

Table S6 List of oligonucleotides

Gene	Primer	Primer pair efficiency		Primer sequence
		<i>N.benth.</i>	<i>S.penn.</i>	
Real-time RT-PCR				
SuRA	ALA745_F ALA745_R	-	100	5'-GCCTCACCCCTTCATCTCATC-3' 5'-TTCAGGTTCCAGAAACGTCGTTAT-3'
TD	TD_F TD_R	-	100	5'-CCGCCGTGAAATCATTGAAT-3' 5'-GCTAATAGGAACAATGACGGAAGTC-3'
<i>IPMS A</i>	QIPMSA_for QIPMSA_rev	96.1	95.4	5'-CAAGTATTAGGTGGCCTCTACACG-3' 5'-CATTAGCTCCAACAATGGCCTTA-3'
<i>IPMS B</i>	QIPMSB_for QIPMSB_rev	100	100	5'-CAAGTACTAGGTGGCCTGTATACA-3' 5'-CATTAGCTCCAACAATGGCCTTG-3'
<i>IPMS C</i>	NbCN199Q_F NbCN199Q_R	97.5	-	5'-CCAAAATACAAGCCGGGAAGA-3' 5'-CTTAGCCAGCTGATGTGCAATT-3'
<i>IPMD</i>	QHYD_for QHYD_rev	88.4	100	5'-TGTCCGATGAAGCATCAATGATTA-3' 5'-TTTATCCTGCCAGCAATATCAG-3'
<i>IPDS-L</i>	DHT_for DHT_rev	100	100	5'-AAGGTATGGATGGATGTATATAGCCTC-3' 5'-GTGTCTTTAGGGCCACCCAAAAC-3'
<i>BCKD E1-β</i>	BOX_F BOX_R	100	-	5'-TCAAGCATACTGACCCAAA-3' 5'-TGAGGACCTCTGCCTCAGAT-3'
<i>BCKD E1-β</i>	BOX_F1 BOX_R1	-	100	5'-TCAAGCATACTGACCCAAA-3' 5'-TGAGGACCTCTGCCTCAGAT-3'
<i>BCKD E2</i>	DHLAT_for DHLAT_rev	100	-	5'-GTGAAGTTCAGAGTGACAAAGCAA-3' 5'-CGATCTTCAAAGTGTCTTCCAA-3'
<i>BCKD E2</i>	DHLAT2_F DHLAT2_R	-	100	5'-TGTCGAAGAGATAAATTGTGATGC-3' 5'-TGAGGACGGGAAGGAAAGTG-3'
<i>KAS III A</i>	K1Q_FOR K1Q_REV	-	95.3	5'-ATTCGGTGGACAAATTAGAGGAT-3' 5'-TCAAGCCTTCTATCATTCTTACCA-3'
<i>KAS I A</i>	K3Q_FOR K3Q_REV	-	100	5'-ATTCGGTGGACAAATTAGAGGAT-3' 5'-TCAAGCCTTCTATCATTCTTACCA-3'
<i>KAS I B</i>	K4QA_FOR K4Q_REV	-	91.8	5'-TCGCATCGTTAACATCGAAAAG-3' 5'-AGTGGAGACGGACGAAATGATA-3'
<i>KAS I C</i>	K5Q_FOR K5Q_REV	-	90.1	5'-GTGTGTTTGGAGAAAATGAGTAGTATTACTT-3' 5'-GCAGAGTTTGCTGTCAATTTCA-3'
<i>ACTIN</i>	Le-Actin-5'Q Le-Actin-3'Q	97.9	100	5'-AAATTGTCAGGGACGTGAAAGAA-3' 5'-TCTCAACAGAAGAGCTGGTCTTTG-3'
<i>GAPDH</i>	Le-GAPDH-5' Le-GAPDH-3'	100	97.2	5'-GGCATTGTTGAGGGTCTCATG-3' 5'-ATGGATGGACCATCAACAGTTTTC-3'
Semi-quantitative PCR/VIGS cloning				
<i>IPMS A and B</i>	IPMS2X_FOR IPMS2X_REV	+	+	5'-GGGGCTCGAGCCTGATACTGTTGGATACAC-3' 5'-GGTTGGATCCGTGTTTTAACATTCCATCC-3'
