

## SUPPLEMENTARY INFORMATION

### **I. Supplementary Figure Legends**

#### **Figure S1. Venus::*UNC-6* expression prior to and during AC invasion.**

**(a-c)** Nomarski images overlaid with Venus::*UNC-6* ( $\Delta$ SP) expression shown in yellow (left), and Venus::*UNC-6* ( $\Delta$ SP) fluorescence alone (right). Venus::*UNC-6* ( $\Delta$ SP) lacks a signal sequence and was thus retained in the cells in which *unc-6* is expressed. Venus::*UNC-6* was retained within the neurons of the ventral nerve cord (VNC, small bracket) under the AC (arrow) prior to invasion at the P6.p 1- and 2-cell stages (large brackets, **a** and **b**), and during the time of invasion at constant levels (**c**). Scale bar in (**a**) is 5 $\mu$ m in this and all subsequent supplementary figures.

**Figure S2. Venus::*UNC-6* expression and localization is not dependent on the vulval cells.** Nomarski image (left), and corresponding fluorescence image (right) of an animal expressing Venus::*UNC-6* at the early L3 stage after laser-mediated ablation of the vulval cells in the early L2 larval stages. Venus::*UNC-6* was expressed normally in the VNC and localized to the basement membrane (arrowhead) under the AC (arrow) as in wild-type animals ( $n = 24/24$  animals). Inset highlights the basement membrane localization of Venus::*UNC-6* (arrowhead) under the AC.

#### **Figure S3. AC-specific expression of *UNC-40::GFP*.**

**(a)** Nomarski image overlaid with *UNC-40::GFP* fluorescence (green). The *unc-40::GFP* construct is driven by the AC-specific regulatory element of the *cdh-3* gene. **(b)** The

corresponding fluorescence image of UNC-40::GFP fluorescence alone. The *cdh-3* AC-specific promoter drives expression solely in the AC (arrow), and not in neighboring cells, including vulval cells (brackets). The asterisk marks autofluorescence from the gut granules.

**Figure S4. The Rac orthologs GFP::MIG-2 and GFP::CED-10 are polarized along the invasive cell membrane of the AC.** (a-e) Nomarski images (left), corresponding fluorescence images (right). Time is post-hatching at 20°C. (a) GFP::MIG-2 expression driven by its endogenous promoter was first expressed in the AC during the late L2 (9.5 hours before invasion) and was not polarized (16/16 animals). (b) MIG-2 was first polarized at the L2 molt (arrowhead; 7/11 animals). (c-e). Polarized MIG-2 was maintained along the invasive cell membrane from the early L3 stage through the time of invasion (20/20 animals for each stage). (f) Projected confocal z-stack showing uterine and vulval expression of GFP::CED-10 driven by its endogenous promoter (left) and the corresponding spectral representation of the fluorescence intensity (right). CED-10 was expressed in all uterine and vulval cells (bracket outlines the 1° VPCs), and was polarized along the basal (invasive) membrane of the AC (arrow) but not in neighboring uterine cells (arrowheads).

**Figure S5. PAR-3::GFP and apical AJM-1::GFP are localized normally in *unc-6* mutants.** (a, b) Nomarski images (left), corresponding fluorescence images (right). (a) PAR-3::GFP was localized to apical and lateral membranes of wild-type ACs prior to invasion (arrowheads), and this polarity was not altered in *unc-6* mutants (b, arrowheads;  $n = 20/20$  animals). (c) In wild-type animals AJM-1::GFP (green) localized to nascent junctions (arrowhead) in the apical region of the AC (expressing *zmp-1*>*mCherry* in magenta). (d) In

*unc-6* mutants apical AJM-1::GFP junctions were formed and positioned normally compared to wild-type animals ( $n \geq 60$  wild-type and *unc-6* animals examined).

**Figure S6. Pan-uterine expression of mCherry:: PLC $\delta^{\text{PH}}$  reveals a unique PI(4,5)P<sub>2</sub> rich invasive membrane domain in the AC.** Fluorescence (left) overlaid on corresponding Normarski image (right). **(a)** Expression of the PI(4,5)P<sub>2</sub> sensor mCherry::PLC $\delta^{\text{PH}}$  in the AC revealed strong polarization at the basal (invasive) cell membrane (arrowhead). **(b)** In contrast, a section lateral to the AC through the neighboring ventral and dorsal uterine cells (VU/DU) expressing PLC $\delta^{\text{PH}}$ ::mCherry, revealed that adjacent uterine cells did not have PI(4,5)P<sub>2</sub> concentrated in basal membranes (arrowheads).

**Figure S7. GFP::MIG-2 is localized normally in the AC of *fos-1(ar105)* animals.**

Nomarski images (left), corresponding fluorescence images (right). In **(a)** wild-type animals and **(b)** *fos-1(ar105)* mutants, MIG-2::GFP was strongly polarized to the invasive cell membrane of the AC (arrowhead;  $n = 20/20$  *fos-1a* mutants examined). **(c)** In *unc-6* animals, MIG-2::GFP polarity was perturbed.

**Video S1. Hemicentin::GFP deposition under the AC in wild-type animals.**

Hemicentin::GFP was localized at low levels along gonadal and ventral epidermal basement membranes, but was deposited at high levels (green, orange arrows) under the AC's invasive cell membrane (expressing *cdh-3*>mCherry::moeABD in magenta) during AC invasion.

