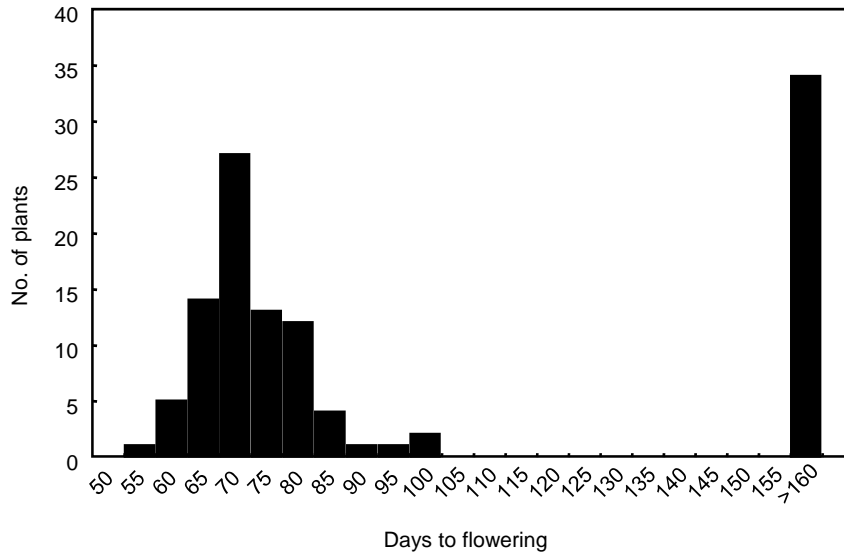
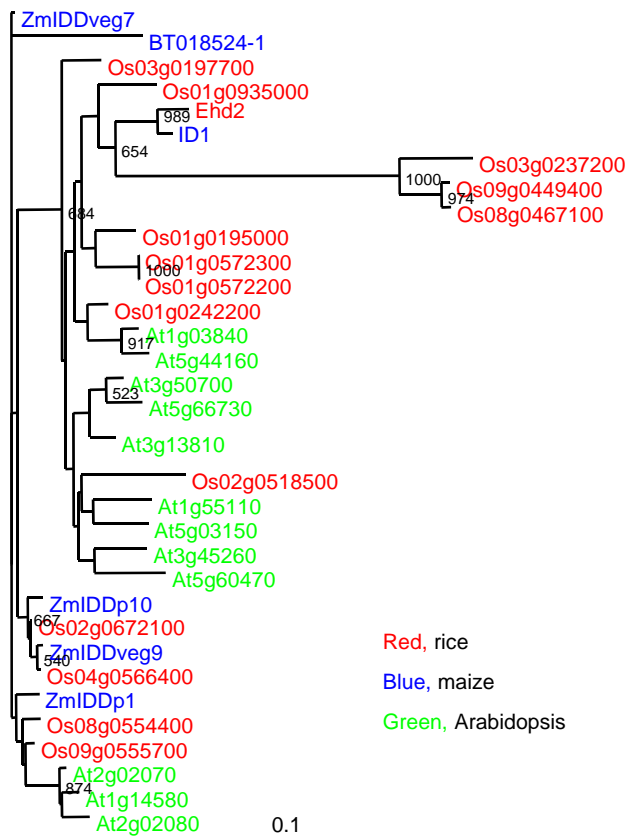


**Supplemental Figure S1.** Distribution of days to flowering under natural-day conditions in BC<sub>1</sub>F<sub>2</sub> population ( $n = 109$ ) from the cross between the *ehd2* mutant and 'Guang Lu Ai 4'. The segregation ratio of normal to late-flowering phenotype was 81 : 28 (the expected ratio 3 : 1,  $\chi^2 = 0.03^{ns}$ ). Days to flowering of the wild-type plants ('Tohoku IL9'), 'Guang Lu Ai 4' and the *ehd2* mutants were  $80.7 \pm 2.5$ ,  $78.3 \pm 2.9$  and  $169.7 \pm 0.6$ , respectively ( $n = 10$ ). The seeds were sown in the middle of June and the plants were grown in a paddy field in Tsukuba, Japan.

