

Supporting Information

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SI Methods

Isolation of DAPK-1 cDNA. To isolate a full-length DAPK-1 cDNA, we designed primers based on an early predicted gene structure for K12C11.4 and used these in RT-PCR from polyA(+) RNA from mixed-stage animals. The purified products were cloned into pST-Blue vector (Novagen) and confirmed by sequencing. The *dapk-1* ORF is 4275 bp and encodes a 1425 residue polypeptide.

RNA Interference of Secretion and Endocytosis Genes. We used postembryonic feeding RNAi to inhibit the following genes: adaptin mu3 (*apm-3*), dynamin (*dyn-1*), Rab1 (*rab-1*), Sec23 (*sec-23*), synaptotagmin (*snt-2*), Vps32 (*vps-32.1*), and Vps54 (*vps-54*), in either *rrf-3(pk1426)* or in *dapk-1(ju469); rrf-3(pk1426)* host strains. Embryos were hatched on plates containing the RNAi bacteria and grown at 25 °C for 3 days before scoring. Because RNAi of some genes (*dyn-1*, *rab-1*, *sec-23*, *vps-32.1*) causes highly penetrant larval

lethality, we titrated the RNAi bacteria between 1:10 to 1:100 with control HT115 bacteria to find RNAi levels compatible with viability to the L4 stage.

Cuticle Fragility Tests. We tested cuticle strength by measuring time to cuticle rupture in bleach. Day old hermaphrodites were placed in freshly made alkaline hypochlorite solution (1N NaOH, 2% hypochlorite) on a microscope slide and the time to cuticle rupture recorded. Wild type worms typically rupture at the vulva, whereas *dapk-1* mutants ruptured at the regions of cuticle thickening at the head or dorsal midline.

Microarray Transcriptional Profiling. RNA was isolated from L4 stage *dapk-1(ju4)* animals and hybridized to oligonucleotide arrays essentially as described (1). Data in SI Table S4 show the top 18.75 percentile (of 303 genes) with median fold up-regulation >1.24 across 3 replicates.

1. Pujol N, et al. (2008) Anti-fungal innate immunity in *C. elegans* is enhanced by evolutionary diversification of antimicrobial peptides. *PLoS Pathog* 4:e1000105.
2. Tereshko V, Teplova M, Brunzelle J, Watterson DM, Egli M (2001) Crystal structures of the catalytic domain of human protein kinase associated with apoptosis and tumor suppression. *Nat Struct Biol* 8:899–907.
3. Michaux G, Gansmuller A, Hindelang C, Labouesse M (2000) CHE-14, a protein with sterol-sensing domain, is required for apical sorting in *C. elegans* ectodermal epithelial cells. *Curr Biol* 10:1098–1107.
4. Wong D, Bazopoulou D, Pujol N, Tavernarakis N, Ewbank JJ (2007) Genome-wide investigation reveals pathogen-specific and shared signatures in the response of *Caenorhabditis elegans* to infection. *Genome Biol* 8:R194.

