

Supplementary Figure legends

Supplementary Figure 1.

(A) Efficiency of XLRP5/6 MO and mLrp5/6 overexpression in *Xenopus* embryos. Cell lysates were injected as indicated and analyzed by Western Blotting with phospho Lrp6 antibody, which also recognizes unphosphorylated Lrp5 and Lrp6, or XLRP5 and XLRP6, respectively. Levels of E-/N-cadherin and total protein were used as a loading control.

(B) Scheme of *Xenopus* injections used in the study.

(C) Effects of XLRP5/6 MOs on ventralization. *Xenopus* embryos were injected as indicated and the ventralization has been scored using DAI index (Kao and Elinson, 1988). Ventralization induced by XLRP5 MO is rescued by mLrp5 and β -catenin.

Supplementary Figure 2. Heart morphology in *Lrp6* and *Wnt5a* mutants at E10.5.

Heart of E10.5 wild type, *Lrp6*^{-/-} and *Wnt5a*^{-/-} mouse embryos was dissected and the morphology was analyzed. No obvious abnormalities were detected at this stage.

Representative examples are shown. RV – right ventricle, LV – left ventricle, RA - right atrium, LA - left atrium, OFT – outflow tract.

Supplementary Materials and Methods

Mouse genotyping

Ear or embryonic tissues were boiled at 95°C for 40 minutes in 100-200 μ l of 25mM NaOH/0,2mM EDTA and an equal volume of 40mM Tris HCl pH 5 was added. 3 μ l of this solution was used for PCRs. *Lrp6* wild type allele was identified with the previously described primers LRP6-U1 and LRP6-D1, while mice with the gene trap insertion (*Lrp6* knockout allele) were recognized with the following primer set: CD4mix forward (5'-

GCACGGATGTCTCAGATCAAGAGG-3') and CD4mix reverse (5'-CGGGATCATCGCTCCCATATATG-3'), with an annealing temperature of 63°C and an amplicon of 108bp. *Wnt5a* WT and null alleles were identified with the following primers: 5a WT For (5'-GACTTCCTGGTGAGGGTGC GTG-3'), Wnt5a WT Rev (5'-GGAGAATGGGCACACAGAATCAAC-3'), Wnt5a null For (5'-GGGAGCCGGTTGGCGCTACCGGTGG-3') and Wnt5a null Rev (5'-GGAGAATGGGCACACAGAATCAAC-3').

Sequences of antisense MOs

XLRP5 MO (5'-ctc cca tgg cct cgt acc cct ctcc), XLRP6 MO (5'-get caa tgc tcc ccc gta acc cgac) or standard control MO directed against *Lrp5* and *Lrp6* were used at 0.8 pmol (all MOs: GeneTools, Philomath, OR, USA). Sequences of XWnt5a and XWnt11 MOs were published before (Pandur et al., 2002; Schambony and Wedlich, 2007).

Antibodies

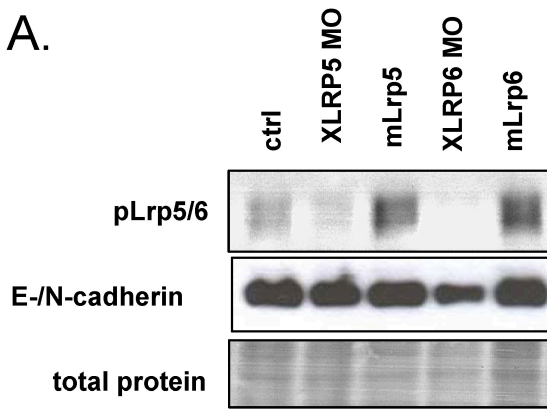
Following antibodies were used: anti P-Ser1490-Lrp6 (#2568), anti Lrp6 (#2560) from Cell Signaling Technologies), anti-Dvl2, anti-Dvl3 (sc-8027), anti-c-Myc (sc-40) (from Santa Cruz Biotechnology), anti-active β -catenin (ABC, #05-389) and anti-Rac1 (-389) from Upstate Biotechnology, anti- β -actin (Ab6276) and anti HA (Ab9110) for IP from Abcam, anti HA (HA.11) for WB from Nordic Biosite, and anti-human IgG Fc fragment (109-005-098, Jackson ImmunoResearch Laboratories). Cadherin levels in *Xenopus* lysates were detected using the mixture of antibodies against E- and N-cadherin (610182 and 610921, BD Biosciences).

