

SUPPLEMENTARY FIGURE LEGENDS

Supplementary Figure 1.

A. *P66^{Shc} expression in adipocytes.* P66^{Shc} protein expression in BAT and WAT (upper panel), BAT-derived pre-adipocytes (BAT ADPC; central panel), and WAT-derived pre-adipocytes (WAT ADPC; lower panel). I: insulin treatment; d: days of treatment.

B. *Effects of p66^{Shc} and p66^{Shc}qq expression in Foxo1 localization.* Foxo1 localization in untreated and insulin treated p66^{Shc}^{-/-} BAT pre-adipocytes infected with empty (Vect), p66^{Shc} or p66^{Shc}qq retroviral vectors.

C. *Trygliceride accumulation in WT and p66^{Shc}^{-/-} BAT pre-adipocytes upon treatment with IBMX/DEX.* White light microscopy pictures of Oil red-stained BAT pre-adipocytes from WT and p66^{Shc}^{-/-} mice, untreated or after treatment with IBMX-dexametasone (IBMX-DEX), and their corresponding OD values (this experiment is representative of 3 that gave comparable results).

Supplementary Figure 2.

Gene regulations in WT and p66^{Shc}^{-/-} BAT pre-adipocytes treated with insulin. Venn diagrams representing overlapping gene regulations in WT and p66^{Shc}^{-/-} BAT pre-adipocytes treated with insulin. The table reports the top 20 insulin-regulated genes in each group. In bold are indicated the genes whose analysis is showed in Figure 3C.

Supplementary Figure 3.

A. *Lipid profile and blood levels of insulin and glucose in WT and p66^{Shc}^{-/-} mice.* Plasma levels of free fatty acids, triglycerides, cholesterol, insulin and glucose in WT and p66^{Shc}^{-/-} mice fed standard (SD) and high fat (HF) diet (n=20 mice per group).

B. *Effect of N-acetylcysteine supplementation on weight gain.* Body weight curves of WT 129Sv male mice fed standard or high fat (HF) diets plus or minus 40 mM NAC in the drinking water (n=12 mice per group).

Supplementary Figure 4.

Model of p66^{Shc}-mediated regulation of TG synthesis in adipocytes. The scheme depicts the role of p66^{Shc} in regulating insulin-mediated TG accumulation in adipocytes, and highlights its role in the integration of insulin signaling (right side of the scheme) and mitochondrial respiration (left circle). Availability of carbon sources (Food) is indicated at the top as major determinant of the levels of circulating insulin. Engagement of insulin receptors in adipocytes leads to p66^{Shc} activation (via phosphorylation, as described in the text; vertical red arrow) and its translocation (indicated by the arrowed curve) within the mitochondrial inter membrane space (in pink). There, p66^{Shc} generates H₂O₂, which selectively potentiates insulin signaling (the red arrow indicates the insulin signaling branch regulated by p66^{Shc}-generated H₂O₂: PTEN-PI3K-AKT-Foxo1 pathway, as described in the text) and contributes to gene expression reprogramming by insulin. Regulated expression of selected target genes (UCP1 and others, see text) leads to respiration coupling, inhibition of β -oxidation and TG accumulation. The circled scheme on the left magnifies the electron transfer chain (green arrowed curves refer to electron flow across complexes I to IV) and the redox reaction between p66^{Shc} (in red) and reduced cytochrome *c* (c) that catalyzes the partial reduction of O₂ to H₂O₂. Emphasis is given to the fact that production of H₂O₂ by p66^{Shc} depends on availability of respiratory substrates, thus allowing cross-talk between the energetic status of adipocytes and the insulin signaling pathway, via p66^{Shc}.

