

Zirconium Bis(Amido) Catalysts for Asymmetric Intramolecular Alkene Hydroamination

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Supporting Information

Index:	Page
General Experimental Details and Procedures	S2
Preparation of Diarylphosphinic Acids	S3
Preparation of Diphosphinic Amide Ligands	S5
Procedures for Cyclization Experiments	S11
Spectral and Chromatographic Data	S15

General Experimental Details.

“Oven-dried” glassware was dried at 180 °C for at least 12 h before use. THF, CH₂Cl₂, and PhMe were dried according to published procedures.¹ Triethylamine, pyridine, F-Ph, and Cl-Ph were distilled from CaH₂ and deoxygenated prior to use. 1,4-Dioxane was distilled from Na/benzophenone ketyl prior to use. For in situ NMR experiments, C₆D₆ and d₈-toluene were sparged with N₂ and stored over 4Å molecular sieves prior to use. Ligands were dried under high vacuum (*ca.* 20 mtorr) for at least 12 h prior to use. Hydroamination substrates were prepared according to literature procedures.² All other reagents and solvents were purchased at highest commercial quality and used as received. Unless otherwise noted, reactions were run using standard Shlenk line techniques. Where noted a nitrogen filled glovebox was used.

General Procedures:

General Procedure 1. Preparation of Diarylphosphinic Acids. In an oven-dried flask protected by an atmosphere of N₂, a rapidly stirred solution of aryl bromide in THF was treated dropwise with a solution of either ¹PrMgCl or ^tBuLi at -78 °C. The solution was stirred for 10 min, warmed to 25 °C and stirred for 30 min. The solution was re-cooled to -78 °C and treated with 0.5 equivalents Cl₂P(O)NMe₂ in one portion. The reaction mixture was warmed to 25 °C and stirred for 12 h. The flask was opened to the atmosphere and treated with 3 equivalents of 6 M aqueous HCl. The resulting suspension was rapidly stirred for 3–12 h until the reaction was completed as judged by TLC or ³¹P NMR analysis³ and then worked up and purified as indicated.

General Procedure 2. Preparation of Diphosphinic Amide Ligands from Chlorodiarylphosphines or Chlorodialkylphosphines. In an oven-dried 20 mL vial protected by an atmosphere of N₂ and equipped with a septum and a magnetic stirbar, R₂PCl (2.1 equiv) was added to a solution of chiral diamine (1 equiv) and Et₃N (5 equiv) in CH₂Cl₂ at 25 °C. The reaction mixture was stirred for 12 h, opened to the atmosphere, and cooled to 0 °C with an ice bath. A solution of either *t*-BuOOH (5.5 M in decane, 3-4 equiv) or H₂O₂ (30% aqueous, 3-4 equiv) was slowly added (note: oxidation is exothermic). The reaction mixture was stirred at 0 °C for 10 min, then at 25 °C for 30 min before being quenched with aqueous Na₂S₂O₃ (1 M, 10 equiv). The resulting biphasic solution was extracted with CH₂Cl₂ and the combined extracts were dried (Na₂SO₄) and concentrated in vacuo. The product was purified as specified below.

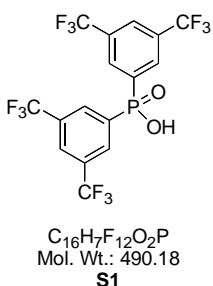
¹ Pangborn, A. B.; Giardello, M. A.; Grubbs, R. H.; Rosen, R. K.; Timmers, F. J. *Organometallics* **1996**, *15*, 1518.

² (a) Tamaru, Y.; Hojo, M.; Higashimura, H.; Yoshida, Z. *J. Am. Chem. Soc.* **1988**, *110*, 3994–4002. (b) Harding, K. E.; Burks, S. R. *J. Org. Chem.* **1981**, *46*, 3920–3922. (c) Hurd, C. D.; Jenkins, W. W. *J. Org. Chem.* **1957**, *22*, 1418–1423.

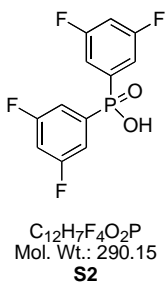
³ Monitoring by ³¹P NMR was readily accomplished by transferring a 200 µL sample of the biphasic solution into an NMR tube containing 0.6 mL CDCl₃ and obtaining a spectrum directly of the solution once the water layer had risen to top of the biphasic solution.

General Procedure 3. Preparation of Diphosphinic Amide Ligands from Diarylphosphinic Acids. In an oven-dried Schlenk flask protected by an atmosphere of N₂ and equipped with a reflux condenser, stirbar and nitrogen inlet, the diarylphosphinic acid was dissolved in an excess of SOCl₂ and then heated at reflux for 30 min. The reaction mixture was cooled to 25 °C, the reflux condenser was replaced with a greased glass stopper and the remaining SOCl₂ was removed in vacuo. The resulting phosphinic chloride was dried under high vacuum (*ca.* 100 mm Hg) for at least 12 h before continuing. The glass stopper was then replaced with a rubber septum and the phosphinic chloride was dissolved in CH₂Cl₂. Triethylamine (2.5 equiv) and chiral diamine (0.5 equiv) were then added and the reaction mixture was stirred for another 12 h. The flask was then opened to the atmosphere and worked up by the addition of saturated aqueous NaHCO₃. Extraction with CH₂Cl₂, followed by drying the combined extracts (Na₂SO₄) and concentration in vacuo provided the crude ligand, which was purified as described below.

Preparation of Diarylphosphinic Acids:



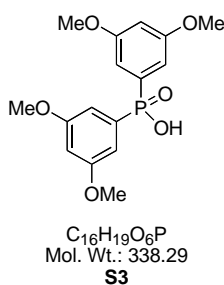
Bis(3,5-bis(trifluoromethyl)phenyl)phosphinic Acid (S1): 3,5-Bis(trifluoromethyl)phenyl bromide (18.09 g, 61.7 mmol, 10.5 mL), isopropyl magnesium chloride (34.1 mL, 61.7 mmol, 1.8 M in Et₂O), *N,N*-dimethylphosphoramidic dichloride (5.0 g, 30.9 mmol, 3.67 mL), 6 M aqueous HCl (30 mL) and THF (150 mL) were combined as described in General Procedure 1. The reaction mixture was then extracted with CH₂Cl₂, and the combined extracts were dried over Na₂SO₄, concentrated in vacuo and purified by precipitation from THF/hexane solution to give 6.50 g (43%) of **S1** as an off-white powder: IR (solid) cm⁻¹ 1281, 1135, 674; ¹H NMR (400 MHz, d₆-DMSO) δ 8.39 (d, *J* = 11.7 Hz, 4 H), 8.23 (s, 4 H), 6.69 (br. s, 1 H); ¹³C NMR (101 MHz, d₆-DMSO) δ 138.5 (d, *J* = 135.7 Hz), 132.2-132.6 (m), 130.9 (qd, *J* = 33.0, 13.2 Hz), 125.8-126.1 (m), 123.4 (q, *J* = 272.9 Hz); ¹⁹F NMR (376 MHz, d₆-DMSO) δ -60.7; ³¹P NMR (162 MHz, d₆-DMSO) δ 14.0; HRMS (ESI⁺) *m/z* 575.9991 [575.9975 calcd for C₁₆H₇F₁₂O₂P (M+2Na-H+CH₃CN)⁺].⁴



Bis(3,5-difluorophenyl)phosphinic Acid (S2): 3,5-Difluorophenyl bromide (7.72 g, 40.0 mmol, 4.6 mL), isopropylmagnesium chloride (22.1 mL, 40 mmol, 1.8 M in Et₂O), *N,N*-dimethylphosphoramidic dichloride (3.8 g, 30.9 mmol, 2.99 mL), 6 M aqueous HCl (20 mL) and THF (120 mL) were combined as described in General Procedure 1. The reaction mixture was extracted with CH₂Cl₂, dried over Na₂SO₄ and concentrated in vacuo. The product was purified by precipitation from CH₂Cl₂/hexane solution to give 1.35 g (10%) of **S2** as a colorless

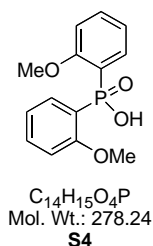
⁴ Acetonitrile from electrospray mass spectrometer.

powder: IR (solid) cm^{-1} 1584, 1421, 1928, 977; ^1H NMR (400 MHz, d_6 -DMSO) δ 8.77 (br. s, 1 H), 7.39-7.57 (m, 6 H); ^{13}C NMR (101 MHz, d_6 -DMSO) δ 162.7 (ddd, $J = 250.3, 20.5, 11.0$ Hz), 139.2 (dt, $J = 138.3, 5.9$ Hz), 114.7 (ddd, $J = 18.3, 10.2, 8.1$ Hz), 107.9 (t, $J = 24.9$ Hz); ^{19}F NMR (376 MHz, d_6 -DMSO) δ -107.1; ^{31}P NMR (162 MHz, d_6 -DMSO) δ 16.9 (t, $J = 6.9$ Hz); HRMS (ESI $^+$) m/z 376.0103 [376.0108 calcd for $\text{C}_{12}\text{H}_7\text{F}_4\text{O}_2\text{P}$ ($\text{M}+2\text{Na}-\text{H}+\text{CH}_3\text{CN}$) $^+$].⁴



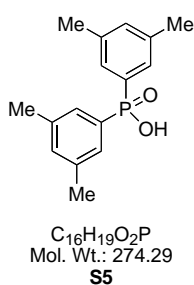
Bis(3,5-dimethoxyphenyl)phosphinic Acid (S3): 3,5-Dimethoxyphenyl bromide (4.34 g, 20 mmol), *t*-BuLi (24.2 mL, 40 mmol, 1.65 M in pentane), *N,N*-dimethylphosphoramidic dichloride (1.61 g, 10 mmol, 1.19 mL), 6 M aqueous HCl (10 mL) and THF (100 mL) were combined as described in General Procedure 1 to give a thick, white heterogeneous suspension. The suspension was filtered, and the precipitate was washed with water (50 mL) and Et₂O (200 mL) to give 2.0 g (74%) of **S3** as a colorless powder: IR (solid) cm^{-1} 1607, 1409, 1199, 1152; ^1H

NMR (400 MHz, d_6 -DMSO) δ 6.78 (dd, $J = 13.2, 2.2$ Hz, 4 H), 6.61 (t, $J = 2.2$ Hz, 2 H), 3.72 (s, 12 H); ^{13}C NMR (101 MHz, d_6 -DMSO) δ 160.8 (d, $J = 18.3$ Hz), 137.3 (d, $J = 134.2$ Hz), 108.9 (d, $J = 10.3$ Hz), 103.4 (d, $J = 1.5$ Hz), 55.8; ^{31}P NMR (162 MHz, d_6 -DMSO) δ 23.0; HRMS (ESI $^+$) m/z 383.0636 [383.0636 calcd for $\text{C}_{16}\text{H}_{19}\text{O}_6\text{P}$ ($\text{M}+2\text{Na}-\text{H}$) $^+$].



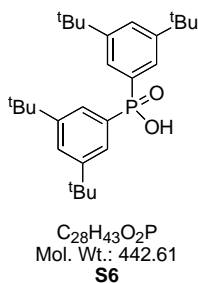
Di(o-methoxyphenyl)phosphinic Acid (S4): *O*-methoxyphenyl bromide (3.74 g, 20 mmol), *t*-BuLi (24.2 mL, 40 mmol, 1.65 M in pentane), *N,N*-dimethylphosphoramidic dichloride (1.61 g, 10 mmol, 1.19 mL), 6 M aqueous HCl (10 mL) and THF (100 mL) were combined as described in General Procedure 1 to give a thick, white heterogeneous suspension. The suspension was filtered, and the precipitate was washed with water (50 mL) and Et₂O (200 mL) to give 2.2 g (79%) of **S4** as a colorless powder: IR (solid) cm^{-1} 1590, 1479, 1275, 1441, 948; ^1H NMR

(400 MHz, d_6 -DMSO) δ 8.78 (br. s, 1 H), 7.71 (ddd, $J = 13.7, 7.3, 1.5$ Hz, 2 H), 7.43 (t, $J = 7.3$ Hz, 2 H), 6.99 (td, $J = 7.1, 1.7$ Hz, 2 H), 6.92 (dd, $J = 7.8, 5.9$ Hz, 2 H), 3.47 (s, 6 H); ^{13}C NMR (101 MHz, d_6 -DMSO) δ 160.8 (d, $J = 3.7$ Hz), 133.7 (d, $J = 6.6$ Hz), 133.4 (d, $J = 1.5$ Hz), 123.7 (d, $J = 141.6$ Hz), 120.3 (d, $J = 11.7$ Hz), 112.0 (d, $J = 7.3$ Hz), 55.8; ^{31}P NMR (162 MHz, d_6 -DMSO) δ 19.1; HRMS (ESI $^+$) m/z 323.0434 [323.0425 calcd for $\text{C}_{14}\text{H}_{15}\text{O}_4\text{P}$ ($\text{M}+2\text{Na}-\text{H}$) $^+$].



Bis(3,5-dimethylphenyl)phosphinic Acid (S5): 3,5-Dimethylphenyl bromide (10.0 g, 54 mmol), *t*-BuLi (65.4 mL, 108 mmol, 1.65 M in pentane), *N,N*-dimethylphosphoramidic dichloride (4.32 g, 27 mmol, 3.21 mL), 6 M aqueous HCl (30 mL) and THF (200 mL) were combined as described in General Procedure 1. The reaction mixture was then extracted with CH₂Cl₂, dried over Na₂SO₄ and concentrated in vacuo. The product was purified by precipitation from CH₂Cl₂/hexane solution to give 5.65 g (76%) of **S5** as a colorless powder: IR (solid) cm^{-1} 2915, 1584, 1275, 1135, 937, 685;

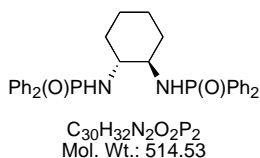
^1H NMR (400 MHz, CD_2Cl_2) δ 12.68 (br. s, 1 H), 7.32 (d, $J = 12.7$ Hz, 4 H), 7.10 (s, 2 H), 2.27 (s, 12 H); ^{13}C NMR (101 MHz, CD_2Cl_2) δ 138.1 (d, $J = 13.9$ Hz), 133.5 (d, $J = 2.9$ Hz), 132.6 (d, $J = 137.9$ Hz), 128.5 (d, $J = 10.3$ Hz), 20.9; ^{31}P NMR (162 MHz, CD_2Cl_2) δ 33.4; HRMS (ESI $^+$) m/z 319.0840 [319.0840 calcd for $\text{C}_{16}\text{H}_{19}\text{O}_2\text{P}$ ($\text{M}+2\text{Na}-\text{H}$) $^+$].



Bis(3,5-di-*t*-Bu-phenyl)phosphinic Acid (S6): 3,5-Di-*t*-Bu-phenyl bromide (7.56 g, 28 mmol), *t*-BuLi (39.2 mL, 56 mmol, 1.43 M in pentane), *N,N*-dimethylphosphoramidic dichloride (2.67 g, 14 mmol, 1.27 mL), 6 M aqueous HCl (15 mL) and THF (100 mL) were combined as described in General Procedure 1. The reaction mixture was then extracted with CH_2Cl_2 , dried over Na_2SO_4 and concentrated in vacuo. The product was purified by flash chromatography (Et $_2$ O/hex) to give 5.44 g (87%) of **S6** as an off-white foamy solid: IR (solid) cm^{-1} 2961, 2856, 1590, 1357, 1141, 960; ^1H NMR (400

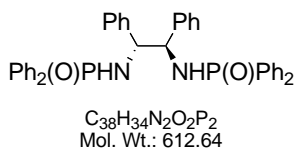
MHz, CD_2Cl_2) δ 7.90 (br. s, 1 H), 7.67 (dd, $J = 13.4, 1.8$ Hz, 4 H), 7.61 (s, 2 H), 1.31 (s, 36 H); ^{13}C NMR (101 MHz, CD_2Cl_2) δ 151.0 (d, $J = 13.2$ Hz), 131.8 (d, $J = 139.0$ Hz), 126.3 (d, $J = 2.2$ Hz), 125.1 (d, $J = 11.7$ Hz), 34.9, 31.1; ^{31}P NMR (162 MHz, d_6 -DMSO) δ 36.5; HRMS (ESI $^+$) m/z 487.2708 [487.2718 calcd for $\text{C}_{28}\text{H}_{43}\text{O}_2\text{P}$ ($\text{M}+2\text{Na}-\text{H}$) $^+$].

Preparation of Diphosphinic Amide Ligands **3c**, **9-12** and **13a-13i**:

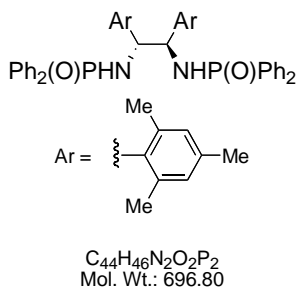


(*R,R*)-Diphosphinic Amide Ligand **3c:** In an oven-dried 20 mL vial protected by an atmosphere of N_2 and equipped with a septum and a magnetic stirbar, diphenylphosphinic chloride (2.07 g, 3.05 mL, 8.75 mmol) was added dropwise to a solution of (*R,R*)-1,2-cyclohexanediamine (500 mg, 4.78 mmol) and Et_3N (2.21 g, 3.05 mL, 21.9 mmol) in CH_2Cl_2 (10 mL) at 0 $^\circ\text{C}$.

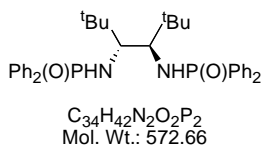
The reaction mixture was warmed to 25 $^\circ\text{C}$ and stirred for 2 h, during which time colorless precipitates were observed. The reaction mixture was quenched by the careful addition of saturated aqueous NaHCO_3 and extracted with CH_2Cl_2 . The combined extracts were dried over Na_2SO_4 and concentrated in vacuo to give a yellow foam. The product was purified by precipitation from EtOAc/hex solution to give 1.66 g (74%) of **3c** as an extremely fluffy white solid: IR (solid) cm^{-1} 3181, 2922, 1434, 1185; ^1H NMR (400 MHz, CD_2Cl_2) δ 7.99 (ddd, $J = 11.9, 6.8, 1.3$ Hz, 4 H), 7.81-7.88 (m, $J = 11.6, 7.1$ Hz, 4 H), 7.46-7.59 (m, 8 H), 7.39 (td, $J = 7.6, 3.0$ Hz, 4 H), 4.42 (t, $J = 7.1$ Hz, 2 H), 2.83-2.97 (m, 2 H), 2.07 (d, $J = 13.4$ Hz, 2 H), 1.58 (d, $J = 7.8$ Hz, 2 H), 1.32 (q, $J = 10.4$ Hz, 2 H), 1.12 (t, $J = 9.9$ Hz, 2 H); ^{13}C NMR (101 MHz, CD_2Cl_2) δ 133.66 (d, $J = 122.9$ Hz), 133.62 (d, $J = 129.2$ Hz), 131.58 (d, $J = 2.9$ Hz), 131.55 (d, $J = 2.9$ Hz), 131.51 (d, $J = 9.5$ Hz), 132.4 (d, $J = 9.5$ Hz), 128.34 (d, $J = 12.4$ Hz), 128.31 (d, $J = 12.4$ Hz), 56.1, 36.0 (d, $J = 4.4$ Hz), 25.0; ^{31}P NMR (162 MHz, CD_2Cl_2) δ 24.3; HRMS (FAB $^+$) m/z 515.2010 [515.2017 calcd for $\text{C}_{30}\text{H}_{32}\text{N}_2\text{O}_2\text{P}_2$ ($\text{M}+\text{H}$) $^+$].



(*R,R*)-Diphosphinic Amide Ligand 9: In an oven-dried 20 mL vial protected by an atmosphere of N_2 and equipped with a septum and a magnetic stirbar, diphenylphosphinic chloride (111 mg, 90 μ L, 470 μ mol) was added dropwise to a solution of (*R,R*)-1,2-diphenylethylenediamine (50 mg, 235 μ mol) and Et_3N (120 mg, 160 μ L, 1.17 mmol) in CH_2Cl_2 (5 mL) at 25 $^\circ C$. The reaction mixture was stirred for 12 h, during which time colorless precipitates were observed. The reaction mixture was quenched by the careful addition of saturated aqueous $NaHCO_3$ and extracted with CH_2Cl_2 . The combined extracts were dried over Na_2SO_4 and concentrated to give a yellow foam. The product was purified by precipitation from CH_2Cl_2 /hexanes solution to give 140 mg (97%) of **9** as a white powder: IR (solid) cm^{-1} 3210, 3056, 1434, 1188, 1108; 1H NMR (500 MHz, CD_2Cl_2) δ 7.77 (dd, $J = 11.3, 7.9$ Hz, 4 H), 7.61 (dd, $J = 11.3, 7.9$ Hz, 4 H), 7.53 (t, $J = 7.0$ Hz, 2 H), 7.33-7.45 (m, 8 H), 7.18-7.28 (m, 4 H), 7.07 (d, $J = 7.3$ Hz, 4 H), 6.88 (d, $J = 6.4$ Hz, 4 H), 6.19 (br. s, 2 H), 4.26-4.37 (m, 2 H); ^{13}C NMR (126 MHz, CD_2Cl_2) δ 132.70 (d, $J = 126.2$ Hz), 132.69 (d, $J = 9.9$ Hz), 131.9 (d, $J = 133.8$ Hz), 131.7 (d, $J = 2.4$ Hz), 131.53 (d, $J = 9.4$ Hz), 131.48 (d, $J = 2.4$ Hz), 128.2 (d, $J = 12.7$ Hz), 128.07, 128.05 (d, $J = 13.2$ Hz), 127.8, 127.5, 126, 61.7; ^{31}P NMR (162 MHz, CD_2Cl_2) δ 27.8; HRMS (FAB $^+$) m/z 613.2188 [613.2173 calcd for $C_{38}H_{34}N_2O_2P_2$ (M+H) $^+$].

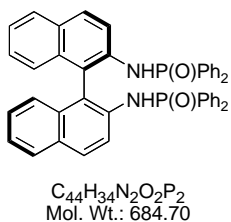


(*R,R*)-Diphosphinic Amide Ligand 10: (*R,R*)-1,2-dimesitylethylenediamine (75 mg, 252 μ mol), chlorodiphenylphosphine (117 mg, 98 μ L, 531 μ mol), Et_3N (128 mg, 176 μ L, 1.26 mmol), aqueous H_2O_2 (30%, 171 μ L, 1.51 mmol) and CH_2Cl_2 (4 mL) were combined according to General Procedure 2 to produce a crude oil. The product was isolated by flash chromatography (65:35 Et_2O /hexanes) to provide 152 mg (87%) of **10** as a colorless solid: IR (solid) cm^{-1} 3199, 3064, 2965, 1617, 1434, 1185; 1H NMR (500 MHz, CD_2Cl_2) δ 7.85 (dd, $J = 11.3, 7.9$ Hz, 4 H), 7.72 (dd, $J = 11.6, 7.9$ Hz, 4 H), 7.55 (t, $J = 7.3$ Hz, 2 H), 7.39-7.50 (m, 10 H), 7.28 (td, $J = 7.6, 3.1$ Hz, 4 H), 6.78 (s, 2 H), 6.30 (s, 2 H), 5.91 (d, $J = 9.2$ Hz, 2 H), 4.80-4.92 (m, 2 H), 2.68 (s, 6 H), 2.14 (s, 6 H), 0.73 (s, 6 H); ^{13}C NMR (125 MHz, CD_2Cl_2) δ 136.9, 136.6, 134.2 (d, $J = 6.6$ Hz), 133.2 (d, $J = 125.7$ Hz), 132.9 (d, $J = 9.4$ Hz), 132.3 (d, $J = 133.9$ Hz), 131.7 (d, $J = 7.5$ Hz), 131.6 (d, $J = 7.5$ Hz), 131.5 (d, $J = 9.4$ Hz), 130.8, 128.32, 128.31 (d, $J = 12.2$ Hz), 128.2 (d, $J = 12.3$ Hz), 53.7 (d, $J = 1.9$ Hz), 21.4, 20.3, 18.5; ^{31}P NMR (202 MHz, CD_2Cl_2) δ 26.7; HRMS (FAB $^+$) m/z 697.3123 [697.3113 calcd for $C_{62}H_{96}N_2O_2P_2$ (M+H) $^+$].



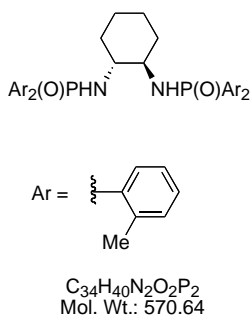
(R,R)-Diphosphinic Amide Ligand 11: (*R,R*)-1,2-di-*t*-Bu-ethylenediamine⁵ (100 mg, 577 μ mol), chlorodiphenylphosphine (380 mg, 320 μ L, 1.73 mmol), Et₃N (290 mg, 400 μ L, 2.88 mmol), *t*-BuOOH (5.5 M in decane, 420 μ L, 2.30 mmol) and CH₂Cl₂ (6 mL) were combined according to

General Procedure 2 to produce a crude oil. The product was isolated by flash chromatography (40:60 EtOAc/hexanes) then crystallized from hexanes to provide 90 mg (27%) of **11** as colorless needles: IR (solid) cm⁻¹ 3350, 3280, 2958, 1442, 1108; ¹H NMR (400 MHz, CD₂Cl₂) δ 7.89 (dd, *J* = 11.6, 10.4 Hz, 4H), 7.84 (dd, *J* = 12.0, 10.4 Hz, 4H), 7.39–7.52 (m, 8H), 7.26 (t, *J* = 7.6 Hz, 4H), 4.63 (dd, *J* = 12.0, 5.2 Hz, 2H), 3.06 (t, *J* = 9.2 Hz, 2H), 0.88 (s, 18H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 135.7 (d, *J* = 127.0 Hz), 132.5 (d, *J* = 129.0 Hz), 132.4 (d, *J* = 10 Hz), 131.5, 131.3 (d, *J* = 9.0 Hz), 131.2, 128.3 (d, *J* = 13.0 Hz), 128.2 (d, *J* = 12 Hz), 58.6 (d, *J* = 4.0 Hz), 36.8 (d, *J* = 9.0 Hz), 28.1; ³¹P NMR (162 MHz, CD₂Cl₂) δ 21.2; HRMS (FAB⁺) *m/z* 573.2801 [573.2800 calcd for C₃₄H₄₂N₂O₂P₂ (M⁺)].



(R,R)-Diphosphinic Amide Ligand 12: (*R*)-BINAM (56 mg, 196 μ mol), chlorodiphenylphosphine (91 mg, 77 μ L, 413 μ mol), Et₃N (100 mg, 137 μ L, 984 μ mol), *t*-BuOOH (5.5 M in decane, 200 μ L) and CH₂Cl₂ (4 mL) were combined according to General Procedure 2 to produce a crude oil. The product was isolated by flash chromatography (50:50 Et₂O/hexanes) and then crystallized

by slow diffusion of hexanes into a toluene solution to provide 94 mg (70%) of **12** as colorless prisms: IR (solid) cm⁻¹ 3354, 3058, 1623, 1583, 1211; ¹H NMR (500 MHz, CD₂Cl₂) δ 7.90 (d, *J* = 6.1 Hz, 2 H), 7.82 (d, *J* = 9.2 Hz, 2 H), 7.62–7.72 (m, 6 H), 7.31–7.50 (m, 18 H), 7.29 (d, *J* = 7.6 Hz, 2 H), 7.12–7.21 (m, 4 H); ¹³C NMR (125 MHz, CD₂Cl₂) δ 138.0, 132.7, 132.2 (d, *J* = 129.0 Hz), 132.1 (d, *J* = 128.0 Hz), 132.0 (d, *J* = 2.8 Hz), 131.9 (d, *J* = 2.4 Hz), 131.6 (d, *J* = 9.9 Hz), 131.0 (d, *J* = 9.9 Hz), 130.0, 129.8, 128.7 (d, *J* = 12.7 Hz), 128.6 (d, *J* = 12.5 Hz), 128.5, 127.5, 124.5, 124.4, 119.0 (d, *J* = 3.8 Hz), 116.9 (d, *J* = 8.0 Hz); ³¹P NMR (162 MHz, CD₂Cl₂) δ 15.8; HRMS (FAB⁺) *m/z* 685.2171 [685.2174 calcd for C₆₂H₉₆N₂O₂P₂ (M+H⁺)].

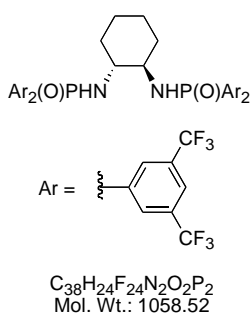


(R,R)-Diphosphinic Amide Ligand 13a: (*R,R*)-1,2-cyclohexanediamine (50 mg, 437 μ mol), chlorodi(*o*-tolyl)phosphine (228 mg, 919 μ mol), Et₃N (443 mg, 610 μ L, 4.37 mmol), *t*-BuOOH (5.5 M in decane, 310 μ L, 1.78 mmol) and CH₂Cl₂ (10 mL) were combined according to General Procedure 2 to produce a crude oil. The product was purified by precipitation from CH₂Cl₂/hexane solution to provide 92 mg (37%) of **13a** as a colorless powder: IR (solid) cm⁻¹ 3467, 3178, 2942, 2853, 1560, 1455, 1163; ¹H NMR (500 MHz, CD₂Cl₂) δ 7.98 (dd, *J* = 13.4, 7.6 Hz, 2 H), 7.59 (dd, *J* = 12.5, 7.9 Hz, 2 H),

7.35–7.44 (m, 4 H), 7.28–7.34 (m, 2 H), 7.08–7.22 (m, 6 H), 4.49–4.56 (m, 2 H), 3.13–3.23 (m, 2 H), 2.32 (s, 6 H), 2.24 (s, 6 H), 2.19 (d, *J* = 12.8 Hz, 2 H), 1.55–1.68 (m, 2 H), 1.40

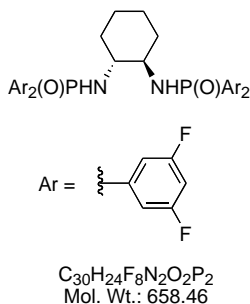
⁵ Roland, S.; Mangeney, P.; Alexakis, A. *Synthesis* **1999**, 228-230.

(q, $J = 10.1$ Hz, 2 H), 1.21 (t, $J = 9.8$ Hz, 2 H); ^{13}C NMR (125 MHz, CD_2Cl_2 , one aromatic signal partially obscured) δ 142.0 (d, $J = 10.8$ Hz), 141.4 (d, $J = 9.9$ Hz), 133.2 (d, $J = 9.9$ Hz), 132.7 (d, $J = 10.4$ Hz), 131.5 (d, $J = 118.2$ Hz), 131.43, 131.42, 131.41 (d, $J = 7.6$ Hz), 131.3 (d, $J = 6.1$ Hz), 125.2 (d, $J = 12.2$ Hz), 125.1 (d, $J = 12.2$ Hz), 56.4 (d, $J = 3.8$ Hz), 35.7 (d, $J = 2.8$ Hz), 25.1, 21.2 (d, $J = 3.8$ Hz), 21.1 (d, $J = 3.8$ Hz); ^{31}P NMR (162 MHz, CD_2Cl_2) δ 26.7; HMRS (FAB $^+$) m/z 571.2652 [571.2643 calcd for $\text{C}_{34}\text{H}_{40}\text{N}_2\text{O}_2\text{P}_2$ (M+H $^+$)].



