Supplemental Data

E2A Proteins Promote Development

of Lymphoid-Primed Multipotent Progenitors

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Figure S1. E2A protein in multipotent progenitors

Intracellular staining for the E2A protein E47 in WT HSCs (LSK Flt3⁻), MPPs (LSK Flt3^{lo}), LMPPs (LSK Flt3^{hi}), CLPs (LSK^{lo}IL-7R α ⁺Flt3⁺), and myeloid progenitors (MyP, LS⁻K expressing Fc γ RII/III), as indicated by the black line. Secondary antibody only control is shown by the dashed line.





Figure S2. E2A proteins are dispensable for the generation of CMP and MEP.

Number of CMPs (LS⁻K Fc γ RII/III^{lo}CD34⁺) and MEPs (LS⁻K Fc γ RII/III^{lo}CD34⁻) in *Tcfe2a^{+/-}* (grey) and *Tcfe2a^{-/-}* (white) mice relative to WT (black; set to 1). A minimum of 6 mice were analyzed in each group; bars represent the mean ± SD; * *p*<0.05.



Figure S3. Comparative analysis of the $Tcfe2a^{+/+}$ and $Tcfe2a^{-/-}$ LMPP transcriptome Clusters show genes that are differentially expressed in replicate samples (one per column) of $Tcfe2a^{+/+}$ and $Tcfe2a^{-/-}$ LMPPs and expression of these genes in $Tcfe2a^{+/+}$ HSCs is shown for comparison. The clustering includes all genes with expression levels >50 in at least one of the four LMPP arrays and differing by 50% (using a lower 90% confidence bound of fold change). Each row corresponds to one unique identifier and numbers are the raw expression values. The right-most clustering shows the lineage

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association of these differentially expressed genes in LT-HSC, preGM, CLP, MkP and preCFU-E (as defined in (Pronk et al., 2007b)). Red indicates high, blue low, and white intermediate expression levels.



Figure S4. $Tcfe2a^{-/-}$ LMPPs have a higher GM clonogenic potential than $Tcfe2a^{+/+}$ LMPPs.

(A) Total number of GM colonies generated per 180 single MPPs or LMPP plated from mice of the indicated *Tcfe2a* genotype. Data from 3 independent experiments is shown. (B) Wright-Giemsa staining of a GM $Tcfe2a^{+/+}$ (left) and $Tcfe2a^{-/-}$ (right) clone. G = granulocyte; M = macrophage. $Tcfe2a^{-/-}$ derived G frequently have an atypical morphology consistent with apoptosis.

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	Population	<u>Markers</u>	Lineage
Thymus	ETP	Lin` <i>c-kit^{high}</i> CD25 ⁻	CD3ε, CD8α, TCRβ, TCRγδ, NK1.1, CD11c, Ter119, CD11b, Ly-6G, B220, CD19
	DN2	Lin ⁻ <i>c-kit</i> ^{high} CD25 ⁺	
	DN3	Lin⁻ <i>c-kit</i> CD25⁺	
Bone	HSC LSK Flt3 ⁻ CD3ε, CD4, CD8	CD3ε, CD4, CD8α, NK1.1,	
Marrow	MPP	LSK FIt3 ^{low}	Ter119, CD11b, Ly-6G, B220 CD3ε, CD4, CD8α, NK1.1, Ter119, Ly-6G, B220, CD19, IgM, IL-7Rα
	LMPP	LSK FIt3 ^{high}	
	CLP	LS ^{low} K ^{low} IL-7Rα⁺Flt3⁺	
	CMP	LS ⁻ K FcgRII/III ^{low} CD34⁺	
	MEP	LS ⁻ K FcgRII/III ^{Iow} CD34 ⁻	
Fetal Liver	MPP	Lin ⁻ <i>c-kit</i> ^{high} CD27 ⁺	Ter119, Ly-6G

Table S1. Surface markers for the progenitor populations studied.

Antibody clones: CD3ε, CD4 (GK1.5), CD8α (53-6.7), TCRβ (H57-597), TCRγδ (UC7-13D5), NK1.1 (PK136), CD11c (HL3), Ter-119, FcγIII/IIR (2.4G2), CD11b (M1/70), Ly-6G (RB6-8C5), B220 (RA3-6B2), CD19 (1D3), IgM (R6-60.2), IL-7Rα (A7R34), CD25 (PC61.5), CD27 (LG. 3A10), c-*kit* (2B8), Sca-1 (D7), Flt3 (A2F10), CD34 (RAM34).

Table S2. QPCR primers.

mGene	Sequence
Hprt	For 5' ACCTCTCGAAGTGTTGGATA
	Rev 5' CAACAACAAACTTGTCTGGA
lkzf1	For 5' CAATGTCGCCAAACGTAAGA
	Rev 5' GTTGATGGCATTGTTGATGG
Sfpi1	For 5' GCTTCCCTTATCAAACCTTGTCCC
	Rev 5' GGCGAATCTTTTTCTTGCTGC
Gfi1	For 5' TCCGAGGGTCCAAACATCG
	Rev 5' TTGAAAGGCAGCGTGTAGGG
Notch1	For 5' GCATTGATGATGTCGCTGGATAC
	Rev 5' GCATACCCCGCTGTTTTT
Ccr9	For 5' CAATCTGGGATGAGCCTAAACAAC
	Rev 5' ACCAAAAACCAACTGCTGCG