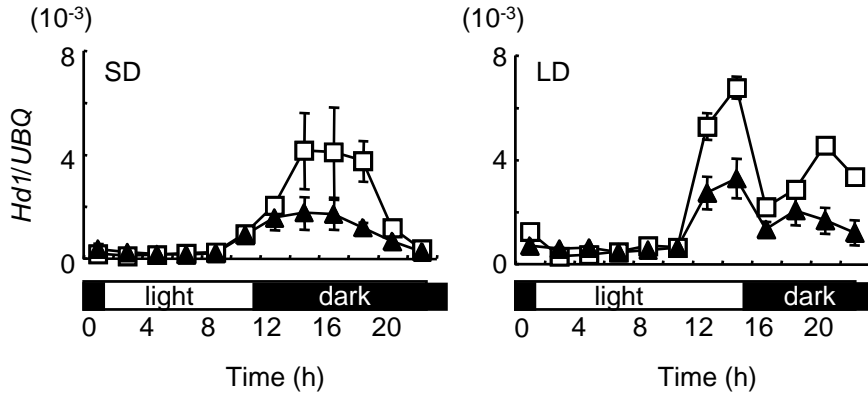




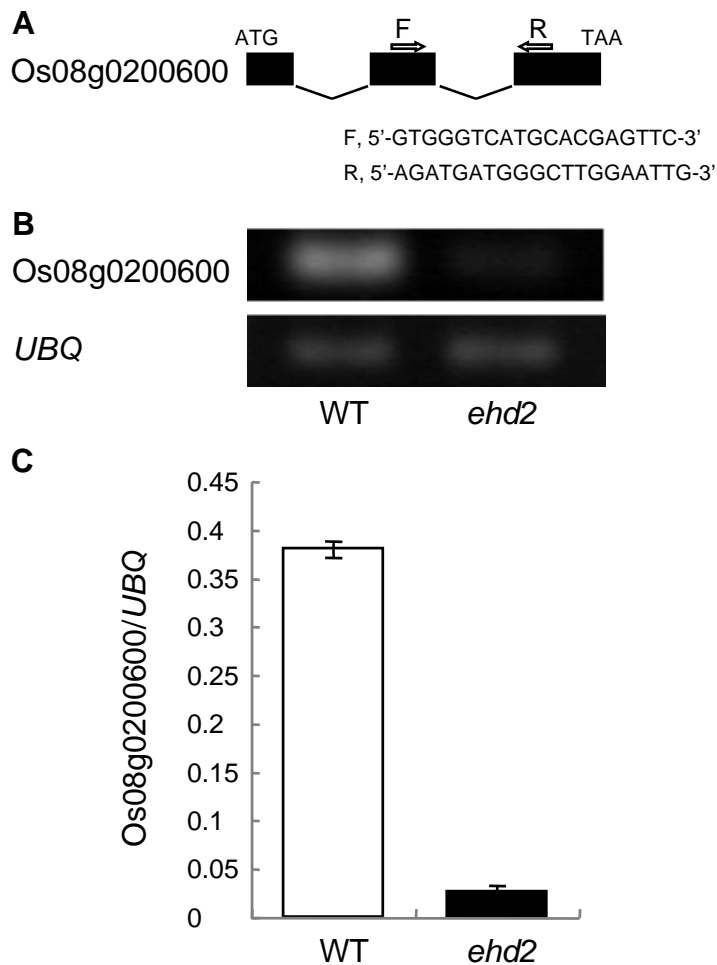
**Supplemental Figure S3.** Phylogenetic comparison of ID-domain amino acid sequences from rice, maize and *Arabidopsis thaliana*. Multiple sequence alignments were produced with CLUSTALW version 1.83 (<http://clustalw.ddbj.nig.ac.jp/top-j.html>) using default settings. The phylogenetic tree was calculated using the neighbor-joining method and bootstrap analysis (1000 replicates) using PHYLIP via the same website and visualized with Treeview version 1.6.6 (<http://taxonomy.zoology.gla.ac.uk/rod/treeview.html>). Bootstrap values for nodes supported in > 50% were shown.



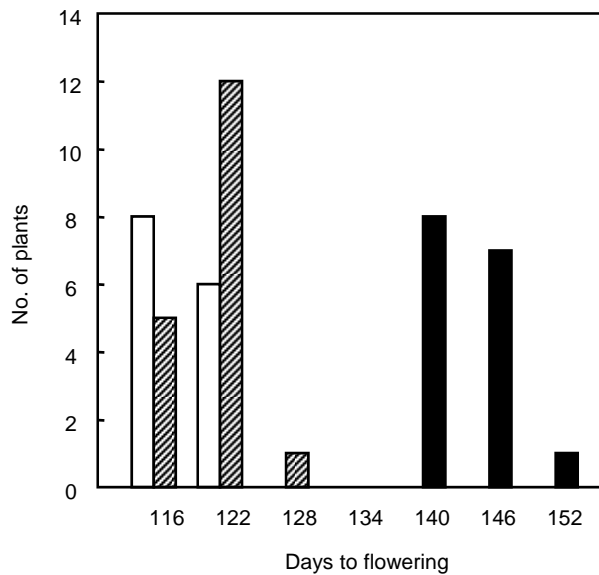
**Supplemental Figure S4.** Diurnal changes in levels of *Hd1* transcripts under SD (10 h light / 14 h dark) and LD (14.5 h light / 9.5 h dark) conditions. UBQ, ubiquitin. Open squares represent wild-type plants; filled triangles represent the *ehd2* mutants. Developing leaves were harvested every 2 h. Values are shown as mean  $\pm$  s.d. of three independent experiments. Data are representative of two independent biological replicates.