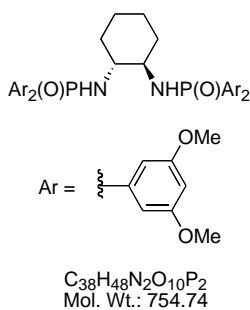
(*R,R*)-Diphosphinic Amide Ligand 13b: Bis(bis(3,5-trifluoromethyl)phenyl)phosphinic acid (1.0 g, 2.04 mmol), SOCl_2 (10 mL), (*R,R*)-1,2-cyclohexanediamine (116 mg, 1.02 mmol), Et_3N (516 mg, 710 μL , 5.10 mmol), and CH_2Cl_2 (10 mL) were combined according to General Procedure 3 to provide a light brown solid. The product was dissolved in Et_2O and filtered through a short plug of silica gel. The silica was washed with Et_2O , and the resulting solution was concentrated. The product was further purified by precipitation from Et_2O /hexanes solution to provide 618 mg (57%) of **13b** as a

colorless powder: IR (solid) cm^{-1} 3174, 2932, 1621, 1456, 1360, 1277, 1119; ^1H NMR (400 MHz, CD_2Cl_2) δ 8.45 (d, $J = 12.0$ Hz, 4 H), 8.32 (d, $J = 11.7$ Hz, 4 H), 8.08 (d, $J = 14.2$ Hz, 4 H), 4.80-4.98 (m, 2 H), 2.80-2.98 (m, 2 H), 1.92 (d, $J = 13.0$ Hz, 2 H), 1.60 (d, $J = 8.3$ Hz, 2 H), 1.33-1.47 (m, 2 H), 1.05 (t, $J = 9.5$ Hz, 2 H); ^{13}C NMR (101 MHz, CD_2Cl_2) δ 135.1 (d, $J = 135.0$ Hz), 134.7 (d, $J = 128.2$ Hz), 132.8-133.1 (m), 132.3 (t, $J = 12.5$ Hz), 131.9 (t, $J = 12.5$ Hz), 131.5-131.8 (m), 126.1-126.5 (m), 122.9 (q, $J = 272.2$ Hz), 122.7 (q, $J = 272.2$ Hz), 56.6, 35.6 (d, $J = 5.9$ Hz), 24.7; ^{31}P NMR (162 MHz, CD_2Cl_2) δ 20.2; ^{19}F NMR (376 MHz, CD_2Cl_2) δ -62.57, -62.65; LRMS (FAB $^+$) m/z 1059 (M+H $^+$).

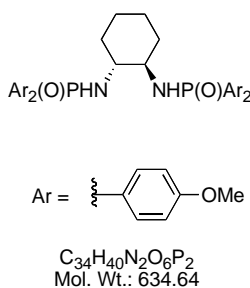


(*R,R*)-Diphosphinic Amide Ligand 13c: Bis(3,5-difluorophenyl)phosphinic acid (300 mg, 1.03 mmol), SOCl_2 (5 mL), (*R,R*)-1,2-cyclohexanediamine (59 mg, 516 μmol), Et_3N (253 mg, 350 μL , 2.5 mmol), and CH_2Cl_2 (5 mL) were combined according to General Procedure 3 to provide a light brown solid. The product was purified by flash chromatography (90:10 hexanes/ Et_2O) to provide 165 mg (49%) of **13c** as a colorless solid: IR (solid) cm^{-1} 3150, 2907, 2852, 1613, 1591,

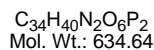
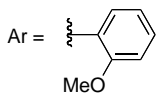
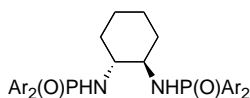
1420, 1298, 1999, 1121; ^1H NMR (400 MHz, CD_2Cl_2) δ 7.51 (ddt, $J = 12.9, 5.6, 2.3$ Hz, 4H), 7.02 (ddt, $J = 12.6, 5.6, 2.3$ Hz, 4H), 6.96-7.09 (m, 4 H), 4.79-4.87 (m, 2 H), 2.91-3.01 (m, 1 H), 2.02 (d, $J = 14.4$ Hz, 2 H), 1.59-1.68 (m, 2 H), 1.38 (q, $J = 9.3$ Hz, 6 H), 1.15 (t, $J = 10.1$ Hz, 2 H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 164.2 (dt, $J = 24.9, 11.7$ Hz), 161.7 (ddd, $J = 21.2, 13.2, 11.0$ Hz), 136.8 (dt, $J = 133.9, 7.3$ Hz), 136.2 (dt, $J = 126.8, 7.3$ Hz), 115.4 (ddd, $J = 18.3, 10.2$ Hz), 114.4 (dt, $J = 18.3, 10.3$ Hz), 107.5 (d, $J = 24.9$ Hz), 107.8 (d, $J = 25.6$ Hz), 56.2, 35.6 (d, $J = 4.4$ Hz), 24.9; ^{31}P NMR (162 MHz, CD_2Cl_2) δ 21.0 (td, $J = 13.8, 5.9$ Hz); ^{19}F NMR (376 MHz, CD_2Cl_2) δ -107.04 (q, $J = 8.2$ Hz), -107.26 (q, $J = 8.3$ Hz); HRMS (FAB $^+$) m/z 659.1255 [659.1264 calcd for $\text{C}_{30}\text{H}_{24}\text{F}_8\text{N}_2\text{O}_2\text{P}_2$ (M+H $^+$)].



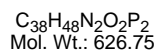
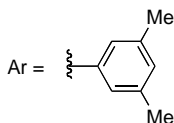
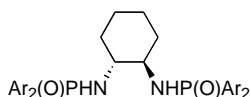
(*R,R*)-Diphosphinic Amide Ligand 13d: Bis(3,5-dimethoxyphenyl)phosphinic acid (676 mg, 2.0 mmol), $SOCl_2$ (10 mL), (*R,R*)-1,2-cyclohexanediamine (114 mg, 1.0 mmol), Et_3N (506 mg, 700 μ L, 5.0 mmol), and CH_2Cl_2 (10 mL) were combined according to General Procedure 3 to provide a light brown solid. The product was purified by flash chromatography ($EtOAc$) to provide 660 mg (89%) of **13d** as a tan foamy solid: IR (film) cm^{-1} 3174, 2930, 2833, 1587, 1447, 1423, 1151; 1H NMR (500 MHz, CD_2Cl_2) δ 7.15 (d, $J = 13.4$ Hz, 4 H), 7.02 (d, $J = 12.8$ Hz, 4 H), 6.61 (s, 2 H), 6.55 (s, 2 H), 4.57-4.67 (m, 2 H), 3.84 (s, 12 H), 3.75 (s, 12 H), 2.89-3.03 (m, 2 H), 2.11 (d, $J = 12.5$ Hz, 2 H), 1.52-1.63 (m, 2 H), 1.35 (q, $J = 9.2$ Hz, 2 H), 1.13 (t, $J = 8.9$ Hz, 2 H); ^{13}C NMR (125 MHz, CD_2Cl_2) δ 160.8 (d, $J = 18.4$ Hz), 135.6 (d, $J = 130.5$ Hz), 135.5 (d, $J = 123.9$ Hz), 109.9 (d, $J = 10.4$ Hz), 109.0 (d, $J = 10.8$ Hz), 103.8, 103.4, 56.2, 55.4, 35.7 (d, $J = 3.8$ Hz), 25.0; ^{31}P NMR (162 MHz, CD_2Cl_2) δ 24.9; HRMS (FAB $^+$) m/z 755.2869 [755.2862 calcd for $C_{38}H_{48}N_2O_{10}P_2$ ($M+H^+$)].



(*R,R*)-Diphosphinic Amide Ligand 13e: Using a modification of General Procedure 3, di(*p*-methoxyphenyl)phosphinic acid (2.5 g, 8.9 mmol) was heated at reflux in $SOCl_2$ (15 mL) for 1 h. After concentration, the resulting oil was distilled bulb-to-bulb (300 $^{\circ}C$, 0.4 mmHg) to provide *ca.* 2 g of the phosphinic chloride as a thick yellow oil, which was used without further purification. A portion of the phosphinic chloride (1.10 g, 3.70 mmol) was transferred to an oven-dried 100 mL round bottom flask via pipet and dissolved in CH_2Cl_2 (15 mL). Triethylamine (750 mg, 1.03 mL, 7.40 mmol) and (*R,R*)-1,2-cyclohexanediamine (212 mg, 1.89 mmol) were then added sequentially, and the reaction mixture was stirred for 12 h. Saturated aqueous $NaHCO_3$ (10 mL) was added to quench the reaction. Extraction with CH_2Cl_2 , followed by drying the combined extracts over Na_2SO_4 and concentration, gave an oily solid. The product was purified by precipitation from CH_2Cl_2/Et_2O solution to provide 1.05 g (89%) of **13e** as a fluffy, colorless solid: IR (solid) cm^{-1} 3192, 2921, 2844, 1595, 1500, 1159, 1122; 1H NMR (500 MHz, $CDCl_3$) δ 7.87 (dd, $J = 11.3, 8.9$ Hz, 4 H), 7.74 (dd, $J = 11.0, 8.9$ Hz, 4 H), 6.98 (d, $J = 6.4$ Hz, 4 H), 6.87 (d, $J = 6.7$ Hz, 4 H), 4.15-4.28 (m, 2 H), 3.85 (s, 6 H), 3.81 (s, 6 H), 2.71-2.87 (m, 2 H), 2.02 (d, $J = 12.5$ Hz, 2 H), 1.50-1.59 (m, 2 H), 1.21-1.31 (m, 2 H), 1.09 (t, $J = 9.5$ Hz, 2 H); ^{13}C NMR (126 MHz, $CDCl_3$) δ 162.2 (d, $J = 2.4$ Hz), 162.1 (d, $J = 2.8$ Hz), 134.1 (d, $J = 10.8$ Hz), 133.3 (d, $J = 10.8$ Hz), 125.4 (d, $J = 131.4$ Hz), 125.0 (d, $J = 135.0$ Hz), 113.7 (d, $J = 8.9$ Hz), 113.6 (d, $J = 8.5$ Hz), 55.9, 55.3, 55.2, 36.0 (d, $J = 3.3$ Hz), 25.1; ^{31}P NMR (162 MHz, $CDCl_3$) δ 24.4; HRMS (FAB $^+$) m/z 635.2441 [635.2440 calcd for $C_{34}H_{40}N_2O_6P_2$ ($M+H^+$)].

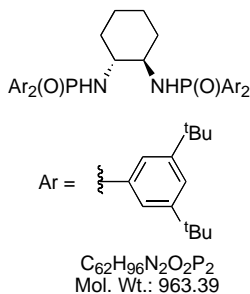


(*R,R*)-Diphosphinic Amide Ligand 13f: Di(*o*-methoxyphenyl)phosphinic acid (556 mg, 2.0 mmol), $SOCl_2$ (10 mL), (*R,R*)-1,2-cyclohexanediamine (114 mg, 1.0 mmol), Et_3N (506 mg, 700 μ L, 5.0 mmol), and CH_2Cl_2 (10 mL) were combined according to General Procedure 3 to provide a light brown solid. The product was purified by flash chromatography, first with $CH_2CH_2/MeOH$, then with acetone/ $EtOAc$ to provide 121 mg (19%) of **13f** as a tan foamy solid: IR (film) cm^{-1} 3367, 2939, 2853, 1591, 1479, 1279, 1243; 1H NMR (500 MHz, CD_2Cl_2) δ 7.87 (dd, $J = 13.7, 7.3$ Hz, 2 H), 7.73-7.80 (m, $J = 14.0, 7.6$ Hz, 2 H), 7.42 (t, $J = 7.3$ Hz, 2 H), 7.37 (t, $J = 7.6$ Hz, 2 H), 7.03 (t, $J = 7.0$ Hz, 2 H), 6.79-6.88 (m, 4 H), 6.74 (t, $J = 7.3$ Hz, 2 H), 4.04-4.19 (m, 2 H), 3.61 (s, 6 H), 3.48 (s, 6 H), 3.15-3.27 (m, 2 H), 2.03 (d, $J = 9.8$ Hz, 2 H), 1.50-1.62 (m, 2 H), 1.15-1.33 (m, 4 H); ^{13}C NMR (125 MHz, CD_2Cl_2) δ 160.2 (d, $J = 3.8$ Hz), 160.0 (d, $J = 3.8$ Hz), 133.6 (d, $J = 7.1$ Hz), 133.4 (d, $J = 7.1$ Hz), 132.5, 125.1 (d, $J = 124.8$ Hz), 123.3 (d, $J = 125.3$ Hz), 120.2 (d, $J = 12.2$ Hz), 120.1 (d, $J = 11.9$ Hz), 110.9 (d, $J = 2.4$ Hz), 110.9 (d, $J = 2.4$ Hz), 55.3, 55.3, 55.2, 35.7, 25.1; ^{31}P NMR (162 MHz, CD_2Cl_2) δ 21.2; HRMS (FAB⁺) m/z 635.2447 [635.2440 calcd for $C_{34}H_{40}N_2O_6P_2$ (M+H⁺)].



(*R,R*)-Diphosphinic Amide Ligand 13g. (This ligand has been prepared in two ways. Both procedures are presented.)

Method A: (*R,R*)-1,2-cyclohexanediamine (706 mg, 6.18 mmol), chlorodi(3,5-dimethylphenyl)phosphine (3.51 g, 12.9 mmol), Et_3N (6.26 g, 8.62 mL, 61.8 mmol), *t*-BuOOH (5.5 M in decane, 4.5 mL, 24.7 mmol) and CH_2Cl_2 (100 mL) were combined according to General Procedure 2 to produce a crude solid. The product was purified by flash chromatography (60:40 Et_2O/CH_2Cl_2) to provide 1.74 g (45%) of **13g** as an off-white, foamy solid. **Method B:** Di(3,5-dimethylphenyl)phosphinic acid (3.0 g, 10.9 mmol), $SOCl_2$ (20 mL), (*R,R*)-1,2-cyclohexanediamine (624 mg, 5.46 mmol), Et_3N (2.76 g, 3.8 mL, 27.3 mmol), and CH_2Cl_2 (50 mL) were combined according to General Procedure 3 to provide a light brown solid. The product was purified by flash chromatography (60:40 Et_2O/CH_2Cl_2) to provide 2.73 g (80%) of **13g** as a colorless solid: IR (solid) cm^{-1} 3189, 2925, 2859, 1603, 1448, 1189; 1H NMR (400 MHz, $CDCl_3$) δ 7.63 (d, $J = 12.4$ Hz, 4 H), 7.47 (d, $J = 12.1$ Hz, 4 H), 7.15 (s, 2 H), 7.10 (s, 2 H), 4.31 (br. s, 2 H), 2.93-3.12 (m, 2 H), 2.38 (s, 12 H), 2.28 (s, 12 H), 2.04 (d, $J = 12.6$ Hz, 2 H), 1.57 (d, $J = 7.8$ Hz, 2 H), 1.24-1.37 (m, 2 H), 1.12 (t, $J = 9.6$ Hz, 2 H); ^{13}C NMR (126 MHz, $CDCl_3$) δ 137.9 (d, $J = 13.2$ Hz), 137.8 (d, $J = 13.4$ Hz), 133.4 (d, $J = 2.4$ Hz), 133.3 (d, $J = 2.8$ Hz), 133.0 (d, $J = 128.1$ Hz), 132.7 (d, $J = 126.0$ Hz), 130.1 (d, $J = 9.9$ Hz), 129.0 (d, $J = 9.9$ Hz), 55.8 (d, $J = 1.4$ Hz), 35.8 (d, $J = 2.4$ Hz), 25.0, 21.3, 21.2; ^{31}P NMR (162 MHz, CD_2Cl_2) δ 26.4; HRMS (FAB⁺) m/z 627.3266 [627.3269 calcd for $C_{38}H_{48}N_2O_6P_2$ (M+H⁺)].



(*R,R*)-Diphosphinic Amide Ligand 13h. Bis(3,5-di-*t*-Bu-phenyl)phosphinic acid (5.44 g, 12.3 mmol), SOCl₂ (20 mL), (*R,R*)-1,2-cyclohexanediamine (701 mg, 6.13 mmol), Et₃N (3.10 g, 4.3 mL, 30.7 mmol), and CH₂Cl₂ (50 mL) were combined according to General Procedure 3 to provide a light brown solid. The product was purified by flash chromatography (50:50 Et₂O/hexanes) to provide 4.90 g (83%) of **13h** as an off-white foamy solid: IR (solid) cm⁻¹ 3189, 2958, 2859, 1602, 1247, 1178, 1119; ¹H NMR (500 MHz, CD₂Cl₂, one aliphatic signal not observed) δ 7.86 (d, *J* =

12.5 Hz, 4 H), 7.77 (d, *J* = 12.2 Hz, 4 H), 7.56 (d, *J* = 10.0 Hz, 4 H), 2.87 (s, 2 H), 2.14 (d, *J* = 12.5 Hz, 2H), 1.55 (m, 2 H), 1.36 (s, 36 H), 1.29 (s, 36 H), 1.08 (t, *J* = 10.0 Hz, 2 H); ¹³C NMR (125 MHz, CD₂Cl₂) δ 150.8 (d, *J* = 12.0 Hz), 150.7 (d, *J* = 12.0 Hz), 133.6 (d, *J* = 32.5 Hz), 132.6 (d, *J* = 37.5 Hz), 126.4 (d, *J* = 9.9 Hz), 125.6 (d, *J* = 10.4 Hz), 125.4 (d, *J* = 2.4 Hz), 56.5 (d, *J* = 2.8 Hz), 35.8 (d, *J* = 3.3 Hz), 34.9 (d, *J* = 7.5 Hz), 31.1 (d, *J* = 3.3 Hz), 25.0; ³¹P NMR (162 MHz, CD₂Cl₂) δ 25.5; HRMS (FAB⁺) *m/z* 963.7011 [963.7025 calcd for C₆₂H₉₆N₂O₂P₂ (M+H⁺)].

Ligand Screening Experiments (Tables 1 and 2):

In a glovebox, ligand (22 μmol, solid) and a solution of M(NMe₂)₄ (20 μmol, 500 μL, 40.0 mM in toluene) were combined in an oven-dried 4 mL (1 dram) vial equipped with an oven-dried stirbar. The vial was sealed⁶ and heated⁷ for 15 min. The vial was cooled to 25 °C and substrate **1** (15 μL, 11.4 mg, 100 μmol) was added. The vial was resealed and heated for the specified time. The vial was cooled to 25 °C, removed from the glovebox, and opened to the atmosphere. While rapidly stirring the reaction mixture, trifluoroacetic anhydride (50 μL) was added, and the resulting solution was stirred for 5 min. Saturated aqueous Na₂CO₃ (200 μL) was then added, and the resulting suspension was stirred for 1 min. This suspension was then transferred to a 4 mL solid phase extraction (SPE) column (containing approx. 1 g anhydrous MgSO₄ atop approx. 0.25 g silica gel)⁸ via pipet,

⁶ Vials were sealed using specific caps designed to withstand elevated pressure (Kontes Mini-Inert Valve, size 21, Kontes Article Number 749110-0021). Other caps did not perform as well.

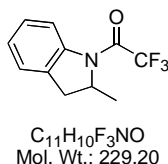
⁷ The reactions were heated in an aluminum block set atop a digitally controlled heating magnetic stirrer. Temperature was controlled using a thermocouple controller placed in a hole bored to the diameter of the thermocouple end. The vials were placed in holes bored to the diameter of the vial (15 mm) and to a depth of the shoulder of the vial (35 mm). This depth proved critical in order to minimize non-heated surface area in the vial and prevent solvent reflux at temperatures lower than that of the aluminum block.

⁸ For convenience, common household English Standard measuring spoons were used to measure packing materials for SPE columns. For the anhydrous MgSO₄, 3/8th of an teaspoon was used; for silica gel, 1/8th teaspoon was used. These materials, silica then MgSO₄, were added to the SFE columns in layers using a small funnel and packed down using vacuum (ca. 15 mm Hg).

filtered and washed with EtOAc (3 mL). The filtrate was collected into 2 mL auto-sampler vials⁹ and analyzed directly by chiral GC. Conversion was determined by comparison of the area of the sum of the two enantiomers to that of the starting material.

Cyclization Experiments, GC Analysis (Table 3, Entries 1-6):

In a glovebox, a solution of ligand **13g** (10 μ mol, 100 μ L, 0.1 M in toluene), a solution of $\text{Zr}(\text{NMe}_2)_4$ (10 μ mol, 100 μ L, 0.1 M in toluene) and 200 μ L of toluene were combined in an oven-dried 4-mL (1 dram) vial equipped with an oven-dried stirbar.¹⁰ The vial was sealed⁶ and heated⁷ for 15 min. The vial was cooled to 25 $^\circ\text{C}$ and a standard solution¹¹ of the substrate amine and hexamethylbenzene in toluene (100 μ L, 1.0 M amine, 100 μ mol amine) was added. The vial was resealed and heated for the specified time. The vial was then cooled to 25 $^\circ\text{C}$, removed from the glovebox, and opened to the atmosphere. To this rapidly stirred solution, trifluoroacetic anhydride (50 μ L) was added, and the resulting solution was stirred for 5 min. Saturated aqueous Na_2CO_3 (200 μ L) was then added, and the resulting suspension was stirred for 1 min. This suspension was then transferred to a 4-mL solid phase extraction (SPE) column (containing approx. 1 g anhydrous MgSO_4 atop approx. 0.25 g silica gel)⁸ via pipet, filtered and washed with EtOAc (2 mL). The filtrate was collected into 2 mL auto-sampler vials⁹ and analyzed directly by chiral GC. For internal standard calibration, 100 μ L of the standard amine solution (see above) was diluted to 500 μ L using toluene and worked up in the same fashion as the reaction mixture. This latter solution was then analyzed under chiral GC conditions identical to those used for analysis of the reaction mixture.



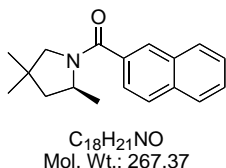
Amide 18, Isolated Yield. A solution of ligand **13g** (88.3 μ mol, 883 μ L, 0.1 M in toluene) and a solution of $\text{Zr}(\text{NMe}_2)_4$ (88.3 μ mol, 883 μ L, 0.1 M in toluene) were combined in an oven-dried 4 mL (1 dram) vial equipped with an oven-dried stir bar. The vial was sealed and heated to 135 $^\circ\text{C}$ for 15 min. The vial was allowed to cool to room temperature, and a solution of substrate amine (442 μ mol, 442 μ L, 1.0 M in toluene) was added. The vial was re-sealed and heated to 135 $^\circ\text{C}$ for 48 h. The vial was then cooled to 25 $^\circ\text{C}$ and removed from the glovebox. The reaction mixture was transferred to a 20 mL scintillation vial. Trifluoroacetic anhydride (185 mg, 123 μ L, 883 μ mol) was added, and the resulting mixture was stirred 15 min at 25 $^\circ\text{C}$. The reaction mixture was quenched by adding saturated Na_2CO_3 (3 mL). The organic layer was washed with brine (1 x 3 mL), dried over MgSO_4 , filtered, and concentrated to provide a brown oil. The product was purified by flash chromatography to yield 88 mg (85%) of **18** as a yellow oil: IR (film) 2971,

⁹ A standard 12-port solid phase extraction vacuum manifold was used.

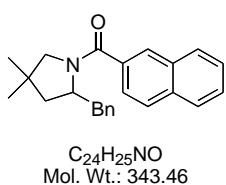
¹⁰ For reactions involving 20 mol% catalyst, 200 μ L each of the ligand and $\text{Zr}(\text{NMe}_2)_4$ were used. No additional solvent was added.

¹¹ The standard solution of amine containing hexamethylbenzene as an internal standard was prepared in a 1.0 mL volumetric flask using amine (1.0 mmol), hexamethylbenzene (15 mg), and toluene (to make 1 mL total volume).

1683, 1482, 1137; ^1H NMR (400 MHz, CDCl_3) δ 8.03 (d, $J = 8$ Hz, 1 H), 7.17-7.21 (m, 2 H), 7.06-7.10 (m, 1 H), 4.72 (m, 1 H), 3.38 (dd, $J = 13$ Hz, 24 Hz, 1 H), 2.63 (d, $J = 16$ Hz, 1 H), 1.24 (d, $J = 6$ Hz, 3 H); ^{13}C NMR (101 MHz, CDCl_3) δ 154.3 (q, $J = 27$ Hz), 140.2, 131.2, 128.3, 127.7, 126.0, 125.4, 119.4, 116.3 (q, $J = 289$ Hz), 56.8, 36.9, 21.9; ^{19}F NMR (376 MHz, C_6D_6) δ -69.9; HRMS (ESI $^+$) m/z 252.0608 [252.0612 calcd for $\text{C}_{11}\text{H}_{10}\text{F}_3\text{NO}$ (M+Na) $^+$].

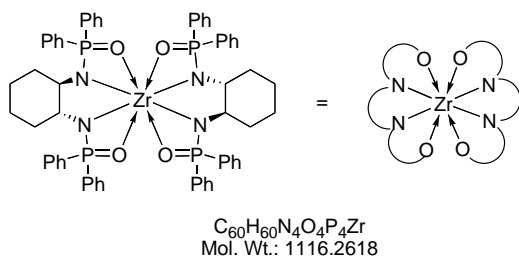


Amide 19. A solution of ligand **13g** (88.3 μmol , 883 μL , 0.1 M in toluene), a solution of $\text{Zr}(\text{NMe}_2)_4$ (88.3 μmol , 883 μL , 0.1 M in toluene) and 883 μL of toluene were combined in an oven-dried 4 mL (1 dram) vial equipped with an oven-dried stir bar. The vial was sealed and heated to 115 $^\circ\text{C}$ for 15 min. The vial was cooled to 25 $^\circ\text{C}$, and a solution of substrate amine (883 μmol , 883 μL , 1.0 M in toluene) was added. The vial was re-sealed and heated at 115 $^\circ\text{C}$ for 48 h. The vial was then cooled to 25 $^\circ\text{C}$ and removed from the glovebox. The reaction mixture was transferred to a 20 mL scintillation vial. Triethylamine (134 mg, 184 μL , 1.33 mmol) and 2-naphthoyl chloride (168 mg, 883 μmol) were added, and the resulting suspension was stirred for 2 h at 25 $^\circ\text{C}$. The reaction mixture was filtered and washed with saturated NaHCO_3 (2 x 3 mL). The organic layer was washed with brine (1 x 3 mL), dried over MgSO_4 , filtered, and concentrated to provide a brown oil. The product was purified by flash chromatography to yield 184 mg (78%) of **19** as grainy, colorless crystals: IR (solid) cm^{-1} 2957, 2926, 2867, 1619, 1406; ^1H NMR (400 MHz, CD_2Cl_2 , multiple rotamers observed in solution; major peaks reported) δ 7.89-8.04 (m, 4 H), 7.52-7.64 (m, 3 H), 4.39 (m, 1 H), 3.39 (d, $J = 12$ Hz, 1 H), 3.22 (d, $J = 12$ Hz, 1 H), 1.98 (dd, $J = 12$ Hz, 7 Hz, 1 H), 1.47 (dd, $J = 16$ Hz, 12 Hz, 1 H), 1.43 (d, $J = 6$ Hz, 3 H), 1.05 (s, 3 H), 0.81 (s, 3 H); ^{13}C NMR (101 MHz, CD_2Cl_2) δ 169.7, 134.8, 133.8, 132.5, 128.4, 128.3, 127.8, 127.6, 127.2, 127.0, 126.5, 62.6, 47.4, 38.2, 25.4, 25.2, 20.0; HRMS (ESI $^+$) m/z 268.1696 [268.1701 calcd for $\text{C}_{18}\text{H}_{21}\text{NO}$ (M+H) $^+$].



Amide 20. A solution of ligand **13g** (88.3 μmol , 883 μL , 0.1 M in toluene) and a solution of $\text{Zr}(\text{NMe}_2)_4$ (88.3 μmol , 883 μL , 0.1 M in toluene) were combined in an oven-dried 4 mL (1 dram) vial equipped with an oven-dried stir bar. The vial was sealed and heated at 135 $^\circ\text{C}$ for 15 min. After cooling to 25 $^\circ\text{C}$, a solution of substrate amine (442 μmol , 442 μL , 1.0 M in toluene) was added. The vial was re-sealed and heated at 135 $^\circ\text{C}$ for 24 h. It was then cooled to 25 $^\circ\text{C}$ and removed from the glovebox. The reaction mixture was transferred to a 20 mL scintillation vial. Triethylamine (134 mg, 184 μL , 1.33 mmol) and 2-naphthoyl chloride (168 mg, 883 μmol) were added, and the resulting suspension was stirred for 2 h at 25 $^\circ\text{C}$. The reaction mixture was filtered and washed with saturated NaHCO_3 (2 x 3 mL). The organic layer was washed with brine (1 x 3 mL), dried over MgSO_4 , filtered, and concentrated to provide a brown oil. The product was purified by flash chromatography to yield 115 mg (76%) of **20** as a yellow oil: IR (film) cm^{-1} 3058, 2956, 2867, 1620, 1407; ^1H NMR (400 MHz, C_6D_6) δ 8.12 (s, 1 H), 7.80 (d, $J = 8$ Hz, 1 H), 7.32-7.64 (m, 3 H), 7.18-7.32 (m, 2 H), 4.74-4.76 (m, 1 H), 3.48 (d, $J = 12$ Hz, 1 H), 3.01-3.06 (m, 1 H), 2.92 (d, $J = 8$ Hz, 1 H), 2.79 (d, $J = 8$ Hz), 1.35-1.42 (m, 2 H), 0.56

(s, 3H), 0.54 (s, 3 H); ^{13}C NMR (126 MHz, CD_2Cl_2) δ 169.8, 138.6, 134.7, 133.9, 132.5, 129.8, 128.5, 128.1, 127.8, 127.6, 127.5, 127.3, 127.0, 126.5, 126.2, 124.7, 63.1, 57.9, 43.9, 39.0, 37.9, 25.2; HRMS (ESI $^+$) m/z 366.1841 [366.1834 calcd for $\text{C}_{24}\text{H}_{25}\text{NO}$ ($\text{M}+\text{Na}$) $^+$].



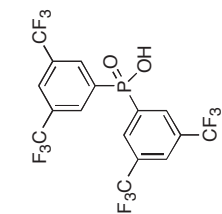
Complex 21a. In a glovebox, (*R,R*)-ligand **3c** (50 mg, 97 μmol), $\text{Zr}(\text{NMe}_2)_4$ (26 mg, 97 μmol) and d_8 -toluene (0.5 mL) were combined in a 4 mL vial equipped with a magnetic stirbar. The resulting suspension was stirred for 30 min, during which time the reaction mixture became homogenous.

The solution was transferred to a medium-walled NMR tube equipped with a teflon valve (J. Young tube). The tube was sealed and then removed from the glovebox. The tube was heated in an oil bath at 150 $^\circ\text{C}$ for 48 h and was occasionally removed to monitor reaction progress by NMR. Once intermediate **20a** was fully converted to **21a** (as judged by NMR), the tube was returned to the glovebox and opened. The solution was transferred to a 4 ml vial and layered with 1 ml of pentane, resulting in formation of microcrystalline **21a**. Decanting of the solvent provided 16 mg (30%) of **21a** as colorless prisms. Recrystallization from hot toluene provided X-ray quality crystals: ^1H NMR (400 MHz, C_6D_6) δ 8.03 (ddd, $J = 20.8, 12.2, 7.3$ Hz, 16 H), 7.26 (t, $J = 7.3$ Hz, 4 H), 7.15 (td, $J = 7.6, 2.2$ Hz, 8 H), 6.97-7.03 (m, 4 H), 6.9 (t, $J = 7.8$ Hz, 8 H), 4.07 (d, $J = 9.5$ Hz, 4 H), 1.79 (d, $J = 11.0$ Hz, 4 H), 1.51-1.67 (m, 4 H), 1.35 (t, $J = 8.6$ Hz, 4 H), 1.08 (t, $J = 9.8$ Hz, 3 H); ^{13}C NMR (126 MHz, C_6D_6 , 2 resonances obscured) δ 137.1 (d, $J = 117.3$ Hz), 135.2 (d, $J = 99.8$ Hz), 133.0 (d, $J = 10.8$ Hz), 132.4 (d, $J = 10.8$ Hz), 130.3 (d, $J = 1.9$ Hz), 130.0 (d, $J = 2.4$ Hz), 65.5 (d, $J = 16.0$ Hz), 35.3, 25.1; ^{31}P NMR (162 MHz, C_6D_6) δ 34.3. Anal. Calcd for $\text{C}_{60}\text{H}_{60}\text{N}_4\text{O}_4\text{P}_4\text{Zr}$: C, 64.56; H, 5.42; N, 5.02. Found: C, 64.49; H, 5.22; N, 5.92.

