

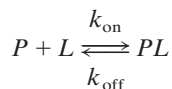
Supporting Information

Vallurupalli et al. 10.1073/pnas.0804221105

Structure Calculation Protocol

Experimental Restraints. In the discussion that follows, the crystal structure (1jo8) of the apo form of the Abp1p SH3 domain (1) was used as the representative ground state (“visible”) structure after (i) addition of the missing N-terminal residue, Ala 1, and (ii) addition of hydrogen atoms [using the hbuild feature of X-PLOR (2)]. This was followed by energy minimization with all heavy atoms fixed.

Structures of the invisible state corresponding to the Ark1p peptide bound form of the Abp1p SH3 domain have been calculated from ^{15}N , ^1HN , $^{13}\text{C}\alpha$, and ^{13}CO chemical shifts, along with anisotropic restraints including ^1HN - ^{15}N , $^1\text{H}\alpha$ - $^{13}\text{C}\alpha$, ^1HN - ^{13}CO RDCs, and ^{13}CO RCSAs. RDC and RCSA values were recorded on several samples, as described in *Materials and Methods*, with amounts of alignment media tailored to the size of the interaction measured. To employ all of the anisotropic restraints in a single structure calculation, the RDC and RCSA values must be scaled appropriately (3). The scaling factors were obtained by calculating the components of the alignment frame of the ground state structure individually for each type of anisotropic restraint (for example, ^1HN - ^{15}N RDCs) by fitting RDCs measured for the (visible) state P of the



exchanging system to the x-ray structure of the P state (apo form of the Abp1p SH3 domain). The D_a values (4) so obtained were subsequently scaled to a common set of bond lengths (^1HN - ^{15}N) and gyromagnetic ratios (^1HN , ^{15}N). Different amounts of alignment media were used to record (i) ^1HN - ^{15}N RDCs, (ii) $^1\text{H}\alpha$ - $^{13}\text{C}\alpha$ RDCs, and (iii) ^1HN - ^{13}CO RDCs and ^{13}CO RCSAs. D_a values of -5.5 , -3.4 , and -15.3 Hz were obtained for *i*, *ii*, and *iii*, respectively, and $^1\text{H}\alpha$ - $^{13}\text{C}\alpha$ and ^1HN - ^{13}CO RDCs were scaled by 5.5/3.4 and 5.5/15.3. Before scaling the ^{13}CO RCSAs by 5.5/15.3, we first offset values to correct for differences in peak positions between aligned and unaligned samples caused by splitting of the lock signal upon alignment. The offset was determined by minimizing the difference between experimental and calculated RCSA values of the ground state structure; this was done by fitting RCSAs of the apo protein (P) to those predicted on the basis of the x-ray structure of the unliganded form of the SH3 domain.

A flowchart summarizing the protocol used for structure determination is provided in Fig. S3, with additional details given in what follows. As described in *Results and Discussion*, regions for which $\Delta\sigma_{\text{RMS}} < 0.05$ were assumed to have the same conformation in both the ground (apo) and “excited” (ligand-bound) states. This was enforced by fixing the coordinates of all of the atoms in these regions (gray in Fig. 2C) during the torsion angle molecular dynamics (TAMD) protocol, as implemented in Xplor-NIH (5). Covalent geometry was maintained throughout all calculations by using the BOND, ANGLE, and IMPROPER (dihedral angle) energy terms. The RAMA torsion angle database term (6), which biases dihedral angles toward those expected from a database of high-resolution structures, was also used. Typically in NMR structure calculations, only a repulsive nonbonded interaction term is used, with the “attractive component” provided through experimentally derived distance restraints and a radius of gyration potential (7). To this point, it is not possible to measure such restraints for an “invisible” state,

and we have therefore used the full Lennard-Jones potential to describe nonbonded interactions. Experimental RDC and RCSA restraints were imposed by using a flat bottom harmonic potential with the width determined by experimental errors (5), with (ϕ, ψ) restraints enforced using the CDIH potential term of X-PLOR (Table S3). We have chosen not to use the alignment tensor values, D_a and R , estimated from the histogram of Fig. S1 as fixed input in structure calculations because we cannot be sure that there is a near-isotropic sampling of interaction vector orientations. Rather, D_a and R were allowed to float during the structure calculation protocol (5). It is clear, however, from inspection of Fig. S1 that $D_a < 0$ (see below), and this was enforced by imposing a torsion angle restraint on the four “atoms” that describe the orientation tensor.

Generating Structures with Random Conformations for Variable Regions 1–3 of Fig. 2C. TAMD is performed on the ground state structure (5) for 200 ps at 5,000 K, followed by a “production” run (250 ps of TAMD, 5,000 K), in which structures are stored every 25 ps. The 10 structures generated by this method have a wide range of conformations for the variable regions (Fig. 3A). Only BOND, ANGLE, IMPROPER, and NONBONDED (Lennard-Jones) potential energy terms were used at this stage.

Initial Generation of (ϕ, ψ) Torsion Angle Restraints from Chemical Shift Data. In total, 57 ^{15}N , 56 ^1HN , 50 $^{13}\text{C}\alpha$, and 49 ^{13}CO chemical shifts of the invisible state were obtained through analysis of relaxation dispersion profiles. (ϕ, ψ) torsion angle restraints were obtained for close to 60% (22/37) of the residues that compose variable regions 1–3 by using the program TALOS (8). Values of (ϕ, ψ) were assigned only if the prediction was deemed “good” by the program. The restraints were imposed with a width of $\max(\pm 2\sigma, \pm 30^\circ)$, where σ is the standard deviation of the TALOS best matches.

Conformation of Each Variable Region. To improve the rate of convergence of the structure calculation protocol, we individually optimized the conformations of each of regions 1–3 using TALOS-derived (ϕ, ψ) dihedral angle restraints, as well as RDC and RCSA restraints via a simulated annealing TAMD protocol. In this procedure all but one of the variable regions are deleted from each of the 10 high-temperature structures, calculated as described above. Each of the structures so obtained is subjected to 100 ps of TAMD at 5,000 K, with weak dihedral ($20 \text{ kcal}\cdot\text{mol}^{-1}\cdot\text{rad}^{-2}$), RDC ($0.002 \text{ kcal}\cdot\text{mol}^{-1}\cdot\text{Hz}^{-2}$), and RCSA ($5 \times 10^{-5} \text{ kcal}\cdot\text{mol}^{-1}\cdot\text{ppb}^{-2}$) restraints. The ANGLE, IMPROPER, and RAMA energy terms (but not the BOND and NONBONDED terms) were scaled at this high-temperature TAMD stage by factors of 0.4, 0.1, and 0.002. The temperature of the system was gradually reduced to 25 K over 798 ps whereas the force constants of the dihedral, RDC and RCSA terms were increased to $400 \text{ kcal}\cdot\text{mol}^{-1}\cdot\text{rad}^{-2}$, $0.4 \text{ kcal}\cdot\text{mol}^{-1}\cdot\text{Hz}^{-2}$, and $0.01 \text{ kcal}\cdot\text{mol}^{-1}\cdot\text{ppb}^{-2}$, respectively; the scaling factors for the ANGLE, IMPROPER and RAMA terms were also increased to 1. The structure at the end of the run was stored for further analysis, and the process was repeated 48 times for each starting structure, with different starting random velocities, to generate a total of 480 structures. The 10 lowest energy structures were selected. If a TALOS dihedral restraint was consistently violated, the restraint was removed and the whole processes repeated again. (This occurred for only one residue, Phe-31.)

Generation of new (ϕ , ψ) Torsion Angle Restraints. The 10 lowest energy structures obtained as described above for each of regions 1–3 were used to generate a new set of (ϕ , ψ) dihedral restraints for residues within the variable regions (color in Fig. 2C). The (ϕ , ψ) restraints were calculated as the mean of the dihedral angles obtained from the generated structures, imposed with a width of $\max(\pm 2\sigma, \pm 30^\circ)$, where σ is the standard deviation of (ϕ , ψ) in the 10 lowest energy structures; restraints were not imposed if 2σ was greater than 50° . Note that this process generates restraints for residues even if there were originally none (based on TALOS calculations from chemical shifts).

Determination of the Invisible, Ark1p-Bound Abp1p SH3 Domain Structure. A TAMM protocol was used to generate structures of the excited state starting from the 10 initial structures obtained

from MD runs at 5,000 K that produce random conformations for regions 1–3 (Fig. 3A, see above). The newly derived and more comprehensive set of (ϕ , ψ) restraints were used, along with the complete set of RDC and RCSA restraints, to produce structures. The TAMM protocol used was very similar to that used to optimize individual variable regions, except that the starting dihedral angle restraint force constant was $5 \text{ kcal}\cdot\text{mol}^{-1}\cdot\text{rad}^{-2}$; 96 structures were generated from each starting structure to produce a total of 960 structures.

All rmsd values quoted in the text were obtained by superimposing the “fixed” regions (gray in Fig. 2C) and subsequently calculating pairwise rmsd values by including only the variable regions (1–3) in the computations.