<i>BCKD E1-β</i>	BOXD1_FOR BOXD1_REV	+	+	5'-GGCGCTCGAGTCGTTCTTATGTATTGGAGAAG-3' 5'-GCCCGGATCCTCCAACAGCTCCATAAGGT-3'
<i>BCKD E1-α I</i>	AOXDN2_FOR AOXDN2_REV	+	-	5'-GGGCTCGAGCATTTTCCGGTAAGTCTG-3' 5'-GCCTGGATCCCTTCTCTCCAGCAGTT-3'
<i>BCKD E1-α I</i>	AOXDC1_FOR AOXDC1_REV	-	+	5'-GCCCTCGAGTAGCCACCATTAACA-3' 5'-GGTTGGATCCTTTAATGGCACTTTGTCTATAA-3'
<i>BCKD E1-α II</i>	AOXDN1_for AOXDN1_rev	-	+	5'-GGGGCTCGAGATTTTCAATCTTTGGTTT-3' 5'-GCCTGGATCCGATAGCTTCTTCCAACT-3'
<i>BCKD E1-α III</i>	AOXDN3_FOR AOXDN3_REV	-	+	5'-GGGCTCGAGCTTTTCCGGCAAGTCTA-3' 5'-GGTTGGATCCCTTCCAAAAGTGGTCA-3'
<i>KAS I A</i>	5KASI946for 6KASI946rev	-	+	5'-CGGGCTCGAGATGCAAGCTATCCAATCTT-3' 5'-CCCTGGATCCCCCACTTTACCAGCT-3'
<i>KAS I B</i>	7KASI966for 8KASI966rev	-	+	5'-GCGCCTCGAGTTGGAAGCAATCCAATCTC-3' 5'-TGCCGGATCCTCCCACTTTACCAGCC-3'
<i>KAS I C</i>	9KASI565for 10KASI565rev	-	+	5'-CGGGCTCGAGTCAATGGTGGATCTCATTTA-3' 5'-GCCTGGATCCTAAACTGATTCCACTCTGC-3'
<i>KAS II A</i>	21KASII174f 22KASII174r	-	+	5'-GGGGCTCGAGTGCTCATCCTGGCAAAC-3' 5'-CCCTGGATCCCATGAATTTATCTGCTCTTTTC-3'
<i>KAS II B</i>	17KASII868f 18KASII868r	-	+	5'-GGGGCTCGAGTCTTATGCTCAAACAGGG-3' 5'-TCCGGGATCCATTCTTTTAGAAAAGTTGGGTA-3'
<i>KAS II C</i>	11KASI820for 12KASI820rev	-	+	5'-GGCCCTCGAGTTGAATCAGATATCGGATCA-3' 5'-GCGGGGATCCGGCCATAATTCATCGATAT-3'
<i>KAS II D</i>	19KASII241f 20KASII241r	-	+	5'-GGGGCTCGAGGGCAGTGCAAGACAAGGT-3' 5'-GCCTGGATCCGATGAATGTCCATTCCG-3'
<i>KAS II E</i>	13KASX458for 14KASX458rev	-	+	5'-GGCCCTCGAGTCAATTTCCAAGCAGTTGTG-3' 5'-GCGGGGATCCGCTCTAAGAAGGAGCGCTT-3'
<i>HR7</i>	HR7_for HR7_rev	-	+	5'-GATAAATACATCAAGTGAAGTCA-3' 5'-CATTTTCCAATCAAAGTAGCATCA-3'
<i>ACTIN</i>	Le-Actin-5' Le-Actin-3'	+	+	5'-TGTGCTGGACTCTGGAGATG-3' 5'-CAGCTTCCATTCCGATCATT-3'
<i>18S</i>	18SrRNA-5' 18SrRNA-3'	+	+	5'-AAACGGCTACCACATCCAAG-3' 5'-TCATTACTCCGATCCCGAAG-3'

List of primers used for RT-PCR analyses and gene cloning. Gene accessions used for primer design are listed in Table S7. Primer pair efficiencies are indicated for real-time RT-PCR primers with cDNA from both species and effectiveness is indicated (+) for other primers.