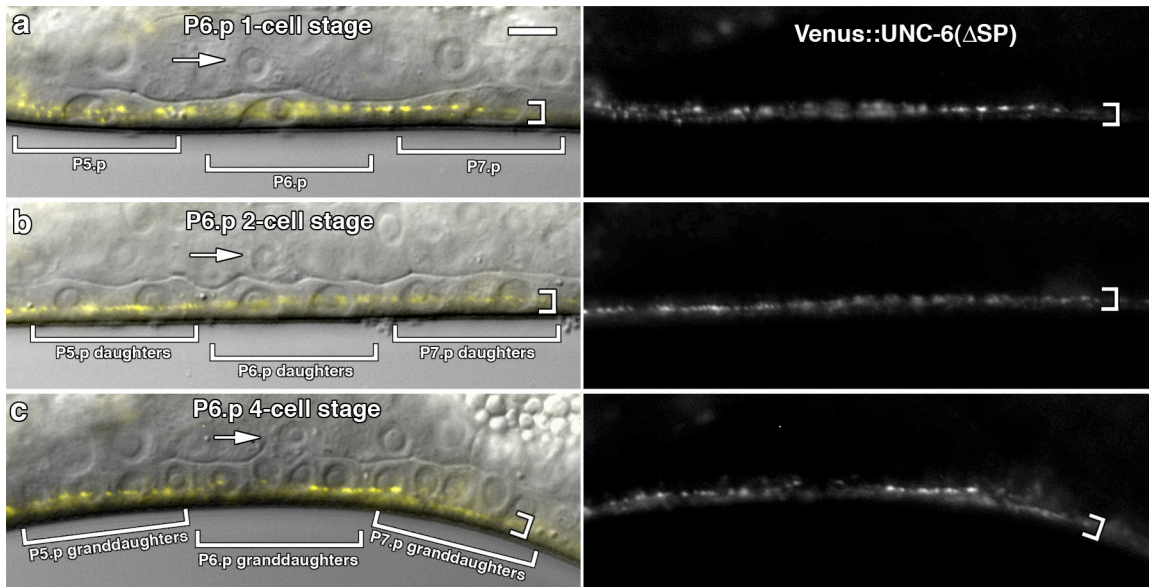
Little hemicentin::GFP was deposited along apical or lateral domains of the AC (note small deposit at white arrowhead).

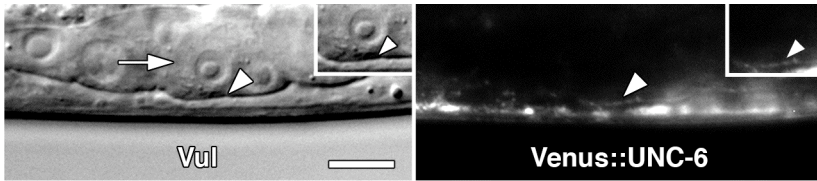
**Video S2. Hemicentin::GFP deposition is perturbed *unc-6* mutants.**

In *unc-6* mutants there was a dramatic decrease in hemicentin deposited under the AC (expressing *cdh-3*>mCherry::moeABD in magenta) at the site of basement membrane contact (green, orange arrows), while apical and lateral accumulations of hemicentin increased (white arrowheads).