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3. Clore GM, Gronenborn AM, Bax A (1998) A robust method for determining the magnitude of the fully asymmetric alignment tensor of oriented macromolecules in the absence of structural information. *J Magn Reson* 113:216–221.
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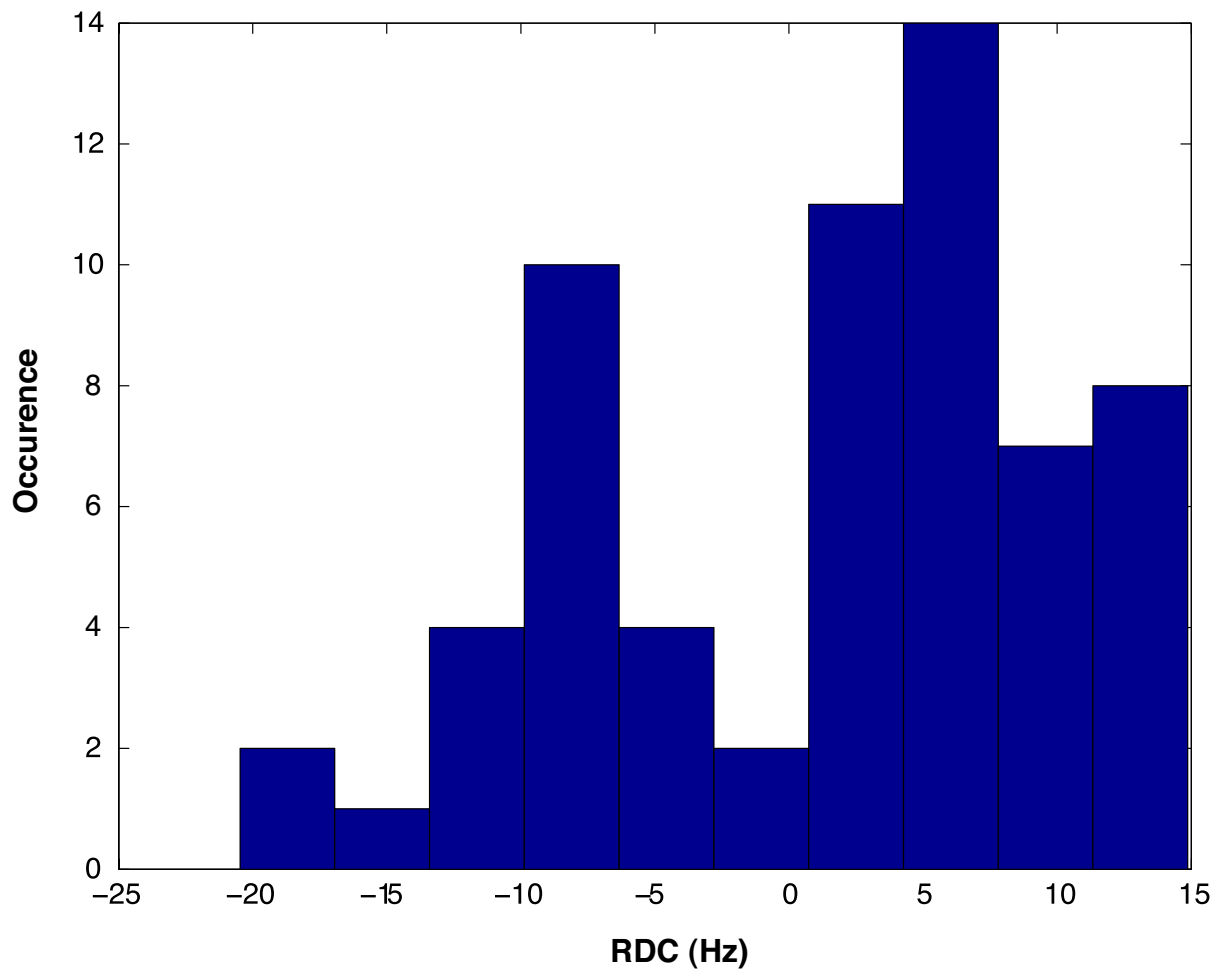


Fig. S1. Distribution of CPMG-derived RDCs for the invisible state. The histogram consists of 24 $^1\text{HN-}^{15}\text{N}$, 2 $^1\text{H}^{\alpha}\text{-}^{15}\text{N}^{\beta 1}$, 21 $^1\text{H}^{\alpha}\text{-}^{13}\text{C}^{\alpha}$, and 16 $^1\text{HN-}^{13}\text{CO}$ RDC values. The $^1\text{H}^{\alpha}\text{-}^{13}\text{C}^{\alpha}$ and $^1\text{HN-}^{13}\text{CO}$ RDCs have been scaled to the $^1\text{HN-}^{15}\text{N}$ values to account for differences in gyromagnetic ratios and distances and different concentrations of alignment media (Pf1 phage) in the samples used for measurements (as described in *Materials and Methods*). It is clear that the right-hand side corresponds to the well sampled edge of the histogram, $D_{\gamma\gamma} = -D_a(1 + 1.5R)$; therefore, $D_a < 0$.

Protocol for Invisible State Structure Determination Using CPMG Derived Chemical Shifts, RDCs and RCSAs.

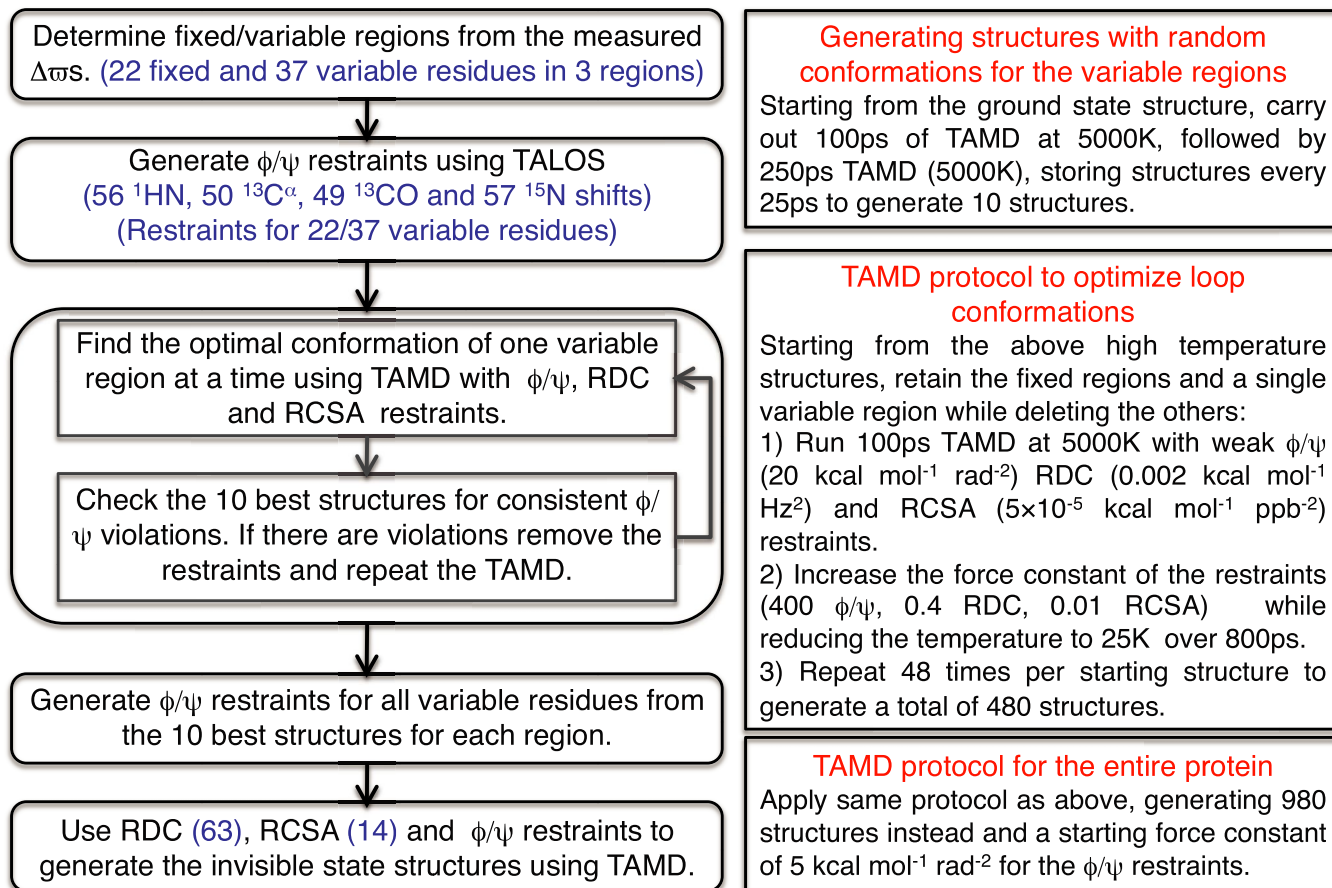


Fig. S3. Flow chart of the structure-determination protocol.

Table S1. Nitrogen chemical shifts (ppm) of the SH3 domain component of the Abp1p SH3–Ark1p peptide complex derived from CPMG relaxation dispersion experiments