Current Data Parameters
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 EXPNO 1
 PROCNO 1
 DU /u
 USER DAW

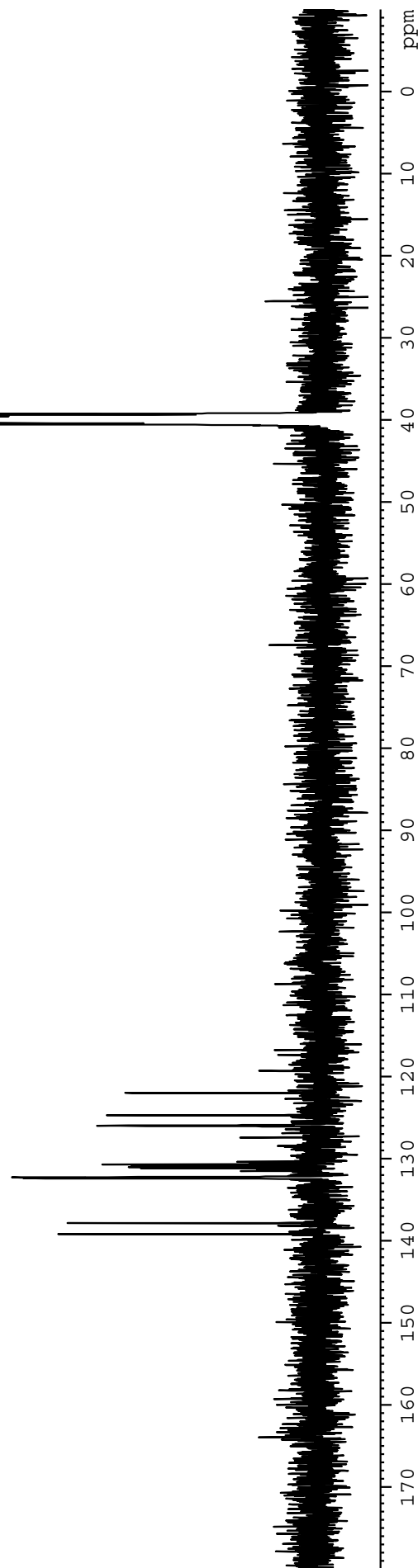
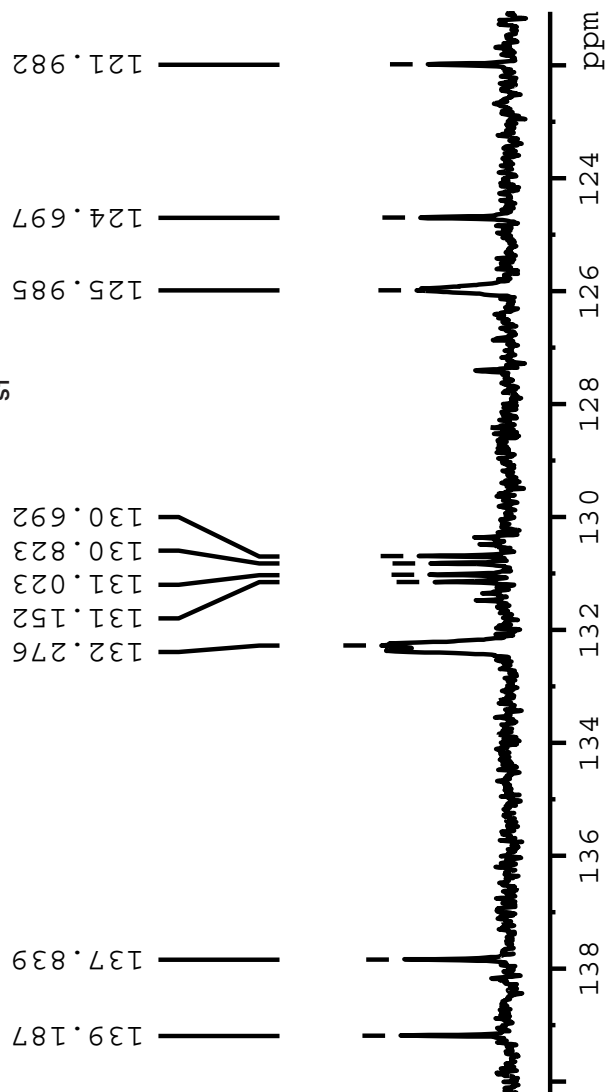
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 DS 0
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3632196 sec
 RG 16384
 DW 20.800 usec
 DE 6.00 usec
 TE 292.6 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8999998 sec
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 MCSRK 0.0150000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -2.00 dB
 SFO1 100.6228298 MHz



139.19
 137.84
 132.28
 131.15
 131.02
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 124.70
 121.98

AVQ-400 QNP Carbon Starting parameters 7/16/03 revised 7



Supporting Information - Watson, Chiu and Bergman

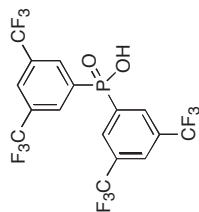
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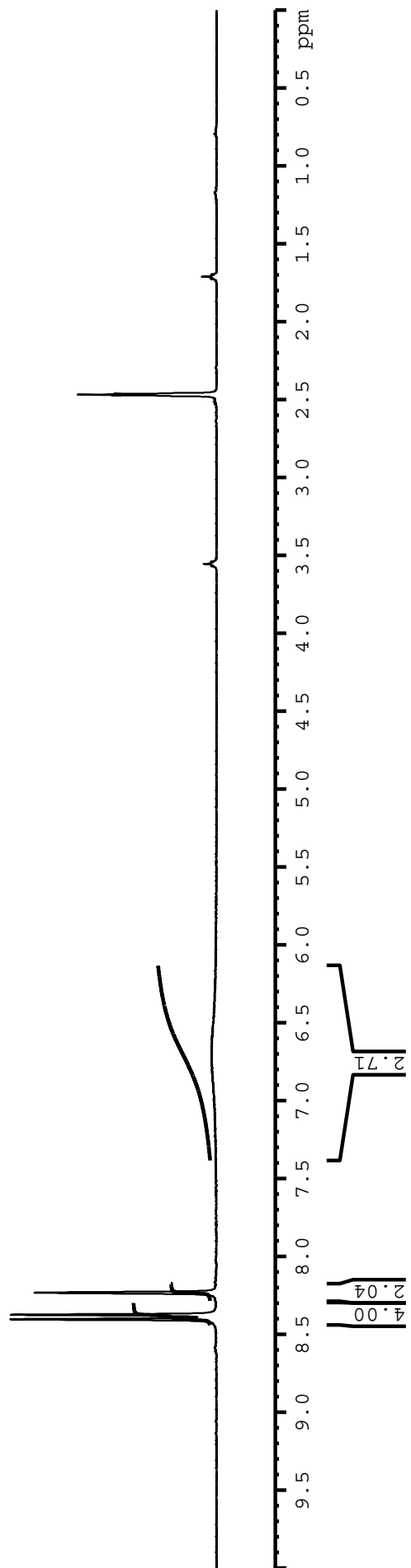
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RG         322.5
DE         62.400 use
TE         6.00 use
TE         292.6 K
D1         1.0000000 sec
MCREST    0.0000000 sec
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F2 - Processing parameters
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C₁₆H₇F₁₂O₂P
Mol. Wt.: 490.18
S1



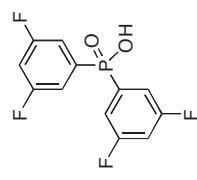

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RG       16384
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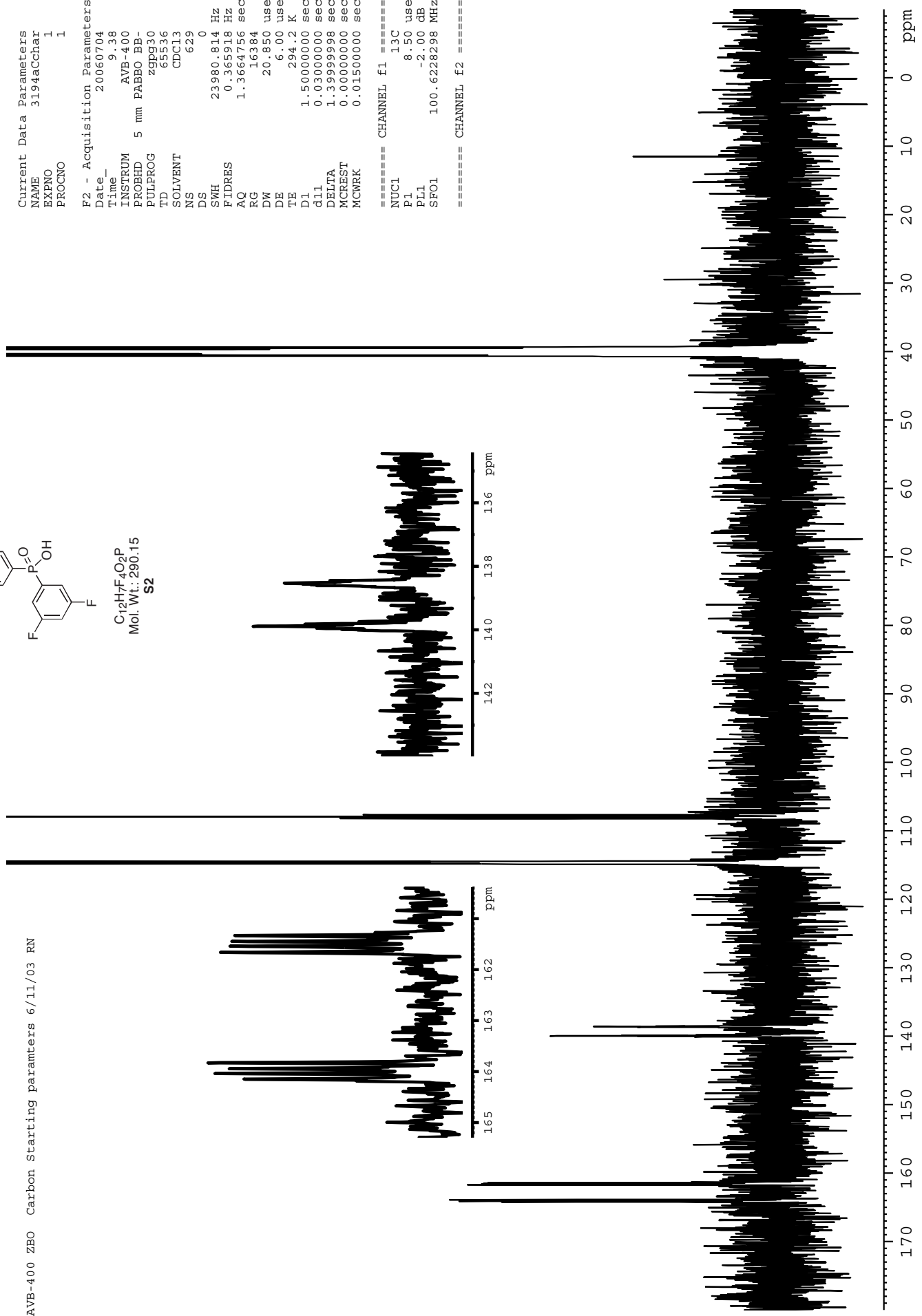
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===== CHANNEL f2 =====
  
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114.82
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 114.56
 114.46
 108.18
 107.93
 107.66

AVB-400 Z80 Carbon Starting parameters 6/11/03 RN



Supporting Information - Watson, Chiu and Bergman

1.768
1.750
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1.161
1.143
1.000

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 PROCNO 1
 DU /u
 USER DAW

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 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 203.2
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 TE 294.1 K
 DI 1.0000000 sec
 MCREST 0.0000000 sec
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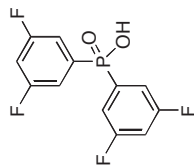
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2.920

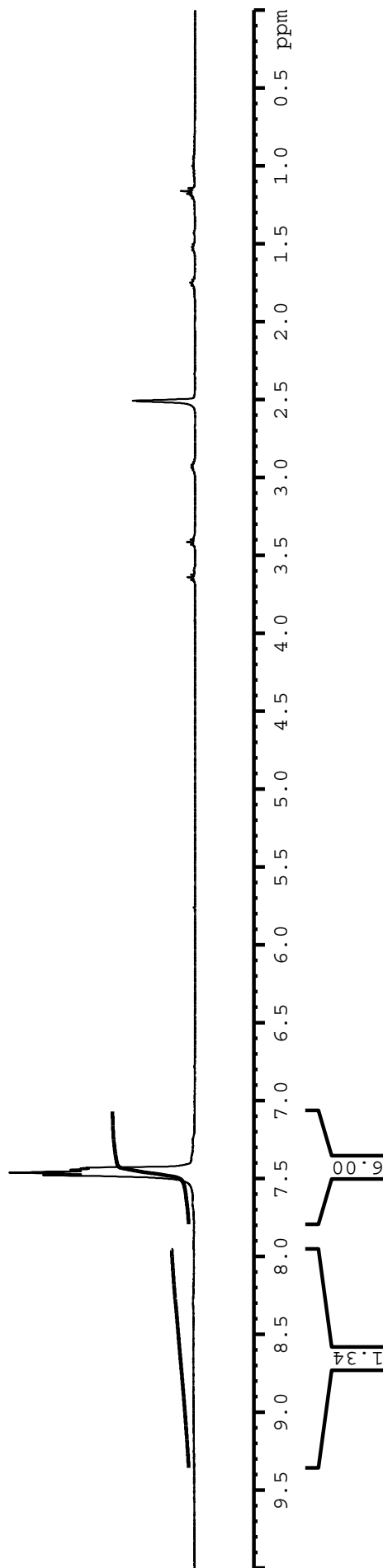
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5.759

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C₁₂H₇F₄O₂P
 Mol. Wt.: 290.15
S2



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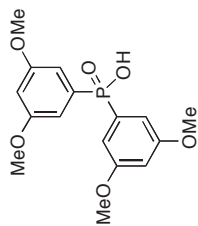
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PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        36
DS        0
SWH       24038.461 Hz
FIDRES    0.366798 Hz
AQ        1.3631988 sec
RG        16384
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TE        292.6 K
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d11       0.03000000 sec
DELTA     1.89999998 sec
MCREST    0.0000000 sec
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PL1       -2.00 dB
SFO1      100.6228298 MHz

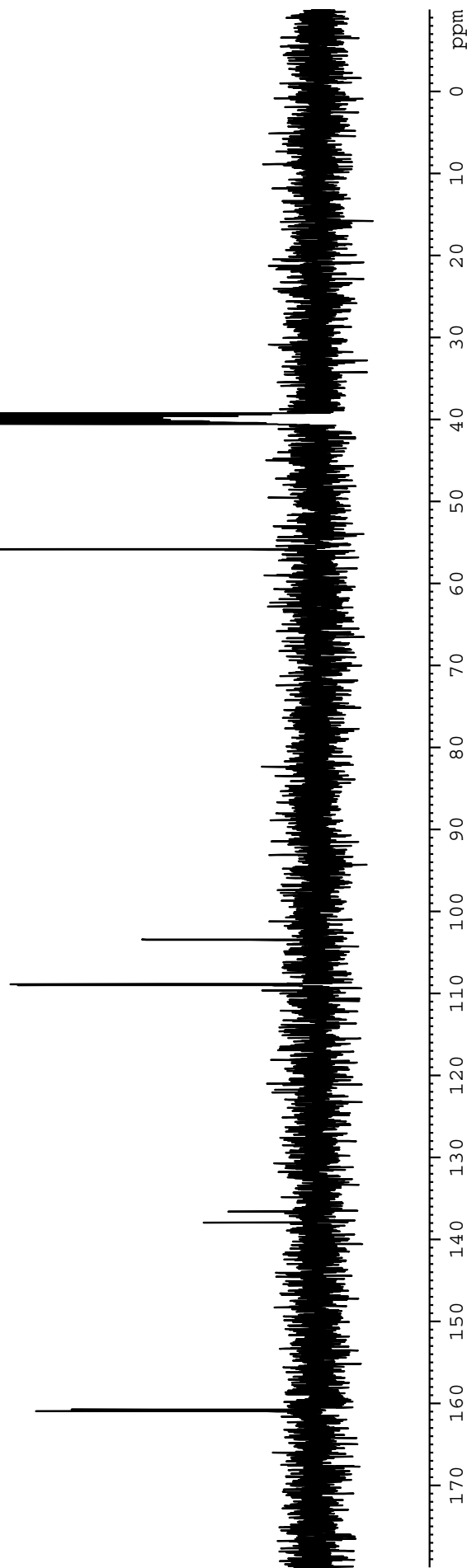
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108.85
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C₁₆H₁₉O₆P
Mol. Wt.: 338.29
S3

AVQ-400 QNP Carbon Starting parameters 7/16/03 revised 7



Supporting Information - Watson, Chiu and Bergman

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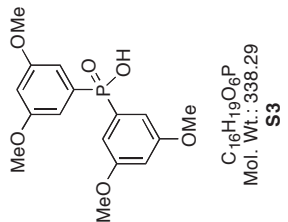
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DU       /u
USER     DAW

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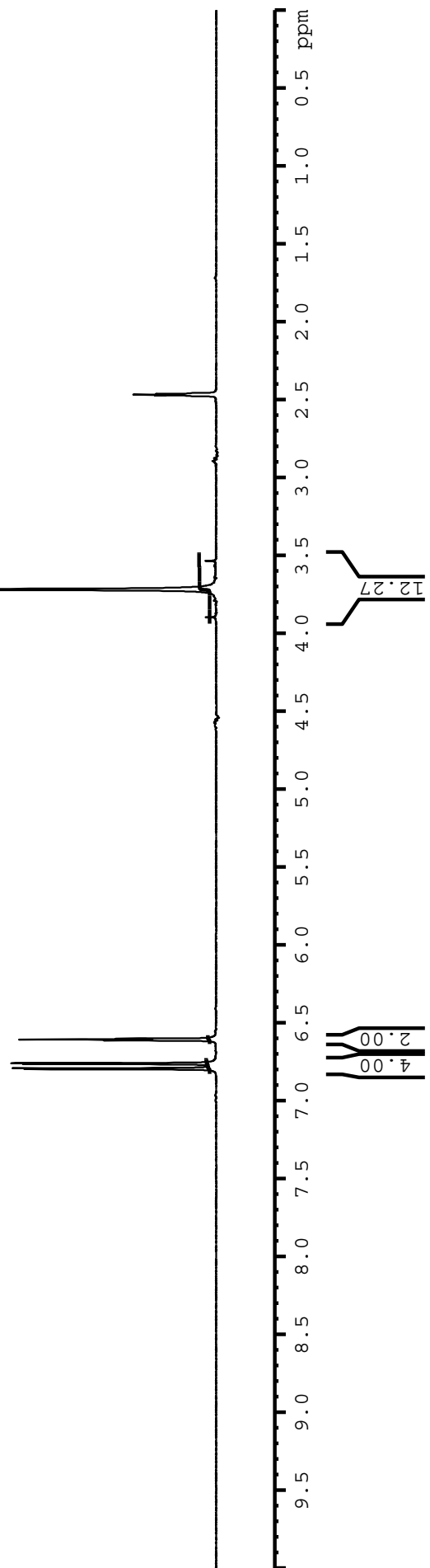
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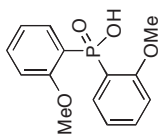
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WDW       RM
    
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3.898
4.576



6.799
6.793
6.766
6.760
6.614
6.609
6.603





C₁₄H₁₅O₄P
 Mol. Wt.: 278.24
S4

Supporting Information - Watson, Chiu and Bergman

S21

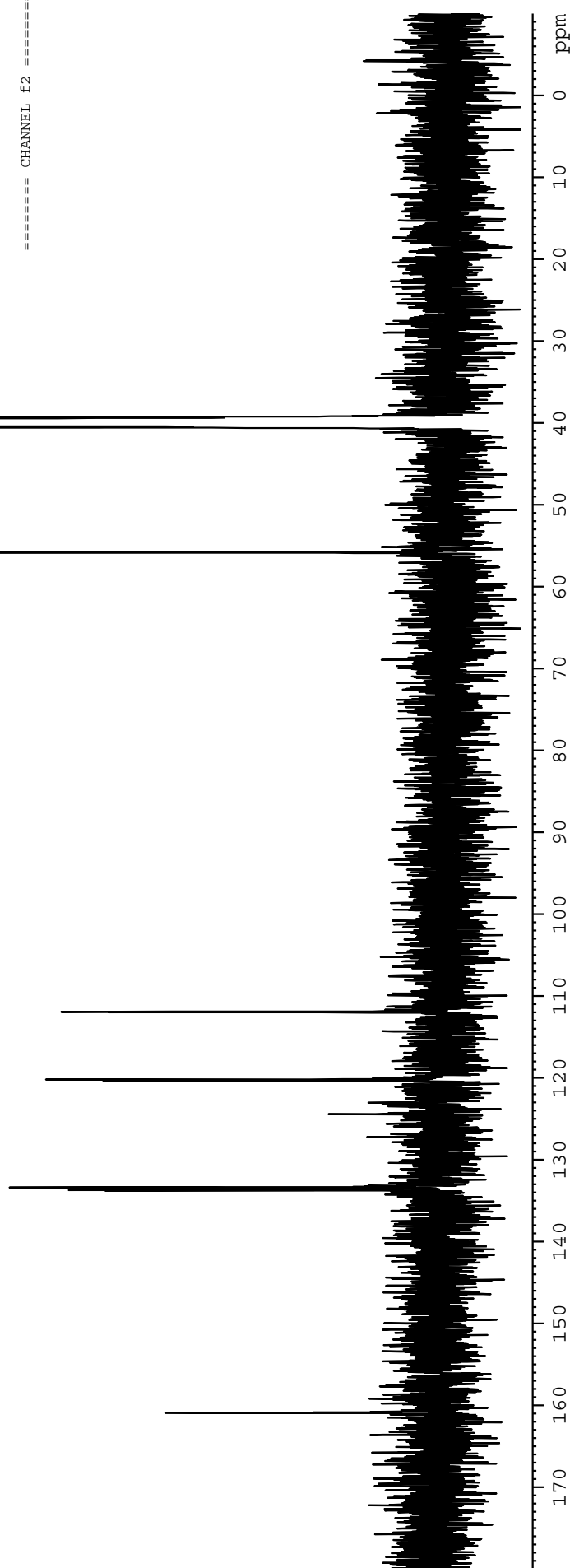
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NS         266
DS         0
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 sec
RG         16384
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DE         6.00 usec
TE         292.5 K
D1         2.0000000 sec
d11        0.03000000 sec
DELTA     1.89999998 sec
MCREST    0.0000000 sec
MCWRK     0.01500000 sec

===== CHANNEL f1 =====
NUC1       13C
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===== CHANNEL f2 =====
    
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Supporting Information - Watson, Chiu and Bergman

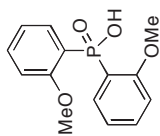
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DU       /u
USER     DAW

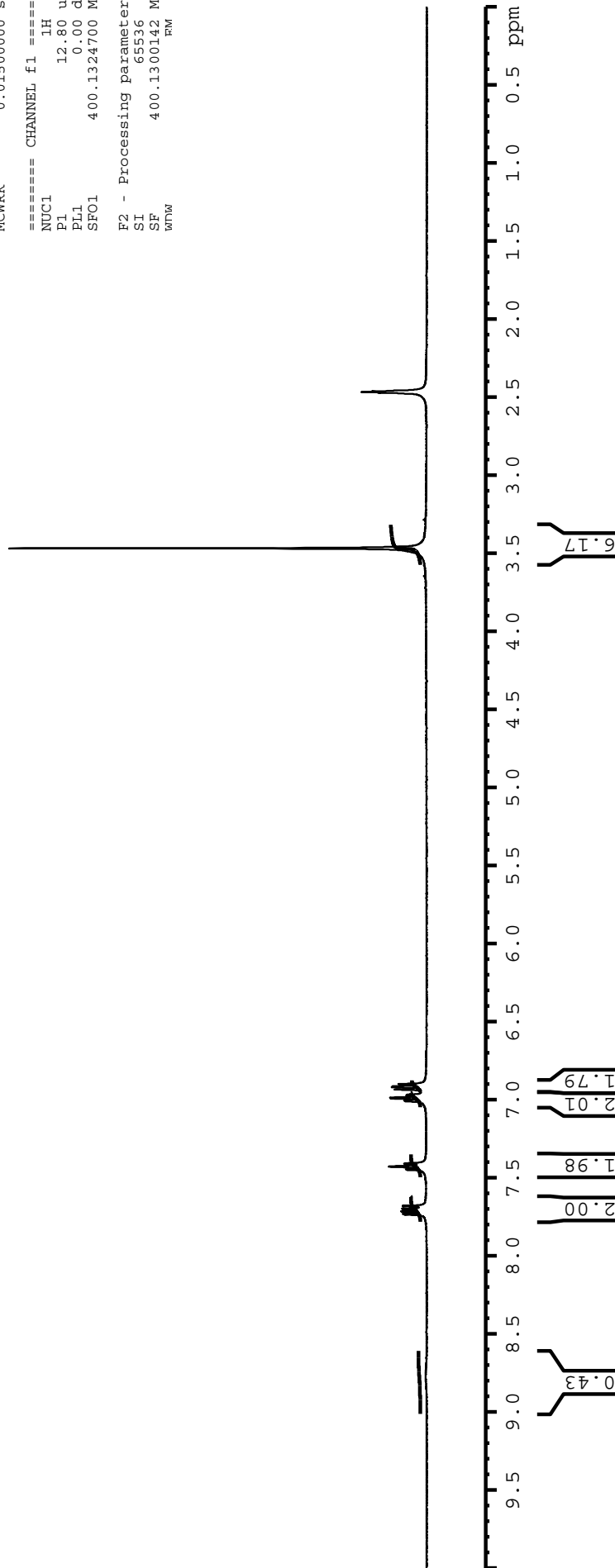
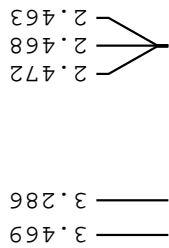
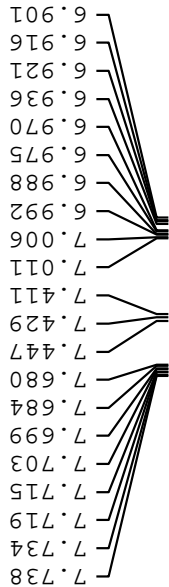
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FIDRES    0.122266 Hz
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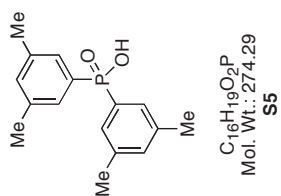
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F2 - Processing parameters
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WDW        RM
    
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C₁₄H₁₅O₄P
Mol. Wt.: 278.24
S4

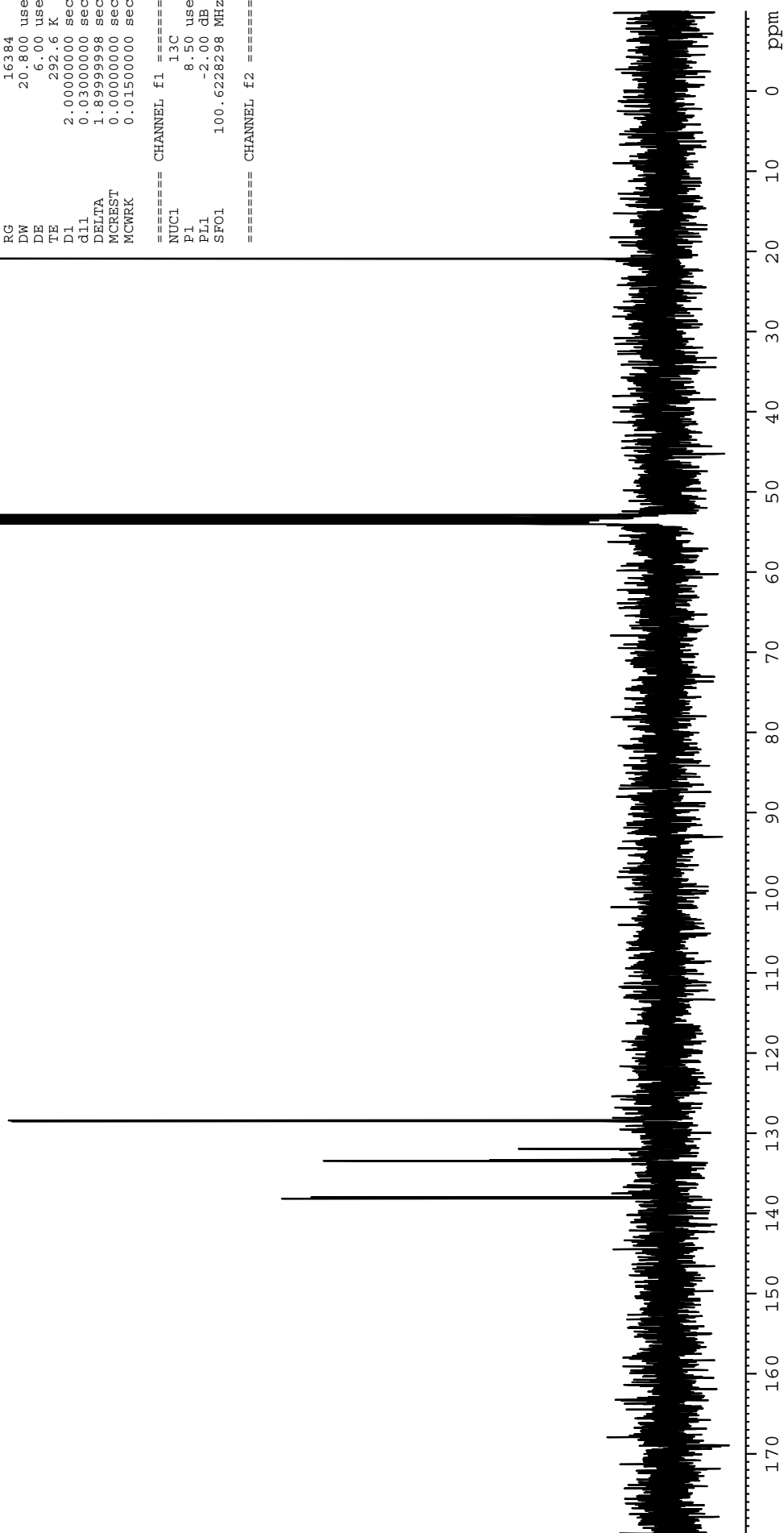




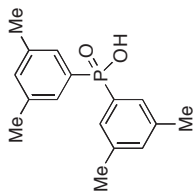
AVQ-400 QNP Carbon Starting parameters 7/16/03 revised 7

138.12
 137.99
 133.46
 133.43
 133.31
 133.31
 131.94
 128.49
 128.39

Current Data Parameters
 NAME 4080Cchar
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20060703
 Time 18.01
 INSTRUM AVQ-400
 PROBD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 42
 DS 0
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 16384
 DW 20.800 usec
 DE 6.00 usec
 TE 292.6 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -2.00 dB
 SFO1 100.6228298 MHz
 ===== CHANNEL f2 =====



Supporting Information - Watson, Chiu and Bergman



C₁₁H₁₄O₂P
Mol. Wt.: 274.29
S5

Current Data Parameters
NAME 4080Hchar
EXPNO 1
PROCNO 1
DU /u
USER DAW

F2 - Acquisition Parameters
Date_ 20060703
Time 17.58
INSTRUM AVO-400
PROBHD 5 mm QNP 1H/13
PULPROG zg30
TD 65536
SOLVENT CDC13
NS 1
DS 0
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0895586 sec
RG 181
DE 62.400 use
TE 292.5 K
DI 1.0000000 sec
MCREST 0.0000000 sec
MCWRK 0.01500000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 12.80 use
PL1 0.00 dB
SFO1 400.1324700 MHz