## II. Supplementary Figures and Tables

**Figure S1**



**Figure S2**

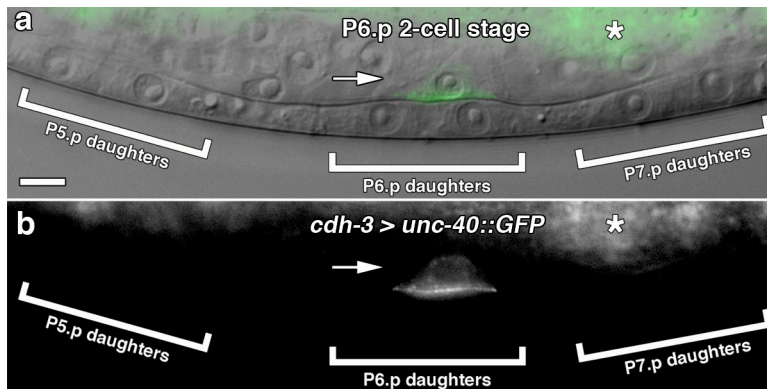
**Figure S3**

Figure S4

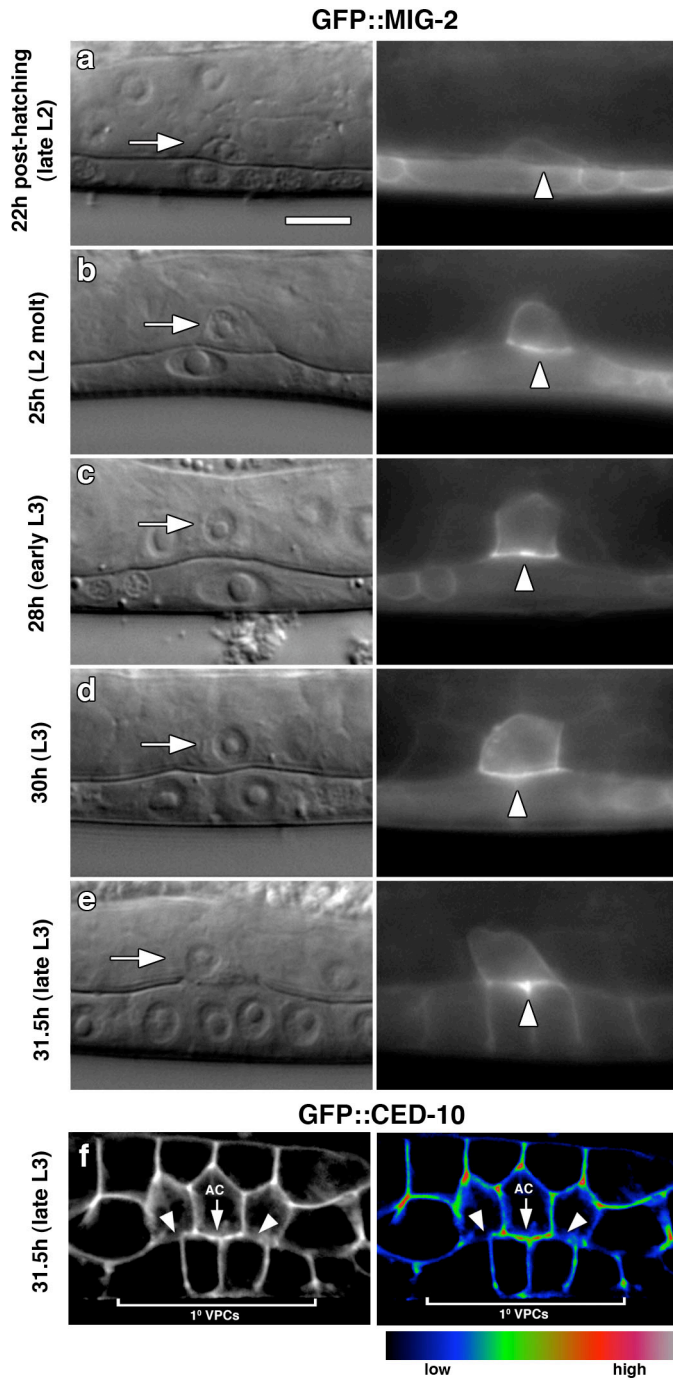




Figure S5

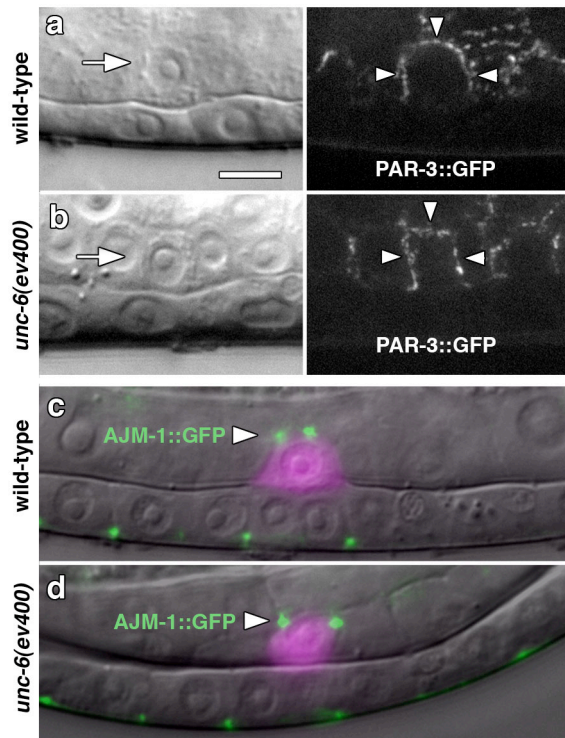


Figure S6

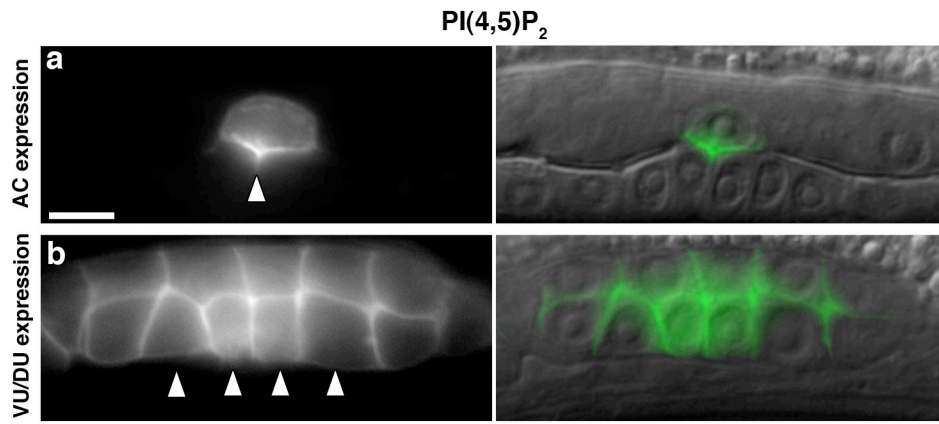
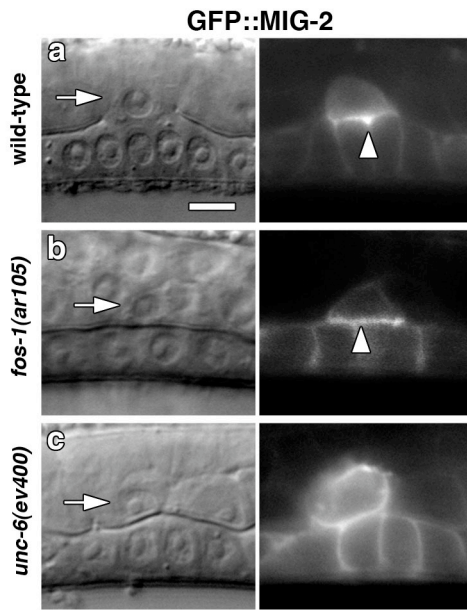