| Residue | ω_{bound} , ppm | | $\Delta\omega = \omega_{\text{free}} - \omega_{\text{bound}}$, ppm | |
|---------|-------------------------------|--------|---|---------------|
| | CPMG | Direct | CPMG | Direct |
| A1 | 126.83* | 126.90 | | |
| P2 | | | | |
| W3 | 116.18 [†] | 116.29 | 0.121 ± 0.044 | -0.113 |
| A4 | 120.85 [†] | 120.71 | 0.152 ± 0.035 | 0.142 |
| T5 | 113.32 ± 0.01 | 113.34 | -0.393 ± 0.015 | -0.407 |
| A6 | 127.52 ± 0.03 | 127.49 | -0.155 ± 0.034 | -0.123 |
| E7 | 125.35* | 125.31 | | |
| Y8 | 113.09 ± 0.01 | 113.08 | 0.850 ± 0.012 | 0.858 |
| D9 | 118.66 ± 0.02 | 118.65 | -0.351 ± 0.017 | -0.338 |
| Y10 | 121.77 ± 0.01 | 121.75 | -0.595 ± 0.012 | -0.574 |
| D11 | 125.75 ± 0.02 | 125.77 | 0.365 ± 0.017 | 0.345 |
| A12 | 123.70 ± 0.02 | 123.70 | 0.278 ± 0.020 | 0.275 |
| A13 | 125.74 ± 0.02 | 125.73 | 0.305 ± 0.019 | 0.316 |
| E14 | 113.06 ± 0.01 | 113.06 | 1.075 ± 0.013 | 1.074 |
| D15 | 117.87 ± 0.02 | 117.75 | 1.964 ± 0.023 | 2.080 |
| N16 | 113.58 ± 0.01 | 113.58 | 0.671 ± 0.012 | 0.676 |
| E17 | 117.77 ± 0.01 | 117.75 | 1.039 ± 0.013 | 1.062 |
| L18 | 122.35 ± 0.01 | 122.35 | 0.517 ± 0.013 | 0.510 |
| T19 | 114.34 ± 0.02 | 114.34 | 0.366 ± 0.016 | 0.365 |
| F20 | 120.59 [†] | 120.51 | 0.114 ± 0.046 | 0.083 |
| V21 | 116.79 [†] | 116.87 | 0.122 ± 0.043 | -0.080 |
| E22 | 120.53* | 120.49 | | |
| N23 | 117.26 [†] | 117.09 | 0.162 ± 0.033 | 0.164 |
| D24 | 122.59 [†] | 122.68 | 0.108 ± 0.049 | -0.093 |
| K25 | 120.27 ± 0.03 | 120.24 | -0.158 ± 0.034 | -0.131 |
| I26 | 130.67 ± 0.03 | 130.69 | 0.159 ± 0.034 | 0.141 |
| I27 | 118.98* | 118.98 | | |
| N28 | 115.82 [†] | 115.64 | 0.177 ± 0.030 | 0.178 |
| I29 | 117.13 [†] | 116.98 | 0.156 ± 0.034 | 0.154 |
| E30 | 124.76 ± 0.01 | 124.78 | 0.452 ± 0.014 | 0.433 |
| F31 | 128.36 ± 0.02 | 128.31 | -2.008 ± 0.024 | -1.956 |
| V32 | 117.38 ± 0.06 | 117.12 | 7.152 ± 0.063 | 7.418 |
| D33 | 120.43 ± 0.02 | 120.46 | -1.781 ± 0.020 | -1.806 |
| D34 | 118.56 ± 0.04 | 118.48 | -3.790 ± 0.046 | -3.703 |
| D35 | 116.47 ± 0.01 | 116.48 | 0.943 ± 0.012 | 0.937 |
| W36 | 122.49 ± 0.01 | 122.52 | 0.544 ± 0.013 | 0.513 |
| W37 | 119.94 ± 0.02 | 119.92 | 1.683 ± 0.019 | 1.702 |
| L38 | 121.00 ± 0.03 | 121.01 | 0.204 ± 0.027 | 0.200 |
| G39 | 109.39 ± 0.01 | 109.46 | -0.572 ± 0.013 | -0.644 |
| E40 | 119.76* | 119.77 | | |
| L41 | 125.87 ± 0.04 | 125.89 | 0.133 ± 0.041 | 0.113 |
| E42 | 129.28* | 129.20 | | |
| K43 | 113.61* | 113.60 | | |
| D44 | 114.24* | 114.28 | | |
| G45 | 109.80* | 109.81 | | |
| S46 | 118.28* | 118.26 | | |
| K47 | 119.48* | 119.55 | | |
| G48 | 108.57 ± 0.03 | 108.57 | 0.161 ± 0.034 | 0.165 |
| L49 | 119.62 ± 0.01 | 119.51 | 0.917 ± 0.012 | 1.021 |
| F50 | 115.17 ± 0.04 | 115.18 | 0.142 ± 0.037 | 0.129 |
| P51 | | | | |
| S52 | 120.57 ± 0.01 | 120.58 | 1.242 ± 0.015 | 1.230 |
| N53 | 116.14 ± 0.01 | 116.10 | -0.869 ± 0.012 | -0.830 |
| Y54 | 119.16* | 119.13 | | |
| V55 | 109.26 ± 0.02 | 109.25 | -0.252 ± 0.022 | -0.243 |
| S56 | 113.84 [†] | 113.92 | 0.099 ± 0.054 | -0.081 |
| L57 | 129.01* | 129.10 | | |
| G58 | 111.54 [†] | 111.66 | 0.142 ± 0.038 | -0.122 |
| N59 | 123.28* | 123.29 | | |

Shifts are compared with those obtained directly when the complex is the major (visible) state in solution. Residues in bold indicate that no sign information about $\Delta\omega$ could be obtained.

*A two-site chemical exchange model did not generate statistically significant improvements in fits over a model of no exchange (constant $R_2(\nu_{\text{CPMG}})$) at the 98% confidence level. Values of the excited state chemical shifts were assumed the same as the ground state.

[†]The sign of $\Delta\omega$ could not be obtained accurately from changes in peak positions among the following four experiments that are used to quantify the sign [Skrynnikov NR, Dahlquist FW, Kay LE (2002) Reconstructing NMR spectra of “invisible” excited protein states using HSQC and HMQC experiments. *J Am Chem Soc* 124:12352–12360]: (HSQC at 11.7T, HSQC at 18.8T, HMQC at 11.7T and HMQC at 18.8T) i.e., $\max(|\omega_{\text{HSQC},500} - \omega_{\text{HMQC},500}|, |\omega_{\text{HSQC},800} - \omega_{\text{HMQC},800}|, |\omega_{\text{HSQC},500} - \omega_{\text{HSQC},800}|) < 3.0$ ppb. However, the absolute value of $\Delta\omega$ is so small that the chemical shift of the bound state can be assumed to be identical to that of the free state.

Table S2. Amide proton chemical shifts (ppm) of the SH3 domain component of the Abp1p SH3–Ark1p peptide complex derived from CPMG relaxation dispersion experiments

| Residue | ω_{bound} (ppm) | | $\Delta\omega = \omega_{\text{free}} - \omega_{\text{bound}}$ (ppm) | |
|---------|-------------------------------|--------|---|--------------|
| | CPMG | Direct | CPMG | Direct |
| A1 | 8.336* | 8.343 | | |
| P2 | | | | |
| W3 | 7.824* | 7.823 | | |
| A4 | 10.086* | 10.084 | | |
| T5 | 8.306 ± 0.004 | 8.302 | −0.046 ± 0.004 | −0.042 |
| A6 | 8.942 ± 0.006 | 8.946 | −0.033 ± 0.006 | −0.037 |
| E7 | 9.410* | 9.400 | | |
| Y8 | 7.449 ± 0.004 | 7.447 | −0.049 ± 0.004 | −0.047 |
| D9 | 8.598 ± 0.004 | 8.590 | −0.155 ± 0.004 | −0.147 |
| Y10 | 8.220 ± 0.005 | 8.223 | 0.041 ± 0.005 | 0.038 |
| D11 | 7.090 ± 0.003 | 7.090 | 0.106 ± 0.003 | 0.106 |
| A12 | 7.709 ± 0.003 | 7.701 | −0.095 ± 0.003 | −0.087 |
| A13 | 8.854 ± 0.005 | 8.857 | 0.227 ± 0.005 | 0.224 |
| E14 | 7.085 ± 0.004 | 7.086 | −0.048 ± 0.004 | −0.049 |
| D15 | 8.671 ± 0.008 | 8.667 | 0.319 ± 0.008 | 0.323 |
| N16 | 7.324 ± 0.013 | 7.351 | 0.606 ± 0.013 | 0.579 |
| E17 | 7.561* | 7.555 | | |
| L18 | 8.756 ± 0.003 | 8.758 | 0.099 ± 0.003 | 0.097 |
| T19 | 7.387 ± 0.004 | 7.385 | −0.048 ± 0.004 | −0.046 |
| F20 | 9.013* | 9.023 | | |
| V21 | 9.253* | 9.279 | | |
| E22 | 8.786† | 8.798 | 0.019 ± 0.009 | 0.012 |
| N23 | 8.905* | 8.900 | | |
| D24 | 8.645* | 8.648 | | |
| K25 | 8.172 ± 0.005 | 8.170 | −0.039 ± 0.005 | −0.037 |
| I26 | 9.321* | 9.322 | | |
| I27 | 9.827* | 9.820 | | |
| N28 | 9.059* | 9.065 | | |
| I29 | 7.898* | 7.899 | | |
| E30 | 9.236 ± 0.008 | 9.243 | −0.024 ± 0.008 | −0.031 |
| F31 | 8.722 ± 0.006 | 8.711 | −0.239 ± 0.006 | −0.228 |
| V32 | 7.730 ± 0.004 | 7.717 | −0.064 ± 0.004 | −0.051 |
| D33 | 8.315 ± 0.009 | 8.342 | 0.354 ± 0.009 | 0.327 |
| D34 | 8.169 ± 0.003 | 8.167 | 0.090 ± 0.003 | 0.092 |
| D35 | 8.720 ± 0.003 | 8.717 | 0.072 ± 0.003 | 0.075 |
| W36 | 7.827 ± 0.008 | 7.848 | 0.348 ± 0.008 | 0.327 |
| W37 | 7.876 ± 0.012 | 7.907 | 0.463 ± 0.012 | 0.432 |
| L38 | 8.358 ± 0.009 | 8.399 | 0.360 ± 0.009 | 0.319 |
| G39 | 8.833 ± 0.003 | 8.841 | −0.107 ± 0.003 | −0.115 |
| E40 | 8.587* | 8.580 | | |
| L41 | 9.521* | 9.500 | | |
| E42 | 8.190* | 8.188 | | |
| K43 | 9.240* | 9.246 | | |
| D44 | 6.822* | 6.820 | | |
| G45 | 8.411* | 8.398 | | |
| S46 | 8.686* | 8.696 | | |
| K47 | 8.374* | 8.371 | | |
| G48 | 8.751 ± 0.005 | 8.746 | −0.037 ± 0.005 | −0.032 |
| L49 | 9.106 ± 0.008 | 9.099 | −0.320 ± 0.008 | −0.313 |
| F50 | 9.234 ± 0.004 | 9.230 | 0.056 ± 0.004 | 0.060 |
| P51 | | | | |
| S52 | 7.870 ± 0.005 | 7.865 | −0.231 ± 0.005 | −0.226 |
| N53 | | 7.967 | | |
| Y54 | 7.792 ± 0.004 | 7.794 | 0.061 ± 0.004 | 0.059 |
| V55 | 7.419 ± 0.008 | 7.413 | −0.022 ± 0.008 | −0.016 |
| S56 | 8.604† | 8.615 | 0.014 ± 0.012 | 0.011 |
| L57 | 9.158† | 9.187 | 0.026 ± 0.007 | 0.029 |
| G58 | 8.487* | 8.495 | | |
| N59 | 7.478* | 7.462 | | |

Shifts are compared with those obtained directly when the complex is the major (visible) state in solution. Signs are measured relative to the ^{15}N shift in a series of double quantum and zero quantum experiments [Korzhnev DM, Neudecker P, Mittermaier A, Orekhov VY, Kay LE (2005) Multiple-site exchange in proteins studied with a suite of six NMR relaxation dispersion experiments: An application to the folding of a Fyn SH3 domain mutant. *J Am Chem Soc* 127:15602–15611].