**Supplemental Figure S5.** Expression analyses of Os08g0200600 (NAM protein domain containing protein). (A) Putative gene structure of Os08g0200600. F and R are primers used for RT-PCR and real-time RT-PCR analyses. Expression of Os08g0200600 shown by RT-PCR (B) and real-time RT-PCR (C). Values are shown as means  $\pm$  s.d. of three independent experiments in (C). UBQ, ubiquitin. Young leaves of the *ehd2* mutants and its wild-type plants (Tohoku IL9) were collected at two hours after dawn from one-month-old plants grown under LD (14.5h light / 9.5 h dark) condition.



**Supplemental Figure S6.** Distribution of days to flowering under natural-day condition in the selfed-progeny ( $n = 48$ ) of the *Tos17*-induced mutants heterozygous for *Ehd2*. The segregation ratio of homozygotes for 'Nipponbare' allele (open boxes), heterozygotes (slashed boxes) and homozygotes for *Tos17*-inserted allele (filled boxes) was 14 : 18 : 16 (the expected ratio 1 : 2 : 1,  $\chi^2 = 3.17^{ns}$ ). Days to flowering of 'Nipponbare' was  $116.9 \pm 1.2$  ( $n = 10$ ). The seeds were sown in the middle of April and the plants were grown in a paddy field in Tsukuba, Japan.



**Supplemental Table S1.** Genetic markers used in the delimitation of candidate genomic region of *Ehd2*

Primer name	Sequence (5' - 3')		Remarks
	Forward	Reverse	
Indel-1	CTAGTTTGTGCAACTAAGCAAGTGG	GTTTCTTAATGTTGCCATGCATGC	-
CAPS-1	AAGCAGTGGGATTGGTTGAC	CCATGACAGCAGGAGAGACA	<i>HindIII</i>
SNP-1	TGAGTGGCAGAGGTTACAACA	GGTGGTGGTCCAAGAAAAGA	Direct sequencing
SSR-1	AGGATTTGCAGGTTTTCAA	TCTAATGCATACATGGTGCT	-
SNP-2	GGCCAGGAGACTCTTATTGG	GTCCCCATGACCATCTTCC	Direct sequencing
CAPS-2	ATTTCCAAGAACTCTACCACCC	CTTGTCTCTGCCACTCACG	<i>BamHI</i>

## Supplemental Table S2. Summary of microarray data

Top three genes that showed the greatest fold-change and *Ehd2* were represented. Young leaves of the *ehd2* mutants and its wild-type plants (Tohoku IL9) were collected at two hours after dawn from one-month-old plants grown under LD (14.5h light / 9.5 h dark) condition. Total RNA was isolated using a Plant RNA Isolation plant Mini Kit (Agilent Technologies). The RNAs (400-ng aliquots) were labeled with a Low RNA Input Linear Amplification/Labeling Kit (Agilent Technologies) according to the manufacturer's instructions. Aliquots of Cy3- and Cy5-labeled cRNAs (1 mg each) of samples from the *ehd2* mutants or its wild-type plants were used for hybridization in an Agilent Rice Oligo Microarray (44K, custom-made; Agilent Technologies). After hybridization, microarray slides were scanned (scanner model G2505B and software G2565BA; Agilent), and data were analyzed using Feature Extraction software (version 9.1; Agilent Technologies) at the default settings. All microarray procedures and data analyses were performed according to the manufacturer's manual. Two independent biological replicates were performed with color-swap experiments. Down, down-regulation by the *ehd2* mutation.

Locus ID	Annotation	Fold-change ( <i>Ehd2/ehd2</i> )		<i>P</i> value	Fold-change ( <i>Ehd2/ehd2</i> )		<i>P</i> value
1st experiment							
Os10g0463400	Early heading date 1 (Ehd1)	12.0	Down	3.48E-13	8.2	Down	2.20E-16
Os08g0200600	NO APICAL MERISTEM (NAM) protein domain containing protein	4.8	Down	6.94E-22	3.6	Down	4.99E-18
Os04g0640700	Alpha-L-arabinofuranosidase/beta-D-xylosidase isoenzyme (ARA-I)	4.4	Down	1.13E-09	2.9	Down	1.87E-08
Os10g0419200	Early heading date 2 (Ehd2)	3.4	-	5.41E-05	2.2	-	3.13E-03
2nd experiment							
Os10g0463400	Early heading date 1 (Ehd1)	12.8	Down	2.10E-04	50.2	Down	1.52E-21
Os08g0200600	NO APICAL MERISTEM (NAM) protein domain containing protein	4.8	Down	5.51E-14	5.7	Down	4.92E-16
Os04g0640700	Alpha-L-arabinofuranosidase/beta-D-xylosidase isoenzyme (ARA-I)	5.9	Down	1.19E-02	3.4	Down	9.93E-11
Os10g0419200	Early heading date 2 (Ehd2)	5.6	-	0.12	25.4	-	3.76E-18