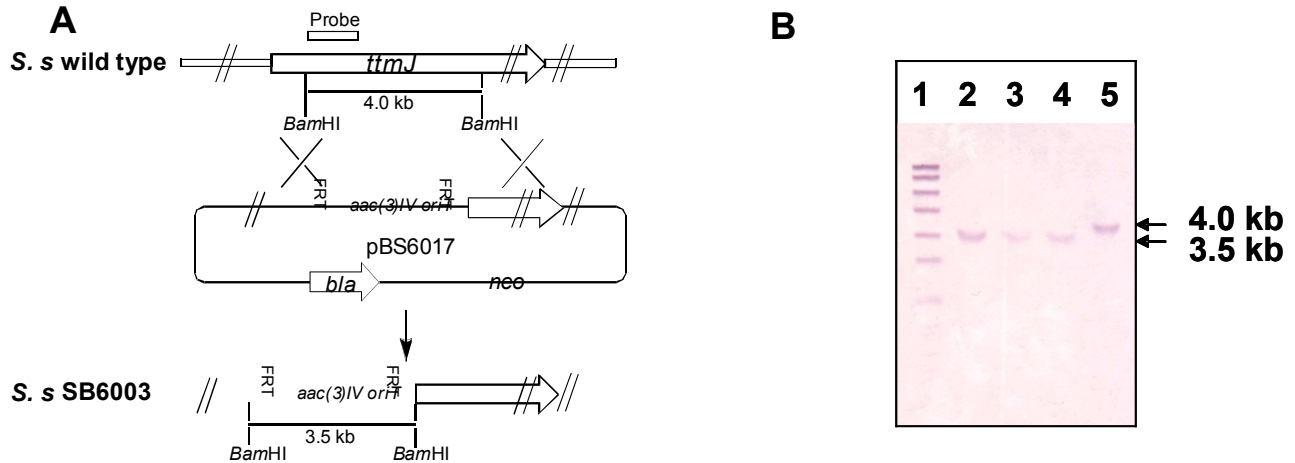


FIGURE S2. Inactivation of *ttmK* by gene replacement. (A) Construction of the Δ *ttmK* gene replacement mutant and restriction maps of *S. spiroverticillatus* wild-type and SB6004 mutant strains showing predicted fragment sizes upon *Bam*HI digestion. (B) Southern analysis of the wild-type (lane 1) and SB6004 (lanes 2, 3 and 4 are three individual isolates) genomic DNAs digested with *Bam*HI using the 853-bp amplified DNA fragment as a probe. Lane 5, molecular weight standard.

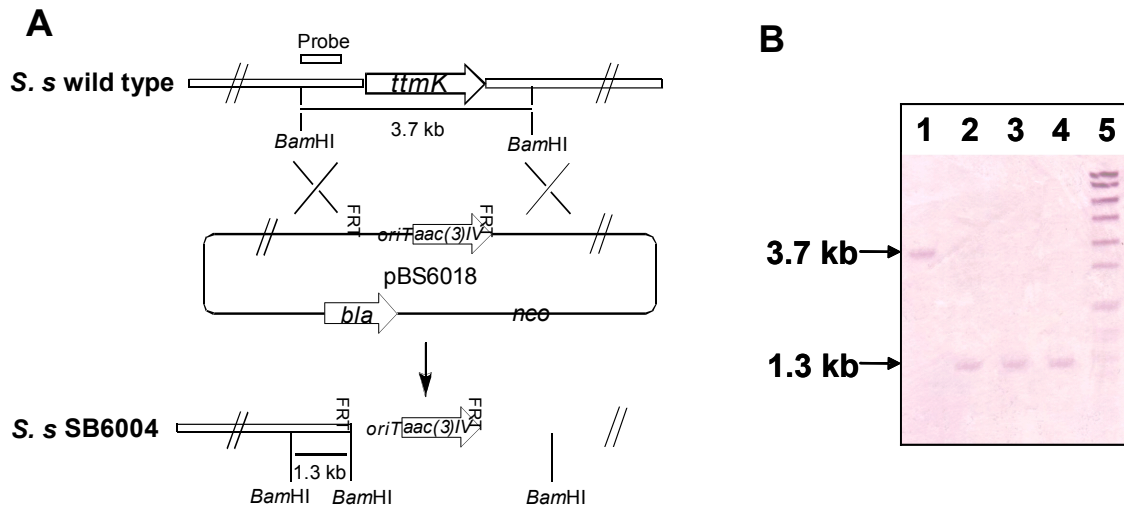


FIGURE S3. Inactivation of *ttmP* by gene replacement. (A) Construction of the *AttmP* gene replacement mutant and restriction maps of *S. spiroverticillatus* wild-type and SB6006 mutant strains showing predicted fragment sizes upon *PvuI* digestion. (B) Southern analysis of the wild-type (lane 2) and SB6006 (lanes 3 and 4 are two individual isolates) genomic DNAs digested with *PvuI* using the 1405-bp amplified DNA fragment as a probe. Lane 1, molecular weight standard.