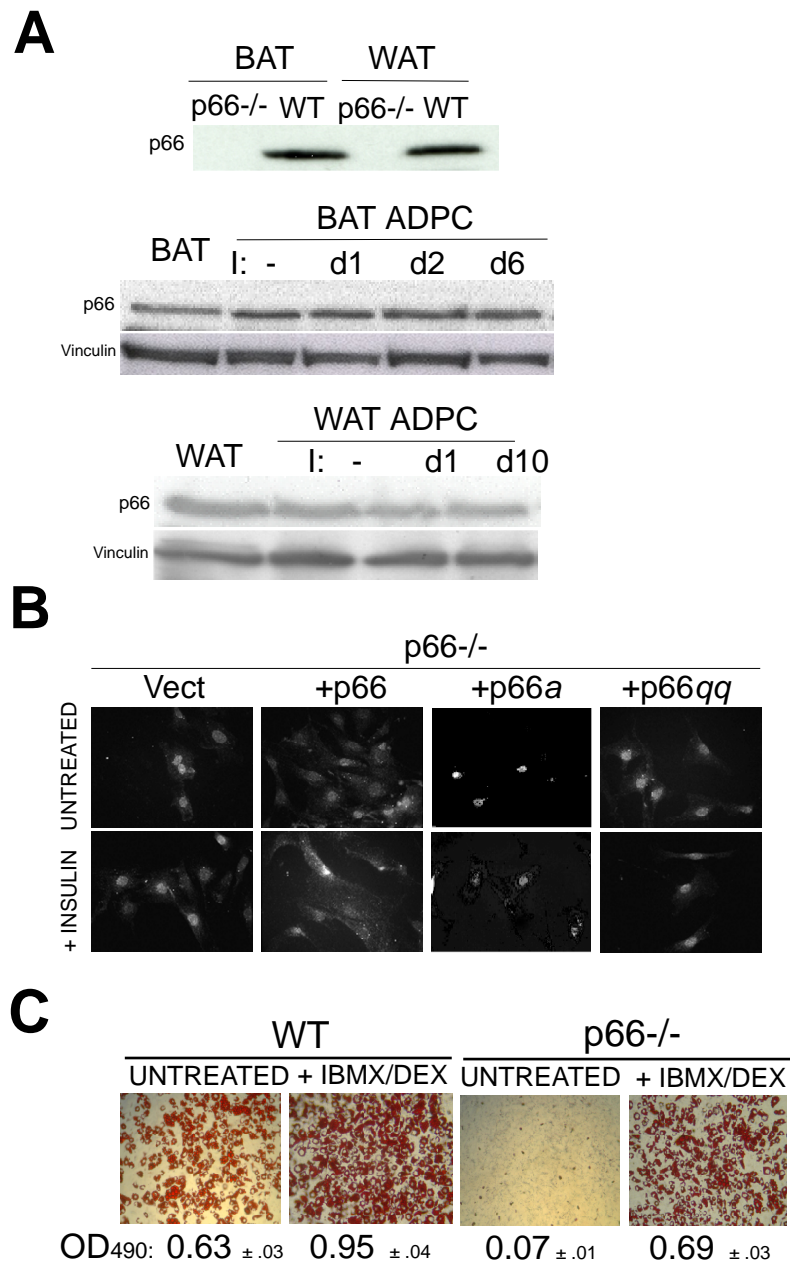
SUPPLEMENTARY REFERENCES

- Ross, SH., Lindsay, Y., Safrany, ST., Lorenzo, O., Villa, F., Toth, R., Clague, MJ., Downes, CP., and Leslie, NR. (2007). Differential redox regulation within the PTP superfamily. *Cell Signal.* **19**, 1521-1530.
- Meng, T.C., Fukada, T., and Tonks, N.K. (2002). Reversible oxidation and inactivation of protein tyrosine phosphatases in vivo. *Mol Cell.* **9**, 387-99
- Chiarugi P. (2005) PTPs versus PTKs: the redox side of the coin. *Free Radic Res.* **39**, 353-64.
- Finkel, T. (2000). Redox-dependent signal transduction. *FEBS Lett.* **476**, 52-4.
- Paravicini, T.M., and Touyz, R.M. (2006). Redox signaling in hypertension. *Cardiovasc Res.* **71**, 247-58
- Song, H., Bao, S., Ramanadham, S., and Turk, J. (2006). Effects of biological oxidants on the catalytic activity and structure of group VIA phospholipase A2. *Biochemistry.* **45**, 6392-406.
- Bossis, G., and Melchior, F. (2006). Regulation of SUMOylation by reversible oxidation of SUMO conjugating enzymes. *Mol Cell.* **21**, 349-57.
- Davis, DA., Newcomb, FM., Moskovitz, J., Wingfield, PT., Stahl, SJ., Kaufman, J., Fales, HM., Levine, RL., and Yarchoan, R. (2000). HIV-2 protease is inactivated after oxidation at the dimer interface and activity can be partly restored with methionine sulphoxide reductase. *Biochem J.* **346**, 305-11.
- Ahn, SG., and Thiele, DJ. (2003). Redox regulation of mammalian heat shock factor 1 is essential for Hsp gene activation and protection from stress. *Genes Dev.* **17**, 516-28.
- Clarkson, RW., Shang, CA., Levitt, LK., Howard, T., and Waters, MJ. (1999). Ternary complex factors Elk-1 and Sap-1a mediate growth hormone-induced transcription of egr-1 (early growth response factor-1) in 3T3-F442A preadipocytes. *Mol Endocrinol.* **13**, 619-31.
- Fajas, L., Miard, S., Briggs, MR., and Auwerx, J. (2003). Selective cyclo-oxygenase-2 inhibitors impair adipocyte differentiation through inhibition of the clonal expansion phase. *J Lipid Res.* **44**, 1652-9.
- Alessi, MC., Poggi, M., and Juhan-Vague, I. (2007). Plasminogen activator inhibitor-1, adipose tissue and insulin resistance. *Curr Opin Lipidol.* **18**, 240-5.
- Xu, Y., Mirmalek-Sani, SH., Yang, X., Zhang, J., and Oreffo, RO. (2006). The use of small interfering RNAs to inhibit adipocyte differentiation in human preadipocytes and fetal-femur-derived mesenchymal cells. *Exp Cell Res.* **312**, 1856-64.