Figure S7



**Table S1. AC invasion in mutant strains with roles in cell motility and axon outgrowth**

<i>C. elegans</i> gene	Encoded Product	Invasion at P6.p 4-cell stage (number of ACs invaded/number observed) <sup>a</sup>	Invasion at P6.p 8-cell stage (number of ACs invaded/number observed)
<i>arf-1.2(ok796)</i>	ADP-ribosylation factor 1 homolog	10/10	10/10
<i>arf-1.2(ok1322)</i>	ADP-ribosylation factor 1 homolog	9/9	4/4
<i>bar-1(ga80)</i>	Armadillo/beta-Catenin/plakoglobin	11/11	9/9
<i>C25F6.4(ok874)</i>	protein tyrosine kinase homolog that is also homologous to human RS1	15/15	16/16
<i>cam-1(gm122)</i>	receptor tyrosine kinase of the immunoglobulin superfamily that is orthologous to human ROR1	7/9	28/35
<i>cam-2(gm124)</i>	uncloned locus that affects migration of canal associated neurons	6/6	5/5
<i>cdh-4(ok1323)</i>	member of the cadherin superfamily	10/10	8/8
<i>cdh-5(hc181)</i>	member of the cadherin superfamily	10/10	7/7
<i>cdh-7(ok428)</i>	contains a cadherin domain	10/10	not observed
<i>ced-2(e1752)</i>	Src homology (SH) 2 and 3-containing adaptor protein	10/10	11/11
<i>ced-5(n1812)</i>	homolog of the human protein DOCK180	10/10	13/13
<i>ced-12(k149)</i>	Regulator of Rac1, required for phagocytosis and cell migration	11/11	12/12
<i>ceh-10(ct78)</i>	Paired-like class of homeodomain proteins	10/10	8/8
<i>ces-1(n1414)</i>	C2H2-type zinc finger transcription factor that is a member of the Snail family of proteins	11/11	11/11
<i>ces-1(n703)</i>	C2H2-type zinc finger transcription factor that is a member of the Snail family of proteins	16/16	14/14
<i>cle-1(cg120)</i>	collagen protein with endostatin domain	9/9	13/13
<i>crb-1(ok931)</i>	homolog of <i>Drosophila</i> CRUMBS	9/9	9/9
<i>daf-1(m40)</i>	TGF-beta type I receptor homolog	8/9	6/6
<i>daf-4(e1364)</i>	type II transforming growth factor-beta (TGF-b) receptors	11/11	14/14
<i>daf-7(e1372)</i>	member of the transforming growth factor beta (TGFbeta) superfamily	12/12	14/14
<i>dgn-2(ok209)</i>	dystroglycan	27/27	20/20
<i>dbl-1(wk70)</i>	member of the transforming growth factor beta (TGFbeta) superfamily	21/21	6/6
<i>dpy-19(e1295)</i>	novel transmembrane protein	7/7	7/7
<i>efn-4(bx80)</i>	member of the ephrin family of ligands	10/10	17/17
<i>egl-15(MT3324)</i>	FGF-like receptor tyrosine kinase	13/13	7/7
<i>egl-17(e1313)</i>	fibroblast growth factor-like protein	13/13	8/8
<i>evl-20(ar103)</i>	a functional ortholog of human ADP-ribosylation factor-like protein 2	not observed	4/4
<i>F11D5.3(ok574)</i>	a putative tyrosine kinase homologous to human RS1	11/11	22/22
<i>flt-1(ok722)</i>	a putative homolog of flectin, an extracellular matrix protein	10/10	8/8
<i>gon-1(e1254)/eDf18</i>	a functional metalloprotease that defines a new sub-family of secreted proteases known as MPT (metalloprotease with thrombospondin type 1 repeats)	8/8	11/11
<i>gpn-1(ok377)</i>	glypican, a heparan sulfate proteoglycan anchored to the cell membrane by a GPI linkage	13/13	14/14
<i>hlh-8(nr2061)</i>	helix-loop-helix protein required for normal muscle development	10/10	18/18
<i>lad-1(ok1244)/sax-7</i>	ortholog of human L1CAM	10/10	13/13
<i>let-756(S2613)</i>	fibroblast growth factor (FGF)-like ligand	9/11	21/21
<i>lon-2(e678)</i>	member of the glypican family of heparan sulfate proteoglycans	13/13	19/19
<i>mab-20(ev574)</i>	semaphorin	9/10	13/13
<i>mab-20(bx24)</i>	semaphorin	10/10	10/10
<i>max-1(ju39)</i>	conserved PH/MyTH4/FERM domain-containing protein	15/15	10/10
<i>mig-1(e1787)</i>	Frizzled-like receptor	10/10	13/13
<i>mig-2(gm103)</i>	Rho family of GTP-binding proteins, similar to Rac	12/15	36/37
<i>mig-6(e1931)</i>	uncloned locus involved in cell migration	9/10	15/15
<i>mig-14(ga62)</i>	homologous to <i>drosophila</i> Wntless	10/10	15/15
<i>mig-15(rh326)</i>	Nck-interacting kinase (NIK)	8/9	10/10