*A two-site chemical exchange model did not generate statistically significant improvements in fits over a model of no exchange [constant $R_2(\nu_{\text{CPMG}})$] at the 98% confidence level. Chemical shifts of the ground and excited states were assumed to be the same.

†The sign of $\Delta\omega$ could not be obtained accurately; however, the absolute value of $\Delta\omega$ is so small that the chemical shift of the bound state can be assumed to be identical to that of the free state.

Table S3. Alpha-carbon ($^{13}\text{C}^\alpha$) chemical shifts (ppm) of the SH3 component of the Abp1p SH3–Ark1p peptide complex derived from CPMG relaxation dispersion experiments

| Residue | ω_{bound} , ppm | | $\Delta\omega = \omega_{\text{free}} - \omega_{\text{bound}}$, ppm | |
|---------|-------------------------------|--------|---|-------------|
| | CPMG | Direct | CPMG | Direct |
| A1 | 50.58 ± 0.02 | 50.69 | 0.08 ± 0.02 | −0.04 |
| P2 | 63.59† | 63.59 | 0.14 ± 0.01 | 0.00 |
| W3 | 54.22† | 54.19 | 0.11 ± 0.01 | 0.03 |
| A4 | 50.89† | 50.80 | 0.11 ± 0.01 | 0.09 |
| T5 | 60.77 ± 0.01 | 60.74 | −0.14 ± 0.01 | −0.12 |
| A6 | 53.18† | 53.14 | 0.07 ± 0.01 | 0.04 |
| E7 | 56.36 ± 0.01 | 56.42 | 0.18 ± 0.01 | 0.11 |
| Y8 | 54.89 ± 0.01 | 54.84 | −0.14 ± 0.01 | −0.09 |
| D9 | 54.41 ± 0.02 | 54.58 | 0.19 ± 0.02 | 0.02 |
| Y10 | 58.85 ± 0.01 | 58.85 | 0.15 ± 0.01 | 0.15 |
| D11 | 52.24† | 52.18 | 0.13 ± 0.01 | 0.06 |
| A12 | 53.54 ± 0.01 | 53.52 | −0.11 ± 0.01 | −0.09 |
| A13 | 52.10† | 52.00 | 0.13 ± 0.01 | 0.10 |
| E14 | 54.35 ± 0.01 | 54.38 | −0.70 ± 0.01 | −0.73 |
| D15 | 56.82 ± 0.01 | 56.80 | −0.22 ± 0.01 | −0.21 |
| N16 | | 51.50 | 0.83 ± 0.01 | 0.82 |
| E17 | 54.89 ± 0.01 | 54.95 | 0.32 ± 0.01 | 0.26 |
| L18 | | | | |
| T19 | 61.73* | 61.74 | | |
| F20 | 55.53 ± 0.01 | 55.51 | 0.10 ± 0.01 | 0.11 |
| V21 | 59.08 ± 0.02 | 59.18 | 0.10 ± 0.02 | 0.00 |
| E22 | 58.51† | 58.53 | 0.10 ± 0.01 | 0.02 |
| N23 | 56.16 ± 0.01 | 56.28 | <i>0.11 ± 0.01</i> | −0.01 |
| D24 | 56.05 ± 0.01 | 56.15 | <i>0.09 ± 0.01</i> | −0.01 |
| K25 | 55.98† | 55.97 | 0.07 ± 0.02 | 0.02 |
| I26 | | | | |
| I27 | | | | |
| N28 | 53.87† | 53.84 | 0.09 ± 0.01 | 0.02 |
| I29 | | | | |
| E30 | 54.48 ± 0.01 | 54.42 | −0.28 ± 0.01 | −0.22 |
| F31 | 54.74 ± 0.02 | 54.65 | −0.13 ± 0.02 | −0.05 |
| V32 | 64.30 ± 0.10 | 64.08 | 1.96 ± 0.10 | 2.18 |
| D33 | 53.36 ± 0.01 | 53.43 | 0.14 ± 0.01 | 0.08 |
| D34 | 57.80 ± 0.01 | 57.78 | −0.56 ± 0.01 | −0.54 |
| D35 | 55.12 ± 0.01 | 55.31 | <i>0.13 ± 0.01</i> | −0.06 |
| W36 | 56.95 ± 0.01 | 56.92 | −0.39 ± 0.01 | −0.36 |
| W37 | 51.56 ± 0.01 | 51.59 | 0.40 ± 0.01 | 0.38 |
| L38 | | | | |
| G39 | 45.86 | 45.88 | | |
| E40 | 52.62† | 52.64 | 0.15 ± 0.01 | −0.02 |
| L41 | | | | |
| E42 | 59.83 ± 0.01 | 59.67 | −0.14 ± 0.01 | 0.03 |
| K43 | 58.60† | 58.57 | 0.10 ± 0.01 | 0.02 |
| D44 | 53.56† | 53.40 | 0.09 ± 0.02 | 0.15 |
| G45 | 46.16 | 46.16 | | |
| S46 | 59.80† | 59.82 | 0.09 ± 0.01 | −0.02 |
| K47 | 54.96 ± 0.01 | 55.01 | 0.10 ± 0.01 | 0.04 |
| G48 | 45.61 | 45.55 | | |
| L49 | | | | |
| F50 | 55.13 ± 0.01 | 55.11 | 0.31 ± 0.01 | 0.32 |
| P51 | 60.74 ± 0.01 | 60.68 | 0.29 ± 0.01 | 0.35 |
| S52 | 60.47 ± 0.01 | 60.45 | −0.43 ± 0.01 | −0.41 |
| N53 | 53.22 ± 0.01 | 53.23 | 0.50 ± 0.01 | 0.48 |
| Y54 | 59.10 ± 0.01 | 59.09 | −0.12 ± 0.01 | −0.11 |
| V55 | 58.51 ± 0.02 | 58.43 | −0.16 ± 0.02 | −0.09 |
| S56 | 56.23† | 56.28 | 0.10 ± 0.01 | −0.05 |
| L57 | | | | |
| G58 | 43.56 | 43.56 | | |
| N59 | 54.62* | 54.62 | | |

Shifts are compared with those obtained directly when the complex is the major (visible) state in solution. Chemical shifts for Leu and Ile are not available (blank rows) due to the labeling scheme employed, which leads to the presence of $^{13}\text{C}^\alpha$ - $^{13}\text{C}^\beta$ scalar couplings [Lundstrom P, et al. (2007) Fractional ^{13}C enrichment of isolated carbons using [1- ^{13}C]- or [2- ^{13}C]-glucose facilitates the accurate measurement of dynamics at backbone C α and side-chain methyl positions in proteins. *J Biomol NMR* 38:199–212]. Although such couplings are also present in the case of Val, they can be “suppressed” during the CPMG element by using selective pulses.

*A two-site chemical exchange model did not generate statistically significant improvements in fits over a model of no exchange (constant $R_2(\nu_{\text{CPMG}})$) at the 98% confidence level. Chemical shifts of the ground and excited state were assumed to be identical.

†The sign of $\Delta\omega$ could not be obtained accurately, ($\max(|\omega_{\text{HSQC},500} - \omega_{\text{HMQC},500}|, |\omega_{\text{HSQC},800} - \omega_{\text{HMQC},800}|, |\omega_{\text{HSQC},500} - \omega_{\text{HSQC},800}|) < 3.0$ ppb; residues marked in bold); however, the absolute value of $\Delta\omega$ is so small that the chemical shift of the bound state can be assumed to be identical to that of the free state. Residues for which the determined sign does not agree with expectations based on measurements recorded directly on the bound state are marked in bold italics.