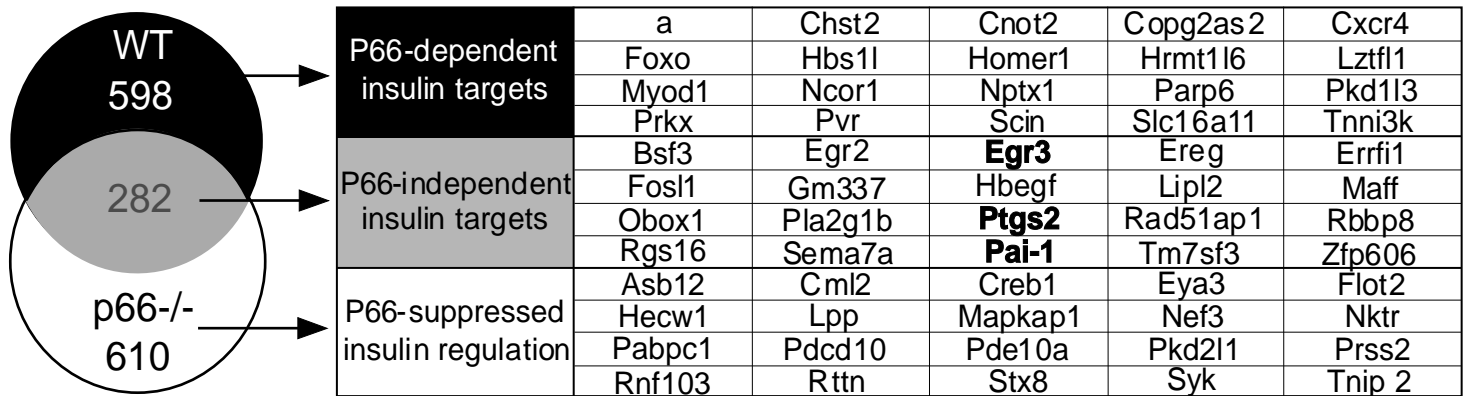
Bonen, A., Tandon, NN., Glatz, JF., Luiken, JJ., and Heigenhauser, GJ. (2006). The fatty acid transporter FAT/CD36 is upregulated in subcutaneous and visceral adipose tissues in human obesity and type 2 diabetes. *Int J Obes (Lond)*. **30**, 877-83.

Balakrishnan, K., Verdile, G., Mehta, PD., Beilby, J., Nolan, D., Galvão, DA., Newton, R., Gandy, SE., and Martins, RN. (2005). Plasma Abeta42 correlates positively with increased body fat in healthy individuals. *J Alzheimers Dis*. **8**, 269-82.

Supplementary Figure 1



Supplementary Figure 2

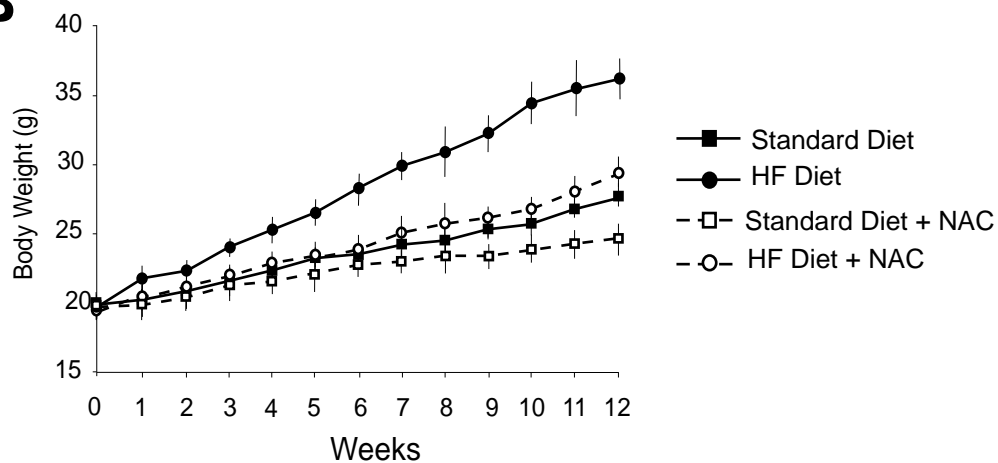


Supplementary Figure 3

A

| | WT SD | WT HFD | p66-/- SD | p66-/- HFD |
|-------------------------|-------------|-------------|-------------|-------------|
| Free Fatty Acid (meq/L) | 0.57 ± 0.12 | 1.24 ± 0.32 | 0.45 ± 0.11 | 1.11 ± 0.28 |
| Triglycerides (mg/dL) | 68 ± 9 | 143 ± 30 | 61 ± 6 | 118 ± 35 |
| Cholesterol (mg/dL) | 53 ± 6 | 112 ± 23 | 55 ± 8 | 103 ± 17 |
| Insulin (ng/dL) | 1.34 ± 0.22 | 1.47 ± 0.18 | 0.93 ± 0.15 | 0.90 ± 0.12 |
| Glucose (mg/dL) | 96 ± 5.4 | 103 ± 7.9 | 89 ± 4.5 | 90 ± 4.3 |

B



Supplementary Figure 4