F2 - Processing parameters
SI 65536
SF 400.1300142 MHz
WUM

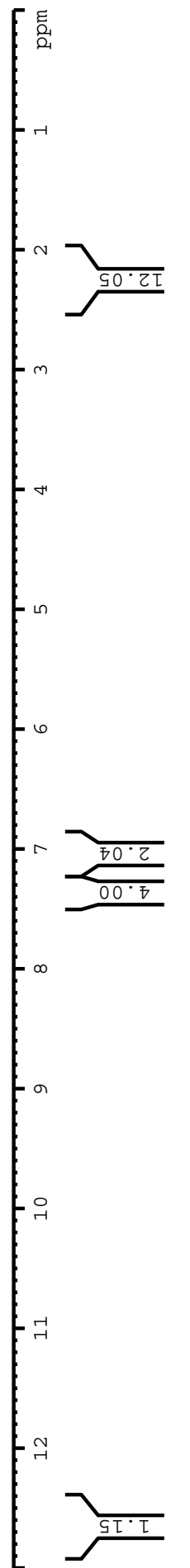
5.334
5.324
5.322
5.319

7.334
7.302
7.102

12.638

2.430
2.305
2.273
2.213
2.113

1.448
1.272
0.889
0.806
0.779



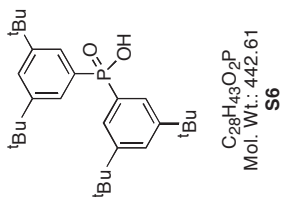

```

Current Data Parameters
NAME      4006aCchar
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20060704
Time      10.14
INSTRUM   AVB-400
PROBHD    5 mm PABBO BB-
PULPROG   zgpg30
TD         65536
SOLVENT   CD2Cl2
NS         202
DS         0
SWH        23980.814 Hz
FIDRES     0.365918 Hz
AQ         1.3664756 sec
RG         16384
DW         20.850 usec
DE         6.00 usec
TE         294.7 K
D1         1.5000000 sec
d11        0.0300000 sec
DELTA     1.39999998 sec
MCREST    0.0000000 sec
MCWRK     0.01500000 sec

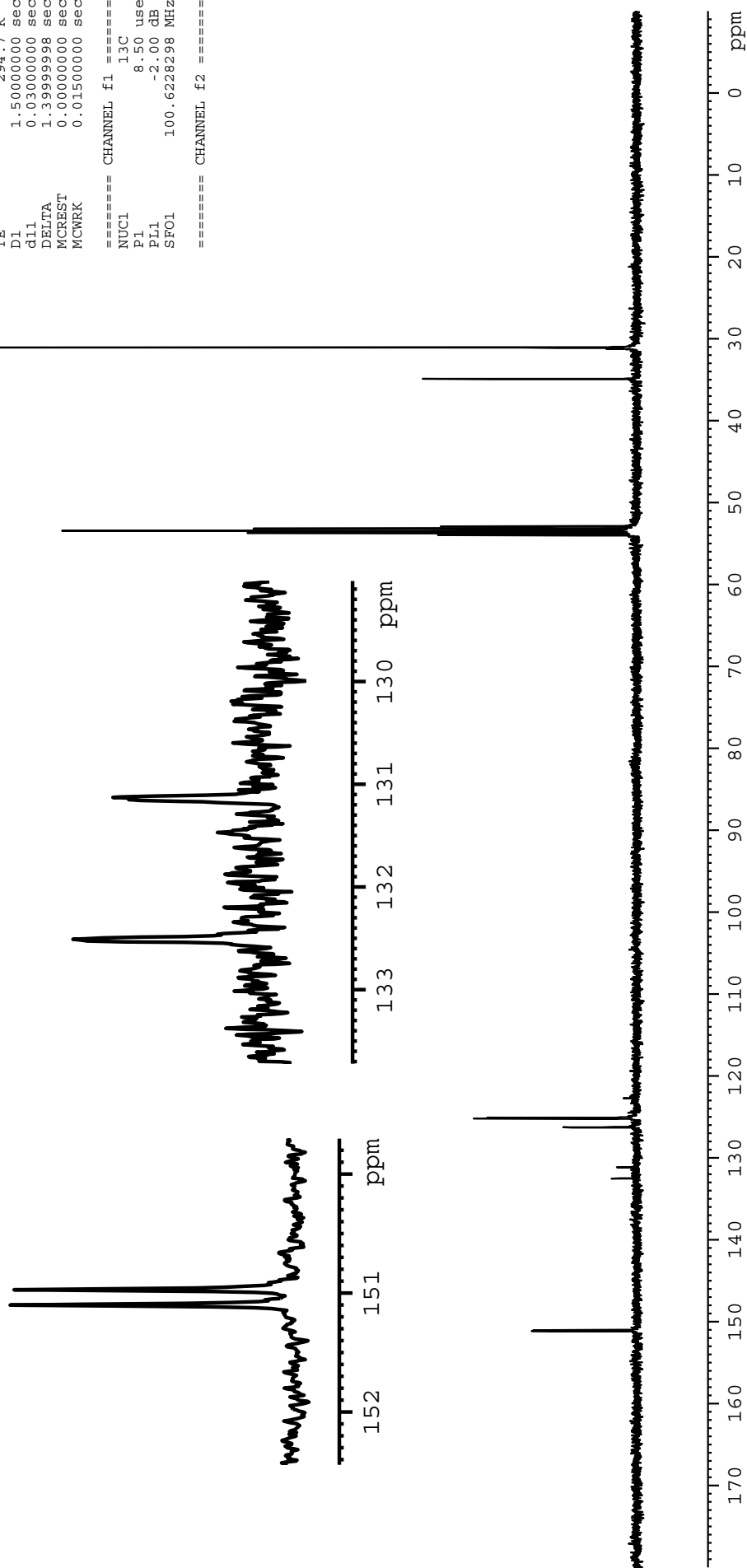
===== CHANNEL f1 =====
NUC1       13C
P1         8.50 usec
PL1        -2.00 dB
SFO1       100.6228298 MHz

===== CHANNEL f2 =====
    
```



126.25
 125.19
 125.08
 122.70

AVB-400 ZBO Carbon Starting paramters 6/11/03 RN



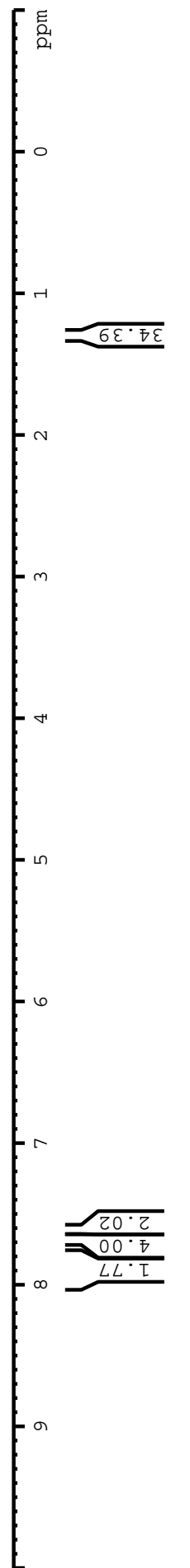
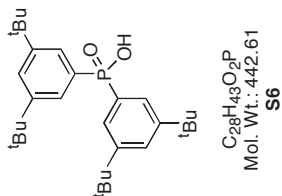
Supporting Information - Watson, Chiu and Bergman

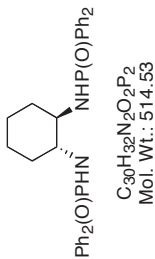


Current Data Parameters
 USER DAW
 NAME 4106ahChar
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060704
 Time 9.27
 INSTRUM AVB-400
 PROBD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 181
 DW 60.400 use
 DE 6.00 use
 TE 294.1 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 seq

==== CHANNEL f1 =====
 NUC1 1H
 P1 8.20 use
 PL1 -3.00 dB
 SFO1 400.1324710 MHz
 F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WDW EM
 SSR 0

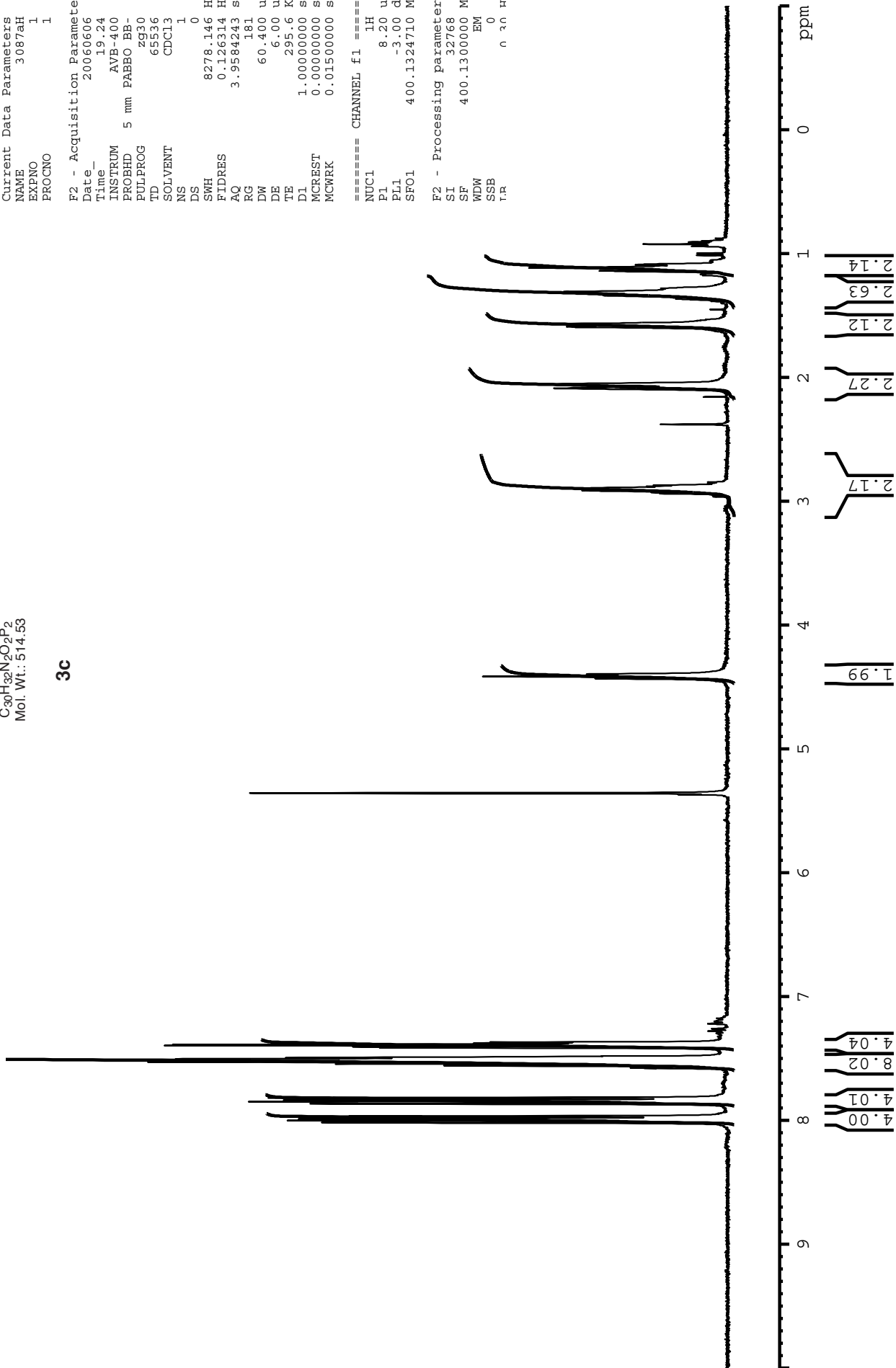




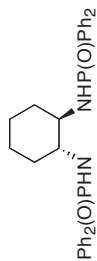
3c

2.158
 2.089
 2.056
 1.591
 1.572
 1.453
 1.334
 1.308
 1.168
 1.139
 1.115
 1.091
 1.015
 0.998
 0.941
 0.925
 0.907

Current Data Parameters
 NAME 3087AH
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20060606
 Time 19.24
 INSTRUM AVB-400
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 181
 DW 60.400 use
 DE 6.00 use
 TE 295.6 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 8.20 use
 PL1 -3.00 dB
 SFO1 400.1324710 MHz
 F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 T.R 0.00 Hz



Supporting Information - Watson, Chiu and Bergman



C₃₀H₃₂N₂O₂P₂
Mol. Wt.: 514.53

3c

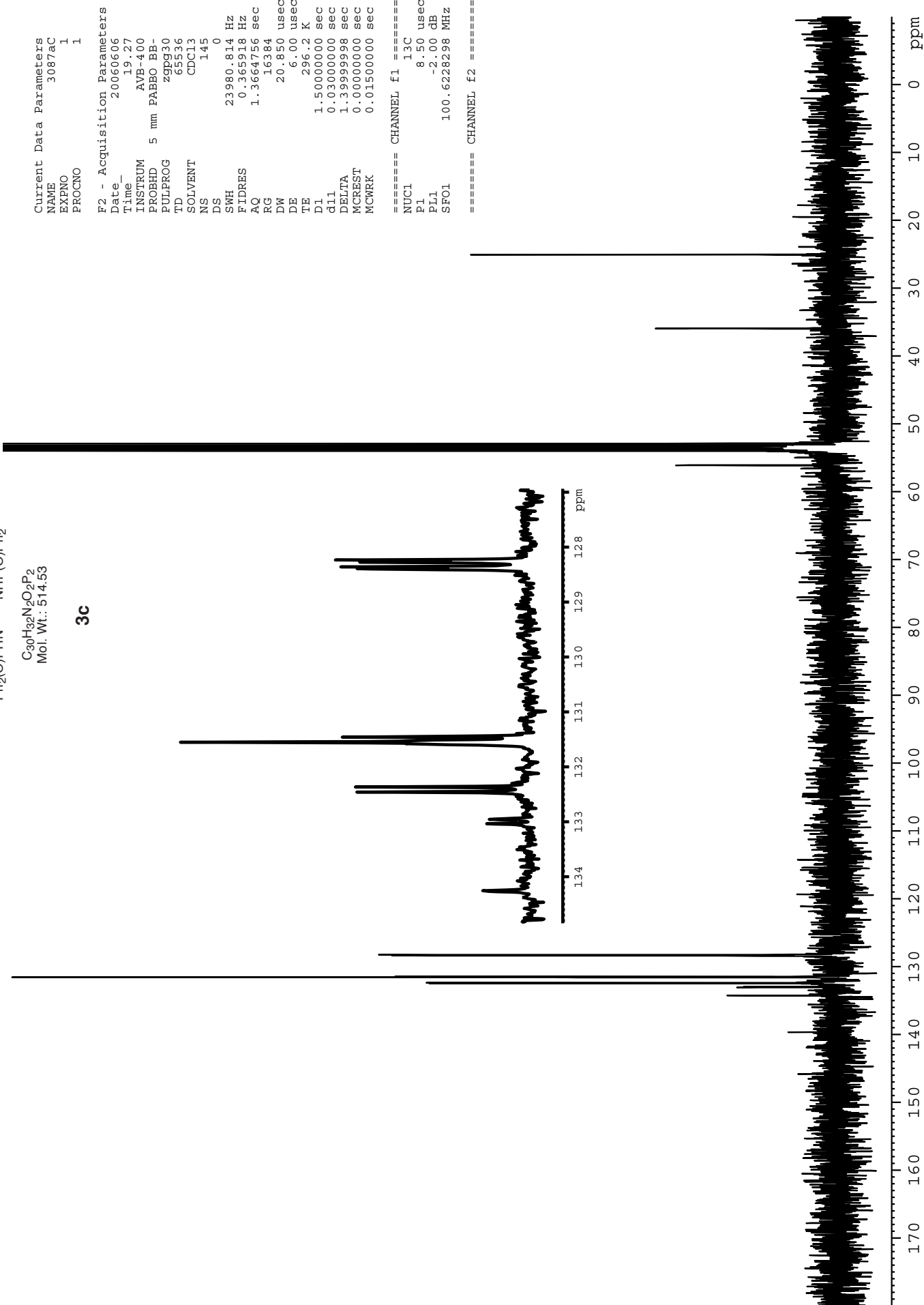
```

Current Data Parameters
NAME          3087aC
EXPNO         1
PROCNO        1

F2 - Acquisition Parameters
Date_         20060606
Time         19.27
INSTRUM       AVB-400
PROBHD        5 mm PABBO BB-
PULPROG       zgpg30
TD            65536
SOLVENT       CDCl3
NS            1.45
DS            0
SWH           23980.814 Hz
FIDRES        0.365918 Hz
AQ            1.3664756 sec
RG            16384
DE            20.850 usec
TE            296.2 K
D1            1.5000000 sec
d11           0.0300000 sec
DELTA         1.39999998 sec
MCREST        0.0000000 sec
MCWRK         0.01500000 sec

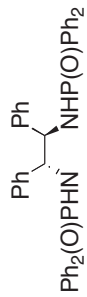
===== CHANNEL f1 =====
NUC1          13C
P1            8.50 usec
PL1           -2.00 dB
SFO1          100.6228298 MHz

===== CHANNEL f2 =====
    
```



13C DRX-500 5mm ZBBO probe
 starting parameters with zgpg30
 uses ns*td0

132.10
132.11
131.48
128.27
128.17
128.09
127.99
127.82
127.48
126.82



$\text{C}_{38}\text{H}_{34}\text{N}_2\text{O}_2\text{P}_2$
 Mol. Wt.: 612.64

9

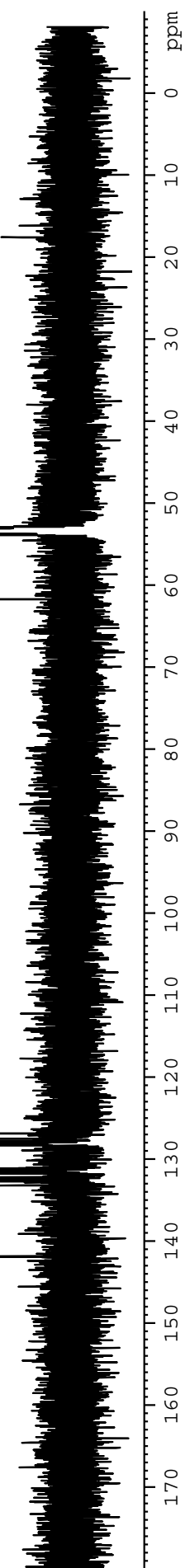
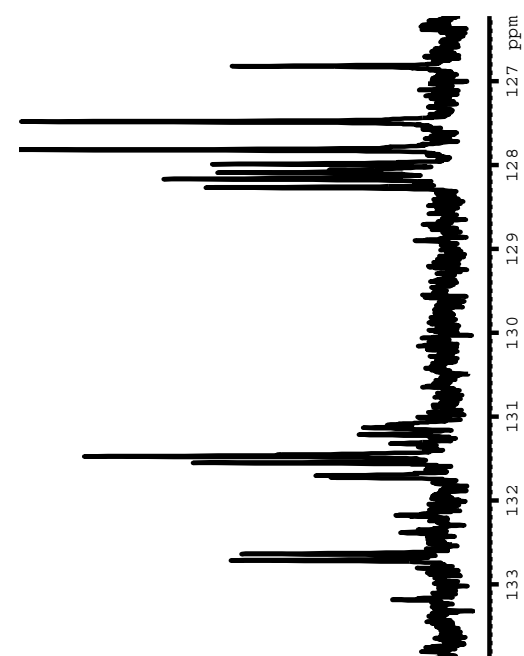
61.65
53.80
53.59
53.37
53.15
52.94

Current Data Parameters
 NAME 3088C
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060606
 Time 21.28
 INSTRUM DRX-500
 PROBD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 131072
 SOLVENT CDCl3
 NS 448
 DS 0
 SWH 30864.197 Hz
 FIDRES 0.235475 Hz
 AQ 2.1234164 sec
 RG 16384
 DW 16.200 usec
 DE 5.00 usec
 TE 293.0 K
 D1 1.5000000 sec
 d11 0.0300000 sec
 DELTA 1.39999998 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.70 usec
 PL1 -3.00 dB
 SFO1 125.7722011 MHz

==== CHANNEL f2 =====

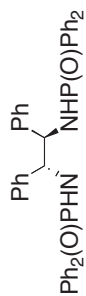


Supporting Information - Watson, Chiu and Bergman

S30

1H starting parameters (zg30)
 DRX-500 zBBO probe

7.785
7.769
7.762
7.746
7.732
7.632
7.615
7.608
7.592
7.526
7.511
7.487
7.412
7.397
7.382
7.232
7.103
7.080
7.066
6.891
6.877
6.186



C₂₈H₃₄N₂O₂P₂
 Mol. Wt.: 612.64

9

Current Data Parameters
 NAME 3088H
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060606
 Time 21.25
 INSTRUM DRX-500
 PROBHD 5 mm BBO BB-IH
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 1
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2768500 sec
 RG 322.5
 DW 50.000 use
 DE 7.11 use
 TE 293.0 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 1H
 P1 12.20 use
 PL1 -5.00 dB
 SFO1 500.1330883 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300127 MHz
 WDW EM
 SSB 0
 LR 0.20 Hz

1.353

4.316

5.335

6.186

6.877

6.891

7.066

7.080

7.103

7.232

7.382

7.397

7.412

7.487

7.511

7.526

7.592

7.608

7.615

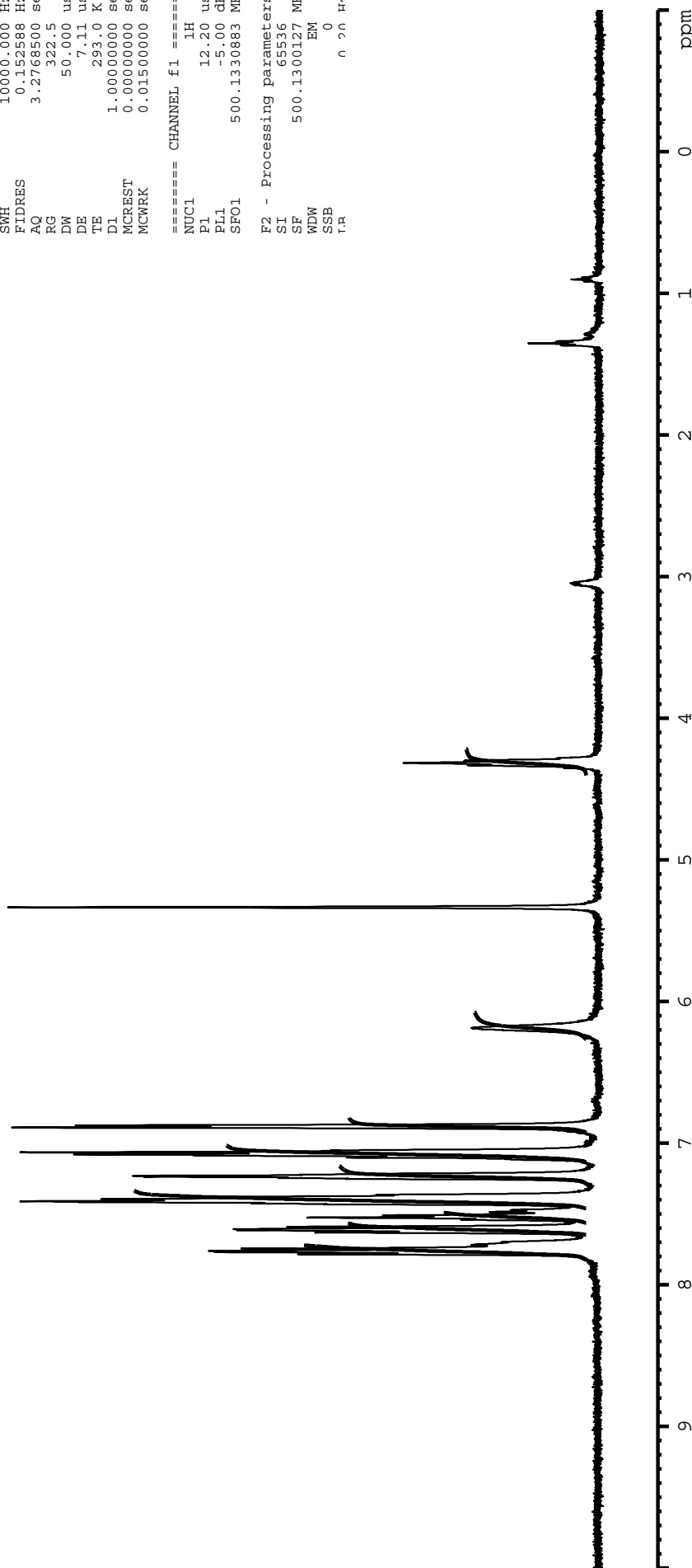
7.632

7.746

7.762

7.769

7.785



2.02

1.87

3.98

6.04

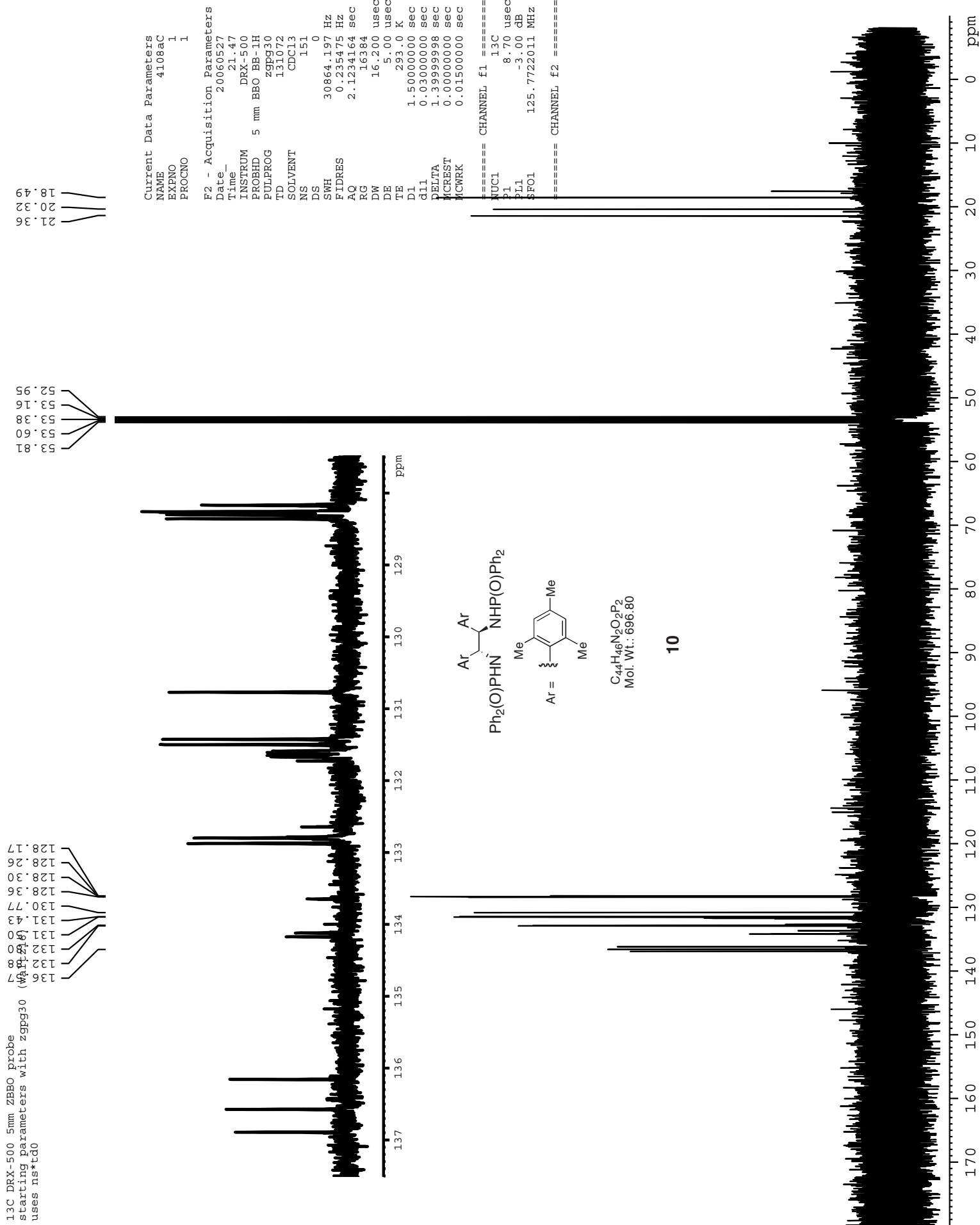
4.14

7.59

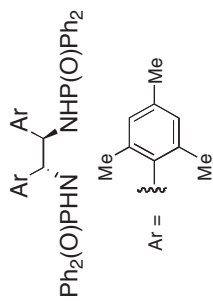
2.39

4.00

4.74



1H starting parameters (zg30)
 DRX-500 zBBO probe
 000004 11-11



C₄₄H₄₆N₂O₂P₂
 Mol. Wt.: 696.80

10

Supporting Information - Watson, Chiu and Bergman

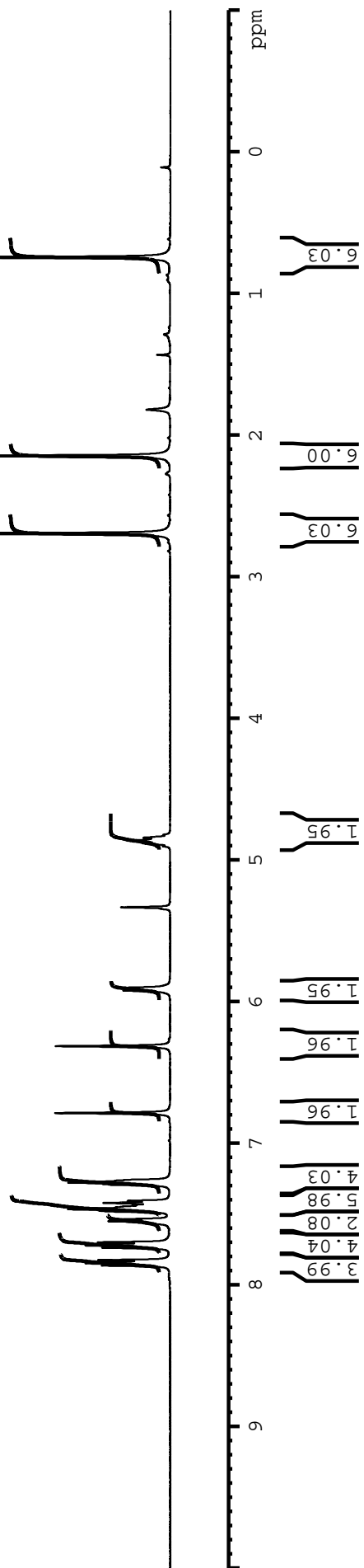
Current Data Parameters
 NAME 4108ah
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060527
 Time_ 21.40
 INSTRUM DRX-500
 PROBHD 5 mm BBO BB-IH
 PULPROG zg30
 TD 65536
 SOLVENT CD2Cl2
 NS 8
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2768500 sec
 RG 114
 DW 50.000 use
 DE 7.11 use
 TE 293.0 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 1H
 P1 12.20 use
 PL1 -5.00 dB
 SFO1 500.1330883 MHz

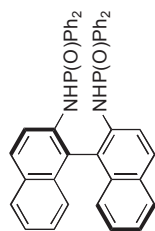
F2 - Processing parameters
 SI 65536
 SF 500.1300127 MHz
 WDW EM
 SSB 0
 LB 0.20 Hz

2.696
 2.273
 2.148
 1.821
 1.435
 1.290
 0.870
 0.746



13C DRX-500 5mm ZBBO probe
 starting parameters with zgpg30 (waltz16)
 uses ns*td0