Table S4. Carbonyl (^{13}C O) chemical shifts (ppm) of the SH3 domain component of the Abp1p SH3–Ark1p peptide complex derived from CPMG relaxation dispersion experiments and compared with those obtained directly when the complex is the major (visible) state in solution

| Residue | ω_{bound} | | $\Delta\omega = \omega_{\text{free}} - \omega_{\text{bound}}$ | |
|---------|-------------------------|--------|---|--------------|
| | CPMG | Direct | CPMG | Direct |
| A1 | | | | |
| P2 | 174.02* | 173.96 | | |
| W3 | 174.29* | 174.29 | | |
| A4 | 175.11* | 175.09 | | |
| T5 | 174.88* | 174.82 | | |
| A6 | 178.43 ± 0.02 | 178.44 | 0.101 ± 0.019 | 0.090 |
| E7 | 174.07 ± 0.02 | 174.01 | 0.132 ± 0.019 | 0.196 |
| Y8 | 173.19 ± 0.01 | 173.19 | 0.255 ± 0.010 | 0.258 |
| D9 | 175.57 ± 0.01 | 175.58 | 0.190 ± 0.012 | 0.175 |
| Y10 | 172.65 ± 0.03 | 172.65 | 0.056 ± 0.034 | 0.056 |
| D11 | 173.25* | 173.26 | | |
| A12 | 178.46 ± 0.04 | 178.44 | −0.048 ± 0.039 | −0.028 |
| A13 | 177.43 ± 0.01 | 177.44 | −0.320 ± 0.010 | −0.324 |
| E14 | 177.41 ± 0.02 | 177.40 | −0.108 ± 0.023 | −0.099 |
| D15 | 175.63 ± 0.01 | 175.64 | 0.424 ± 0.010 | 0.414 |
| N16 | 175.51* | 175.52 | | |
| E17 | 176.57* | 176.61 | | |
| L18 | | | | |
| T19 | 174.45* | 174.38 | | |
| F20 | 174.89* | 174.90 | | |
| V21 | 174.97* | 174.97 | | |
| E22 | 176.64* | 176.69 | | |
| N23 | 175.12* | 175.13 | | |
| D24 | 175.50* | 175.47 | | |
| K25 | 175.86 ± 0.03 | 175.86 | −0.059 ± 0.032 | −0.056 |
| I26 | 174.90* | 174.90 | | |
| I27 | 174.92* | 174.91 | | |
| N28 | 174.95* | 174.93 | | |
| I29 | 176.11 ± 0.02 | 176.11 | 0.086 ± 0.023 | 0.086 |
| E30 | | 176.50 | 0.865 ± 0.024 | 0.905 |
| F31 | 176.67 ± 0.01 | 176.67 | −0.464 ± 0.011 | −0.464 |
| V32 | 174.34 ± 0.04 | 174.34 | 1.756 ± 0.040 | 1.757 |
| D33 | 173.69 ± 0.02 | 173.66 | 0.838 ± 0.021 | 0.871 |
| D34 | 177.49 ± 0.01 | 177.50 | 0.231 ± 0.011 | 0.220 |
| D35 | | 174.99 | 0.778 ± 0.019 | 0.751 |
| W36 | 172.55 ± 0.01 | 172.54 | 0.411 ± 0.010 | 0.421 |
| W37 | 172.54 ± 0.01 | 172.54 | −0.382 ± 0.010 | −0.374 |
| L38 | | | | |
| G39 | 177.66† | 177.67 | 0.057 ± 0.033 | −0.011 |
| E40 | 175.66† | 175.61 | 0.081 ± 0.024 | 0.056 |
| L41 | | | | |
| E42 | 178.54 † | 178.53 | 0.053 ± 0.035 | 0.008 |
| K43 | 176.76* | 176.75 | | |
| D44 | 177.22* | 177.22 | | |
| G45 | 174.47* | 174.47 | | |
| S46 | 172.62 ± 0.08 | 172.61 | −0.024 ± 0.077 | −0.017 |
| K47 | 176.87* | 176.91 | | |
| G48 | 171.50 ± 0.02 | 171.50 | −0.115 ± 0.017 | −0.112 |
| L49 | | | | |
| F50 | | | | |
| P51 | 178.44 ± 0.01 | 178.47 | 0.324 ± 0.011 | 0.294 |
| S52 | 174.52 ± 0.01 | 174.50 | 0.607 ± 0.013 | 0.619 |
| N53 | 175.20 ± 0.01 | 175.18 | 0.481 ± 0.011 | 0.493 |
| Y54 | 175.49 ± 0.01 | 175.50 | −0.282 ± 0.010 | −0.292 |
| V55 | 173.78* | 173.81 | | |
| S56 | 175.46* | 175.44 | | |
| L57 | | | | |
| G58 | 172.43† | 172.43 | 0.035 ± 0.054 | 0.003 |
| N59 | | | | |

^{13}C O shift values for Leu and His could not be obtained (blank rows) due to the labeling scheme employed [Hansen DF, Vallurupalli P, Lundstrom P, Neudecker P, Kay LE (2008) Probing chemical shifts of invisible states of proteins with relaxation dispersion nmr spectroscopy: How well can we do? *J Am Chem Soc* 2667–2675]. Shifts for residues preceding Pro are not available (blank rows) because of the HNC0 type of experiment used [Vallurupalli P, Hansen DF, Kay LE (2008) Probing structure in invisible protein states with anisotropic NMR chemical shifts. *J Am Chem Soc* 130:2734–2735.].

*A two-site chemical exchange model did not generate statistically significant improvements in fits over a model of no exchange (constant $R_2(\nu_{\text{CPMG}})$) at the 98% confidence level. Chemical shifts in the ground and excited states were assumed to be identical.

†The sign of $\Delta\omega$ could not be obtained accurately; however, the absolute value of $\Delta\omega$ is so small that the chemical shift of the bound state can be assumed to be identical to that of the free state.

Table S5. Residual dipolar couplings of the SH3 domain component of the Abp1p SH3–Ark1p peptide complex derived from CPMG relaxation dispersion experiments and compared with those obtained directly [IPAP (1, 2)] when the complex is the major (visible) state in solution

| Residue | $^1\text{D}(^{15}\text{N}-^1\text{HN})$, Hz | | $^1\text{D}(^{13}\text{C} - ^1\text{H}^\alpha)$, Hz | | $^2\text{D}(^{13}\text{CO} - ^1\text{HN})$, Hz | |
|-------------------|--|-------------|--|-------------|---|-------------|
| | CPMG | Direct | CPMG | Direct | CPMG | Direct |
| A1 | | 5.8 ± 0.1 | | −13.0 ± 0.1 | | |
| P2 | | | | −10.5 ± 0.2 | | 3.5 ± 0.5 |
| W3 | | | | −13.1 ± 0.1 | | −8.7 ± 0.3 |
| A4 | | 14.4 ± 0.1 | | −17.0 ± 0.1 | | 4.1 ± 1.7 |
| T5 | 10.5 ± 1.5 | 11.8 ± 0.6 | −9.8 ± 4.3 | −9.5 ± 0.2 | | −10.0 ± 0.5 |
| A6 | | 2.9 ± 0.6 | | −11.0 ± 0.1 | −6.2 ± 1.9 | −5.3 ± 0.3 |
| E7 | | 6.7 ± 0.7 | | −5.3 ± 0.4 | −2.3 ± 3.8 | 2.7 ± 0.5 |
| Y8 | −10.8 ± 1.5 | −12.7 ± 0.1 | 10.2 ± 4.7 | 7.1 ± 0.2 | 8.0 ± 1.8 | 7.0 ± 0.3 |
| D9 | −17.0 ± 1.7 | −16.1 ± 0.5 | | | 8.8 ± 2.6 | 8.6 ± 0.8 |
| Y10 | −8.1 ± 1.5 | −9.7 ± 0.6 | 14.2 ± 3.5 | 20.9 ± 0.3 | | 4.0 ± 0.3 |
| D11 | −7.9 ± 1.6 | −9.5 ± 0.1 | | 7.0 ± 0.2 | | 7.4 ± 0.3 |
| A12 | −0.3 ± 2.1 | −1.5 ± 0.1 | −15.8 ± 4.9 | −19.3 ± 0.1 | −11.3 ± 2.4 | −9.1 ± 0.4 |
| A13 | 9.7 ± 1.8 | 11.6 ± 0.5 | | −2.4 ± 0.1 | 8.0 ± 2.0 | 5.3 ± 0.3 |
| E14 | 4.5 ± 1.5 | 3.8 ± 0.1 | −4.8 ± 5.4 | −10.7 ± 0.2 | | −3.0 ± 1.1 |
| D15 | 12.1 ± 2.7 | 3.2 ± 0.2 | −4.8 ± 2.8 | −5.8 ± 0.1 | −13.1 ± 3.4 | −7.5 ± 1.0 |
| N16 | −4.9 ± 1.5 | −6.7 ± 0.6 | | 7.0 ± 0.2 | | 6.2 ± 0.4 |
| E17 | 7.0 ± 1.5 | 5.9 ± 0.2 | −15.0 ± 4.2 | −12.4 ± 0.5 | | |
| L18 | 13.7 ± 1.5 | 14.8 ± 0.1 | | 14.8 ± 0.5 | | |
| T19 | −10.3 ± 1.5 | −12.5 ± 0.1 | | 8.4 ± 0.3 | | 9.1 ± 0.5 |
| F20 | | −15.6 ± 0.1 | −11.2 ± 4.3 | −6.8 ± 0.1 | −1.7 ± 2.6 | −1.2 ± 0.3 |
| V21 | | −18.3 ± 0.1 | | 14.4 ± 0.3 | | 14.1 ± 0.3 |
| E22 | | −13.3 ± 0.2 | | −14.6 ± 0.2 | −11.1 ± 3.7 | −7.9 ± 1.1 |
| N23 | | 10.8 ± 0.6 | | −20.7 ± 0.1 | | −3.3 ± 0.9 |
| D24 | | −11.6 ± 0.1 | | −4.5 ± 0.2 | 5.3 ± 3.1 | 3.0 ± 1.0 |
| K25 | | 9.5 ± 0.6 | | −10.9 ± 0.3 | | −10.5 ± 0.4 |
| I26 | 5.7 ± 4.0 | 10.2 ± 0.2 | | −19.1 ± 0.5 | | 0.4 ± 0.5 |
| I27 | | 16.5 ± 0.6 | | −17.4 ± 0.5 | | 0.4 ± 0.5 |
| N28 | | 6.3 ± 1.1 | | −1.1 ± 0.2 | | −1.6 ± 1.0 |
| I29 | | 8.6 ± 0.1 | | | | −2.6 ± 0.6 |
| E30 | 3.0 ± 1.5 | 2.2 ± 0.6 | −5.4 ± 2.9 | −4.9 ± 0.3 | | |
| F31 | −8.2 ± 2.3 | −9.1 ± 0.6 | | 0.0 ± 0.0 | −6.7 ± 2.5 | −10.3 ± 0.3 |
| V32 | | | | 18.1 ± 0.3 | | 6.8 ± 0.3 |
| D33 | 2.1 ± 1.7 | 6.1 ± 0.6 | 0.1 ± 4.1 | −2.4 ± 0.2 | | −5.2 ± 1.1 |
| D34 | 6.7 ± 3.8 | 4.4 ± 0.6 | 11.2 ± 3.7 | 4.6 ± 0.5 | −9.1 ± 2.6 | −5.6 ± 1.1 |
| D35 | 5.5 ± 1.5 | 5.1 ± 0.1 | | −4.1 ± 0.2 | | 9.8 ± 1.3 |
| W36 | 4.3 ± 1.6 | 3.6 ± 0.6 | 5.0 ± 2.6 | 2.1 ± 0.3 | −3.6 ± 3.4 | −4.3 ± 0.3 |
| W37 | −5.0 ± 2.7 | −3.1 ± 0.7 | −5.2 ± 2.8 | 1.1 ± 0.2 | 14.9 ± 1.7 | 10.6 ± 0.3 |
| L38 | | 5.9 ± 0.6 | | | | |
| G39 | | 5.1 ± 0.1 | | | | 0.3 ± 0.3 |
| E40 | | 3.9 ± 2.3 | | −11.4 ± 0.4 | | −8.7 ± 0.4 |
| L41 | | 12.3 ± 0.5 | | | | |
| E42 | | 2.0 ± 0.1 | | | | 12.4 ± 0.3 |
| K43 | | −6.8 ± 0.6 | | 1.0 ± 0.1 | | −6.2 ± 1.3 |
| D44 | | 6.0 ± 0.2 | | −8.1 ± 0.2 | | |
| G45 | | 8.7 ± 0.1 | | | | 12.8 ± 0.3 |
| S46 | | −1.2 ± 0.1 | | −1.0 ± 0.1 | −2.9 ± 2.7 | −2.1 ± 0.3 |
| K47 | | 12.7 ± 0.3 | | −12.1 ± 0.2 | −9.6 ± 2.3 | −9.3 ± 0.9 |
| G48 | | 10.9 ± 0.7 | | | −0.9 ± 2.0 | −1.4 ± 0.3 |
| L49 | 8.9 ± 1.5 | 14.1 ± 1.1 | | | | |
| F50 | | 4.5 ± 0.6 | −3.5 ± 3.0 | −1.2 ± 0.3 | | |
| P51 | | | | −3.6 ± 0.6 | | −6.7 ± 1.1 |
| S52 | 5.2 ± 1.5 | 5.4 ± 0.6 | | 3.9 ± 0.2 | 6.7 ± 3.1 | 1.9 ± 0.5 |
| N53 | 6.0 ± 1.5 | 6.9 ± 0.1 | −15.3 ± 3.4 | −13.1 ± 0.2 | 6.0 ± 2.1 | 3.3 ± 0.4 |
| Y54 | | −4.3 ± 0.1 | 25.4 ± 4.4 | 24.4 ± 0.3 | −5.5 ± 1.7 | −3.1 ± 0.3 |
| V55 | | 8.8 ± 0.6 | | −8.6 ± 0.4 | −4.5 ± 2.2 | −3.5 ± 0.3 |
| S56 | | 10.5 ± 0.2 | | −11.9 ± 0.1 | | −8.8 ± 0.3 |
| L57 | | 2.5 ± 0.1 | | 17.3 ± 0.5 | | |
| G58 | | 5.2 ± 0.1 | | 9.6 ± 76.6 | | −0.5 ± 0.3 |
| N59 | | 1.2 ± 0.5 | | 0.8 ± 0.1 | | |
| W36 ^{SC} | −11.6 ± 1.4 | −14.9 ± 0.3 | | | | |
| W37 ^{SC} | 7.2 ± 1.5 | 10.7 ± 0.3 | | | | |