<i>mig-17(k113)</i>	secreted metalloprotease that is a member of the ADAM (A Disintegrin And Metalloprotease) protein family	10/10	13/13
<i>mom-2(or42)</i>	member of the Wnt family of secreted signaling glycoproteins	6/6	20/20
<i>mom-5(or57)</i>	Frizzled-like receptor	2/5	7/8
<i>pkc-3(RNAi)</i>	Serine/threonine protein kinase, atypical Protein Kinase C	15/15	no data
<i>pld-1(ok986)</i>	phospholipase D1	10/10	2/2
<i>plx-1(nc37)</i>	plexin ortholog, semaphorin receptor	10/10	10/10
<i>ptp-3(op147)</i>	receptor-like tyrosine phosphatase	10/10	15/15
<i>qid-7(mu533)</i>	uncloned locus that affects Q neuroblast polarization and migration	11/11	10/10
<i>qid-8(mu342)</i>	uncloned locus that affects Q neuroblast polarization and migration	10/10	13/15
<i>rac-2(ok326)</i>	Rho family GTPase that is one of three C. elegans Rac-related proteins	10/10	7/7
<i>rig-4(ok1160)</i>	Immunoglobulin C-2 Type/fibronectin type III domains	10/10	5/5
<i>sax-3(ky123)</i>	homologous to Drosophila roundabout	11/11	10/10
<i>sdn-1(ok449)</i>	a type I transmembrane heparan sulfate proteoglycan, syndecan	13/13	5/5
<i>slt-1(eh15)</i>	homolog of Drosophila slit	16/16	11/11
<i>slt-1(ok255)</i>	homolog of Drosophila slit	80/81	59/59
<i>sma-6(wk7)</i>	serine/threonine protein kinase that is orthologous to type I TGF-beta receptors	10/10	15/15
<i>smp-1(ev715)</i>	semaphorin	10/11	12/12
<i>smp-2(ev709)</i>	semaphorin	10/10	10/10
<i>syg-1(ky652)</i>	a novel transmembrane protein	9/9	12/12
<i>syg-2(ky671)</i>	transmembrane protein, immunoglobulin superfamily	10/10	4/4
<i>tag-150(gk261)</i>	Guanine nucleotide exchange factor for Rho and Rac GTPases	10/10	11/11
<i>unc-3(e151)</i>	a protein with homology to immunoglobulin (Ig) domain-containing transcription factors	10/10	10/10
<i>unc-5(e53)</i>	a netrin receptor required for dorsal cell and axon migration	10/10	22/22
<i>unc-6(ev400)</i>	netrin ortholog, secreted guidance molecule that regulates pioneer axons and mesodermal cells	4/20	14/18
<i>unc-14(e57)</i>	an activity required for both axonogenesis and sex myoblast migration	8/10	11/11
<i>unc-18(e81)</i>	an ortholog of Saccharomyces cerevisiae SEC1	10/11	12/12
<i>unc-30(e191)</i>	Pitx homeodomain transcription factor family member	10/10	10/10
<i>unc-33(mn407)</i>	CRMP/TOAD/Ulip/DRP homologue	10/10	10/11
<i>unc-34(e315)</i>	EVH1 domain-containing protein that is the sole C. elegans Enabled/VASP homologue	6/10	12/12
<i>unc-34(e566)</i>	EVH1 domain-containing protein that is the sole C. elegans Enabled/VASP homologue	5/8	5/5
<i>unc-39(e257)</i>	homeodomain transcription factor that belongs to the Six4/5 family	10/10	11/11
<i>unc-40(e271)</i>	A netrin receptor required for guiding dorsal and ventral cell and axon migrations	6/21	13/18
<i>unc-44(e1197)</i>	ankyrin-like protein	10/10	10/10
<i>unc-44(e362)</i>	ankyrin-like protein	10/10	2/2
<i>unc-51(e369)</i>	serine/threonine kinase involved in autophagy	9/10	13/13
<i>unc-51(e1189)</i>	serine/threonine kinase involved in autophagy	10/12	10/10
<i>unc-53(e404)</i>	orthologous to human NAV1, NAV2/RAINB1, and NAV3	11/11	7/7
<i>unc-53(n152)</i>	orthologous to human NAV1, NAV2/RAINB1, and NAV3	10/10	13/13
<i>unc-60(e723)</i>	orthologs of actin depolymerizing factor/cofilin	15/15	16/16
<i>unc-60(su158)</i>	orthologs of actin depolymerizing factor/cofilin	37/39	30/30
<i>unc-71(e541)</i>	ADAM, a disintegrin and metalloprotease-containing transmembrane protein	10/10	10/10
<i>unc-73(e936)</i>	guanine nucleotide exchange factor (GNEF) similar to the trio protein	not observed	39/39
<i>unc-73(gm40)</i>	guanine nucleotide exchange factor (GNEF) similar to the trio protein	6/11	22/22
<i>unc-76(e911)</i>	coiled-coil protein that belongs to the FEZ (fasciculation and elongation protein; zygin/zeta-1) family of proteins	9/11	10/10
<i>unc-78(gk27)</i>	homolog of actin-interacting protein 1	26/26	27/27
<i>unc-97(su10)</i>	LIM domain-containing protein of the PINCH family	10/10	14/14
<i>unc-115(e2225)</i>	Actin-binding LIM Zn-finger protein Limatin involved in axon guidance	10/10	15/15
<i>unc-115(ky275)</i>	Actin-binding LIM Zn-finger protein Limatin involved in axon guidance	67/67	68/68
<i>unc-115(mn481)</i>	Actin-binding LIM Zn-finger protein Limatin	8/11	24/24

<i>unc-129(ev554)</i>	member of the transforming growth factor beta (TGFbeta) superfamily	5/5	11/11
<i>vab-1(e2)</i>	ephrin receptor	10/10	10/10
<i>vab-8(e1017)</i>	novel protein containing an atypical kinesin-like motor domain	10/10	10/10
<i>vab-9(e1744)</i>	claudin homolog	8/9	13/13
<i>ver-1(ok859)</i>	Fibroblast/platelet-derived growth factor receptor and related receptor tyrosine kinases	10/10	4/4
<i>ver-2(ok897)</i>	Fibroblast/platelet-derived growth factor receptor and related receptor tyrosine kinases	10/10	8/8
<i>ver-3(gk227)</i>	Fibroblast/platelet-derived growth factor receptor and related receptor tyrosine kinases	11/11	13/13
<i>ver-4(ok1079)</i>	Fibroblast/platelet-derived growth factor receptor and related receptor tyrosine kinases	10/10	5/5
<i>zig-1(ok784)</i>	secreted 2-immunoglobulin-domain protein	9/9	5/5
<i>zig-2(ok696)</i>	secreted 2-immunoglobulin-domain protein	9/9	11/11
<i>zig-3(gk33)</i>	secreted 2-immunoglobulin-domain protein	10/10	4/4
<i>zig-4(gk4)</i>	secreted 2-immunoglobulin-domain protein	10/10	4/4
<i>zig-5(ok1065)</i>	secreted 2-immunoglobulin-domain protein	11/11	10/10
<i>zig-6(ok723)</i>	secreted 2-immunoglobulin-domain protein	10/10	3/3
<i>zig-8(ok561)</i>	secreted 2-immunoglobulin-domain protein	9/9	4/4

<sup>a</sup> ACs showing any degree of invasion were scored as invaded and invasion was scored over the entire range of the 4-cell stage, including the L3 molt. In contrast, Table S2 4-cell stage animals were only scored at beginning of the 4-cell stage at the mid-to-late L3 stage, and partial invasions were scored.