124.47
 124.40
 118.96
 118.90
 116.99
 116.91
 116.83
 116.75



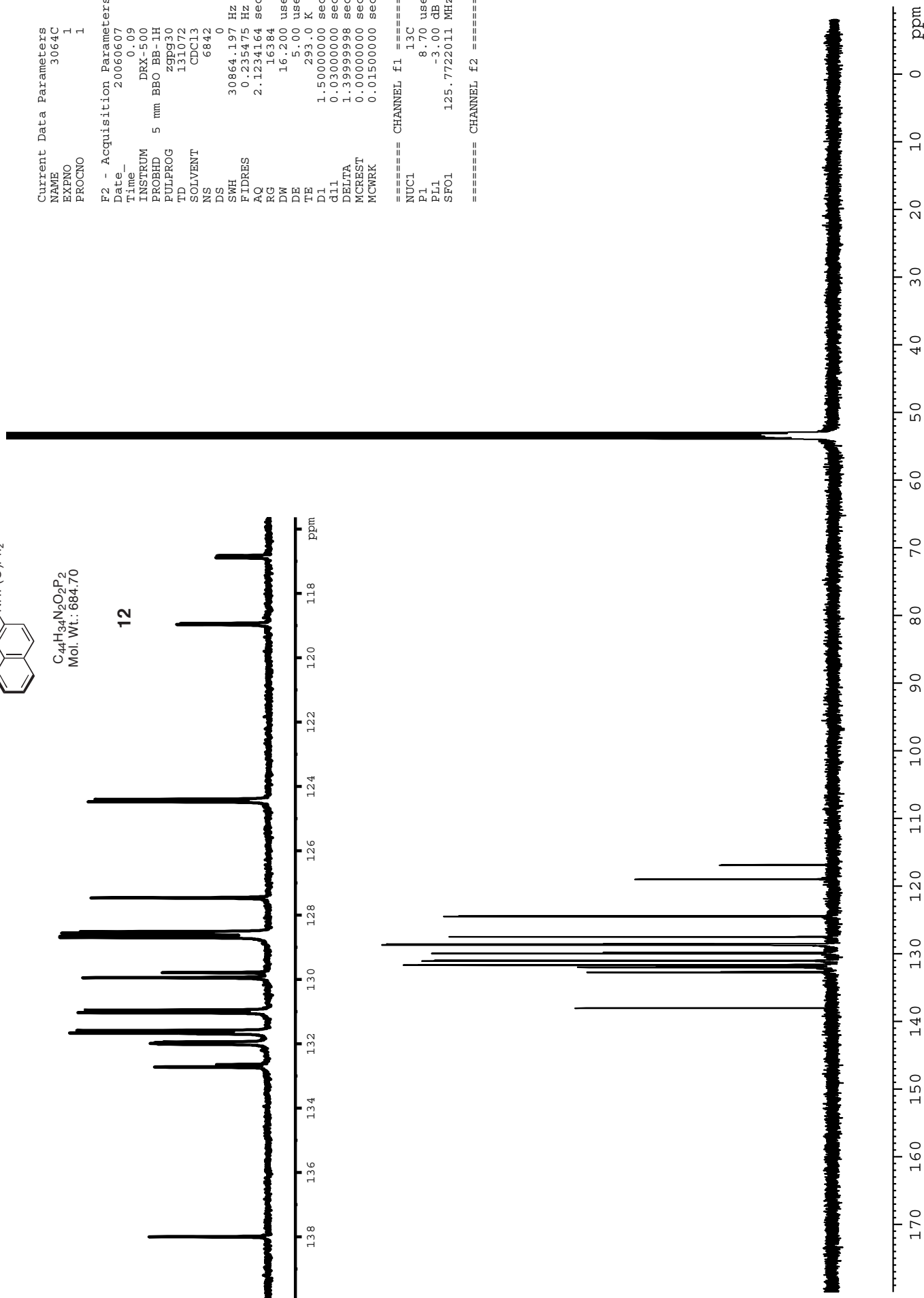
12
 $C_{44}H_{34}N_2O_2P_2$
 Mol. Wt.: 684.70

Current Data Parameters
 NAME 3064C
 EXPNO 1
 PROCNO 1

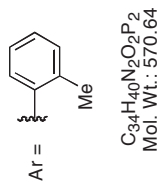
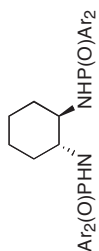
F2 - Acquisition Parameters
 Date_ 20060607
 Time 0.09
 INSTRUM DRX-500
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 131072
 SOLVENT CDCl3
 NS 6842
 DS 0
 SWH 30864.197 Hz
 FIDRES 0.235475 Hz
 AQ 2.1234164 sec
 RG 16384
 DW 16.200 usec
 DE 5.00 usec
 TE 293.0 K
 D1 1.5000000 sec
 d11 0.03000000 sec
 DELTA 1.39999998 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.70 usec
 PL1 -3.00 dB
 SFO1 125.7722011 MHz

==== CHANNEL f2 =====



13C DRX-500 5mm ZBBO probe
 starting parameters with zgpg30 (wat-zf1b)
 uses ns*td0



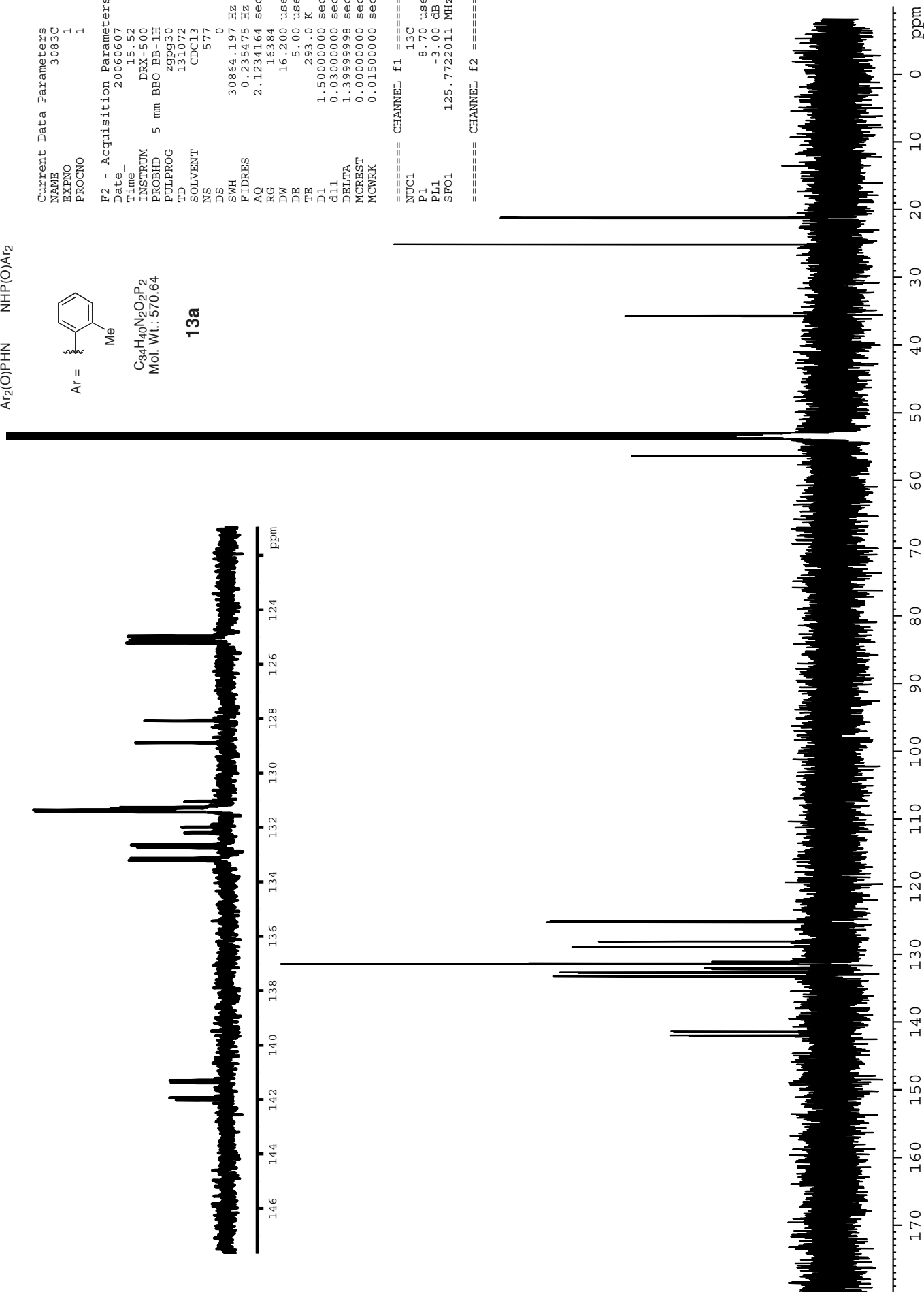
13a

Current Data Parameters
 NAME 3083C
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060607
 Time 15.52
 INSTRUM DRX-500
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 131072
 SOLVENT CDC13
 NS 577
 DS 0
 SWH 30864.197 Hz
 FIDRES 0.235475 Hz
 AQ 2.1234164 sec
 RG 16384
 DW 16.200 usec
 DE 5.00 usec
 TE 293.0 K
 D1 1.5000000 sec
 d11 0.0300000 sec
 DELTA 1.39999998 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.70 usec
 PL1 -3.00 dB
 SFO1 125.7722011 MHz

==== CHANNEL f2 =====



Supporting Information - Watson, Chiu and Bergman

S38

1H starting parameters (zg30)
 DRX-500 zBBO probe

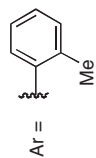
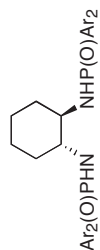
8.004
 7.989
 7.977
 7.961
 7.611
 7.595
 7.586
 7.569
 7.412
 7.397
 7.381
 7.366
 7.305
 7.257
 7.242
 7.196
 7.182
 7.154
 7.113

2.357
 2.319
 2.237
 2.199
 2.173
 1.883
 1.865
 1.619
 1.385
 1.214

3.174

4.520

5.336



C₃₄H₄₀N₂O₂P₂
 Mol. Wt.: 570.64

13a

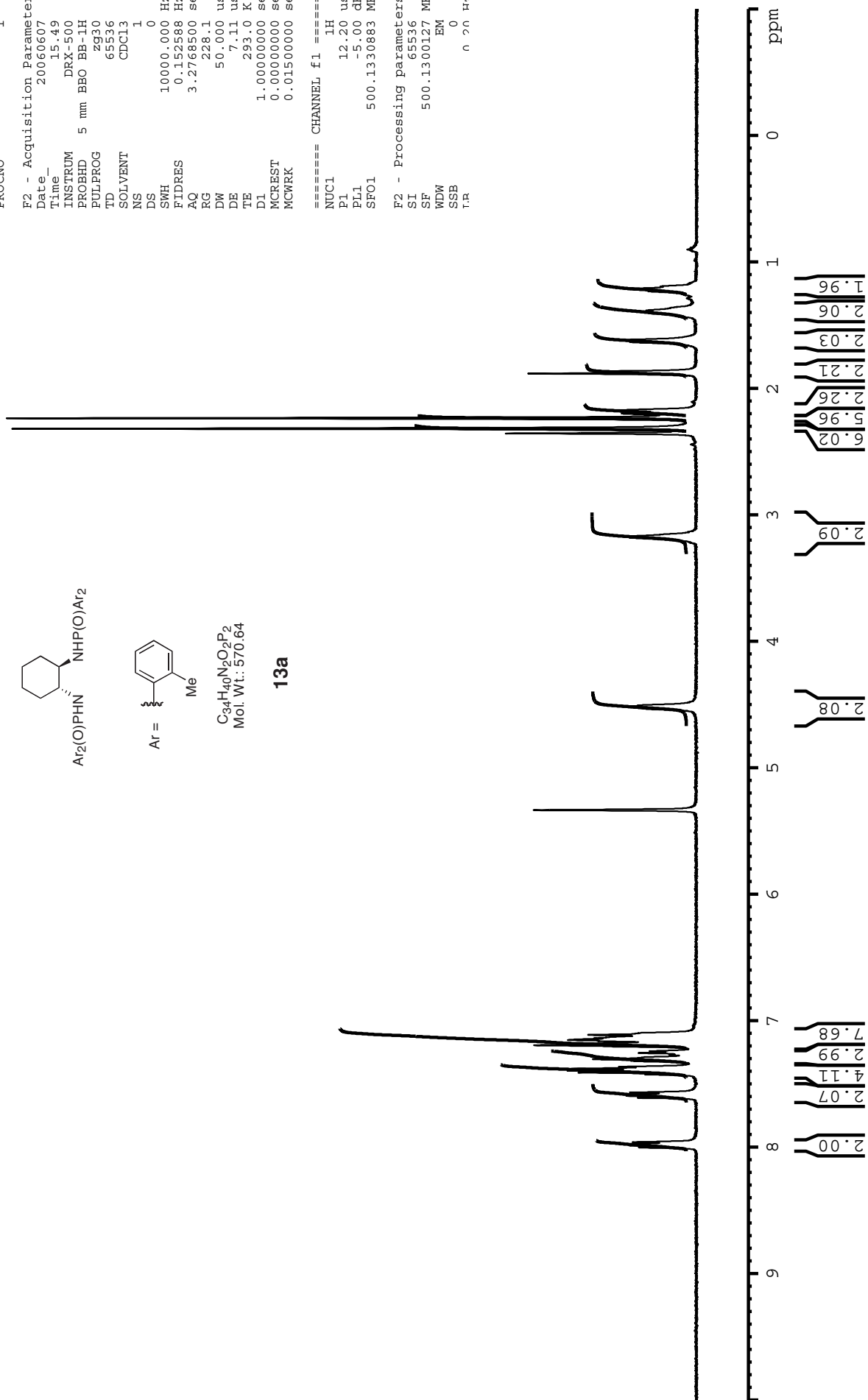
Current Data Parameters
 NAME 3083H
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060607
 Time 15.49
 INSTRUM DRX-500
 PROBHD 5 mm BBO BB-IH
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃

NS 1
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2768500 sec
 RG 228.1
 DW 50.000 use
 DE 7.11 use
 TE 293.0 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 1H
 P1 12.20 use
 PL1 -5.00 dB
 SFO1 500.1330883 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300127 MHz
 WDW EM
 SSB 0
 LR 0.20 Hz



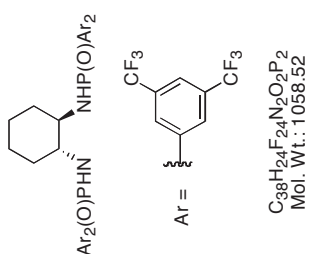
Supporting Information - Watson, Chiu and Bergman

Current Data Parameters
 NAME 3189AH
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060606
 Time 22.02
 INSTRUM AVQ-400
 PROBHD 5 mm QNP 1H/13
 PULPROG zg30
 TD 65536
 SOLVENT CD2Cl2
 NS 1
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 256
 DW 62.400 usec
 DE 6.00 usec
 TE 293.2 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

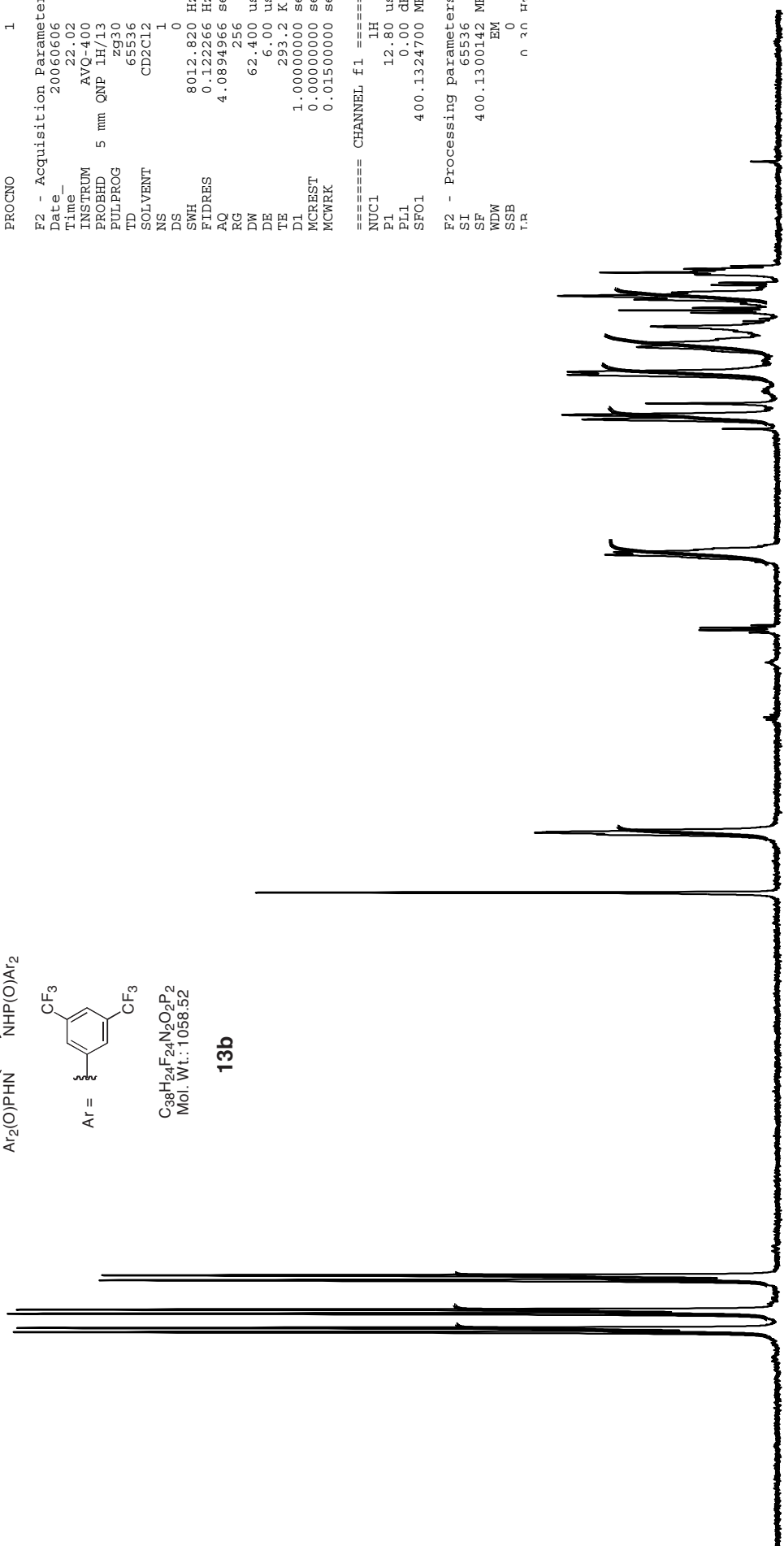
==== CHANNEL f1 =====
 NUC1 1H
 P1 12.80 usec
 PL1 0.00 dB
 SFO1 400.1324700 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300142 MHz
 WDM EM
 SSB 0
 LR 0 30 Hz



8.470
8.440
8.338
8.309
8.098
8.063

5.322
4.893



ppm

9 8 7 6 5 4 3 2 1 0

4.00
4.03
4.02

1.92

2.04
2.12
2.09
1.96

Supporting Information - Watson, Chiu and Bergman

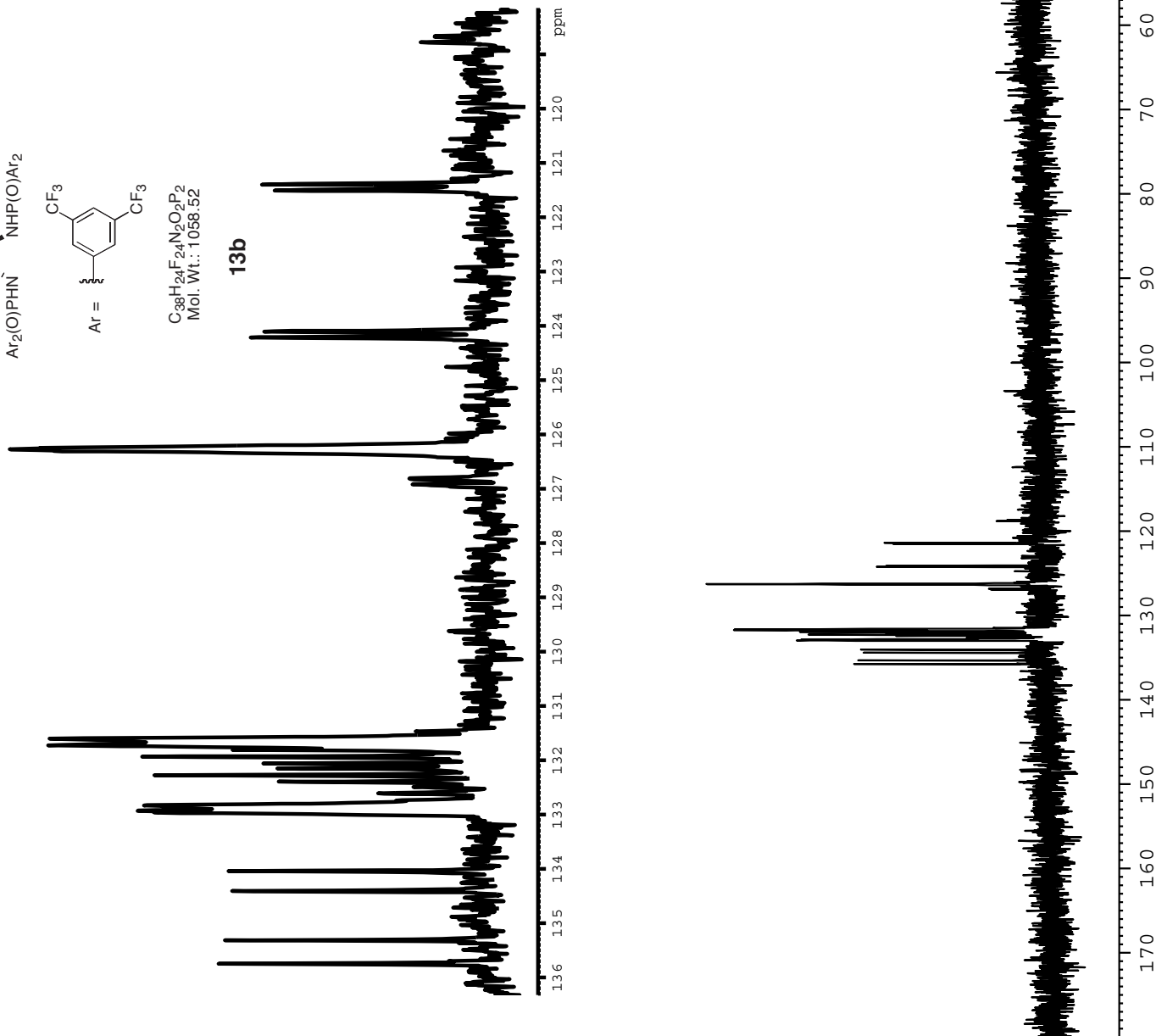
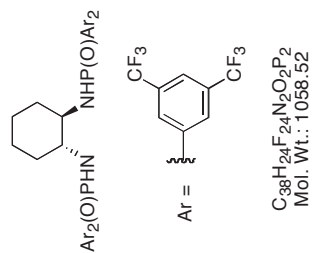
S40

Current Data Parameters
 NAME 3189aC
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060606
 Time 22.09
 INSTRUM AVQ-400
 PROBHD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 829
 DS 0
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 16384
 DW 20.800 usec
 DE 6.00 usec
 TE 293.5 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.89999998 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -2.00 dB
 SFO1 100.6228298 MHz

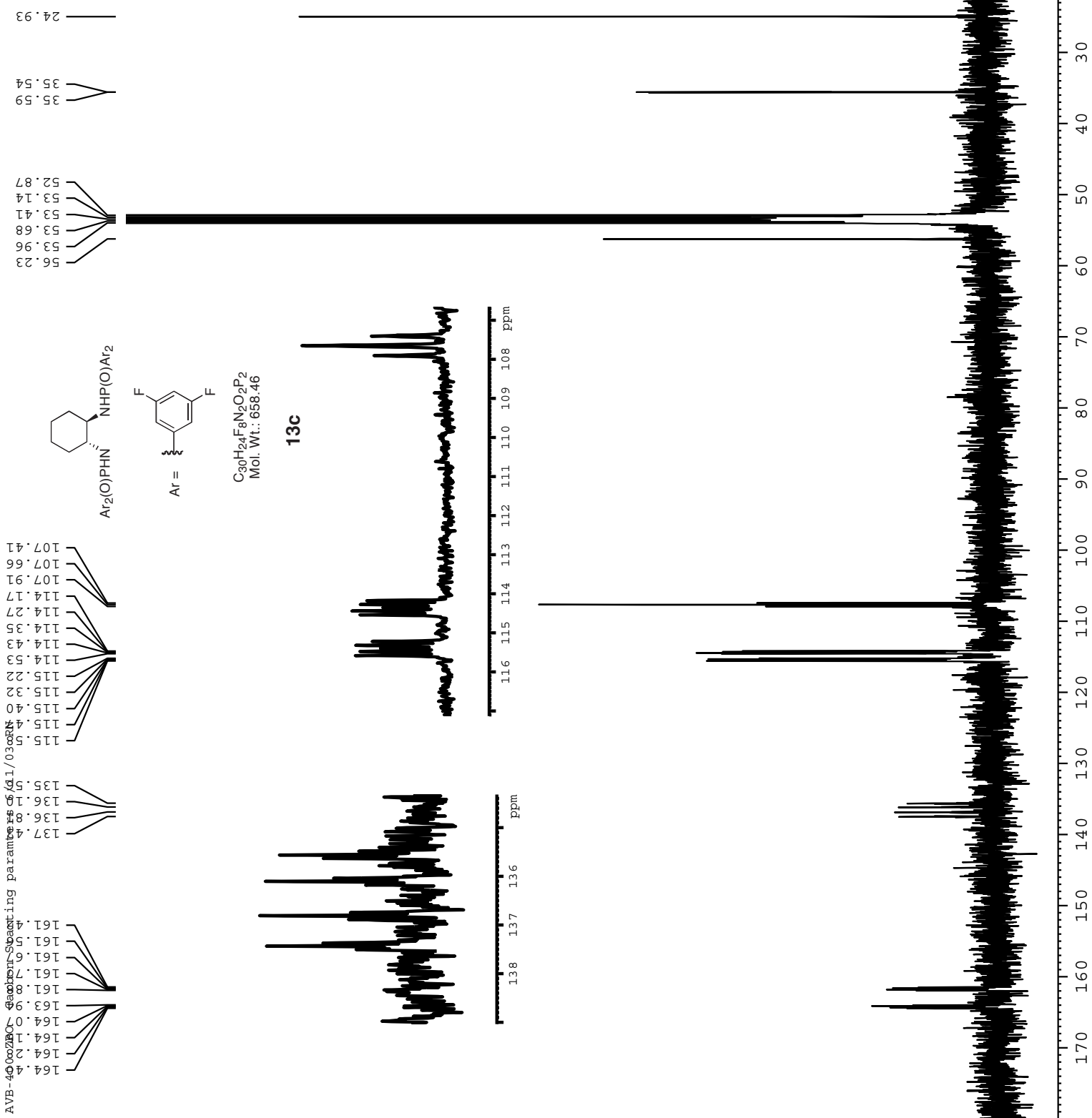
==== CHANNEL f2 =====



Current Data Parameters
 NAME 4119aC
 EXPNO 1
 PROCNO 1
 DU /u
 USER DAW

F2 - Acquisition Parameters
 Date_ 20060610
 Time_ 12.34
 INSTRUM AVB-400
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1586
 DS 0
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664756 sec
 RG 16384
 DW 20.850 usec
 DE 6.00 usec
 TE 295.1 K
 D1 1.5000000 sec
 d11 0.0300000 sec
 DELTA 1.3999998 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -2.00 dB
 SFO1 100.6228298 MHz



Supporting Information - Watson, Chiu and Bergman

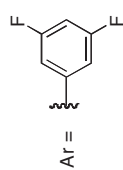
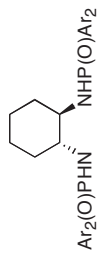
7.520
7.515
7.506
7.501
7.497
7.492
7.488
7.482
7.376
7.371
7.362
7.358
7.353
7.344
7.339
7.330
7.326
7.321
7.078
7.072
7.066
7.056
7.051
7.045
7.035
7.029
7.023
7.019
7.013
7.003
6.997
6.991
6.981
6.975
6.970
5.361
5.358
5.356
4.829
3.478
3.461
2.972
2.964
2.955
2.929
2.156
2.039
2.004
1.790
1.655
1.635
1.447
1.397
1.364
1.267
1.206
1.188
1.177
1.171
1.153
1.128

Current Data Parameters
 NAME 4119AH
 EXPNO 1
 PROCNO 1
 DU /u
 USER DAW

F2 - Acquisition Parameters
 Date_ 20060610
 Time 12.32
 INSTRUM AVB-400
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CD2Cl2
 NS 1
 DS 0
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 256
 DW 60.400 usec
 DE 6.00 usec
 TE 294.5 K
 DT 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

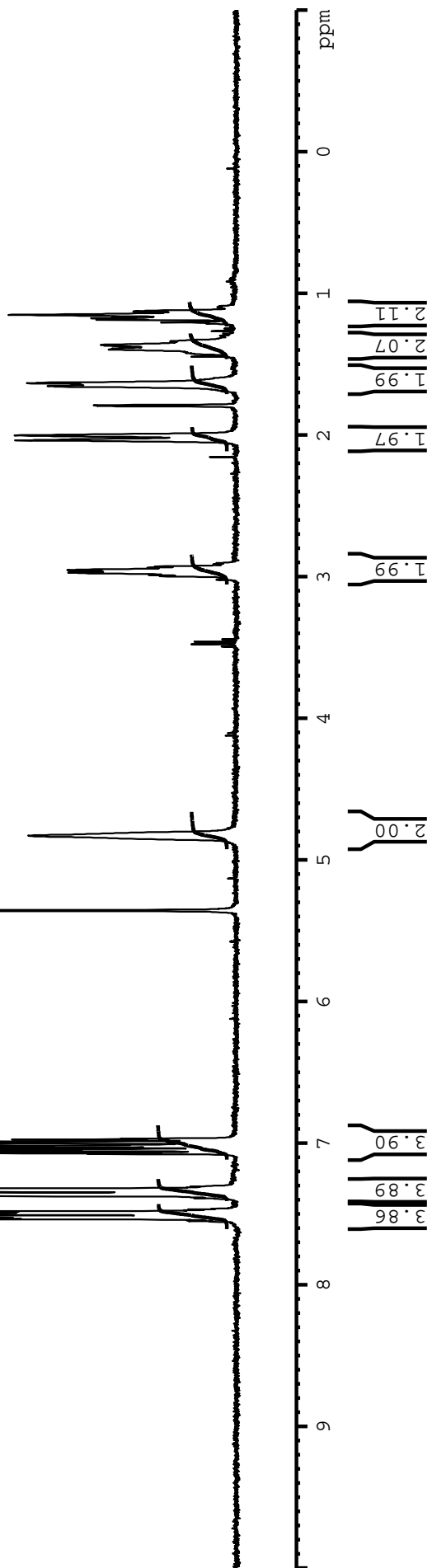
==== CHANNEL f1 =====
 NUC1 1H
 P1 8.20 usec
 PL1 -3.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WTMW RM

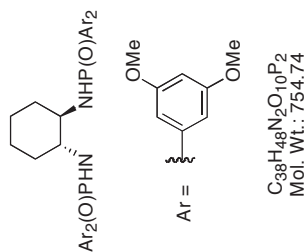
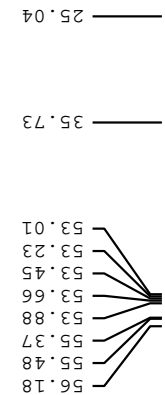
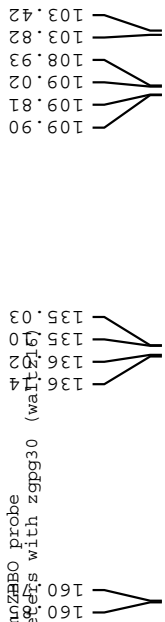


C₃₀H₂₄F₈N₂O₂P₂
 Mol. Wt.: 658.46

13c



13C DRX-500 5mmQNPZABO probe
 starting parameters with zgpg30 (wat-216)
 uses ns*td0



13d

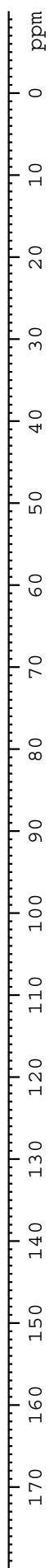
```

Current Data Parameters
NAME           4120c
EXPNO         1
PROCNO        1

F2 - Acquisition Parameters
Date_         20060606
Time_        21.59
INSTRUM      DRX-500
PROBHD       5 mm BBO BB-1H
PULPROG      zgpg30
TD           131072
SOLVENT      CDCl3
NS           158
DS           0
SWH          30864.197 Hz
FIDRES       0.235475 Hz
AQ           2.1234164 sec
RG           16384
DE           16.200 usec
TE           293.0 K
D1           1.5000000 sec
d11          0.0300000 sec
DELTA        1.39999998 sec
MCREST       0.0000000 sec
MCWRK        0.01500000 sec

===== CHANNEL f1 =====
NUC1          13C
P1            8.70 usec
PL1          -3.00 dB
SFO1         125.7722011 MHz

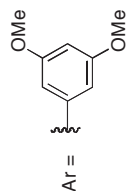
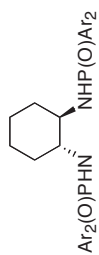
===== CHANNEL f2 =====
    
```