All residual dipolar couplings are measured in Pf1 phage alignment media. Blanks indicate that data cannot be obtained due to the labeling scheme or the nature of the experiment performed.

1. Yang D, Nagayama K (1996) A sensitivity-enhanced method for measuring heteronuclear long-range coupling constants from the displacement of signals in two 1D subspectra. *J Magn Reson Ser A* 118:117–121.
2. Ottiger M, Delaglio F, Bax A (1998) Measurement of J and dipolar couplings from simplified two-dimensional NMR spectra. *J Magn Reson* 131:373–378.

Table S6. ^{13}C RCSA values of the SH3 domain component of the Abp1p SH3–Ark1p peptide complex derived from CPMG relaxation dispersion experiments and compared with values obtained directly when the complex is the major (visible) state in solution

| Residue | RCSA(^{13}C), ppb | |
|-------------------|------------------------------|--------------|
| | CPMG | Direct |
| A1 | | |
| P2 | | -17 ± 3 |
| W3 | | -120 ± 2 |
| A4 | | -81 ± 2 |
| T5 | | 35 ± 8 |
| A6 | -153 ± 22 | -111 ± 2 |
| E7 | -66 ± 24 | -40 ± 9 |
| Y8 | 35 ± 15 | 39 ± 1 |
| D9 | -82 ± 16 | -60 ± 9 |
| Y10 | | 71 ± 0 |
| D11 | | -118 ± 4 |
| A12 | | 108 ± 1 |
| A13 | -212 ± 16 | -158 ± 2 |
| E14 | | -88 ± 6 |
| D15 | 67 ± 15 | 73 ± 4 |
| N16 | | -70 ± 6 |
| E17 | | |
| L18 | | |
| T19 | | -27 ± 8 |
| F20 | | -142 ± 3 |
| V21 | | -48 ± 1 |
| E22 | | 44 ± 9 |
| N23 | | -106 ± 1 |
| D24 | | -92 ± 1 |
| K25 | | 74 ± 4 |
| I26 | | -128 ± 6 |
| I27 | | -70 ± 3 |
| N28 | | 21 ± 4 |
| I29 | | 44 ± 2 |
| E30 | | |
| F31 | 150 ± 19 | 129 ± 2 |
| V32 | -119 ± 56 | -114 ± 1 |
| D33 | | 2 ± 1 |
| D34 | -5 ± 15 | 2 ± 9 |
| D35 | | 25 ± 9 |
| W36 | -154 ± 17 | -87 ± 1 |
| W37 | -62 ± 14 | -54 ± 0 |
| L38 | | |
| G39 | | -34 ± 3 |
| E40 | | 32 ± 2 |
| L41 | | |
| E42 | | -17 ± 1 |
| K43 | | 102 ± 2 |
| D44 | | |
| G45 | | -106 ± 0 |
| S46 | | 145 ± 0 |
| K47 | | 95 ± 5 |
| G48 | | 68 ± 1 |
| L49 | | |
| F50 | | |
| P51 | | 149 ± 7 |
| S52 | -269 ± 24 | -186 ± 6 |
| N53 | 79 ± 16 | 58 ± 5 |
| Y54 | 0 ± 14 | 3 ± 2 |
| V55 | | 144 ± 3 |
| S56 | | -107 ± 1 |
| L57 | | |
| G58 | | 14 ± 1 |
| N59 | | |
| W36 ^{5C} | | |
| W37 ^{5C} | | |

Blanks indicate that data cannot be obtained due to the labeling scheme employed and/or the nature of the experiment.

Table S7. Residual dipolar couplings of the SH3 domain component of the Abp1p SH3 – Ark1p peptide complex derived from CPMG relaxation dispersion experiments and compared with dipolar couplings obtained directly (IPAP) when the complex is the major (visible) state in solution

| Residue | $^1D(^{15}N - ^1HN)$, Hz | |
|-------------------|---------------------------|------------|
| | CPMG | Direct |
| A1 | | -2.2 ± 0.1 |
| P2 | | |
| W3 | | 1.2 ± 0.1 |
| A4 | | -1.7 ± 0.1 |
| T5 | -13.9 ± 1.8 | -4.6 ± 0.1 |
| A6 | -10.1 ± 3.4 | -4.4 ± 0.1 |
| E7 | | 2.0 ± 0.1 |
| Y8 | 17.5 ± 1.1 | 6.5 ± 0.1 |
| D9 | 15.5 ± 1.4 | 6.3 ± 0.1 |
| Y10 | -8.3 ± 1.1 | -4.1 ± 0.1 |
| D11 | -5.4 ± 2.0 | -1.9 ± 0.1 |
| A12 | -10.0 ± 1.6 | -4.3 ± 0.1 |
| A13 | -6.7 ± 1.8 | -1.0 ± 0.1 |
| E14 | 0.8 ± 1.0 | 0.9 ± 0.1 |
| D15 | 4.3 ± 1.7 | 2.6 ± 0.1 |
| N16 | -4.4 ± 1.1 | -3.0 ± 0.1 |
| E17 | -1.9 ± 1.0 | 0.3 ± 0.1 |
| L18 | -13.0 ± 1.2 | -4.6 ± 0.1 |
| T19 | -2.3 ± 1.9 | -1.8 ± 0.1 |
| F20 | | -2.7 ± 0.1 |
| V21 | | -1.0 ± 0.1 |
| E22 | | 0.7 ± 0.1 |
| N23 | | -5.5 ± 0.1 |
| D24 | | 0.1 ± 0.1 |
| K25 | | -5.5 ± 0.1 |
| I26 | | |
| I27 | | -0.2 ± 0.1 |
| N28 | | 1.8 ± 0.1 |
| I29 | | 4.2 ± 0.1 |
| E30 | -1.6 ± 1.3 | -0.3 ± 0.1 |
| F31 | 1.5 ± 1.8 | 1.5 ± 0.1 |
| V32 | | -2.5 ± 0.1 |
| D33 | 1.0 ± 1.2 | 2.0 ± 0.1 |
| D34 | -5.8 ± 2.5 | 1.1 ± 0.1 |
| D35 | 12.0 ± 1.0 | 6.5 ± 0.1 |
| W36 | 2.8 ± 1.4 | 2.5 ± 0.1 |
| W37 | 6.1 ± 1.3 | 2.9 ± 0.1 |
| L38 | | -0.8 ± 0.1 |
| G39 | | |
| E40 | | -1.2 ± 0.1 |
| L41 | | 3.7 ± 0.1 |
| E42 | | -0.9 ± 0.1 |
| K43 | | -1.4 ± 0.1 |
| D44 | | -5.4 ± 0.1 |
| G45 | | 2.7 ± 0.1 |
| S46 | | 5.8 ± 0.1 |
| K47 | | 2.0 ± 0.1 |
| G48 | | |
| L49 | 5.3 ± 1.0 | 2.3 ± 0.1 |
| F50 | -3.1 ± 4.0 | -0.9 ± 0.1 |
| P51 | | |
| S52 | 4.2 ± 1.2 | 2.6 ± 0.1 |
| N53 | 4.5 ± 1.2 | 2.7 ± 0.1 |
| Y54 | | 8.4 ± 0.1 |
| V55 | | -3.3 ± 0.1 |
| S56 | | -5.7 ± 0.1 |
| L57 | | -4.7 ± 0.1 |
| G58 | | 4.0 ± 0.1 |
| N59 | | 1.7 ± 0.1 |
| W36 ^{SC} | -2.5 ± 1.2 | -0.9 ± 0.1 |
| W37 ^{SC} | -13.5 ± 1.7 | -4.8 ± 0.1 |