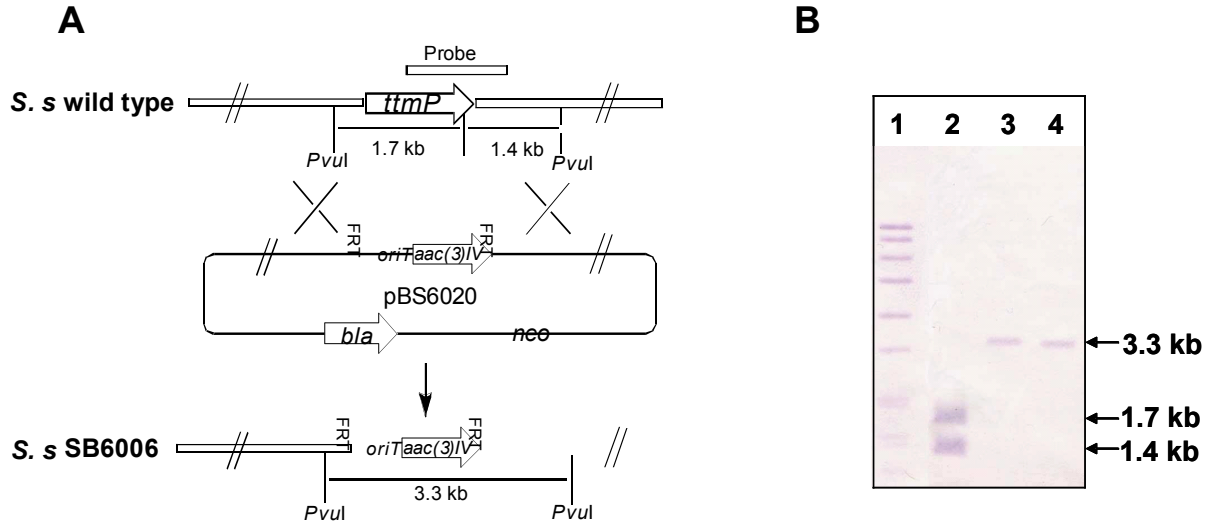


FIGURE S4. Inactivation of *ttmR* by gene replacement. (A) Construction of the *AttmR* gene replacement mutant and restriction maps of *S. spiroverticillatus* wild-type and SB6007 mutant strains showing predicted fragment sizes upon *SmaI* digestion. (B) Southern analysis of the wild-type (lane 8) and SB6007 (lanes 2-7 are six individual isolates) genomic DNAs digested with *SmaI* using the 1490-bp amplified DNA fragment as a probe. Lane 1, molecular weight standard.

