Supporting Information

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SI Methods

Immunohistochemistry. Immunohistochemistry was performed using 4-μm-thick formalin-fixed, paraffin-embedded tissue sections. Briefly, slides were soaked in xylene, passed through graded alcohols, and put in distilled water. Slides then were pretreated with either 1.0 mM EDTA, pH 8.0 (for CD3), or 1× Dako Target Retrieval Solution pH 9 (for CD56) (DAKO) in a steam pressure cooker (Decloaking Chamber; BioCare Medical) per the manufacturer's instructions, followed by washing in distilled water. All further steps were performed at room temperature in a hydrated chamber. Slides were pretreated with Peroxidase Block (DAKO) for 5 min to quench endogenous peroxidase activity. Polyclonal rabbit anti-human CD3 antibody (DAKO) was applied 1:250, or monoclonal mouse anti-human CD56 antibody (clone 123C3; DAKO) was applied 1:100 in antibody diluent (DAKO) for 1 h, primary rabbit anti-gal1 antibody (Gabriel Rabinovich, Instituto de Biología y Medicina Experimental, Consejo Nacional de Investigaciones Científicas y Técnicas, Buenos Aires, Argentina) (1) was applied at a 1:10,000 dilution in antibody diluent for 1 h. Slides were washed in 50 mM Tris·Cl, pH 7.4, and anti-rabbit or mouse HRP-conjugated antibody (Envision detection kit; DAKO) was applied for 30 min. After further washing, immunoperoxidase staining was developed using a DAB chromogen kit (DAKO) per the manufacturer's instructions and was counterstained with Harris hematoxylin. TUNEL method was used according to the manufacturer's instructions (ApopTaG 7100 Kit; Chemicon/Millipore).

Western Blot. 1×10^6 FACS-sorted dNKs and $CD56^{bright}$ and $CD56^{dim}$ pNKs were lysed in reducing SDS/PAGE loading buffer (NuPAGE LDS sample buffer; Invitrogen) by boiling for 10 min. Proteins were separated in 12% polyacrylamide gels and transferred to nitrocellulose filters. Filters were reveled with anti-gal1 mAb (NCL GAL1; Novocastra Laboratories Ltd.) or anti-β-actin antibodies followed by an HRP-conjugated anti-mouse Fc antibody, and were developed by ECL (Amersham).

RT-PCR. Flow-sorted cells were washed with PBS, and the cell pellets were frozen at −80°C in TRIzol (GIBCO–BRL). Total RNA was isolated according to the TRIzol manufacturer's instructions, with addition of 5 μg linear polyacrylamide (Genelute LPA; Sigma–Aldrich) to aid visualization of RNA pellets. A total of 100 ng RNA from each sample was used for first-strand cDNA synthesis using SuperScript First Strand Synthesis System (Invitrogen) according to the manufacturer's protocol. A total of 2 μL cDNA template was used for PCR (Platinum TaqDNA polymerase; Invitrogen). A total of 20 μL of each product was visualized on 1–2% agarose gels. Primers used were as follows: C2GnT, forward: ATGCTGAGGACGTTGCTGCGAA, reverse: TCAGTGTTTTAATGTCTCCAAAGCT; and β-actin, forward: GTTGCTATCCAGGCTGTGCT, reverse: CTCCTTAATGTCACGCACGA.

1. Rodig SJ, *et al.* (2008) AP1-dependent galectin-1 expression delineates classic hodgkin and anaplastic large cell lymphomas from other lymphoid malignancies with shared molecular features. *Clin Cancer Res* 14:3338–3344.