Supporting Information - Watson, Chiu and Bergman

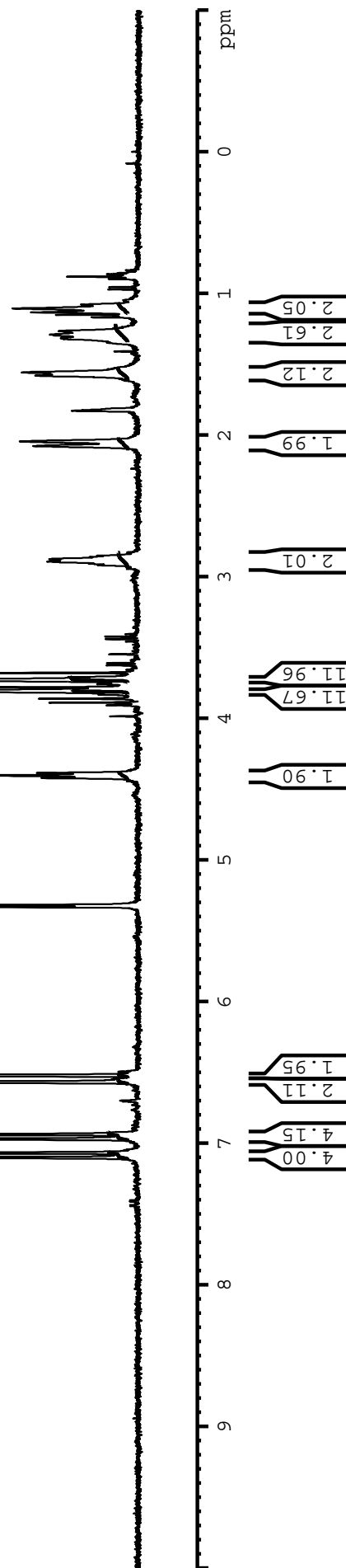
0.883
0.956
0.973
1.107
1.135
1.153
1.170
1.170
1.292
1.411
1.560
1.829
2.046
2.079
2.895
3.424
3.442
3.549
3.611
3.627
3.681
3.709
3.732
3.783
3.810
3.862
3.890
3.909
3.987
4.387
4.405
4.423
5.322
5.334
5.518
6.518
6.523
6.529
6.565
6.570
6.576
6.937
6.943
6.970
6.976
7.067
7.073
7.100
7.106

Current Data Parameters
 NAME 4120AH
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20060601
 Time_ 15.11
 INSTRUM AVQ-400
 PROBDH 5 mm QNP 1H/13
 PULPROG zg30
 TD 65536
 SOLVENT CD2Cl2
 NS 1
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 256
 DW 62.400 usec
 DE 6.00 usec
 TE 293.4 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 12.80 usec
 PL1 0.00 dB
 SFO1 400.1324700 MHz
 F2 - Processing parameters
 SI 65536
 SF 400.1300142 MHz
 WDW EM
 SSB 0
 GB 0
 PR 0
 AR 0
 AS 0
 SR 0
 SSB 0
 T.R 0



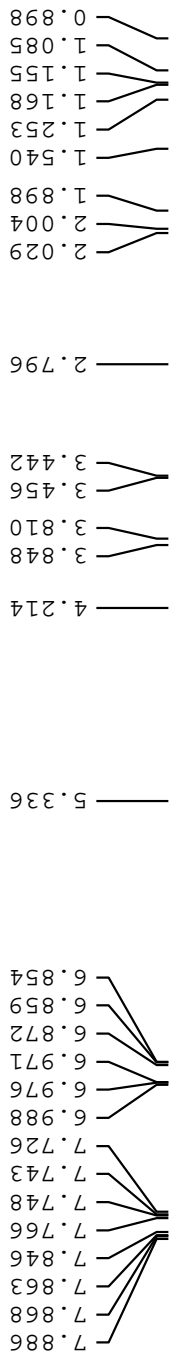
C₃₈H₄₈N₂O₁₀P₂
 Mol. Wt.: 754.74

13d



Supporting Information - Watson, Chiu and Bergman

1H starting parameters (zg30)
 DRX-500 zBBO probe
 =====

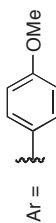
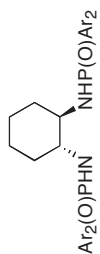


Current Data Parameters
 NAME 3146H
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060606
 Time_ 20.48
 INSTRUM DRX-500
 PROBHD 5 mm BBO BB-IH
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2768500 sec
 RG 322.5
 DW 50.000 usec
 DE 7.11 usec
 TE 293.0 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

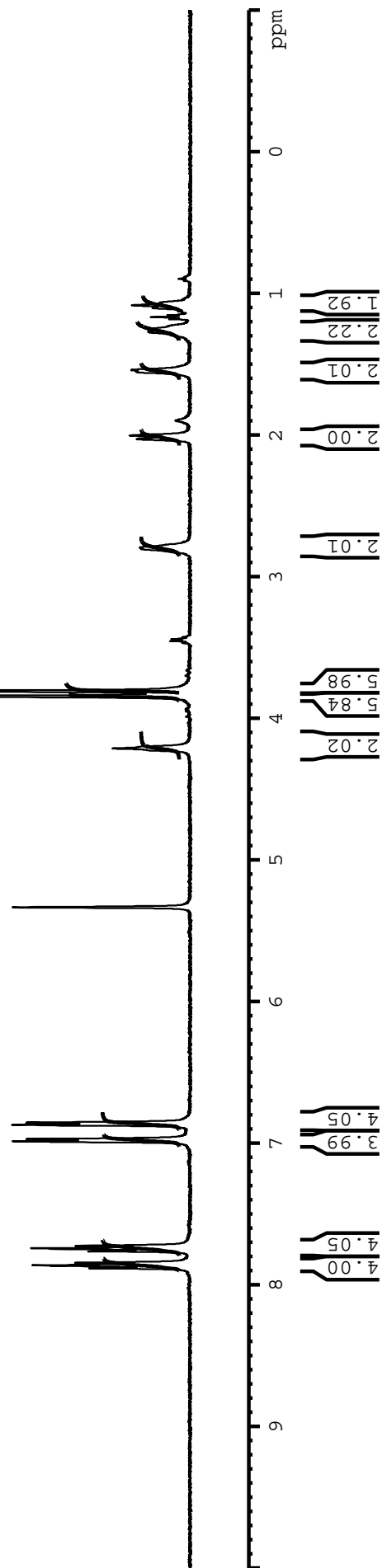
==== CHANNEL f1 =====
 NUC1 1H
 P1 12.20 usec
 PL1 -5.00 dB
 SFO1 500.1330883 MHz

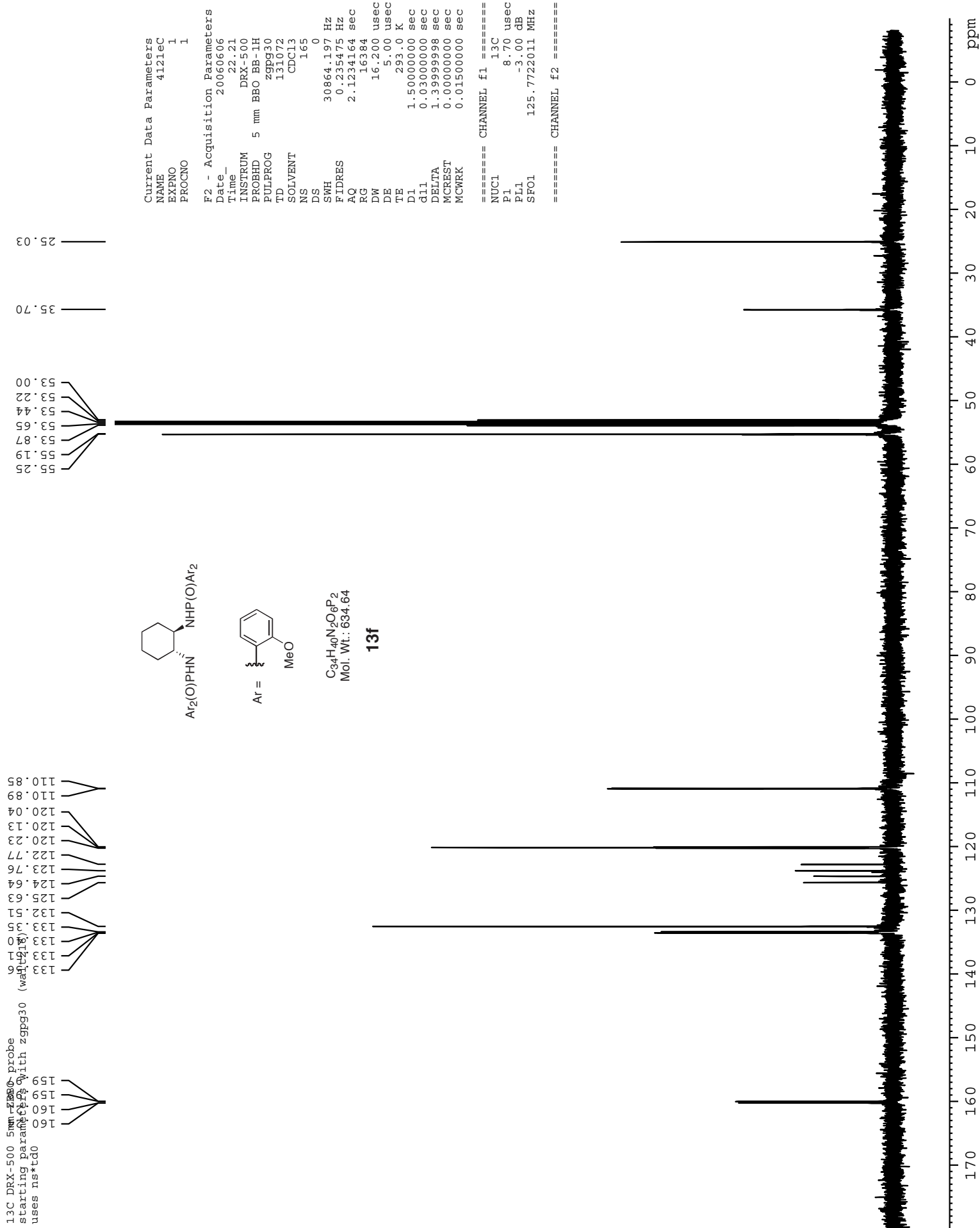
F2 - Processing parameters
 SI 65536
 SF 500.1300127 MHz
 WDW EM
 SSB 0
 LR 0 20 Hz



C₃₄H₄₀N₂O₆P₂
 Mol. Wt.: 634.64

13e





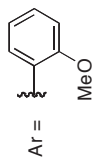
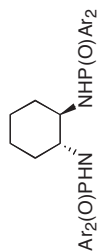
Supporting Information - Watson, Chiu and Bergman

1H starting parameters (zg30)

DRX-500 zBBO probe

=====

7.8888
7.8888
7.8733
7.8600
7.8455
7.7844
7.7699
7.7566
7.7411
7.7436
7.7421
7.4066
7.3833
7.3688
7.3522
7.0422
7.0288
6.8322
6.7555
6.7399
6.7288
6.1033
5.3488
4.1155
3.7277
3.6077
3.5244
3.5022
3.4788
3.2166
2.6399
2.1344
2.1222
2.0411
2.0211
1.8844
1.5722
1.2444



C₃₄H₄₀N₂O₆P₂
Mol. Wt.: 634.64

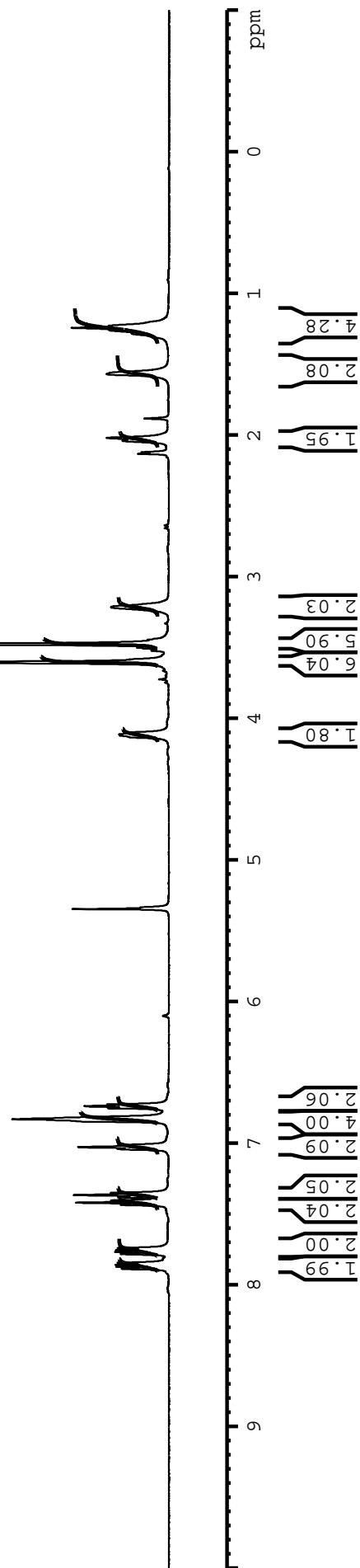
13f

Current Data Parameters
NAME 4121eh
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20060606
Time 22.19
INSTRUM DRX-500
PROBHD 5 mm BBO BB-IH
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 1
DS 0
SWH 10000.000 Hz
FIDRES 0.152588 Hz
AQ 3.2768500 sec
RG 45.3
DW 50.000 usec
DE 7.11 usec
TE 293.0 K
D1 1.0000000 sec
MCREST 0.0000000 sec
MCWRK 0.01500000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 12.20 usec
PL1 -5.00 dB
SFO1 500.1330883 MHz

F2 - Processing parameters
SI 65536
SF 500.1300127 MHz
WDW EM
SSB 0
LR 0.20 Hz



Supporting Information - Watson, Chiu and Bergman

Current Data Parameters
 NAME 3112AH
 EXPNO 1
 PROCNO 1
 DU /u
 USER DAW

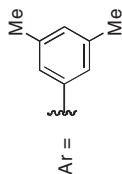
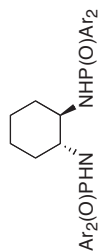
F2 - Acquisition Parameters
 Date_ 20060206
 Time 14.19
 INSTRUM AVB-400
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 322.5
 DW 60.400 usec
 DE 6.00 usec
 TE 294.6 K
 DL 1.0000000 sec
 MCREST 0.0000000 sec
 MCVRK 0.0150000 sec

==== CHANNEL f1 =====
 NUC1 1H
 P1 8.20 usec
 PL1 -3.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WTMW RM

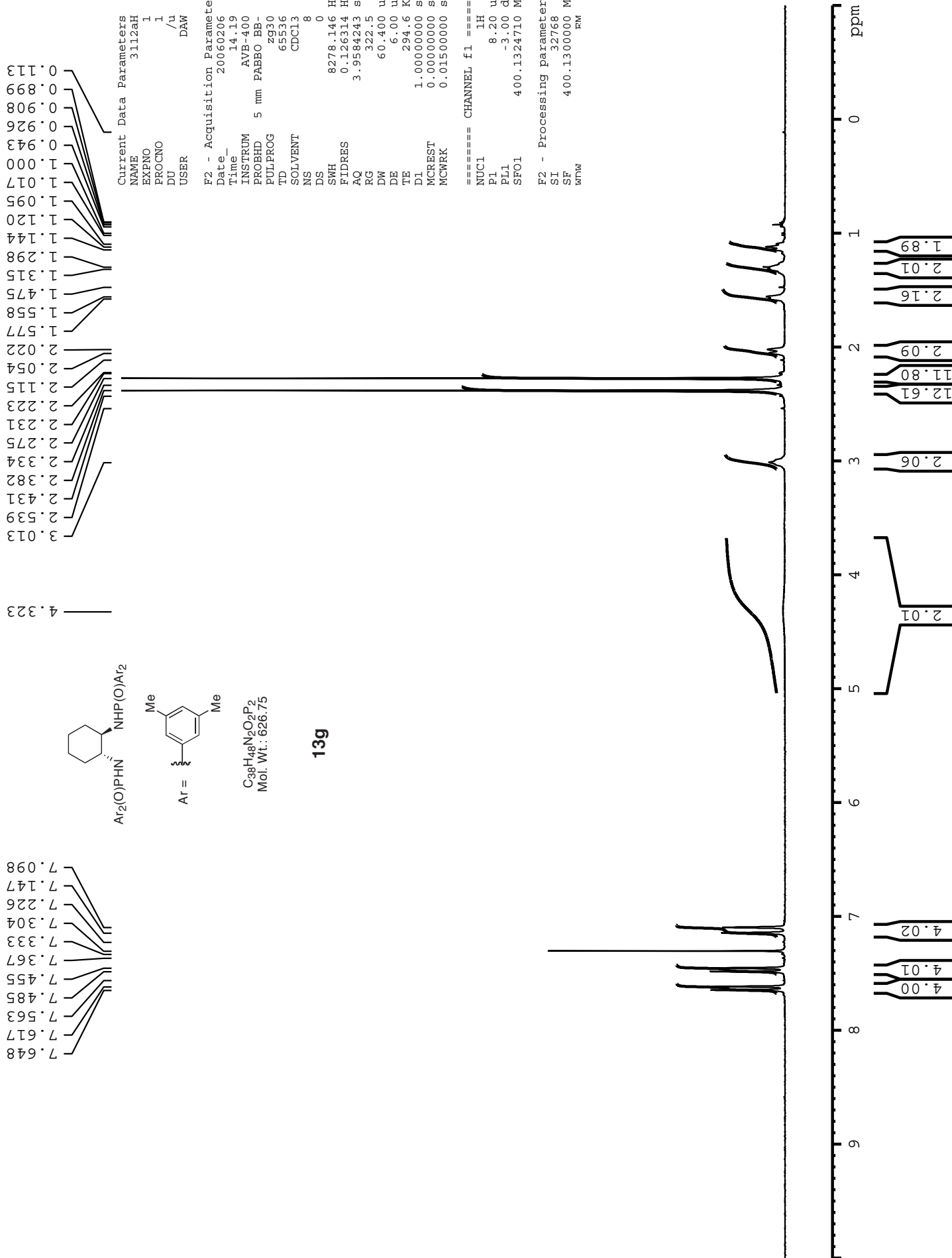
7.648
7.617
7.563
7.485
7.455
7.367
7.333
7.304
7.226
7.147
7.098

4.323



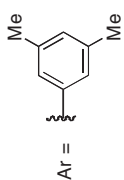
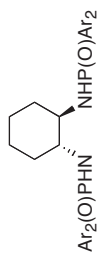
C₃₈H₄₈N₂O₂P₂
 Mol. Wt.: 626.75

13g



13C DRX-500 5mm ZBBO probe
 starting parameters with zgpg30 (waltz16)
 uses ns*td0

130.0
 128.0
 129.0
 128.0
 96.0



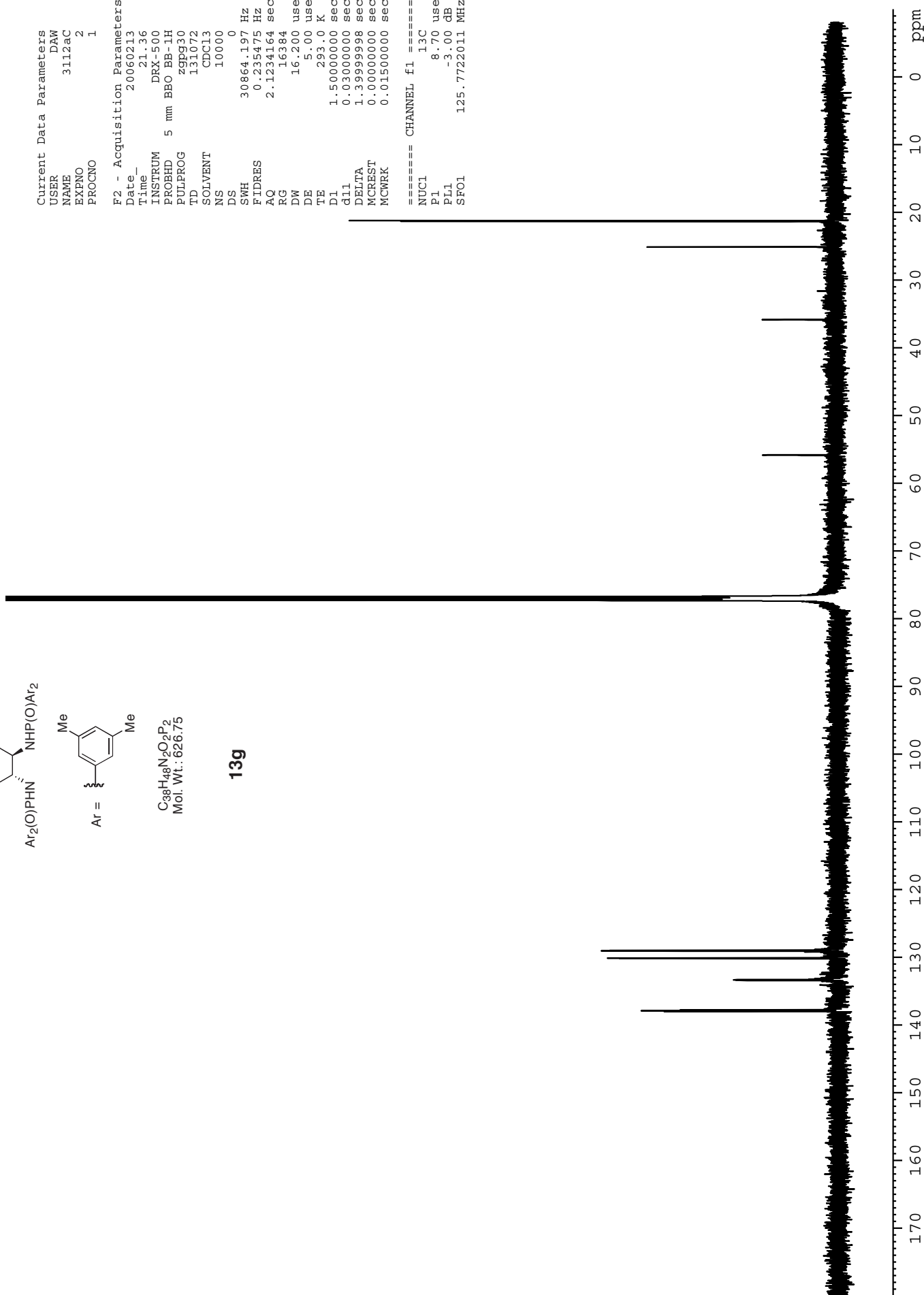
C₃₈H₄₈N₂O₂P₂
 Mol. Wt.: 626.75

13g

Current Data Parameters
 USER DAW
 NAME 3112aC
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060213
 Time_ 21.36
 INSTRUM DRX-500
 PROBED 5 mm BBO BB-IH
 PULPROG zgpg30
 TD 131072
 SOLVENT CDCl₃
 NS 10000
 DS 0
 SWH 30864.197 Hz
 FIDRES 0.235475 Hz
 AQ 2.1234164 sec
 RG 16384
 DW 16.200 usec
 DE 5.00 usec
 TE 293.0 K
 D1 1.5000000 sec
 d11 0.0300000 sec
 DELTA 1.3999998 sec
 MCREST 0.0000000 sec
 MCMRK 0.015000000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.70 usec
 PL1 -3.00 dB
 SFO1 125.7722011 MHz



Supporting Information - Watson, Chiu and Bergman

S52

1H starting parameters (zg30)
 DRX-500 zBBO probe

7.877
7.851
7.780
7.755
7.573
7.553

5.335
4.429

2.873

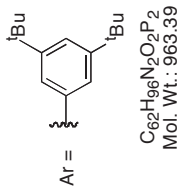
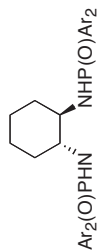
2.150
2.125
2.023
1.539
1.484
1.361
1.286
1.158
1.083
0.992
0.979
0.904

Current Data Parameters
 NAME 4015aH
 EXPNO 1
 PROCNO 1

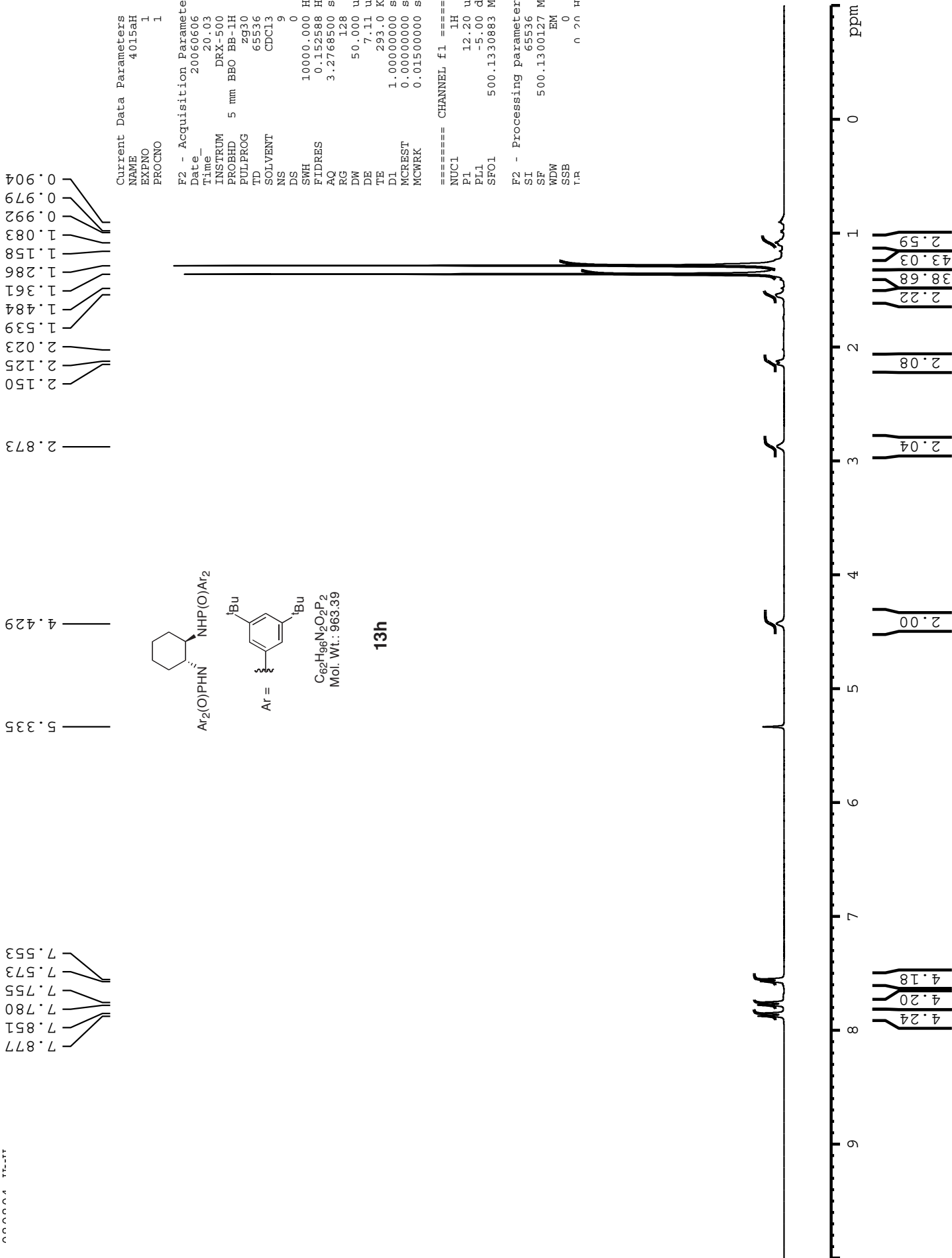
F2 - Acquisition Parameters
 Date_ 20060606
 Time_ 20.03
 INSTRUM DRX-500
 PROBHD 5 mm BBO BB-IH
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 9
 DS 0
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2768500 sec
 RG 128
 DW 50.000 usec
 DE 7.11 usec
 TE 293.0 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

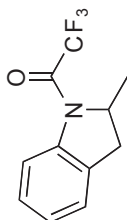
==== CHANNEL f1 =====
 NUC1 1H
 P1 12.20 usec
 PL1 -5.00 dB
 SFO1 500.1330883 MHz

F2 - Processing parameters
 SI 65536
 SF 500.1300127 MHz
 WDW EM
 SSB 0
 LB 0.20 Hz



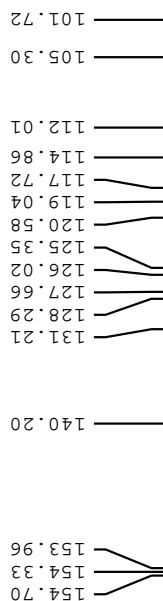
13h





C₁₁H₁₀F₃NO
Mol. Wt.: 229.20

18



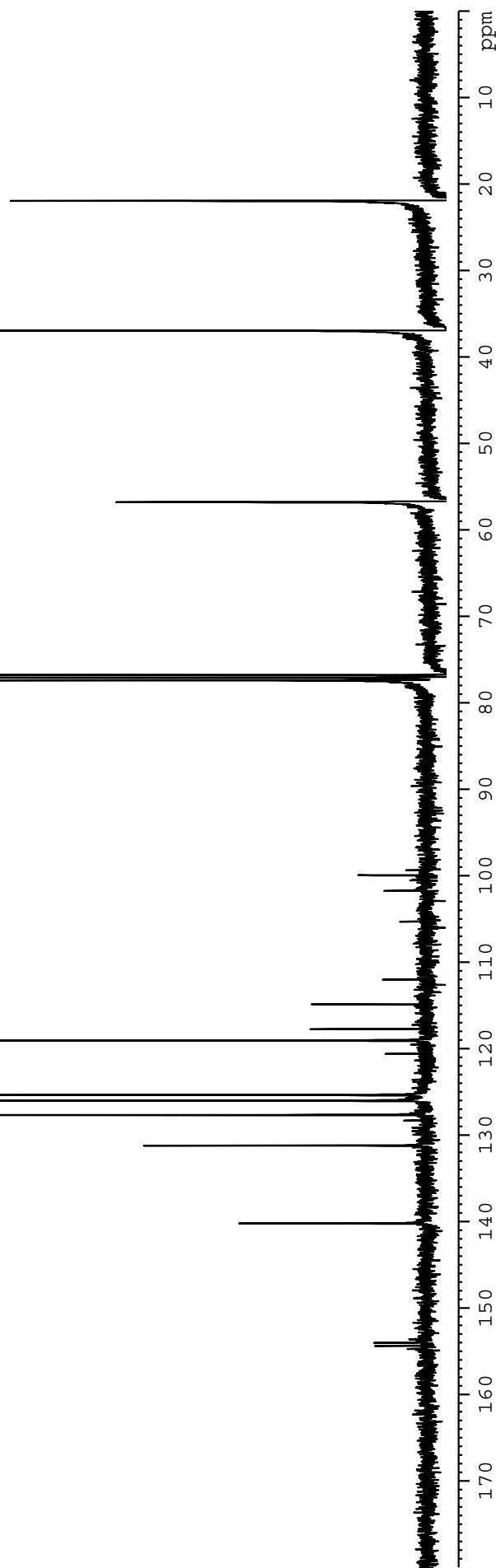
AVQ-400 QNP Carbon Starting parameters 7/16/03 revised 7

Current Data Parameters
 NAME mc3p039pdt
 EXPNO 13
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060707
 Time 18.44
 INSTRUM AVQ-400
 PROBD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 77
 DS 0
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 25.4
 DW 20.800 usec
 DE 6.00 usec
 TE 293.0 K
 D1 2.0000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.50 usec
 PL1 -2.00 dB
 SFO1 100.6228298 MHz