**Table S2. Timing and degree of AC invasion into the vulval epithelium**

Genotype/Treatment	ACs showing full, partial or no invasion							
	P6.p 4-cell stage (mid-to-late L3 stage)				P6.p 8-cell stage (early L4 stage)			
	% Full Invasion	% Partial Invasion	% No Invasion	n =	% Full Invasion	% Partial Invasion	% No Invasion	n =
wild-type (N2)	100	0	0	>100	100	0	0	>100
<u>unc-6/site of action</u>								
<i>unc-6(ev400)</i>	0	26	74	54	52	26	22 <sup>h</sup>	54
<i>ghIs8[unc-6&gt;Venus::unc-6]; unc-6(ev400)<sup>d</sup></i>	91	4	5	55	100	0	0	53
<i>unc-6(ev400); ghEx15[glr-1p&gt;Venus::unc-6; tph-1p&gt;GFP]<sup>b</sup></i>	90	4	6	50	96	4	0	50
<i>unc-6(ev400); ghEx13[egl-17&gt;Venus::unc-6]<sup>c</sup></i>	8	22	70	50	60	16	25	58
<i>rde-1(ne219); ghEx11[egl-17&gt;rde1::mRFP]; unc-6(RNAi)<sup>d</sup></i>	100	0	0	51	100	0	0	34
<i>lin-3(n1059)/lin-3(n378) (Vul)<sup>e</sup></i>	21	0	79	52	19	0	81 <sup>i</sup>	52
<i>lin-3(n1059)/lin-3(n378); unc-6(ev400)</i>	0	0	100	50	0	0	100 <sup>j</sup>	50
<i>laser killed P3.p-P8.p(Vul); unc-6(ev400)</i>	0	0	100	20	0	0	100 <sup>k</sup>	20
<i>kyIs299 [hs&gt;unc-6::HA]<sup>f</sup></i>	50	26	24	50	nd	nd	nd	nd
<i>N2(mock heat shock)</i>	98	2	0	51	nd	nd	nd	nd
<u>Netrin receptors/unc-40/site of action</u>								
<i>unc-40(e271)</i>	2	31	67	54	58	25	17 <sup>l</sup>	53
<i>unc-40(e271); unc-6(ev400)</i>	2	29	69	51	57	21	21 <sup>m</sup>	70
<i>unc-40(e271); qyIs66[cdh-3&gt;unc-40::gfp]</i>	96	2	2	54	98	2	0	53
<i>unc-5(e51)</i>	100	0	0	58	100	0	0	64
<u>Intracellular effectors</u>								
<i>mig-10(ct41)</i>	100	0	0	59	100	0	0	81
<i>ced-10(n1993)</i>	98	2	0	61	100	0	0	57
<i>mig-2(mu28)</i>	100	0	0	54	100	0	0	64
<i>ced-10(n1993); mig-2(mu28)</i>	24	10	66	29	69	5	25	55
<i>unc-34(gm104)</i>	64	16	20	50	100	0	0	50
<i>qyIs23[cdh-3&gt;mCherry::PLC<math>\delta^{PH}</math>]<sup>g</sup></i>	85	13	2	55	100	0	0	50
<i>qyIs23[cdh-3&gt;mCherry::PLC<math>\delta^{PH}</math>]; mig-2(mu28)</i>		18	12	114	99	1	0	110
<i>qyIs23[cdh-3&gt;mCherry::PLC<math>\delta^{PH}</math>]; unc-34(gm104)</i>	41	18	41	51	89	9	2	56
<u><i>fos-1</i> pathway interaction</u>								
<i>unc-40(e271); rrf-3(pk1426)</i>	8	27	65	51	50	22	28	50
<i>unc-40(e271); rrf-3(pk1426); fos-1(RNAi)</i>	4	11	85	54	12	12	77	52
<i>rrf-3(pk1426); fos-1(RNAi)</i>	10	8	82	50	16	26	58	50

<sup>a</sup> Venus::unc-6 driven by its own promoter. <sup>11</sup>

<sup>b</sup> Ventral nerve cord specific expression of Venus::unc-6 driven by *glr-1* promoter. <sup>11</sup>

<sup>c</sup> 1° VPC specific expression of Venus::unc-6 driven by the *egl-17* promoter. <sup>11</sup>

<sup>d</sup> Targeted RNAi mediated knockdown of *unc-6* in the 1° VPCs. <sup>11</sup>

<sup>e</sup> A similar percentage of ACs have been shown to invade when vulval cells are removed by ablation. <sup>5</sup>

<sup>f</sup> 2h heat shock at 30°C followed by 4h recovery at 15°C.

<sup>g</sup> *qyIs23[cdh-3>mCherry::PLC $\delta^{PH}$ ]* binds and sequesters PI(4,5)P<sub>2</sub> in the AC, thus reducing its levels there.

