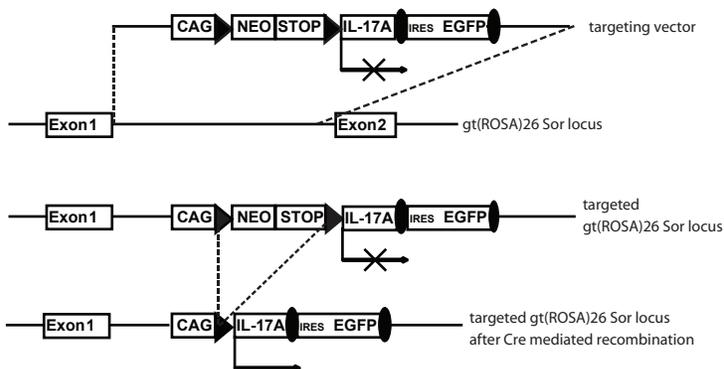
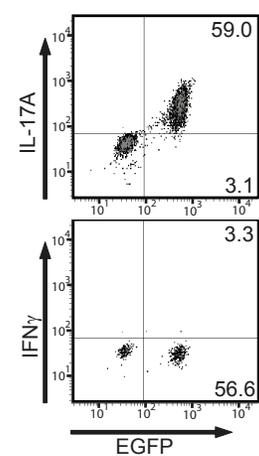


# Supplemental Figure 1

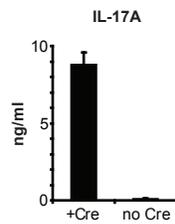
**A**



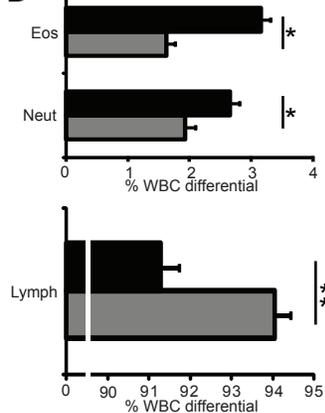
**B**



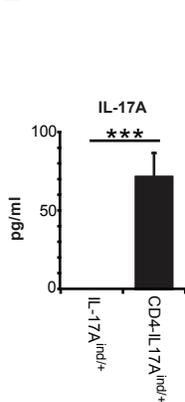
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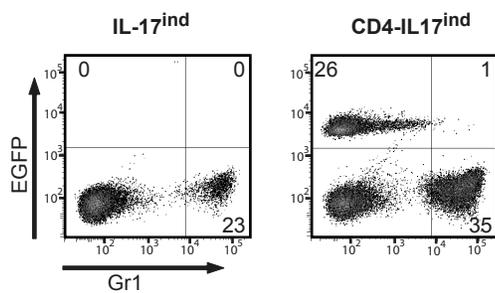
**D**



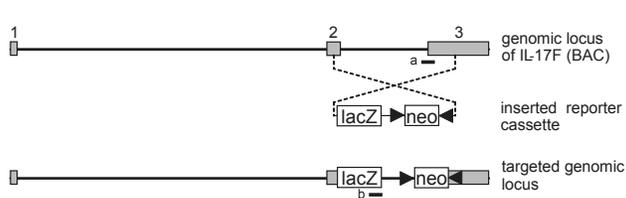
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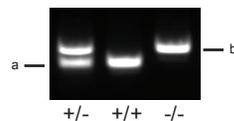
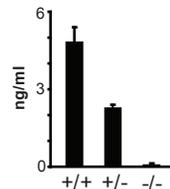
**F**