Quantitative-PCR

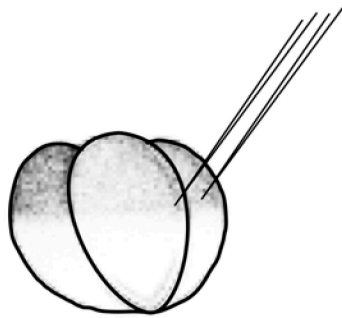
cDNA was generated as described previously (Castelo-Branco et al., 2003) RNA from ventral midbrains of *lrp6*^{+/+}, *lrp6*^{+/-} and *lrp6*^{-/-} mouse embryos at E11.5 and E13.5 (where both Lrp6 and Wnt5a are known to be expressed) was extracted using RNeasy Mini Kit (Qiagen). 1 µg of RNA was reverse-transcribed using SuperScript II Reverse Transcriptase (Invitrogen) and random primers (Invitrogen). Wnt5a primers used in this study have been described previously (Castelo-Branco et al., 2003), Lrp6 primers were as follows: forward 5-GCTACAAATGGCAAAGAGAATGC-3, reverse CAGTATACAAGCCATGACCAAACA.

Supplementary Figure 1

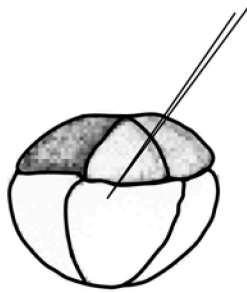
A.



B.

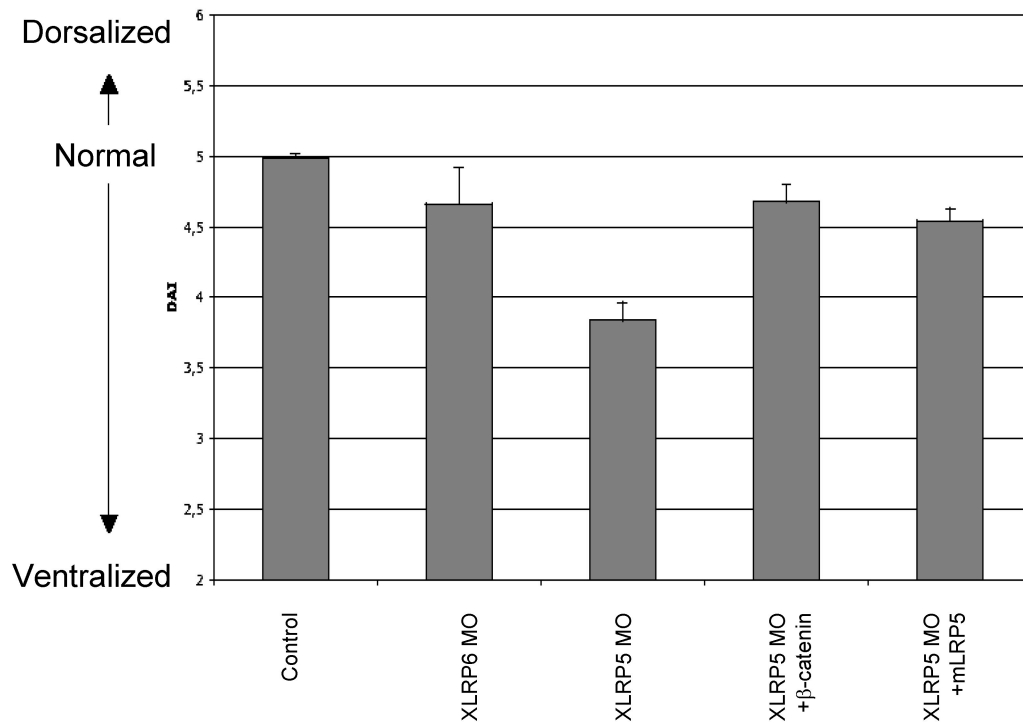


injection in 4-cell stage embryo
 targets Dorsal Marginal Zone
 affects primary axis formation (ventralization) and CE movements
 Figures 2, 3, and Suppl Figure 1



injection in 8-cell stage embryo
 targets presumptive cardiac mesoderm
 affects cardiac development, but avoids earlier effects on primary axis and CE
 Figure 4

C.



Supplementary
Figure 2

