



(A) Comparison of the boric acid induced swelling rates of negative control oocytes (uninjected or DEPC-water "mock" injected oocytes) and NIP5;1- and NIP6;1- expressing oocytes was done as described in the Methods. Error bars show the SEM (n=7 oocytes for each value); (B) Comparison of the boric acid-induced oocyte swelling properties of NIP5;1 and NIP6;1 in the absence (-Hg) and the presence of 1 mM HgCl₂ (+Hg) are shown. The error bars show the SEM (n=6 oocytes for each value).



Supplemental Figure 2. FLAG-tag Western blot of *Xenopus* oocyte lysates.

5 (**A**) and 20 (**B**) μg of lysate protein from NIP5;1- as well as NIP6;1- expressing oocytes were separated on a 12.5% SDS-PAGE gel, blotted to PVDF, and probed with the anti-FLAG antibody. Lane 1: uninjected oocyte lysate, lane 2: NIP5;1lysate, lane 3: NIP6;1- lysate. The position of the molecular mass markers (kDa) are indicated to the left of the blot.



Supplemental Figure 3. Plant growth under high B conditions.

Wild-type plants and the *NIP6;1*T-DNA insertion lines were grown hydroponically for 28 days supplied with 100 μ M B under long-day conditions (16 h/ 8 h light/dark cycle). Bar=50 mm.



Supplemental Figure 4. GUS Staining in *Pro-NIP6;1-GUS* transgenic seedlings.

Pro-NIP6;1-GUS transgenic plants were grown on solid medium for 10 days supplied with 100 μ M B and stained with 5-bromo-4-chloro-3-indolyl-b-D-glucuronic acid. (A) Whole shoot, (B) Close-up of the nodal area. Bar=1 mm in (A), bar=500 μ m in (B).



Supplemental Figure 5. Plant growth of NIP6;1T-DNA insertion lines for 14 days.

Wild-type plants and the *NIP6;1* T-DNA insertion lines were grown hydroponically for 14 days supplied with 0.1 and 100 μ M B under short-day conditions (10 h/14 h light/dark cycle). Bar =10 mm.