All residual dipolar couplings were measured in PEG/hexanol alignment media.

Table S8. (ϕ, ψ) dihedral angle restraint list derived using TALOS written as an input file

```

! Made on 1-Apr-08
! 1. A 4 Phi -130.03 +/- 60.06 (-190.09 to -69.97)
!assign (resid 3 and name C) (resid 4 and name N)
!      (resid 4 and name CA) (resid 4 and name C) 1.0 -130.03 60.06 2
! 2. T 5 Phi -117.83 +/- 48.00 (-165.83 to -69.83)
assign (resid 4 and name C) (resid 5 and name N)
      (resid 5 and name CA) (resid 5 and name C) 1.0 -117.83 48.00 2
! 3. A 6 Phi -67.10 +/- 31.62 (-98.72 to -35.48)
assign (resid 5 and name C) (resid 6 and name N)
      (resid 6 and name CA) (resid 6 and name C) 1.0 -67.10 31.62 2
! 4. Y 8 Phi -130.81 +/- 42.24 (-173.05 to -88.57)
assign (resid 7 and name C) (resid 8 and name N)
      (resid 8 and name CA) (resid 8 and name C) 1.0 -130.81 42.24 2
! 5. D 11 Phi -135.67 +/- 30.00 (-165.67 to -105.67)
assign (resid 10 and name C) (resid 11 and name N)
      (resid 11 and name CA) (resid 11 and name C) 1.0 -135.67 30.00 2
! 6. A 12 Phi -81.95 +/- 51.74 (-133.69 to -30.21)
assign (resid 11 and name C) (resid 12 and name N)
      (resid 12 and name CA) (resid 12 and name C) 1.0 -81.95 51.74 2
! 7. N 16 Phi -93.66 +/- 39.46 (-133.12 to -54.20)
assign (resid 15 and name C) (resid 16 and name N)
      (resid 16 and name CA) (resid 16 and name C) 1.0 -93.66 39.46 2
! 8. E 17 Phi -102.23 +/- 60.22 (-162.45 to -42.01)
assign (resid 16 and name C) (resid 17 and name N)
      (resid 17 and name CA) (resid 17 and name C) 1.0 -102.23 60.22 2
! 9. T 19 Phi -121.67 +/- 41.32 (-162.99 to -80.35)
assign (resid 18 and name C) (resid 19 and name N)
      (resid 19 and name CA) (resid 19 and name C) 1.0 -121.67 41.32 2
! 10. F 20 Phi -131.09 +/- 30.00 (-161.09 to -101.09)
!assign (resid 19 and name C) (resid 20 and name N)
!      (resid 20 and name CA) (resid 20 and name C) 1.0 -131.09 30.00 2
! 11. V 21 Phi -130.04 +/- 39.86 (-169.90 to -90.18)
!assign (resid 20 and name C) (resid 21 and name N)
!      (resid 21 and name CA) (resid 21 and name C) 1.0 -130.04 39.86 2
! 12. N 23 Phi -68.39 +/- 30.00 (-98.39 to -38.39)
!assign (resid 22 and name C) (resid 23 and name N)
!      (resid 23 and name CA) (resid 23 and name C) 1.0 -68.39 30.00 2
! 13. K 25 Phi -118.75 +/- 50.18 (-168.93 to -68.57)
!assign (resid 24 and name C) (resid 25 and name N)
!      (resid 25 and name CA) (resid 25 and name C) 1.0 -118.75 50.18 2
! 14. I 27 Phi -134.26 +/- 35.12 (-169.38 to -99.14)
!assign (resid 26 and name C) (resid 27 and name N)
!      (resid 27 and name CA) (resid 27 and name C) 1.0 -134.26 35.12 2
! 15. I 29 Phi -105.58 +/- 53.22 (-158.80 to -52.36)
assign (resid 28 and name C) (resid 29 and name N)
      (resid 29 and name CA) (resid 29 and name C) 1.0 -105.58 53.22 2
! 16. E 30 Phi -126.79 +/- 43.02 (-169.81 to -83.77)
assign (resid 29 and name C) (resid 30 and name N)
      (resid 30 and name CA) (resid 30 and name C) 1.0 -126.79 43.02 2

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Table S8. Continued.

! 17. F 31 Phi -96.67 +/- 30.00 (-126.67 to -66.67)
assign (resid 30 and name C) (resid 31 and name N)
(resid 31 and name CA) (resid 31 and name C) 1.0 -96.67 30.00 2

! 18. V 32 Phi -75.83 +/- 34.68 (-110.51 to -41.15)
assign (resid 31 and name C) (resid 32 and name N)
(resid 32 and name CA) (resid 32 and name C) 1.0 -75.83 34.68 2

! 19. D 34 Phi -62.47 +/- 30.00 (-92.47 to -32.47)
assign (resid 33 and name C) (resid 34 and name N)
(resid 34 and name CA) (resid 34 and name C) 1.0 -62.47 30.00 2

! 20. D 35 Phi -78.92 +/- 30.00 (-108.92 to -48.92)
assign (resid 34 and name C) (resid 35 and name N)
(resid 35 and name CA) (resid 35 and name C) 1.0 -78.92 30.00 2

! 21. W 36 Phi -112.20 +/- 46.22 (-158.42 to -65.98)
assign (resid 35 and name C) (resid 36 and name N)
(resid 36 and name CA) (resid 36 and name C) 1.0 -112.20 46.22 2

! 22. W 37 Phi -145.51 +/- 30.00 (-175.51 to -115.51)
assign (resid 36 and name C) (resid 37 and name N)
(resid 37 and name CA) (resid 37 and name C) 1.0 -145.51 30.00 2

! 23. L 38 Phi -118.11 +/- 69.94 (-188.05 to -48.17)
assign (resid 37 and name C) (resid 38 and name N)
(resid 38 and name CA) (resid 38 and name C) 1.0 -118.11 69.94 2

! 24. E 40 Phi -118.46 +/- 37.88 (-156.34 to -80.58)
assign (resid 39 and name C) (resid 40 and name N)
(resid 40 and name CA) (resid 40 and name C) 1.0 -118.46 37.88 2

! 25. L 41 Phi -83.58 +/- 30.00 (-113.58 to -53.58)
assign (resid 40 and name C) (resid 41 and name N)
(resid 41 and name CA) (resid 41 and name C) 1.0 -83.58 30.00 2

! 26. E 42 Phi -58.03 +/- 30.00 (-88.03 to -28.03)
assign (resid 41 and name C) (resid 42 and name N)
(resid 42 and name CA) (resid 42 and name C) 1.0 -58.03 30.00 2

! 27. K 43 Phi -68.82 +/- 30.00 (-98.82 to -38.82)
assign (resid 42 and name C) (resid 43 and name N)
(resid 43 and name CA) (resid 43 and name C) 1.0 -68.82 30.00 2

! 28. D 44 Phi -103.94 +/- 46.72 (-150.66 to -57.22)
assign (resid 43 and name C) (resid 44 and name N)
(resid 44 and name CA) (resid 44 and name C) 1.0 -103.94 46.72 2

! 29. G 45 Phi 86.87 +/- 30.00 (56.87 to 116.87)
assign (resid 44 and name C) (resid 45 and name N)
(resid 45 and name CA) (resid 45 and name C) 1.0 86.87 30.00 2

! 30. L 49 Phi -123.88 +/- 39.38 (-163.26 to -84.50)
assign (resid 48 and name C) (resid 49 and name N)
(resid 49 and name CA) (resid 49 and name C) 1.0 -123.88 39.38 2

! 31. P 51 Phi -75.76 +/- 42.62 (-118.38 to -33.14)
assign (resid 50 and name C) (resid 51 and name N)
(resid 51 and name CA) (resid 51 and name C) 1.0 -75.76 42.62 2

! 32. S 52 Phi -68.49 +/- 30.00 (-98.49 to -38.49)
assign (resid 51 and name C) (resid 52 and name N)
(resid 52 and name CA) (resid 52 and name C) 1.0 -68.49 30.00 2

! 33. S 56 Phi -105.94 +/- 41.94 (-147.88 to -64.00)

Table S8. Continued.

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assign (resid 55 and name C) (resid 56 and name N)
      (resid 56 and name CA) (resid 56 and name C) 1.0 -105.94 41.94 2
! 34. L 57 Phi -90.82 +/- 54.94 (-145.76 to -35.88)
!assign (resid 56 and name C) (resid 57 and name N)
!      (resid 57 and name CA) (resid 57 and name C) 1.0 -90.82 54.94 2
! 35. A 4 Psi 150.15 +/- 30.00 (120.15 to 180.15)
!assign (resid 4 and name N) (resid 4 and name CA)
!      (resid 4 and name C) (resid 5 and name N) 1.0 150.15 30.00 2
! 36. T 5 Psi 137.27 +/- 34.46 (102.81 to 171.73)
assign (resid 5 and name N) (resid 5 and name CA)
      (resid 5 and name C) (resid 6 and name N) 1.0 137.27 34.46 2