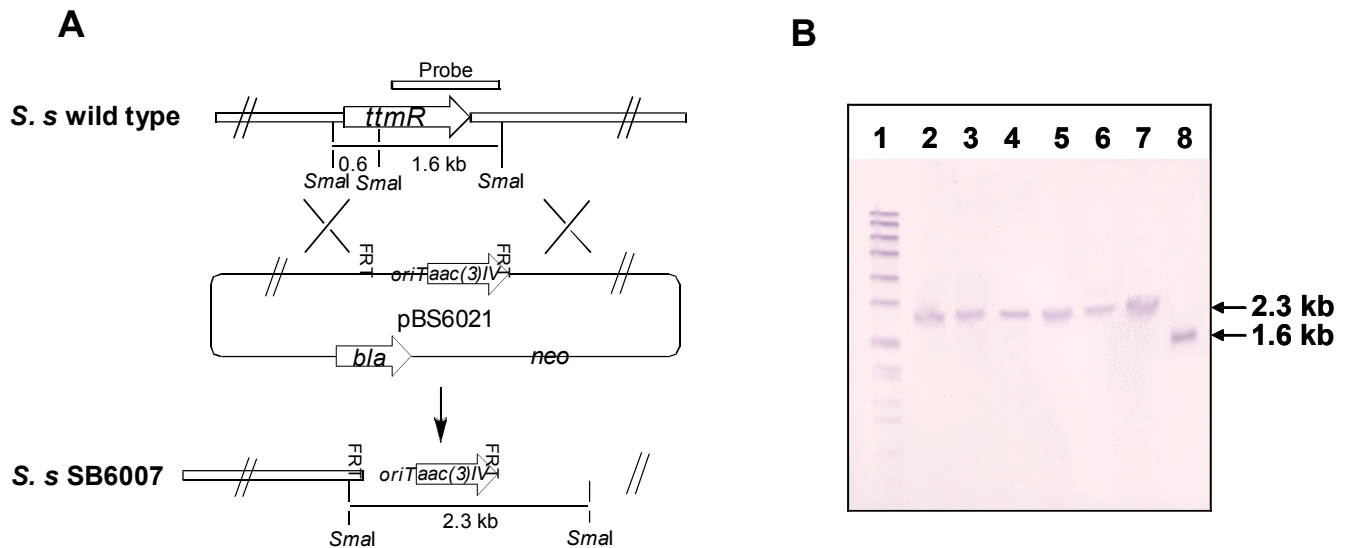


FIGURE S5. Inactivation of *ttnS* by gene replacement. (A) Construction of the $\Delta ttnS$ gene replacement mutant and restriction maps of *S. spiroverticillatus* wild-type and SB6008 mutant strains showing predicted fragment sizes upon *ApaI* digestion. (B) Southern analysis of the wild-type (lane 3) and SB6008 (lanes 1 and 2 are two individual isolates) genomic DNAs digested with *ApaI* using the 659-bp amplified DNA fragment as a probe.

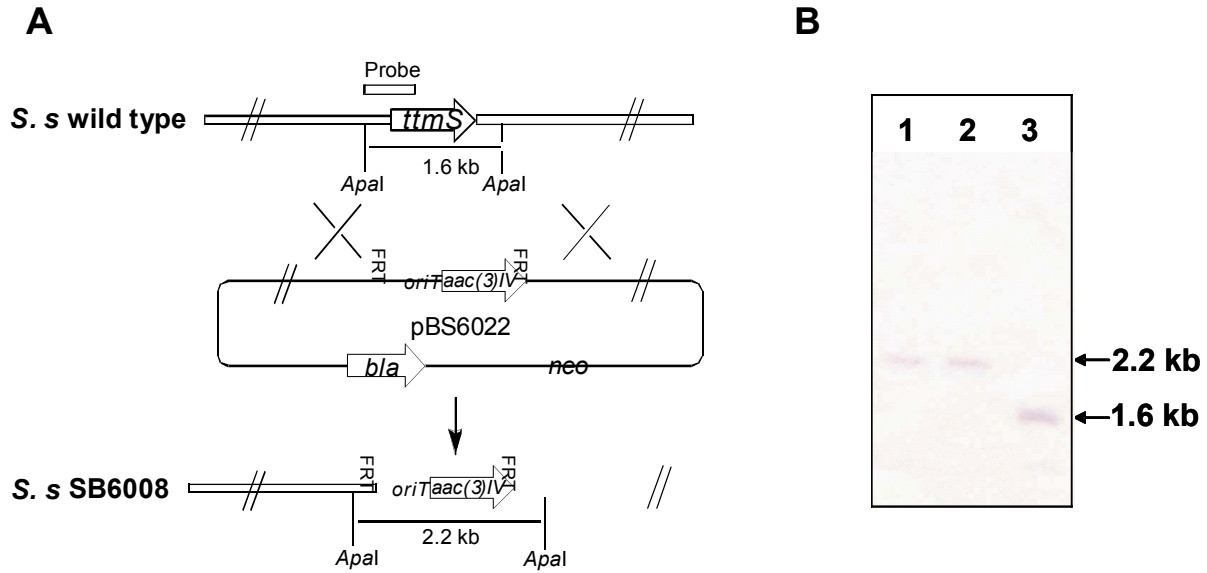


TABLE S3. Expression constructs for complementation to the *AtmK* and *AtmS* mutants

mutant strain	gene mutated	primers used to make the expression constructs ^a	construct	complemented strain
-	-	<i>ErmE</i> *p-FP: 5'-TAGAATTCGTGATGCTAGTCGCGGTTGATC- <i>ErmE</i> *p-RP:5'- TAGAATTCGTAATCATGCATTATCTCCTTCTCGCTGGATCCTA CCAACCGG-3'	pBS6027 ^b	-
SB6004	<i>AtmK</i>	<i>tmK</i> FP3: 5'-CCAATGCATATGGTGGAGACGTCGTCC-3' <i>tmK</i> RP3: 5'-GCTCTAGATCAGTCCGCCGGTACGC-3'	pBS6024 ^c	SB6009
SB6008	<i>AtmS</i>	<i>tmS</i> FP3: 5'-CCAATGCATGACGGACGCGCGAA-3' <i>tmS</i> RP3: 5'-GCTCTAGATCACGCGCCACCACCG-3'	pBS6026 ^c	SB6011

^a*Eco*RI, *Nsi*I and *Xba*I restriction sites are underlined.

^bThis construct was generated by inserting the *ErmE** promoter amplified from pWHM79 (2) at the *Eco*RI site of pBS8004 (3).

^cThese constructs were generated by inserting the PCR-amplified *tmK* or *tmS* fragment from pBS6004 into *Nsi*I and *Xba*I sites of pBS6027.

References

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- S2. Shen, B., and Hutchinson, C. R. (1996) *Proc. Natl. Acad. Sci. USA* **93**, 6600-6604
- S3. Tao, M., Wang, L., Wendt-Pienkowski, E., George, N. P., Zhang, G., Coughlin, J. M., and Shen, B. (2007) *Mol. Biosyst.* **3**, 60-74