Conservation of the AK2 protein sequence in various orthologs

	1	10	20	30	40	50	60
	I	I	I	I	I	I	
hAK2A	MAPS	-VPAAEPEYP	K-GIRAVLLO	SPPGAGKGTQAI	PRLAENFCVC	HLATGDMLRA	MVASG
mAK2A	MAP N- -	-V L A S EPEIP	K-GIRAVLLO	SPPGAGKGTQAI	PKLAENFCVC	HLATGDMLRA	MVASG
zAK2A	MAPST	QEDDTVSGIR	K-GIRAILLO	SPPGAGKGTQAI	PKLAEKYCVC	HLATGDMLRA	MVASG
dAK2	MAPNA	AVP VERYEPE	NIGINAILLO	GPPG S GKGTQAI	PLLKEKFCVO	CHL S TGDMLRA	EISSG
	P5						
hAK2A	SELGK	KLKATMDAGK	LVSDEMVVEI	JIEKNLETPLCI	KNGFLLDGF	RTVROAEMLD	DLMEK
mAK2A	SELGK	KLKATMDAGK	LVSDEMVVEI	.IEKNLETP S CI	KNGFLLDGFI	RTVROAEMLD	DLMEK
zAK2A	SELGQ	RLKETMDAGK	LVSDEMVVEI	IDNNLDTPACI	KNGFLLDGFI	R TV K OAEMLD	DLMEK
dAK2	SKLGA	ELK KV MDAGK	LVSD DL VVEN	IDSNLDKPEC	KNGFLLDGFI	RTV V DAE K LD	t l ld k
	AMP bir	nding doma	in		Adenylat	e Kinase	
			_		1	P1, P2	
hAK2A	RKEKLI	OSVIEFSIPD	SLLIRRITGF	RLIHPKSGRSY	HEEFNPPKEI	MKDDITGEPL	IRRSD
mAK2A	RKEKLI	OSVIEFSI Q D	SLLIRRITGF	RLIHPKSGRSYI	HEEFNPPKEI	MKDDITGEPL	IRRSD
zAK2A	r s ekli	OSVIEFS VD D	SLL V RRI C GF	RLIH QP SGRSYI	HEEF H PPKE H	IMKDD V TGEPL	IRRSD
dAK2	RK TN LDAVIEFAIDDSLL V RRITGRLIHQASGRSYHEEFAPPKKPMTDDVTGEPLIRRSD						
	LID domain						
	P4						
hAK2A	DNEKAI	LKI R LQAYHT	QTTPLIEYYF	RKRGIHSAIDAS	SQTPDVVFAS	ILAAFSKATC	KDLVM
mAK2A	DNEKALK TRLE AYHTQTTPL v EYYRKRGIH C AIDASQTPD i VFASILAAFSKATCKDLVM						
zAK2A	DNETTLRSRLESYHRQTSPLVQYYSARGLHTAIDASQSTDLVFASILAAFSAATCKDLVY						
dAK2	DN AE AI	LK KR L E AYH K	QTKPLVDYYC	L RG L H FKV DA	AKKSSDVFS	IDSIFQRKRP	AQIQL
hAK2A	FI						
mAK2A	FI						
zAK2A	FV						
dAK2							

Alignment of human AK2 (hAK2), murine AK2 (mAK2), zebrafish AK2 (zAK2) and drosophila AK2 (dAK2) protein sequences. Amino acids affected by a mutation are denoted in red.



Phenotype of bone marrow cells from an RD patient

(a) May-Grünwald/Giemsa staining of BM cells from patient P2 at diagnosis (left panel) and after methylcellulose culture (right panel). In both conditions, only a few myeloblast cells could be identified. In the control cord blood (CB) culture, myeloid precursors and polynuclear cells were detected.

(b) Granulocyte and monocyte differentiation of patient P2's BM cells. CD34+ cells isolated from patient P2's BM or from control cord blood (CB) cells were cultured in the presence of SCF, FLT-3L and GM-CSF (all at 100ng/ml) or G-CSF (100ng/ml) for 2 weeks. At the end of the culture, flow cytometry analysis was performed to determine the proportion of neutrophils (CD15⁺ CD11b⁺) and monocytes (CD14⁺ CD11b⁺).





AK2A and *AK2B* expression were quantified by real-time RT-PCR in various bone marrow haematopoietic populations as compared to a SV40-transformed fibroblast cell line (Fibro). The cell populations analyzed were sorted by flow cytometry to isolate : CD3⁺ T-cell (T), CD19⁺ B-cell (B), CD16⁺CD56⁺ NK-cell (NK), CD34⁺ progenitors (CD34), CD15⁺CD11b⁻ promyelocytes (PML), CD15⁺CD11b⁺ polynucleated neutrophils (PN) and CD14⁺ monocytes (Mono). Data correspond to the mean of two independent adult bone marrow-derived populations.

Complementation of the neutrophil differentiation defect by restoration of *AK2* expression in P6 BM cells



Mononuclear cells isolated from the bone marrow (BM) of patient P6 were transduced with either two bicistronic lentivirus vectors (encoding *AK2*A and *AK2*B, together with GFP : AK2A+B+GFP) or mock transduced. Mock transduced cord-blood-derived CD34⁺ cells were used as a positive control. After lentiviral transduction, 250,000 mononuclear cells from P6's BM were seeded in semi-solid medium for each condition. As a control experiment, 1,000 CD34⁺ cord blood cells were seeded in semi-solid medium.

1

1,300

Differentiated

myeloid cells Absolute cell

20

123.700

65

95,500

(a) After 13 days, CFU-G/GM colonies were collected and cytospin preparations were analyzed by May-Grunwald/Giemsa staining. Representative pictures of various stages of neutrophil maturation are shown at magnifications of 50x (left panel) or 100x (right panel).

(b) Enumeration of myeloid subpopulations among CFU-G/GM colonies. Myeloid precursors (blasts, myeloblasts and promyelocytes) and differentiated myeloid cells (myelocytes, metamyelocytes and granulocytes) are represented in pourcentage and in absolute cell numbers.

Inhibition of neutrophil differentiation after AK2 knock-down



CD34⁺ CB progenitors were transduced three times with a lentivirus encoding GFP together with a shRNA directed against AK2 (shAK2) or with a shRNA corresponding to a scramble version as a control (shScramble). At day 3 (d3), the transduced GFP positive cells were sorted and culture in the presence of G-CSF. Eight days later (d3+8), we evaluated neutrophil differentiation (CD15⁺CD11b⁺ count) by flow cytometry analysis (**a**). These results were combined with the analysis of May-Grunwald/Giemsa cytospin preparations. Using both flow cytometry and morphological analysis, we enumerated respectively the absolute numbers of mature granulocytes CD15⁺CD11b⁺ as well as differentiated myeloid cells (myelocytes, metamyelocytes, granulocytes) obtained in both shScramble and shAK2 conditions (**b**).