**G**

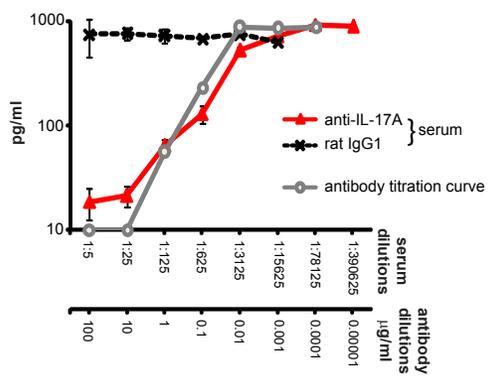


**IL-17F**

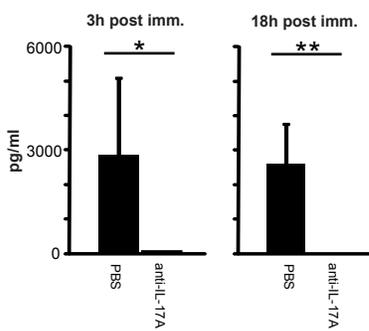


# Supplemental Figure 2

A



B



**Supplemental Figure 1** (A) Generation of the IL-17A<sup>ind</sup> allele was carried out using homologous recombination in embryonic stem cells (C57BL/6-Bruce 4). Our conditional ,knock-in' approach introduced the targeting construct into the endogenous gt(ROSA)26Sor locus. Upon Cre-mediated recombination, a lox-P-flanked transcriptional STOP cassette is excised 5' of an IL-17A cDNA insert and an IRES-EGFP element, allowing a dual expression of IL-17A and EGFP under the control of the chicken  $\beta$ -actin (CAG) promoter. (B)  $1 \times 10^6$  whole lymph node cells from IL-17A<sup>ind/+</sup> mice were cultured for 4 h with or without (data not shown) Tat-Cre protein to induce IL-17A and EGFP expression in the presence of anti-CD3, anti-CD28 and IL-2. After 36 h, cells were briefly cultured with Brefeldin-A and stained for CD4 and either IL-17A or IFN $\gamma$ . Percentages of IL-17A<sup>+</sup>EGFP<sup>+</sup> and IFN- $\gamma$ <sup>+</sup>EGFP<sup>+</sup> cells are given in the quadrants. No EGFP was detected in wild type mice also treated with Tat-Cre (data not shown). (C) LN cells were cultured as in B, and supernatants extracted after 36 h. IL-17A secretion was measured by ELISA. (D) Peripheral blood was extracted from IL-17A<sup>ind/+</sup> and CD4-IL17A<sup>ind/+</sup> mice and subjected to routine WBC differential. Significant increase in neutrophil and eosinophil counts, and a decrease in peripheral blood lymphocytes are shown. (CD4-IL17A<sup>ind/+</sup> n=10, IL-17A<sup>ind/+</sup> n=9). (E) Serum from IL-17A<sup>ind/+</sup> and CD4-IL17A<sup>ind/+</sup> mice was extracted at d14 after immunization with MOG/CFA. Shown are levels of IL-17A detectable in serum isolated from peripheral blood (n=5). Spleen cells from the same mice were stained for Gr1. Quadrant statistics are shown. (F) Spleen-derived single cell suspension were obtained from the indicated genotypes d14 after MOG/CFA immunization. Percentages of Gr1<sup>+</sup> cells are shown in the histograms. (G) IL-17F gene-targeting strategy for the generation of the IL-17F deficient mouse strain. A reporter gene/resistance cassette (*lacZ* gene and a Ub1-Em7 promoter driven neomycin resistance gene, *neo*, flanked by loxP sites, ►) was introduced into *il17f* exon 2 and 3 of a BAC containing the IL-17F gene locus. 129S6SvEv/C57B/6F1 embryonic stem cells were

targeted with a BAC carrying the replaced *il17f* gene locus and were subsequently screened as described elsewhere (41). “a” and “b” represent amplicons for the wild type and targeted allele respectively. Complete loss of IL-17F in the IL-17<sup>null</sup> mouse was confirmed by ELISA on supernatant of T<sub>H</sub>17 polarized splenocytes.

**Supplemental Figure 2 Specification of the IL-17A blocking capacity of *in vivo* applied antagonistic antibody** (A) WT or IL-17A<sup>-/-</sup> mice (n=3) were immunized with MOG/CFA and pertussis toxin. On day 0 and 4 either 200mg of antagonistic anti-IL-17A antibody or isotype control antibody (rat IgG1) was administered intraperitoneally and mice are bled on day 7. The recovered serum was titrated in an IL-17A specific ELISA onto 1000 pg of IL-17A protein standard. The serum was serially diluted as shown in the graph. To estimate the anti-IL-17A titer in the serum the neutralizing antibody was itself in parallel titrated onto the same amount of IL-17A protein standard. IL-17A levels were measured after a 1 hour pre-incubation of the serum/antibody with the IL-17A standard. The graph shows the pooled data of all anti-IL-17A and rat IgG1 sera with SEM indicated. (B) Two groups of CD4-IL17A<sup>ind/+</sup> littermate mice were immunized with MOG/CFA (n=3). After 10 days, either 100µg/mouse of anti-IL-17A or sham antibody was injected intravenously. Mice were subsequently bled and serum was isolated either 3h or 18h after the antibody treatment. IL-17A serum concentrations were measured by flow cytometric cytokine analysis. Statistical significances are indicated in the graph (one-tailed T test).

**Supplemental Table 1 Homeostatic leukocyte composition in spleen and thymus of naïve CD4-IL17A<sup>ind/+</sup> and control mice.**

SPLEEN	Percentage	
	CD4-IL17A <sup>ind/+</sup>	IL-17A <sup>ind/+</sup>
<b>T<sub>H</sub> cells (CD4<sup>+</sup>)</b>	18	19
<b>CTL (CD8<sup>+</sup>)</b>	9	11
<b>T<sub>Reg</sub> cells (FoxP3<sup>+</sup>)</b>	2	2
<b>NK cells (NK1.1<sup>+</sup>)</b>	5	5
<b>B cells (B220<sup>+</sup>)</b>	59	60
<b>Macrophages (CD11b<sup>+</sup>)</b>	6	5
<b>DC (CD11c<sup>+</sup>)</b>	6	4

THYMUS	Percentage	
	CD4-IL17A <sup>ind/+</sup>	IL-17A <sup>ind/+</sup>
<b>T<sub>H</sub> (CD4<sup>+</sup>)</b>	15	16
<b>CTL (CD8<sup>+</sup>)</b>	6	7
<b>Double positive T<sub>H</sub> cells</b>	73	73

**Supplemental Table 2 Homeostatic leukocyte composition in spleen and thymus of naïve IL-17F deficient and control mice.**

SPLEEN	Percentage		
	+/+	+/-	-/-
<b>T<sub>H</sub> cells (CD4<sup>+</sup>)</b>	14	16	14
<b>CTL (CD8<sup>+</sup>)</b>	20	21	23
<b>T<sub>Reg</sub> cells (FoxP3<sup>+</sup>)</b>	3	3	2
<b>NK cells (CD3<sup>-</sup>, NK1.1<sup>+</sup>)</b>	3	4	3
<b>NK T cells (CD3<sup>+</sup>, NK1.1<sup>+</sup>)</b>	2	3	2
<b>B cells</b>	38	38	37
<b>Macrophages (CD11b<sup>+</sup>)</b>	4	4	4
<b>DC (CD11c<sup>+</sup>)</b>	4	4	3

THYMUS	Percentage		
	+/+	+/-	-/-
<b>T<sub>H</sub> (CD4<sup>+</sup>)</b>	14	15	13
<b>CTL (CD8<sup>+</sup>)</b>	5	6	4
<b>Double positive T<sub>H</sub> cells</b>	78	78	80