<sup>h-m</sup> The number of AC's that detached from the basement membrane at the L4 stage was as follows (number of detached/number that failed to invade): h = 2/12; i = 11/42; j = 47/50; k = 18/20 l = 0/9; m = 2/15.

nd = not determined

**Table S3. Polarized marker localization in wild-type and mutant ACs**

Genotype	Marker	Classification of marker localization patterns in wild-type and various mutant ACs											
		P6.p 1-cell stage				P6.p 2-cell stage				P6.p 4-cell stage			
		Polarized (%±SE)	Apicolateral Accumulation (%±SE)	No Polarity (%±SE)	N=	Polarized (%±SE)	Apicolateral Accumulation (%±SE)	No Polarity (%±SE)	N=	Polarized (%±SE)	Apicolateral Accumulation (%±SE)	No Polarity (%±SE)	N=
wild-type	mCherry::PLC $\delta^{PH}$	85±8	15±8	0	20	86±8	14±8	0	21	95±5	0	5±5	20
wild-type	GFP::MIG-2	86±7	13±7	0	22	100	0	0	22	100	0	0	23
wild-type	mCherry::moeABD	100	0	0	18	100	0	0	21	95±5	5±5	0	21
wild-type	UNC-40::GFP	90±7	5±5	5±5	20	100	0	0	21	90±6	10±6	0	20
<i>unc-6(ev400)</i>	mCherry::PLC $\delta^{PH}$	22±10	38±12	38±12	18	15±9	53±12	32±11	19	0	69±12	31±12	16
<i>unc-6(ev400)</i>	mCherry::moeABD	29±11	53±12	18±10	17	29±11	71±11	0	17	18±10	76±11	6±6	17
<i>unc-6(ev400)</i>	UNC-40::GFP	21±10	32±11	47±12	19	5±5	42±12	53±12	20	0	25±10	75±10	20
<i>unc-6(ev400)</i>	GFP::MIG-2	33±11	38±11	29±10	21	14±8	43±11	43±11	21	25±10	40±11	35±11	20
Vul <sup>a</sup>	mCherry::PLC $\delta^{PH}$	84±9	16±9	0	19	90±7	10±7	0	20	73±12	27±12	0	15
Vul <sup>a</sup>	UNC-40::GFP	94±6	6±6	0	17	86±8	14±8	0	21	94±6	6±6	0	17
Vul <sup>a</sup>	GFP::MIG-2	95±5	5±5	0	20	87±7	13±7	0	23	76±10	24±10	0	21
Vul <sup>a</sup>	mCherry::moeABD	88±9	6±6	6±6	16	86±8	14±7	0	21	90±7	10±6	0	20
<i>hs&gt;unc-6::HA<sup>b</sup></i>	UNC-40::GFP	nd	nd	nd	nd	nd	nd	nd	nd	33±13	33±13	33±13	15
wild-type (mock) <sup>c</sup>	UNC-40::GFP	nd	nd	nd	nd	nd	nd	nd	nd	93±7	7±7	0	15

<sup>a</sup> Vul (vulvaless) animals were of the genotype *lin-3(n1059)/lin-3(n378)*.

<sup>b</sup> Heat shock directed expression of *unc-6::HA* from the integrated transgene *kyIs299*, 2h heat shock at 30°C followed by 4h recovery at 15°C.

<sup>c</sup> wild-type animals expressing UNC-40::GFP in the AC were subjected to an identical experimental regimen to confirm that heat shock alone does not cause defects in UNC-40::GFP localization.