==== CHANNEL f2 =====



Supporting Information - Watson, Chiu and Bergman

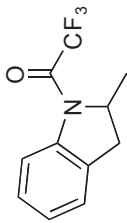
0.043
8.023
7.384
7.260
7.205
7.187
7.167
7.098
7.079
7.061
4.751
4.734
4.717
4.700
4.684
3.397
3.365
3.345
3.326
3.307
2.650
2.611
2.257
1.949
1.724
1.403
1.387
1.246
1.230
1.137
1.120
1.102
1.084
1.068
0.797

Current Data Parameters
NAME mc3p039pdt
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20060707
Time 18.41
INSTRUM AVQ-400
PROBHD 5 mm QNP 1H/13
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 8
DS 0
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894966 sec
RG 45.3
DW 62.400 use
DE 6.00 use
TE 292.7 K
D1 1.0000000 sec
MCREST 0.0000000 sec
MCWRK 0.01500000 sec

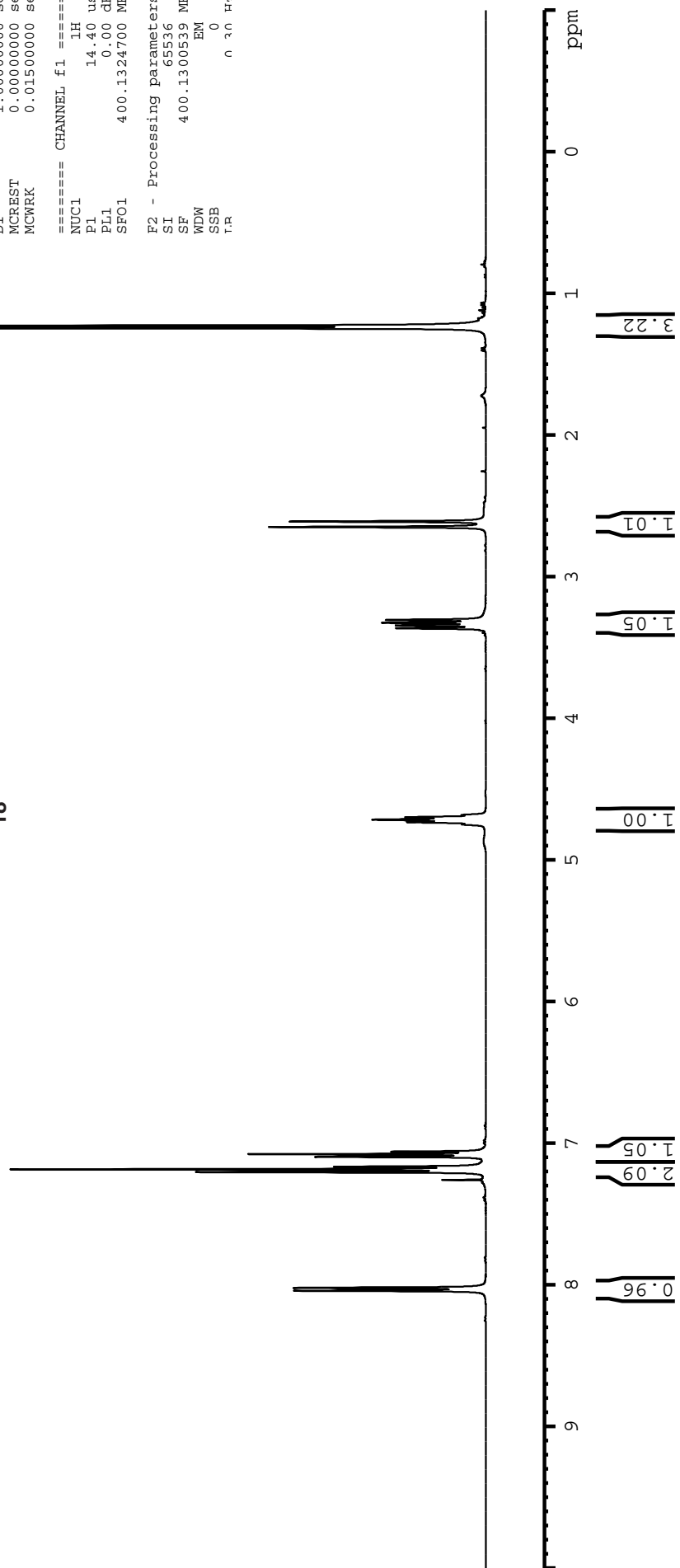
==== CHANNEL f1 =====
NUC1 1H
P1 14.40 use
PL1 0.00 dB
SFO1 400.1324700 MHz

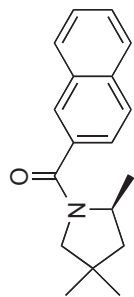
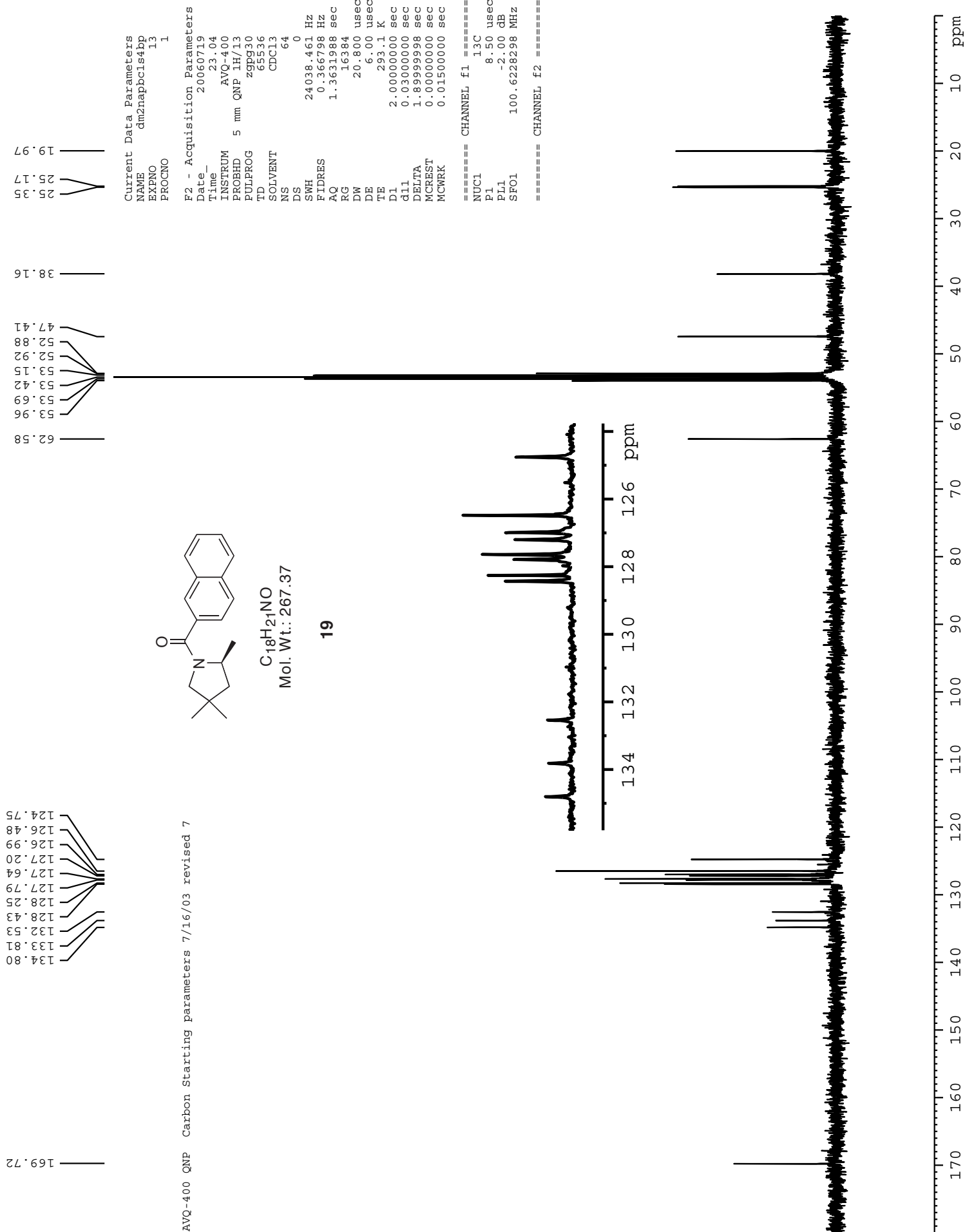
F2 - Processing parameters
SI 65536
SF 400.1300539 MHz
WDW EM
SSB 0
LR 0 20 Hz



C₁₁H₁₀F₃NO
Mol. Wt.: 229.20

18





C₁₈H₂₁NO
 Mol. Wt.: 267.37

19

AVO-400 QNP Carbon Starting parameters 7/16/03 revised 7

Supporting Information - Watson, Chiu and Bergman

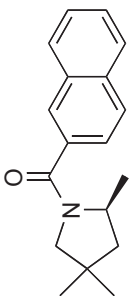
7.934
7.920
7.908
7.908
7.894
7.885
7.819
7.706
7.682
7.644
7.623
7.595
7.592
7.584
7.578
7.567
7.564
7.561
7.553
7.546
7.543
7.541
7.529
7.524
7.524
7.370
5.324
5.322
5.319
4.426
4.411
4.394
4.387
4.378
4.371
4.354
4.339
4.181
3.754
3.551
3.405
3.379
3.252
3.202
3.157
3.131
2.935
2.005
2.001
1.987
1.982
1.974
1.969
1.955
1.951
1.815
1.594
1.578
1.496
1.472
1.465
1.438
1.422
1.344
1.289
1.277
1.262
1.199

Current Data Parameters
 NAME dm2napbcis4bp
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060719
 Time 23.00
 INSTRUM AVQ-400
 PROBHD 5 mm QNP 1H/13
 PULPROG zg30
 TD 65536
 SOLVENT CD2Cl2
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 50.8
 DE 62.400 use
 TE 292.9 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

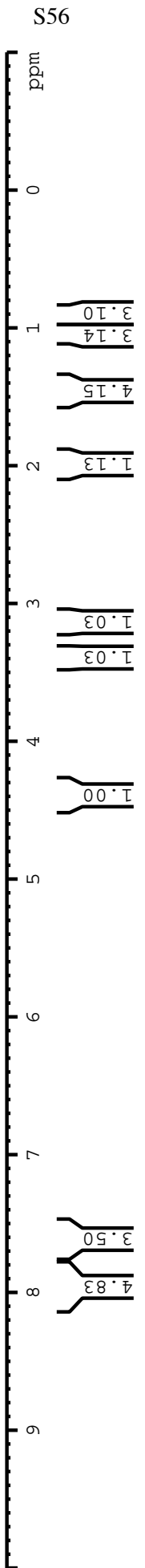
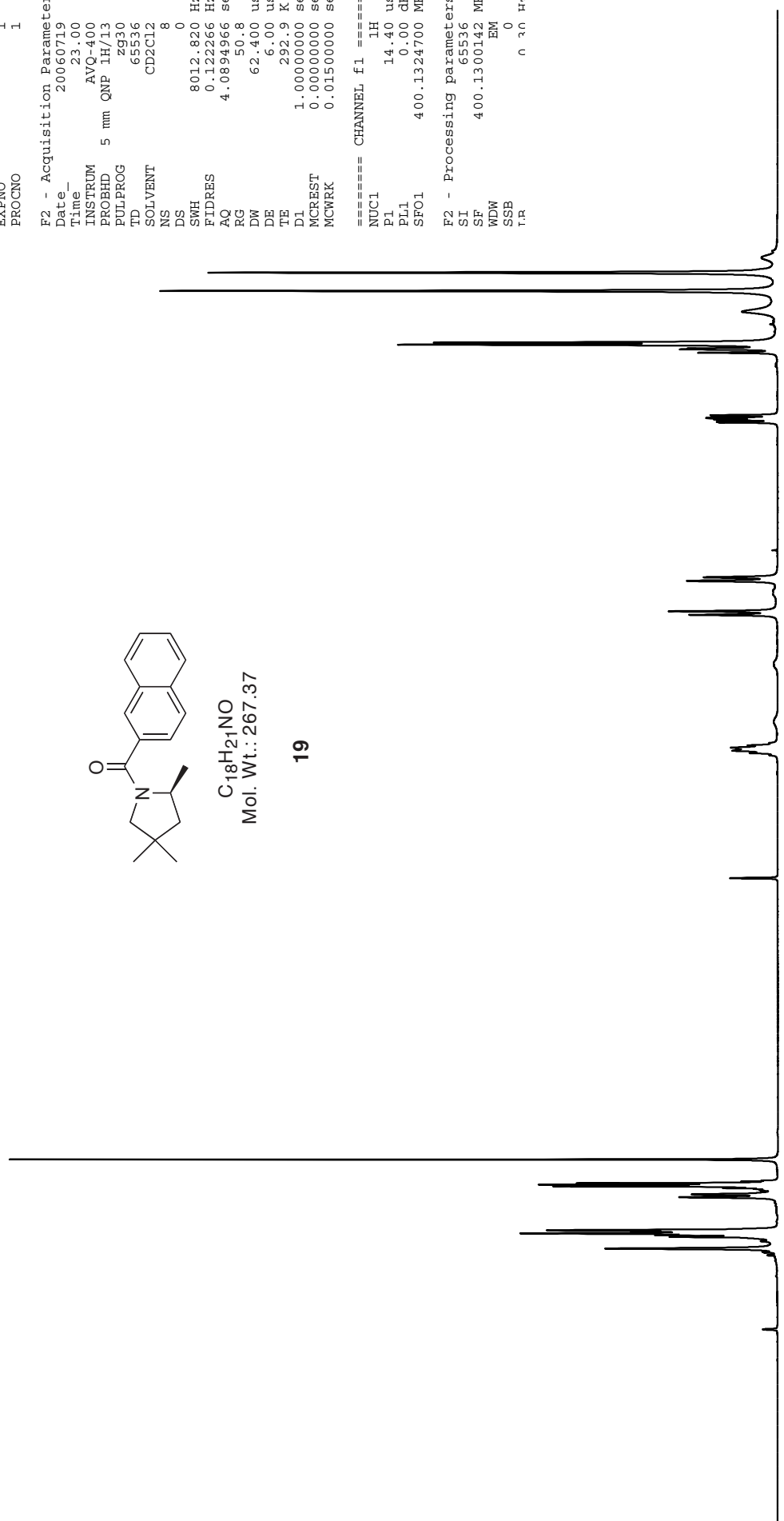
==== CHANNEL f1 =====
 NUC1 1H
 P1 14.40 use
 PL1 0.00 dB
 SFO1 400.1324700 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300142 MHz
 WDW EM
 SSB 0
 T.R 0 30 Hz



C₁₈H₂₁NO
 Mol. Wt.: 267.37

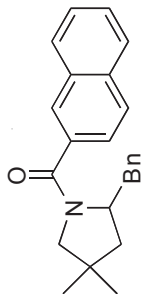
19



Supporting Information - Watson, Chiu and Bergman

S57

13C DRX-500 5mm ZBBO probe
 starting parameters with zgpg30 (waltz16)
 uses ns*td0
 012504 HH



C₂₄H₂₅NO
 Mol. Wt.: 343.46

20

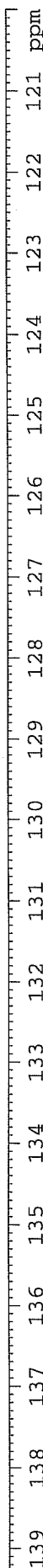
Current Data Parameters
 NAME bn2nabc2s3bp3
 EXPNO 13
 PROCNO 1
 DU /u
 USER labrat

F2 - Acquisition Parameters
 Date_ 20060722
 Time 16.28
 INSTRUM DRX-500
 PROBHD 5 mm BBO BB-LH
 PULPROG zgpg30
 TD 131072
 SOLVENT CDCl3
 NS 608
 DS 0
 SMH 30864.197 Hz
 FIDRES 0.235475 Hz
 AQ 2.1234164 sec
 RG 16384
 DW 16.200 usec
 DE 5.00 usec
 TE 293.0 K
 D1 1.5000000 sec
 d11 0.0300000 sec
 DELTA 1.3999998 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.70 usec
 PL1 -3.00 dB
 SF01 125.7722011 MHz

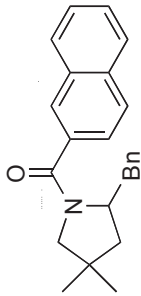
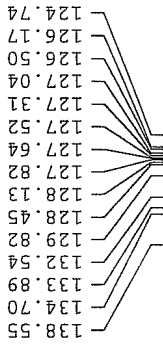
==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 97.00 usec
 PL2 -5.00 dB
 PL12 15.50 dB
 PL13 21.50 dB
 SF02 500.1321560 MHz

F2 - Processing parameters
 SI 131072
 SF 125.7577971 MHz
 WDW EM
 SSB 0
 LB 0.75 Hz
 GB 0
 PC 4.00



Supporting Information - Watson, Chiu and Bergman

13C DRX-500 5mm ZBBO probe
 starting parameters with zgpg30 (waltz16)
 uses ns*td0
 012504 HvH



C₂₄H₂₅NO
 Mol. Wt.: 343.46

20

Current Data Parameters
 NAME: C bn2nbc23bp3
 EXPNO: 13
 PROCNO: 1
 DU: /u
 USER: labrat

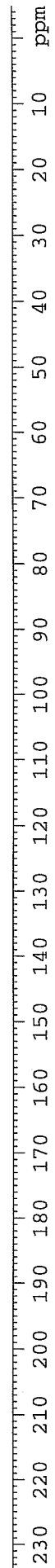
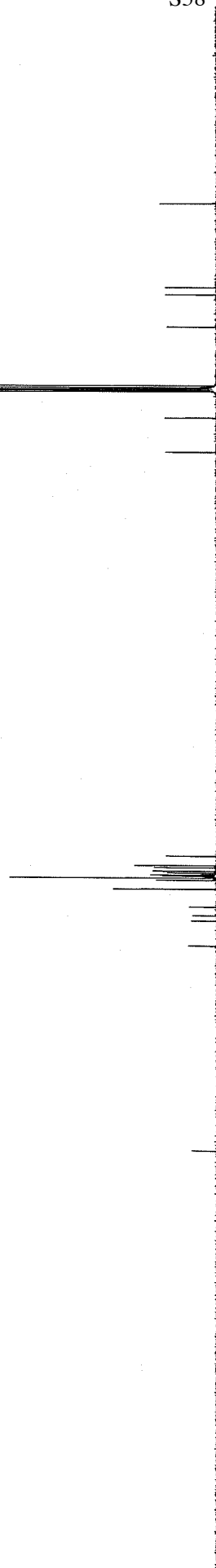
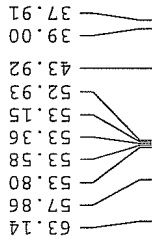
F2 - Acquisition Parameters

Date_ 20060722
 Time 16.28
 INSTRUM DRX-500
 PROBHD 5 mm BBO BB-1H
 PULPROG zgpg30
 TD 131072
 SOLVENT CDCl3
 NS 592
 DS 0
 SWH 30864.197 Hz
 FIDRES 0.235475 Hz
 AQ 2.1234164 sec
 RG 16384
 DE 16.200 usec
 TE 293.0 K
 D1 1.5000000 sec
 d11 0.0300000 sec
 DELTA 1.3999998 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.70 usec
 PL1 -3.00 dB
 SFO1 125.7722011 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 97.00 usec
 PL2 -5.00 dB
 PL12 15.50 dB
 PL13 21.50 dB
 SFO2 500.1321560 MHz

F2 - Processing parameters
 SI 131072
 SF 125.7577971 MHz
 WDM EM
 SSB 0
 LB 0.75 Hz
 GB 0
 PC 4.00



Supporting Information - Watson, Chiu and Bergman

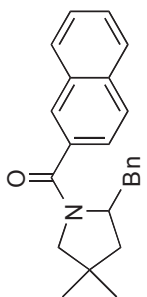
8.116
7.993
7.811
7.790
7.644
7.614
7.593
7.572
7.318
7.301
7.266
7.253
7.248
7.239
7.229
7.224
7.211
7.197
7.179
7.150
7.124
6.954
4.759
4.740
3.491
3.461
3.061
3.039
3.009
2.934
2.909
2.804
2.779
1.966
1.415
1.382
1.372
1.365
1.349
1.317
1.264
1.227
0.979
0.960
0.933
0.891
0.791
0.737
0.564
0.543

Current Data Parameters
 NAME bn2napc2s3bp2
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060722
 Time 14.17
 INSTRUM AVQ-400
 PROBHD 5 mm QNP 1H/13
 PULPROG zg30
 TD 65536
 SOLVENT C6D6
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 128
 DW 62.400 use
 DE 6.00 use
 TE 294.3 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

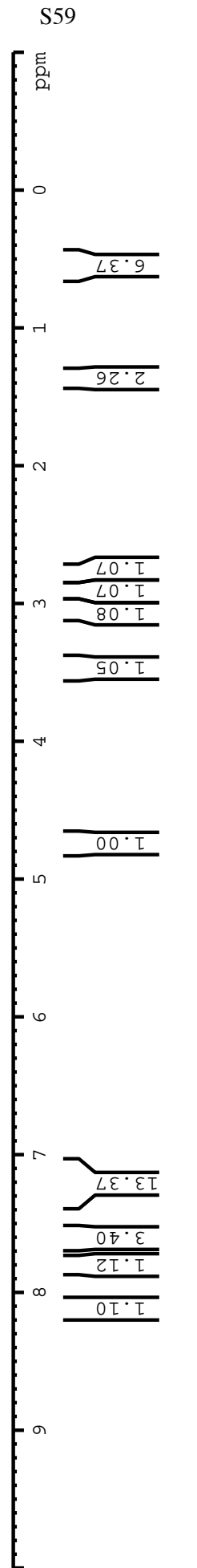
==== CHANNEL f1 =====
 NUC1 1H
 P1 14.40 use
 PL1 0.00 dB
 SFO1 400.1324700 MHz

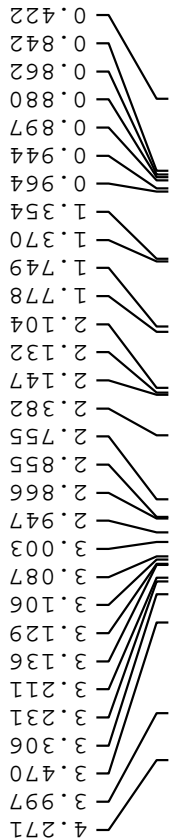
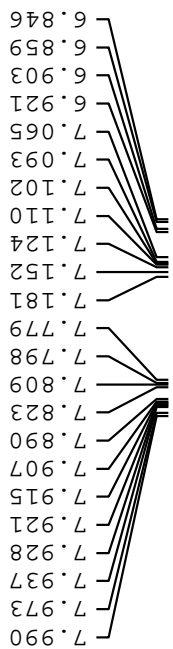
F2 - Processing parameters
 FI 65536
 SF 400.1300461 MHz
 TDW EM
 ASB 0
 IR 0 2x Hz



C₂₄H₂₅NO
 Mol. Wt.: 343.46

20

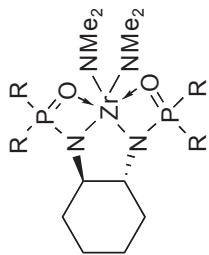




Current Data Parameters
 USER DAW
 NAME 3170aH13515m
 EXPNO 1
 PROCNO 1

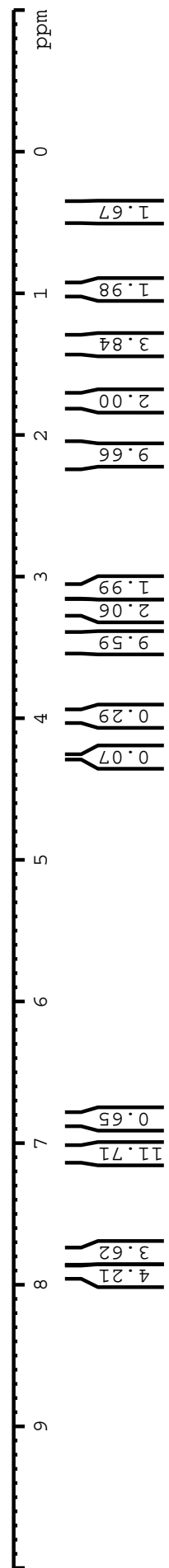
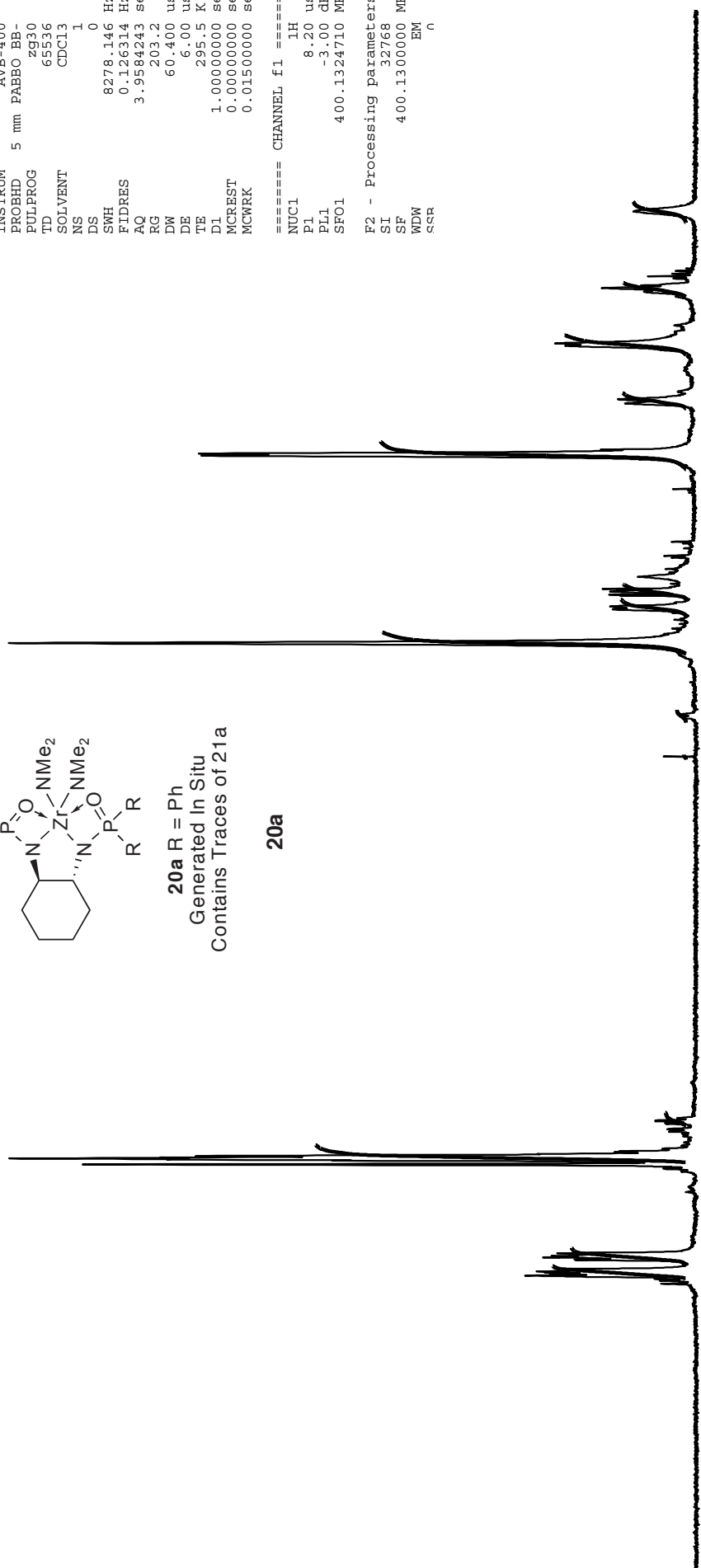
F2 - Acquisition Parameters
 Date_ 20060214
 Time 14.09
 INSTRUM AVB-400
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 1
 DS 0
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 203.2
 DW 60.400 use
 DE 6.00 use
 TE 295.5 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 1H
 P1 8.20 use
 PL1 -3.00 dB
 SFO1 400.1324710 MHz
 F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WDW EM
 SSR n



20a R = Ph
 Generated In Situ
 Contains Traces of 21a

20a



Supporting Information - Watson, Chiu and Bergman

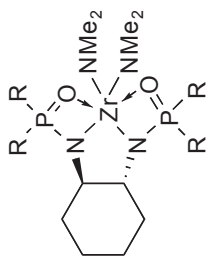
0.291
1.101
1.458
1.622
1.671
1.759
1.782
1.817
1.880
2.024
2.039
2.074
2.104
2.115
2.142
2.181
2.196
2.276
2.361
2.523
2.755
2.856
2.972
3.053
3.087
3.130
3.160
3.339
3.568

Current Data Parameters
NAME 3168AH456d
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20060220
Time 12.34
INSTRUM AVB-400
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT C6D6
NS 1
DS 0
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 228.1
DW 60.400 usec
DE 6.00 usec
TE 294.6 K
D1 1.0000000 sec
MCREST 0.0000000 sec
MCWRK 0.01500000 sec

==== CHANNEL f1 =====
NUC1 1H
P1 8.20 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz

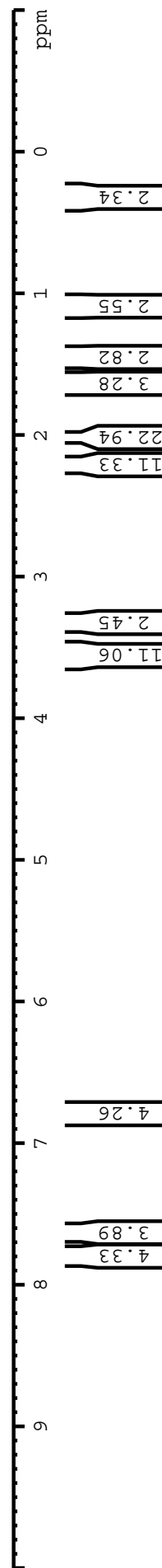
F2 - Processing parameters
SI 32768
SF 400.1300000 MHz
WDW EM
SSB 0
LR 0.30 Hz