! 37. A 6 Psi 125.78 +/- 30.00 (95.78 to 155.78)
assign (resid 6 and name N) (resid 6 and name CA)
      (resid 6 and name C) (resid 7 and name N) 1.0 125.78 30.00 2
! 38. Y 8 Psi 156.16 +/- 30.00 (126.16 to 186.16)
assign (resid 8 and name N) (resid 8 and name CA)
      (resid 8 and name C) (resid 9 and name N) 1.0 156.16 30.00 2
! 39. D 11 Psi 135.65 +/- 50.50 (85.15 to 186.15)
assign (resid 11 and name N) (resid 11 and name CA)
      (resid 11 and name C) (resid 12 and name N) 1.0 135.65 50.50 2
! 40. A 12 Psi 132.53 +/- 30.00 (102.53 to 162.53)
assign (resid 12 and name N) (resid 12 and name CA)
      (resid 12 and name C) (resid 13 and name N) 1.0 132.53 30.00 2
! 41. N 16 Psi -14.30 +/- 44.94 (-59.24 to 30.64)
assign (resid 16 and name N) (resid 16 and name CA)
      (resid 16 and name C) (resid 17 and name N) 1.0 -14.30 44.94 2
! 42. E 17 Psi 147.36 +/- 37.56 (109.80 to 184.92)
assign (resid 17 and name N) (resid 17 and name CA)
      (resid 17 and name C) (resid 18 and name N) 1.0 147.36 37.56 2
! 43. T 19 Psi 140.56 +/- 40.66 (99.90 to 181.22)
assign (resid 19 and name N) (resid 19 and name CA)
      (resid 19 and name C) (resid 20 and name N) 1.0 140.56 40.66 2
! 44. F 20 Psi 156.05 +/- 30.00 (126.05 to 186.05)
!assign (resid 20 and name N) (resid 20 and name CA)
!      (resid 20 and name C) (resid 21 and name N) 1.0 156.05 30.00 2
! 45. V 21 Psi 160.92 +/- 30.00 (130.92 to 190.92)
!assign (resid 21 and name N) (resid 21 and name CA)
!      (resid 21 and name C) (resid 22 and name N) 1.0 160.92 30.00 2
! 46. N 23 Psi -29.64 +/- 39.08 (-68.72 to 9.44)
!assign (resid 23 and name N) (resid 23 and name CA)
!      (resid 23 and name C) (resid 24 and name N) 1.0 -29.64 39.08 2
! 47. K 25 Psi 122.43 +/- 30.00 (92.43 to 152.43)
!assign (resid 25 and name N) (resid 25 and name CA)
!      (resid 25 and name C) (resid 26 and name N) 1.0 122.43 30.00 2
! 48. I 27 Psi 163.68 +/- 34.02 (129.66 to 197.70)
!assign (resid 27 and name N) (resid 27 and name CA)
!      (resid 27 and name C) (resid 28 and name N) 1.0 163.68 34.02 2
! 49. I 29 Psi 142.42 +/- 30.00 (112.42 to 172.42)

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Table S8. Continued.

| | | | | | | |
|--------|--|------------------------|-----|--------|-------|---|
| assign | (resid 29 and name N) | (resid 29 and name CA) | | | | |
| | (resid 29 and name C) | (resid 30 and name N) | 1.0 | 142.42 | 30.00 | 2 |
| ! | 50. E 30 Psi 134.52 +/- 30.00 (104.52 to 164.52) | | | | | |
| assign | (resid 30 and name N) | (resid 30 and name CA) | | | | |
| | (resid 30 and name C) | (resid 31 and name N) | 1.0 | 134.52 | 30.00 | 2 |
| ! | 51. F 31 Psi 152.00 +/- 50.76 (101.24 to 202.76) | | | | | |
| assign | (resid 31 and name N) | (resid 31 and name CA) | | | | |
| | (resid 31 and name C) | (resid 32 and name N) | 1.0 | 152.00 | 50.76 | 2 |
| ! | 52. V 32 Psi -15.21 +/- 50.08 (-65.29 to 34.87) | | | | | |
| assign | (resid 32 and name N) | (resid 32 and name CA) | | | | |
| | (resid 32 and name C) | (resid 33 and name N) | 1.0 | -15.21 | 50.08 | 2 |
| ! | 53. D 34 Psi -42.99 +/- 30.00 (-72.99 to -12.99) | | | | | |
| assign | (resid 34 and name N) | (resid 34 and name CA) | | | | |
| | (resid 34 and name C) | (resid 35 and name N) | 1.0 | -42.99 | 30.00 | 2 |
| ! | 54. D 35 Psi -10.97 +/- 30.00 (-40.97 to 19.03) | | | | | |
| assign | (resid 35 and name N) | (resid 35 and name CA) | | | | |
| | (resid 35 and name C) | (resid 36 and name N) | 1.0 | -10.97 | 30.00 | 2 |
| ! | 55. W 36 Psi 135.84 +/- 60.06 (75.78 to 195.90) | | | | | |
| assign | (resid 36 and name N) | (resid 36 and name CA) | | | | |
| | (resid 36 and name C) | (resid 37 and name N) | 1.0 | 135.84 | 60.06 | 2 |
| ! | 56. W 37 Psi 162.21 +/- 30.00 (132.21 to 192.21) | | | | | |
| assign | (resid 37 and name N) | (resid 37 and name CA) | | | | |
| | (resid 37 and name C) | (resid 38 and name N) | 1.0 | 162.21 | 30.00 | 2 |
| ! | 57. L 38 Psi 153.85 +/- 30.00 (123.85 to 183.85) | | | | | |
| assign | (resid 38 and name N) | (resid 38 and name CA) | | | | |
| | (resid 38 and name C) | (resid 39 and name N) | 1.0 | 153.85 | 30.00 | 2 |
| ! | 58. E 40 Psi 140.85 +/- 35.72 (105.13 to 176.57) | | | | | |
| assign | (resid 40 and name N) | (resid 40 and name CA) | | | | |
| | (resid 40 and name C) | (resid 41 and name N) | 1.0 | 140.85 | 35.72 | 2 |
| ! | 59. L 41 Psi 117.23 +/- 30.00 (87.23 to 147.23) | | | | | |
| assign | (resid 41 and name N) | (resid 41 and name CA) | | | | |
| | (resid 41 and name C) | (resid 42 and name N) | 1.0 | 117.23 | 30.00 | 2 |
| ! | 60. E 42 Psi -37.44 +/- 30.00 (-67.44 to -7.44) | | | | | |
| assign | (resid 42 and name N) | (resid 42 and name CA) | | | | |
| | (resid 42 and name C) | (resid 43 and name N) | 1.0 | -37.44 | 30.00 | 2 |
| ! | 61. K 43 Psi -25.36 +/- 38.82 (-64.18 to 13.46) | | | | | |
| assign | (resid 43 and name N) | (resid 43 and name CA) | | | | |
| | (resid 43 and name C) | (resid 44 and name N) | 1.0 | -25.36 | 38.82 | 2 |
| ! | 62. D 44 Psi -3.51 +/- 30.00 (-33.51 to 26.49) | | | | | |
| assign | (resid 44 and name N) | (resid 44 and name CA) | | | | |
| | (resid 44 and name C) | (resid 45 and name N) | 1.0 | -3.51 | 30.00 | 2 |
| ! | 63. G 45 Psi -5.29 +/- 31.68 (-36.97 to 26.39) | | | | | |
| assign | (resid 45 and name N) | (resid 45 and name CA) | | | | |
| | (resid 45 and name C) | (resid 46 and name N) | 1.0 | -5.29 | 31.68 | 2 |
| ! | 64. L 49 Psi 145.22 +/- 30.00 (115.22 to 175.22) | | | | | |
| assign | (resid 49 and name N) | (resid 49 and name CA) | | | | |
| | (resid 49 and name C) | (resid 50 and name N) | 1.0 | 145.22 | 30.00 | 2 |
| ! | 65. P 51 Psi 127.81 +/- 48.14 (79.67 to 175.95) | | | | | |

Table S8. Continued.

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assign (resid 51 and name N) (resid 51 and name CA)
      (resid 51 and name C) (resid 52 and name N) 1.0 127.81 48.14 2
! 66. S 52 Psi -21.12 +/- 30.00 (-51.12 to 8.88)
assign (resid 52 and name N) (resid 52 and name CA)
      (resid 52 and name C) (resid 53 and name N) 1.0 -21.12 30.00 2
! 67. S 56 Psi 126.84 +/- 30.00 (96.84 to 156.84)
assign (resid 56 and name N) (resid 56 and name CA)
      (resid 56 and name C) (resid 57 and name N) 1.0 126.84 30.00 2
! 68. L 57 Psi 133.59 +/- 30.00 (103.59 to 163.59)
!assign (resid 57 and name N) (resid 57 and name CA)
!      (resid 57 and name C) (resid 58 and name N) 1.0 133.59 30.00 2

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