**Table S4. Primer sequences and templates used for PCR fusions**

Primer Sequence	Primer Type	Amplicon	Template
5' TAA TgT gAg TTA gCT CAC TCA TTA gg 3'	forward	<i>cdh-3/zmp-1</i> > promoters	<i>cdh-3</i> >, pPD107.94/mk62-63; <i>zmp-1</i> >, pPD107.94/mk50-51
5' AAC gAT ggA TAC gCT AAC AAC TTg g 3'	forward nested	<i>cdh-3/zmp-1</i> > promoters	<i>cdh-3</i> >, pPD107.94/mk62-63; <i>zmp-1</i> >, pPD107.94/mk50-51
5' TTT CTg AgC TCg gTA CCC TCC AAg 3'	reverse	<i>cdh-3/zmp-1</i> > promoters	<i>cdh-3</i> >, pPD107.94/mk62-63; <i>zmp-1</i> >, pPD107.94/mk50-51
5' TAG gCT TTT CCg TAT AgC ATC CTC 3'	forward	<i>fos-1a</i> > promoter	cosmid F29G9
5' gCC CAA CTC TAg TCA TTT CTA gC 3'	forward nested	<i>fos-1a</i> > promoter	cosmid F29G9
5' TCC ACT CTC TTA TAT AgC AgA ggT gC 3'	reverse	<i>fos-1a</i> > promoter	cosmid F29G9
5' CTT ggA ggg TAC CgA gCT Cag AAA ggT ACC Atg AgT AAA ggA gAA g 3'	<i>cdh-3/zmp-1</i> extension, forward	<i>GFP::unc-34</i>	<i>punc-86</i> > <i>GFP::unc-34</i>
5' Cgg gAA gCT AgA gTA AgT AgT TCg CC 3'	reverse	<i>GFP::unc-34</i>	<i>punc-86</i> > <i>GFP::unc-34</i>
5' CTC TCA Agg ATC TTA CCg CTg TTg 3'	reverse nested	<i>GFP::unc-34</i>	<i>punc-86</i> > <i>GFP::unc-34</i>
5' CTT ggA ggg TAC CgA gCT Cag AAA ATg ATT TTg CgA CAT TTC gg 3'	<i>cdh-3/zmp-1</i> extension, forward	<i>unc-40::GFP</i>	<i>punc-86</i> > <i>unc-40::GFP</i>
5' gTg CCA CCT gAC gTC TAA g 3'	reverse	<i>unc-40::GFP</i>	<i>punc-86</i> > <i>unc-40::GFP</i>
5' gTA Cgg CCg ACT AgT Agg AAA CAg T 3'	reverse nested	<i>unc-40::GFP</i>	<i>punc-86</i> > <i>unc-40::GFP</i>
5' CTT ggA ggg TAC CgA gCT Cag AAA ATg gTC TCA AAg ggT gAA g 3'	<i>cdh-3/zmp-1</i> extension, forward	<i>mCherry::moeABD</i>	pJWZ6
5' CAg gAA ACA gCT ATg ACC ATg 3'	reverse	<i>mCherry::moeABD</i>	pJWZ6
5' gCC gCT CTA gAA TCA TCg TTC 3'	reverse nested	<i>mCherry::moeABD</i>	pJWZ6
5' CTT ggA ggg TAC CgA gCT Cag AAA ATg gCT CAA ACA AAg CCg ATT gCC 3'	<i>cdh-3/zmp-1</i> extension, forward	<i>mCherry::PLCδ<sup>PH</sup></i>	pAA173
5' TTC gAg CgA Agg TCg CTT TTT ggT C 3'	reverse	<i>mCherry::PLCδ<sup>PH</sup></i>	pAA173
5' TTg AAA TCg AgT TgC AAg CgC gCT CC 3'	reverse nested	<i>mCherry::PLCδ<sup>PH</sup></i>	pAA173
5' CTT ggA ggg TAC CgA gCT Cag AAA ATg gCT CAA ACA AAg CCg ATT gCC 3'	<i>cdh-3/zmp-1</i> extension, forward	<i>mCherry</i>	pAA64
5' TTC gAg CgA Agg TCg CTT TTT ggT C 3'	reverse	<i>mCherry</i>	pAA64
5' TTg AAA TCg AgT TgC AAg CgC gCT CC 3'	reverse nested	<i>mCherry</i>	pAA64
5' gCA CCT CTg CTA TAT AAg AgA gTg gAA Tgg CTC AAA CAA AgC CgA TTg CC 3'	<i>fos-1a</i> extension, forward	<i>mCherry::PLCδ<sup>PH</sup></i>	pAA173
5' TTC gAg CgA Agg TCg CTT TTT ggT C 3'	reverse	<i>mCherry::PLCδ<sup>PH</sup></i>	pAA173
5' TTg AAA TCg AgT TgC AAg CgC gCT CC 3'	reverse nested	<i>mCherry::PLCδ<sup>PH</sup></i>	pAA173
5' TgT AAA ACg ACg gCC AgT 3'	forward	<i>unc-6</i> > promoter	<i>pVns-unc-6</i>
5' TgT AAA ACg ACg gCC AgT 3'	forward nested	<i>unc-6</i> > promoter	<i>pVns-unc-6</i>
5' gTT CTT CTC CTT TAC TgT TTg TgT gAA Agg gTg Taa AgT ggA 3'	reverse	<i>unc-6</i> > promoter	<i>pVns-unc-6</i>
5' TCC ACT TTA CAC CCT TTC ACA CAA ACA TgA gTA AAg gAg AAg gAg AAg AACTTT TCA CTg g 3'	<i>unc-6</i> promoter extension, forward	<i>venus::unc-6(ΔSP)</i>	<i>pVns-unc-6</i>
5' CAg gAA ACA gCT ATg ACC ATg 3'	reverse	<i>venus::unc-6(ΔSP)</i>	<i>pVns-unc-6</i>
5' ATg ACC ATg ATT ACg CCA AgC gC 3'	reverse nested	<i>venus::unc-6(ΔSP)</i>	<i>pVns-unc-6</i>

**Table S5. Extrachromosomal array and integrated strain generation**

<b><i>Ex</i> Designation</b>	<b><i>Is</i> Designation</b>	<b>PCR Fusion created</b>	<b>Injected Concentration</b>	<b>Co-Injection Marker(s)</b>
<i>qyEx27</i>	<i>qyIs23, qyIs24, qyIs25</i>	<i>cdh-3&gt;mCherry::PLCδ<sup>PH</sup></i>	0.01ng/μl	<i>unc-119</i>
<i>qyEx39</i>	<i>qyIs50</i>	<i>cdh-3&gt;mCherry::moeABD</i>	2ng/μl	<i>unc-119</i>
<i>qyEx40</i>	<i>qyIs66, qyIs67</i>	<i>cdh-3&gt;unc-40::GFP</i>	2ng/μl	<i>unc-119 + myo-2&gt;YFP</i>
<i>qyEx30</i>	<i>qyIs37</i>	<i>zmp-1&gt;unc-40::GFP</i>	2ng/μl	<i>unc-119</i>
<i>qyEx42</i>	<i>qyIs61</i>	<i>cdh-3&gt;unc-34::GFP</i>	1ng/μl	<i>unc-119</i>
<i>qyEx60</i>	not integrated	<i>fos-1a&gt;mCherry::PLCδ<sup>PH</sup></i>	0.25ng/μl	<i>pha-1</i>
<i>qyEx68</i>	<i>qyIs7</i>	<i>zmp-1&gt;mCherry</i>	1.0ng/μl	<i>unc-119</i>
<i>qyEx3</i>	<i>qyIs17</i>	pGK41( <i>lam-1::GFP</i> )	10ng/μl	<i>unc-119</i>
<i>qyEx19</i>	<i>qyIs27</i>	pPR80( <i>GFP::ced-10</i> )	75ng/μl	<i>unc-119</i>
<i>qyEx78</i>	not integrated	<i>Venus::unc-6(ΔSP)</i>	15ng/μl	<i>unc-119</i>