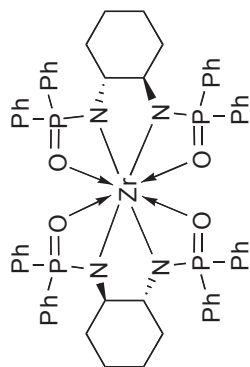
20b R = 3,5-C₆H₃Me₂
Generated In Situ

20b

7.803
7.771
7.657
7.626
7.152
6.821
6.777



¹³C DRX-500 5mm ZBBO probe
starting parameters with zgpg30 (waltz16)
uses ns*td0



$C_{60}H_{60}N_4O_4P_4Zr$
Mol. Wt.: 1116.2618

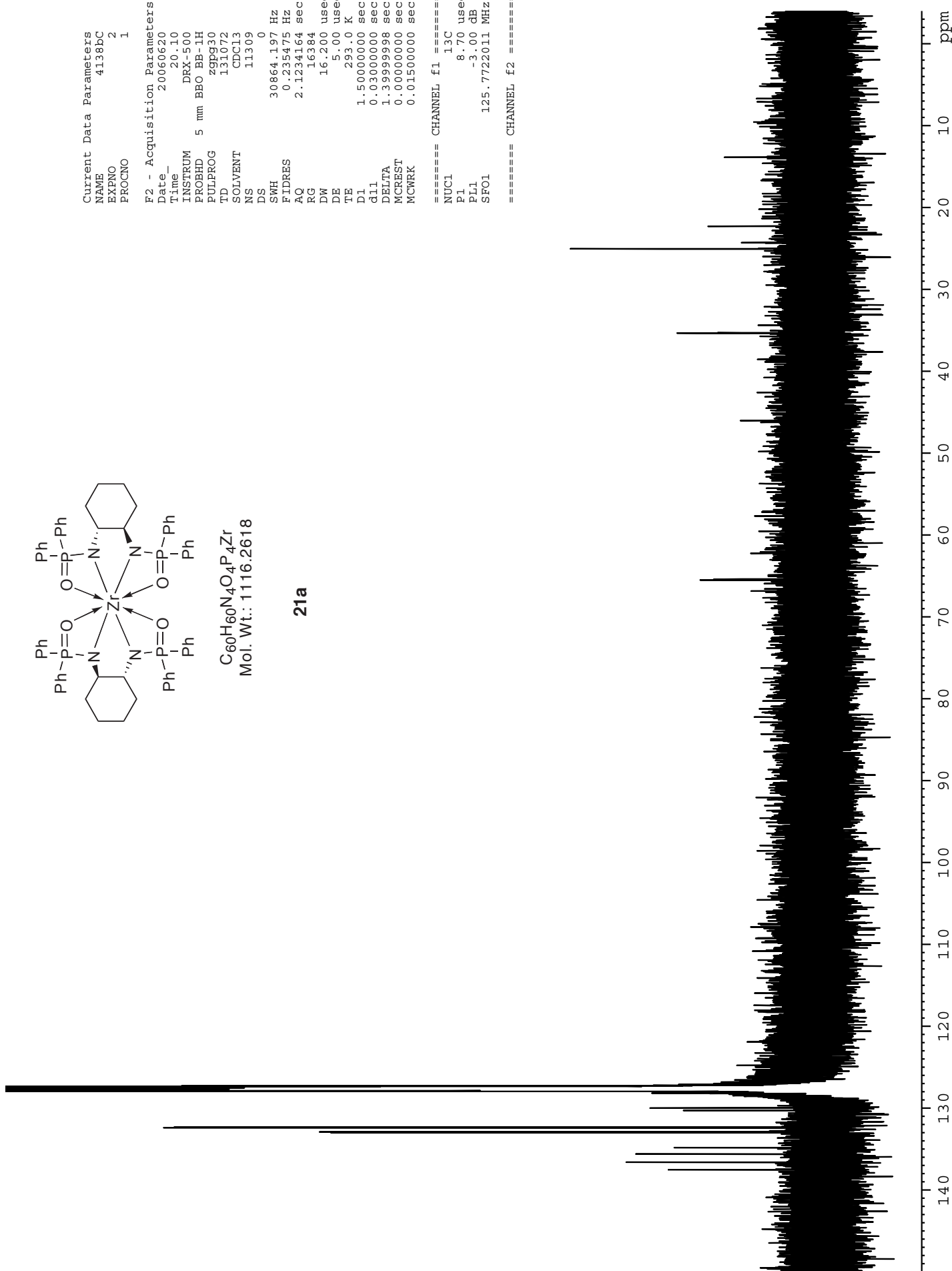
21a

```
Current Data Parameters
NAME          4138bC
EXPNO        2
PROCNO       1

F2 - Acquisition Parameters
Date_        20060620
Time         20.10
INSTRUM      DRX-500
PROBHD       5 mm BBO BB-1H
PULPROG      zgpg30
TD           131072
SOLVENT      CDCl3
NS           11309
DS           0
SWH          30864.197 Hz
FIDRES       0.235475 Hz
AQ           2.1234164 sec
RG           16384
DE           16.200 usec
TE           293.0 K
D1           1.5000000 sec
d11          0.03000000 sec
DELTA        1.39999998 sec
MCREST       0.0000000 sec
MCWRK        0.01500000 sec

===== CHANNEL f1 =====
NUC1         13C
P1           8.70 usec
PL1         -3.00 dB
SFO1        125.7722011 MHz

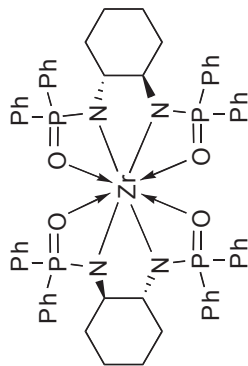
===== CHANNEL f2 =====
```



Supporting Information - Watson, Chiu and Bergman

4.083
4.059

1.779
1.606
1.353
1.083



$C_{60}H_{60}N_4O_4P_4Zr$
Mol. Wt.: 1116.2618

21a

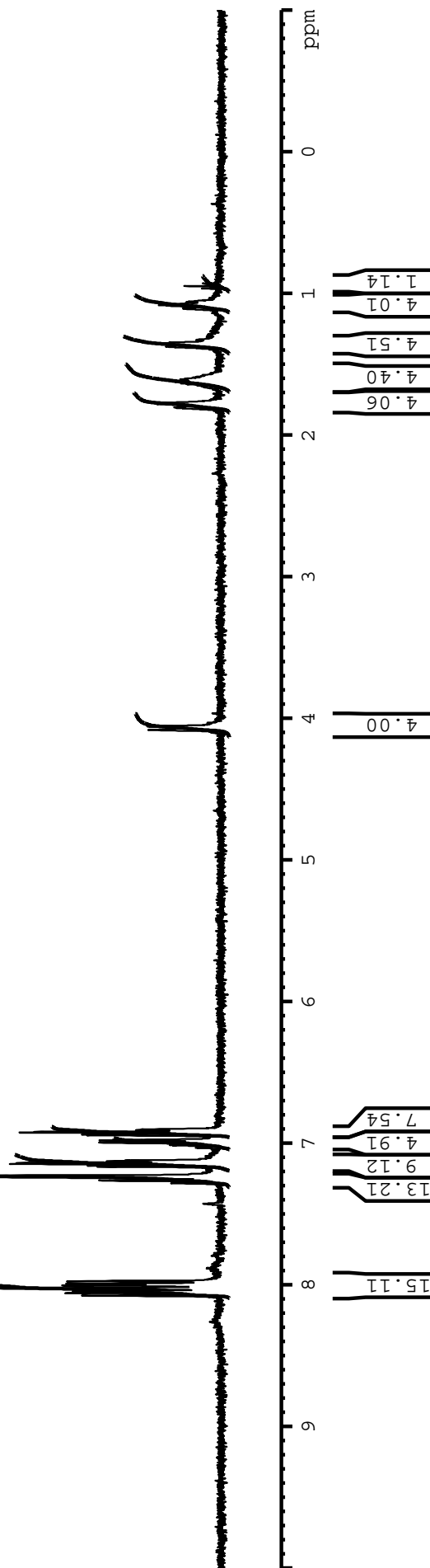
```

Current Data Parameters
NAME      4138BH
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20060620
Time     14.14
INSTRUM  AVQ-400
PROBHD   5 mm QNP 1H/13
PULPROG  zg30
TD        65536
SOLVENT  CDCl3
NS        1
DS        0
SWH       8012.820 Hz
FIDRES    0.122266 Hz
AQ         4.0894966 sec
RG         382
DE         62.400 use
TE         6.00 use
TE        293.4 K
D1         1.0000000 sec
MCREST    0.0000000 sec
MCWRK     0.0150000 sec

===== CHANNEL f1 =====
NUC1      1H
P1        12.80 use
PL1       0.00 dB
SFO1      400.1324700 MHz

F2 - Processing parameters
SI         65536
SF         400.1300142 MHz
WDW        EM
SSB        0
LR         0.30 Hz
    
```



Supporting Information - Watson, Chiu and Bergman

```

Current Data Parameters
NAME      3169aH15024h
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20060216
Time     14.39
INSTRUM  AVB-400
PROBHD   5 mm PABBO BB-
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       1
DS       0
SWH      8278.146 Hz
FIDRES   0.126314 Hz
AQ        3.9584243 sec
RG        287.4
DW        60.400 usec
DE        6.00 usec
TE        294.8 K
D1        1.0000000 sec
MCREST   0.0000000 sec
MCWRK    0.01500000 sec

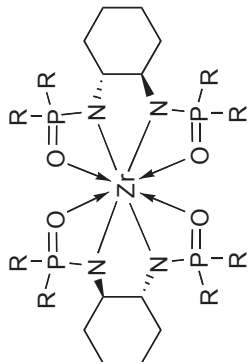
===== CHANNEL f1 =====
NUC1      1H
P1        8.20 usec
PL1       -3.00 dB
SFO1      400.1324710 MHz

F2 - Processing parameters
SI        32768
SF        400.1300000 MHz
WDW       EM
SSB       0
LR        0.30 Hz
    
```

0.115
0.881
1.126
1.361
1.882
1.949
1.994
2.022
2.039
2.095
2.176
2.181
2.199
2.260
2.496
2.524
2.558
3.438

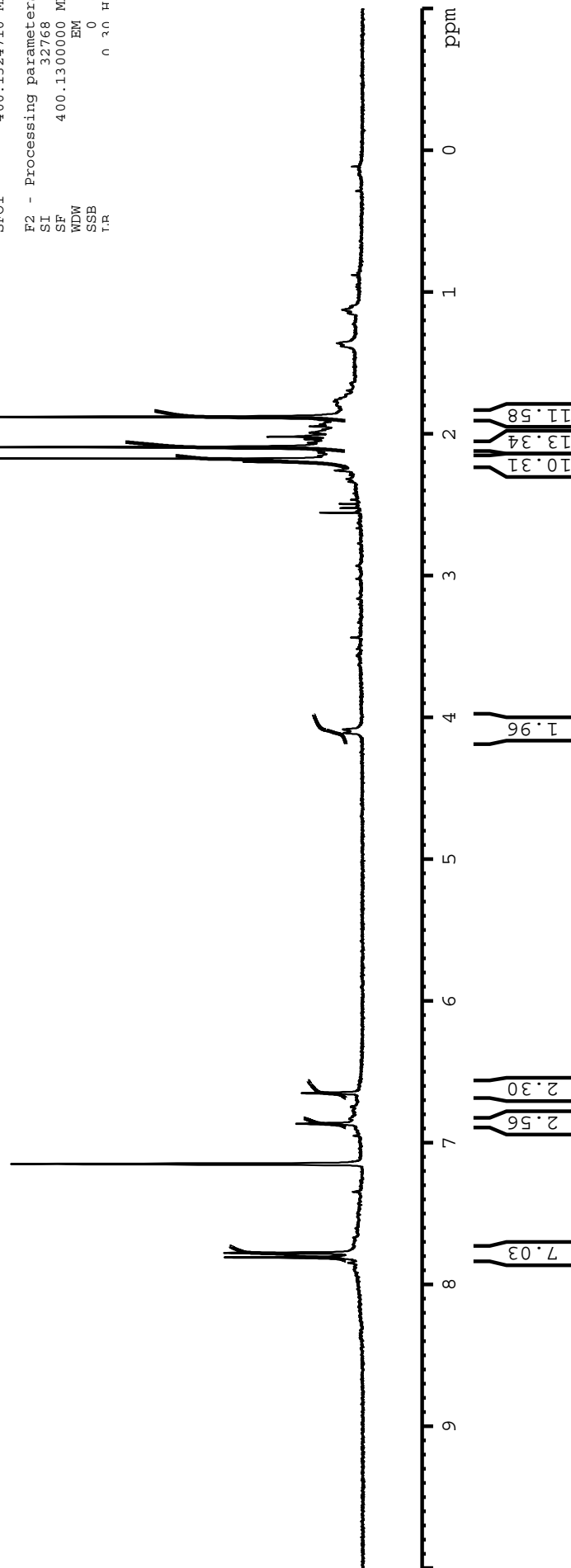
4.086

7.810
7.779
7.152
6.868
6.652



21b R = 3,5-C₆H₃Me₂
Generated In Situ

21b

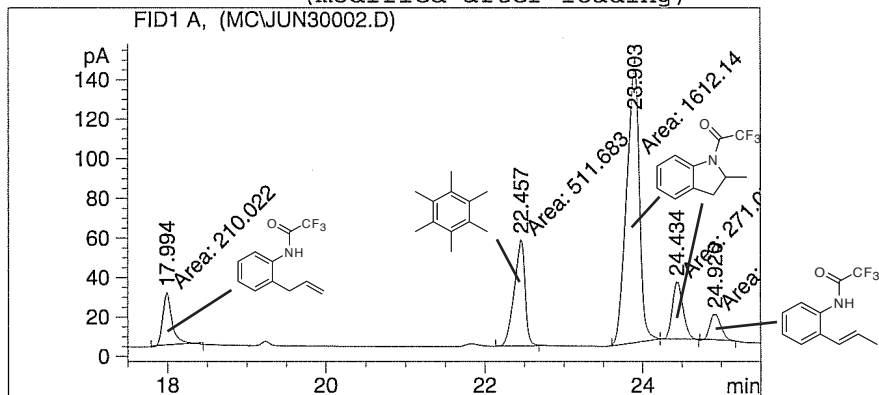



```

=====
Injection Date   : 6/30/06 11:39:42 AM      Seq. Line   :    2
Sample Name     : AN1048 - Watson, Chiu and Bergman  Vial        :    2
Acq. Operator   : MC                          Inj         :    1
                                           Inj Volume  : 2 µl

Acq. Method     : D:\HPCHEM\1\METHODS\HA_SUBFM.M
Last changed    : 6/16/06 10:32:54 PM by MC
Analysis Method : D:\HPCHEM\1\METHODS\DYW.M
Last changed    : 7/23/06 10:16:15 PM by MC
                  (modified after loading)
    
```

S65



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	1.592	BP	0.0152	6.52783	6.82050	0.00247
2	1.652	VV S	0.0277	1.38883e5	6.56908e4	52.45353
3	1.943	VB S	0.0255	1.22989e5	8.02763e4	46.45082
4	2.356	PP	0.0260	10.88994	6.67721	0.00411
5	2.479	BP	0.0269	4.44667	2.74909	0.00168
6	3.214	PB	0.0385	148.28819	53.24548	0.05601
7	17.994	MM	0.1326	210.02168	26.40137	0.07932
8	22.457	MM	0.1586	511.68347	53.78419	0.19325
9	23.903	MM	0.1863	1612.14307	144.22220	0.60888
10	24.434	MM	0.1560	271.07623	28.95608	0.10238
11	24.920	MM	0.1614	125.91330	13.00030	0.04756

Totals : 2.64773e5 1.46303e5

Results obtained with enhanced integrator!

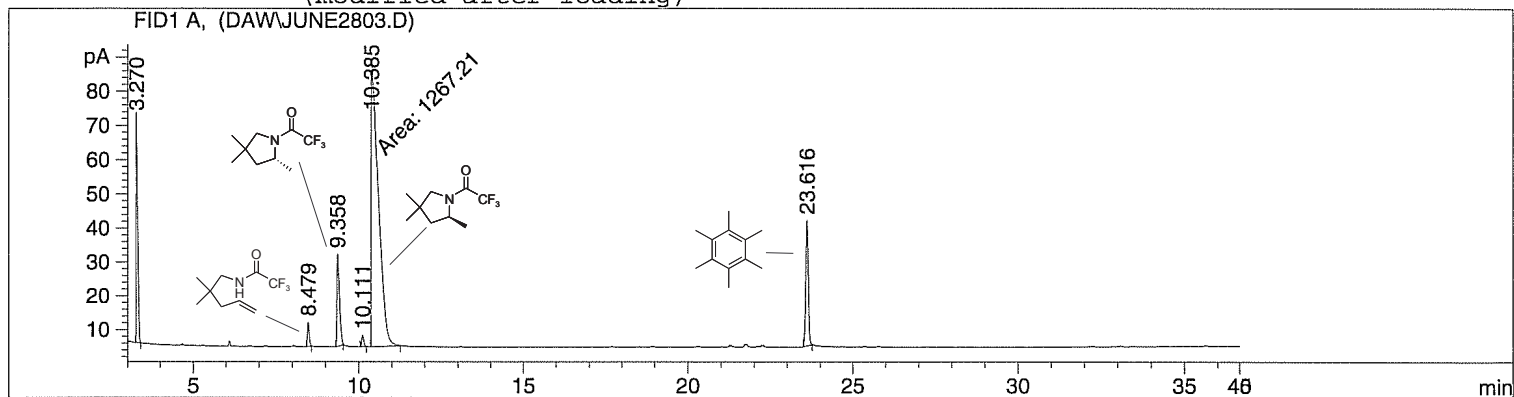
*** End of Report ***

```

=====
Injection Date   : 6/28/06 4:14:45 PM          Seq. Line   :    1
Sample Name     : Supporting Information 176    Vial        :    3
Acq. Operator   : DAW                        Inj         :    1
                                           Inj Volume  : 2 µl

Acq. Method     : D:\HPCHEM\1\METHODS\X_SUBA.M
Last changed    : 6/9/06 10:01:56 AM by MC
Analysis Method : D:\HPCHEM\1\METHODS\SHUTDWN.M
Last changed    : 6/29/06 9:22:36 AM by DAW
                  (modified after loading)
=====

```



```

=====
                          Area Percent Report
=====

```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	1.571	PV	0.0110	2.62242	3.84290	0.00115
2	1.593	VV	0.0111	2.74232	3.93816	0.00120
3	1.618	VV	0.0118	26.58545	35.18530	0.01165
4	1.652	VB S	0.0284	1.56971e5	6.93598e4	68.79561
5	1.952	PB S	0.0255	6.92840e4	4.14704e4	30.36496
6	2.371	PP	0.0280	6.25669	3.65211	0.00274
7	3.270	PB	0.0442	228.19263	66.27210	0.10001
8	8.479	PB	0.0570	26.70593	7.08096	0.01170
9	9.358	PB	0.0717	141.81641	27.26009	0.06215
10	10.111	PB	0.0665	14.35946	3.38737	0.00629
11	10.385	MM T	0.2495	1267.21167	84.64605	0.55538
12	23.616	BB	0.0806	198.84915	37.03812	0.08715

```
Totals :                      2.28171e5  1.11102e5
```

Results obtained with enhanced integrator!

```

=====
*** End of Report ***
=====

```

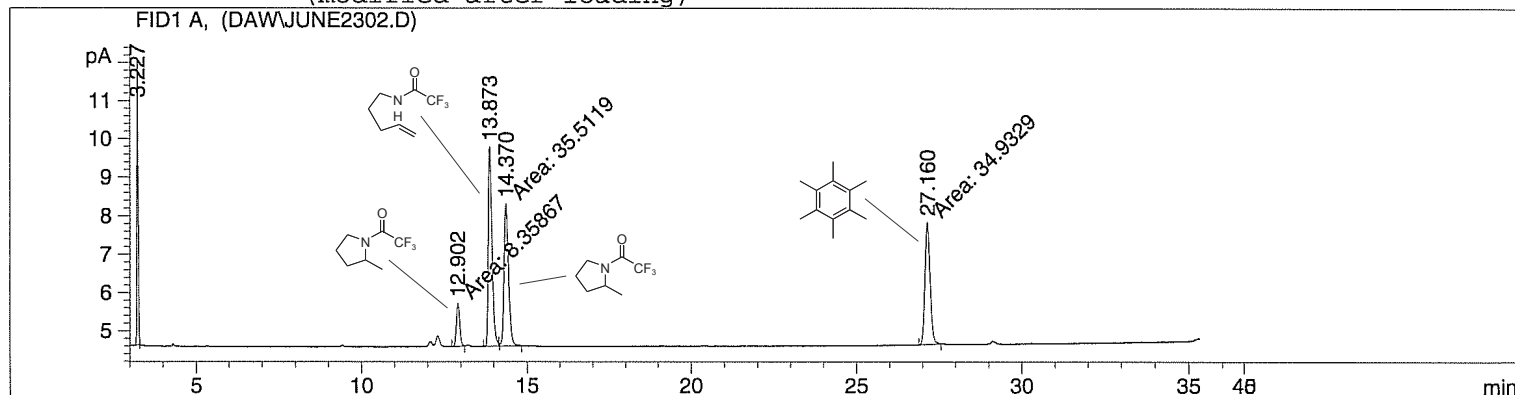
```

=====
Injection Date   : 6/23/06 10:33:13 AM          Seq. Line   :    2
Sample Name     : Supporting Information 1       Vial        :    8
Acq. Operator  : DAW                          Inj         :    1
                                           Inj Volume  : 2 µl
                                           Actual Inj Volume : 0.1 µl

Different Inj Volume from Sequence !
Acq. Method     : D:\HPCHEM\1\METHODS\X_SUBE.M
Last changed    : 6/21/06 3:26:59 PM by dg
Analysis Method : D:\HPCHEM\1\METHODS\SHUTDWN.M
Last changed    : 6/23/06 11:58:56 AM by DAW
                 (modified after loading)
=====

```

S67



```

=====
Area Percent Report
=====

```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000

```

Signal 1: FID1 A,

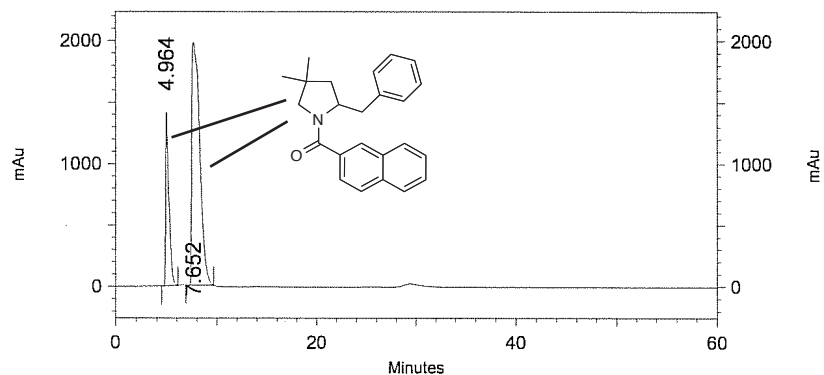
Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	0.692	BP	0.0115	7.55765	10.37646	0.03686
2	0.786	BB S	0.0165	1.29078e4	1.11437e4	62.95304
3	1.008	PB S	0.0117	7446.75293	1.00331e4	36.31869
4	3.227	BB	0.0420	20.50775	7.48248	0.10002
5	12.902	MM T	0.1226	8.35867	1.13644	0.04077
6	13.873	MM R	0.1363	42.45444	5.19316	0.20706
7	14.370	MM T	0.1594	35.51193	3.71375	0.17320
8	27.160	MM T	0.1816	34.93287	3.20565	0.17037

```
Totals :                2.05039e4  2.12079e4
```

```
Results obtained with enhanced integrator!
```

```
*** End of Report ***
```

Sample ID: bn2nc2s3c
Filename:
C:\EZStart\Projects\Default\Data\MC\bn2nc2s3c7-23-2006 10-29-52
PMOD9010IP60min15mL.met.dat Method:
C:\EZStart\Projects\Default\Method\MC\OD9010IP60min15mL.met
Injection volume: 5 uL



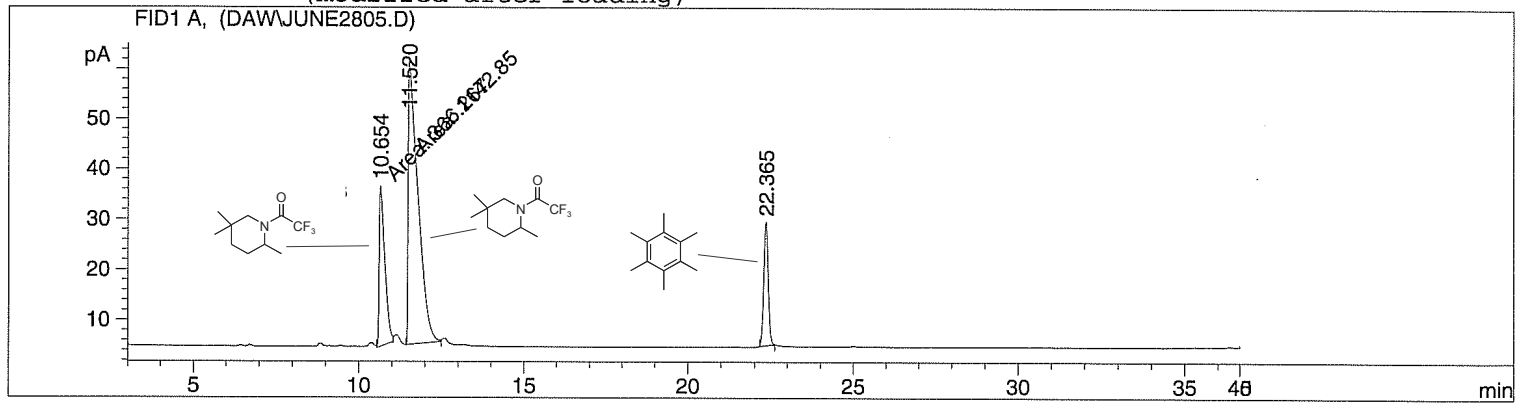
3: 280 nm, 4 nm Results

Retention Time	Area	Area Percent
4.964	31663378	22.336
7.652	110098768	77.664

```

=====
Injection Date   : 6/28/06 5:46:12 PM           Seq. Line   :    3
Sample Name     : DAW41480                     Vial        :    5
Supporting Information: Watson, Chiu and Bergman Inj         :    1
Acq. Operator   : DAW                          Inj Volume  : 2 µl
=====
Acq. Method     : D:\HPCHEM\1\METHODS\X_SUBB.M
Last changed    : 6/9/06 10:03:13 AM by MC
Analysis Method : D:\HPCHEM\1\METHODS\SHUTDWN.M
Last changed    : 6/29/06 9:21:46 AM by DAW
                  (modified after loading)
    
```

S69



Area Percent Report

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	0.727	BP S	0.0175	1.75196e5	1.41437e5	70.38090
2	0.876	VB S	0.0184	7.17398e4	6.50759e4	28.81977
3	1.096	BP	0.0206	6.31058	5.04526	0.00254
4	1.169	BP	0.0190	1.73517	1.55988	0.00070
5	1.496	BP	0.0222	1.66162	1.12613	0.00067
6	1.695	BB	0.0326	227.40259	95.95634	0.09135
7	10.654	MM T	0.1909	366.26709	31.97196	0.14714
8	11.520	MM T	0.3327	1142.85339	57.24689	0.45911
9	22.365	BB	0.1194	243.50099	24.61148	0.09782

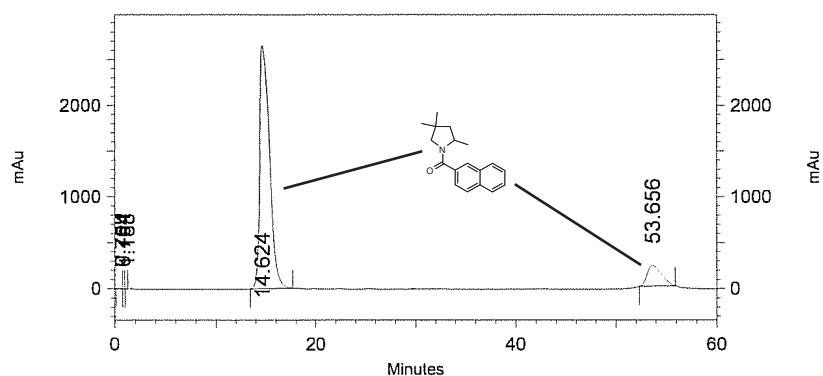
Totals : 2.48926e5 2.06731e5

Results obtained with enhanced integrator!

*** End of Report ***

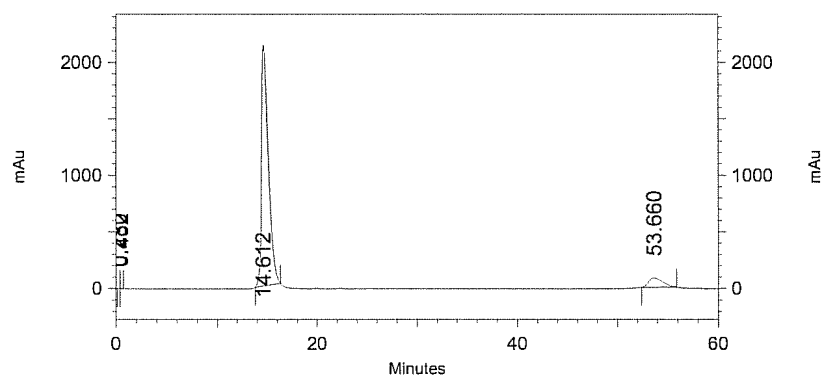
Sample ID:	dm2nap2c1s4a
Filename:	
C:\EZStart\Projects\Default\Data\MC\dm2nap2c1s4a7-20-2006 8-04-49	
PMMCWH9010IP2mL.met.dat	Method:
C:\EZStart\Projects\Default\Method\MC\MCWH9010IP2mL.met	
Injection volume:	5 uL

Description: {Data Description}



1: 230 nm, 4 nm Results

Retention Time	Area	Area Percent
0.280	42145	0.020
0.764	12103	0.006
1.188	6015	0.003
14.624	183288888	89.144
53.656	22260222	10.826



2: 240 nm, 4 nm Results

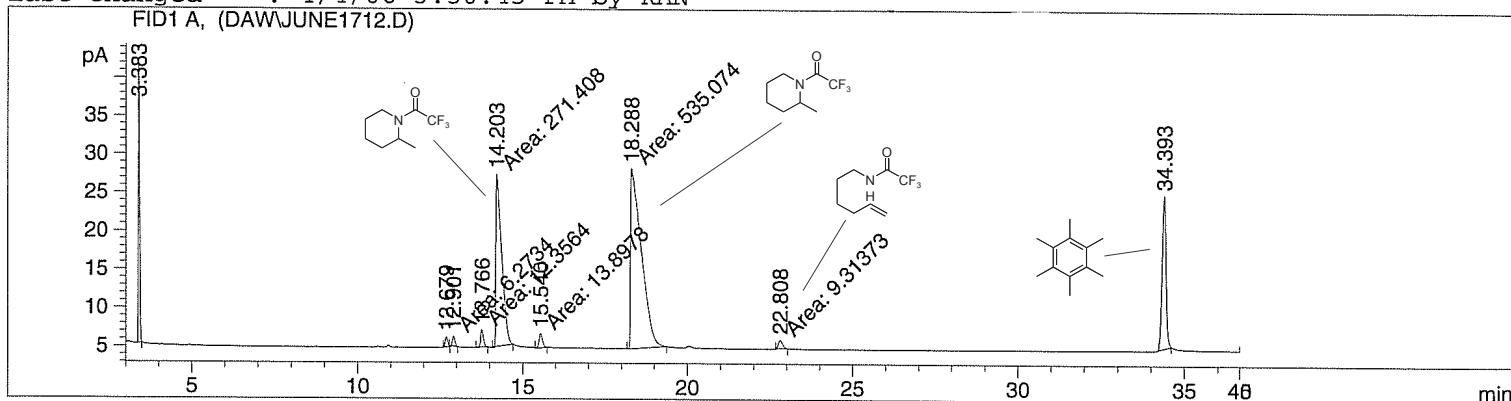
Retention Time	Area	Area Percent
0.280	2239	0.002
0.432	3770	0.003
14.612	102679324	92.651
53.660	8137998	7.343


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Injection Date   : 6/17/06 4:11:57 PM           Seq. Line   :    3
Sample Name     : DAW4133f                     Vial        :    4
Supporting Information: Watson, Chiu and Bergman Inj         :    1
Acq. Operator   : DAW                          Inj Volume  : 2 µl

Acq. Method     : D:\HPCHEM\1\METHODS\X_SUBD.M
Last changed    : 6/9/06 10:05:13 AM by MC
Analysis Method : D:\HPCHEM\1\METHODS\GORIN.M
Last changed    : 4/4/06 3:50:43 PM by KAN
    
```

S72



Area Percent Report

```

Sorted By      : Signal
Multiplier    : 1.0000
Dilution      : 1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	1.580	BV	0.0186	7.89778	5.91077	0.00575
2	1.628	VP	0.0161	46.23575	41.20645	0.03367
3	1.677	VB S	0.0334	7.13816e4	3.56321e4	51.98667
4	1.961	PB S	0.0211	6.47520e4	4.70268e4	47.15844
5	2.388	PB	0.0265	6.55903	3.92605	0.00478
6	3.383	PB	0.0391	96.69431	36.30813	0.07042
7	12.679	MM R	0.0876	6.78294	1.29008	0.00494
8	12.901	MM T	0.0821	6.27340	1.27406	0.00457
9	13.766	MM T	0.0930	12.35639	2.21524	0.00900
10	14.203	MM T	0.2018	271.40753	22.41681	0.19766
11	15.540	MM T	0.1245	13.89779	1.86032	0.01012
12	18.288	MM T	0.3799	535.07355	23.47433	0.38969
13	22.808	MM T	0.1448	9.31373	1.07188	0.00678
14	34.393	BB	0.1259	161.33351	19.82506	0.11750

Totals : 1.37307e5 8.28196e4

Results obtained with enhanced integrator!

*** End of Report ***