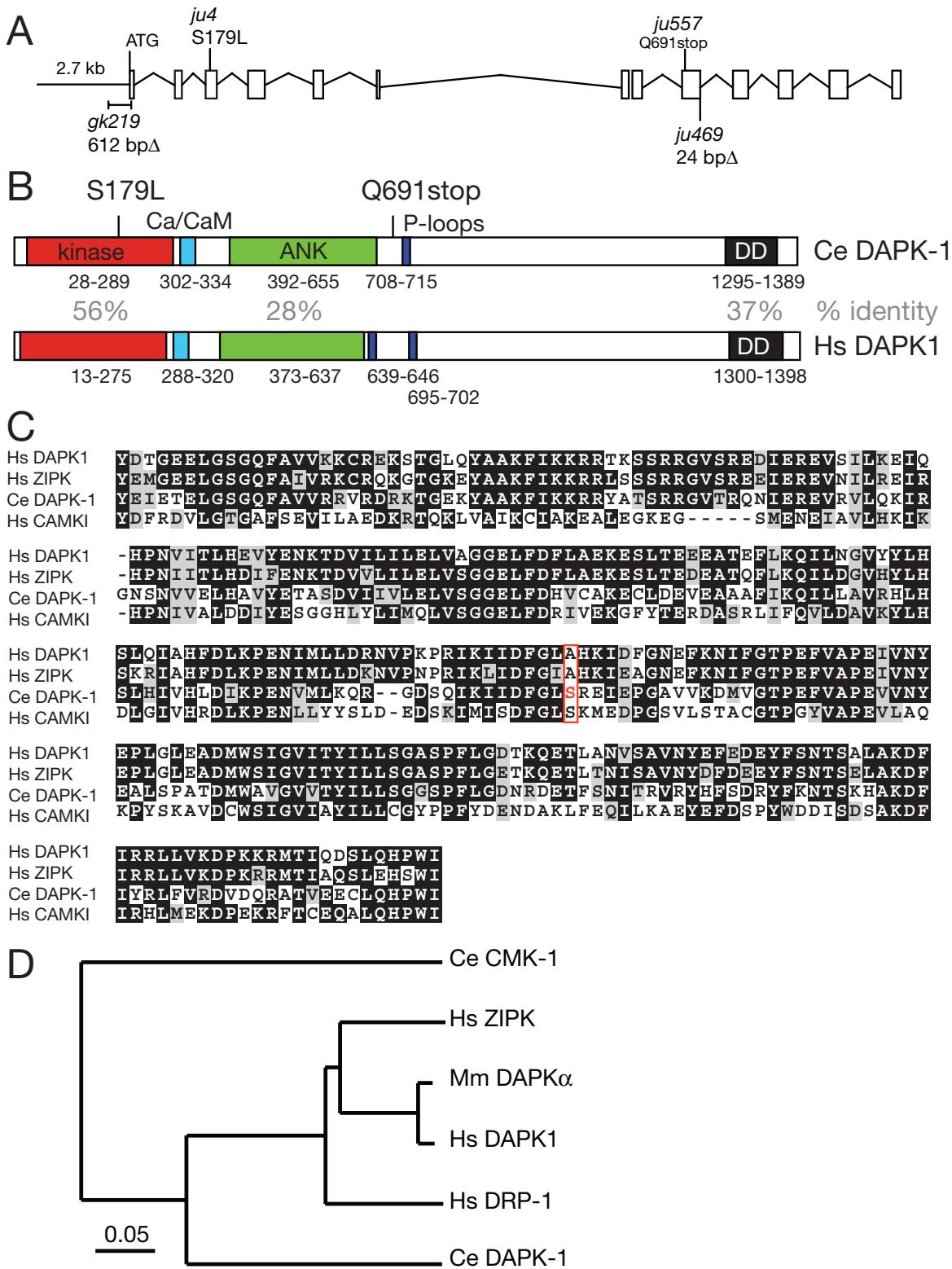


Fig. S1. *dapk-1* encodes the *Caenorhabditis elegans* ortholog of death-associated protein kinase (DAPK). (A) *C. elegans* *dapk-1* gene structure showing positions of mutations and the 2.7-kb promoter used in rescuing construct pcZ767; *ju4* results in the missense alteration S179L, and is predicted to affect the substrate binding groove of the kinase domain (2); *gk219* deletes 516 bp of the 5' UTR and 96 bp of exon 1 (codons 1–32); *ju469* is a 24 bp deletion of the last 10 bp of exon 9 and 14 bp of intron 9; *ju469* is predicted to truncate DAPK-1 after residue 862; *ju557* is a nonsense mutation (Q691stop) in exon 9 after the ANK repeats. (B) Comparison of *C. elegans* DAPK-1 to human DAPK1, showing kinase domains (red), Ca²⁺/calmodulin regulatory domain (blue), ANK repeats (green), P-loops (purple), death domain (black), and percentage identity. DAPK-1 is overall 33% identical to human DAPK1. (C) Clustal 2.0 alignment of DAPK-1 kinase domain with human DAPK1, human ZIP kinase and human CaM kinase I. S179 (red) corresponds to A165 in human DAPK. (D) Phylogenetic tree (ClustalW and njplot) of kinase domains of *C. elegans* DAPK-1, *C. elegans* CaM kinase I/CMK-1, human DAPK1, human ZIP kinase, human DRP-1 and mouse DAPK. DAPK-1 is the only *C. elegans* member of the DAPK family.

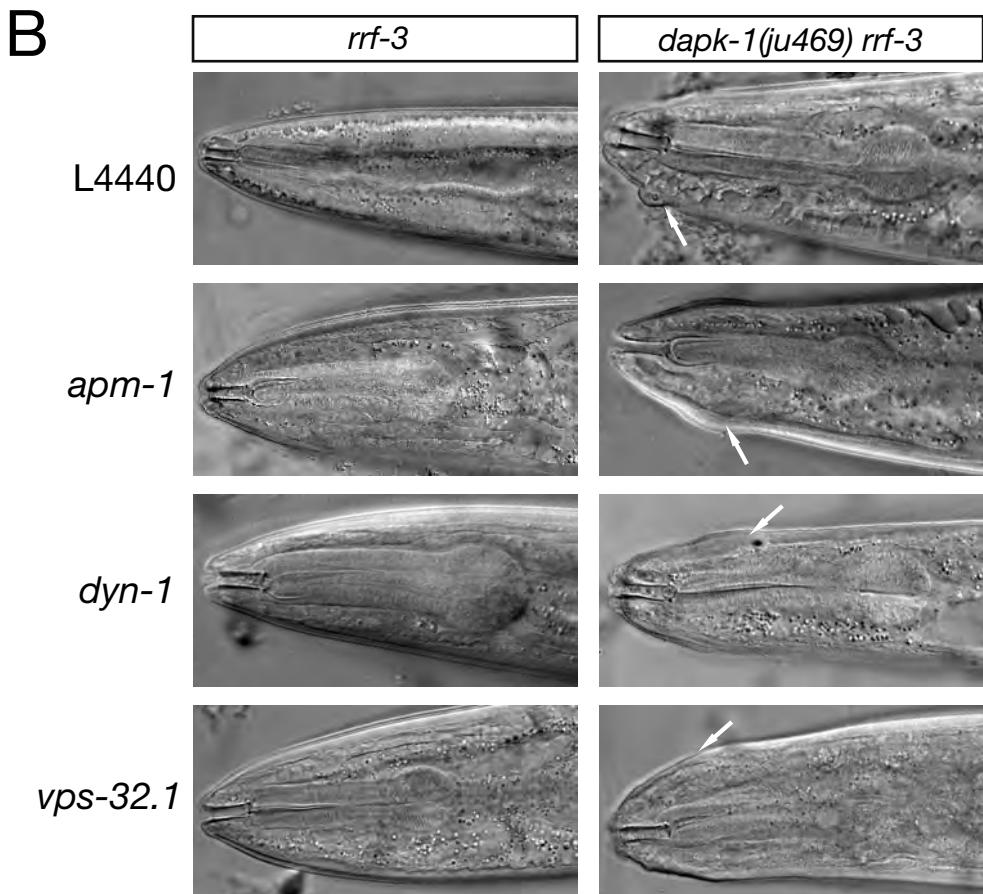
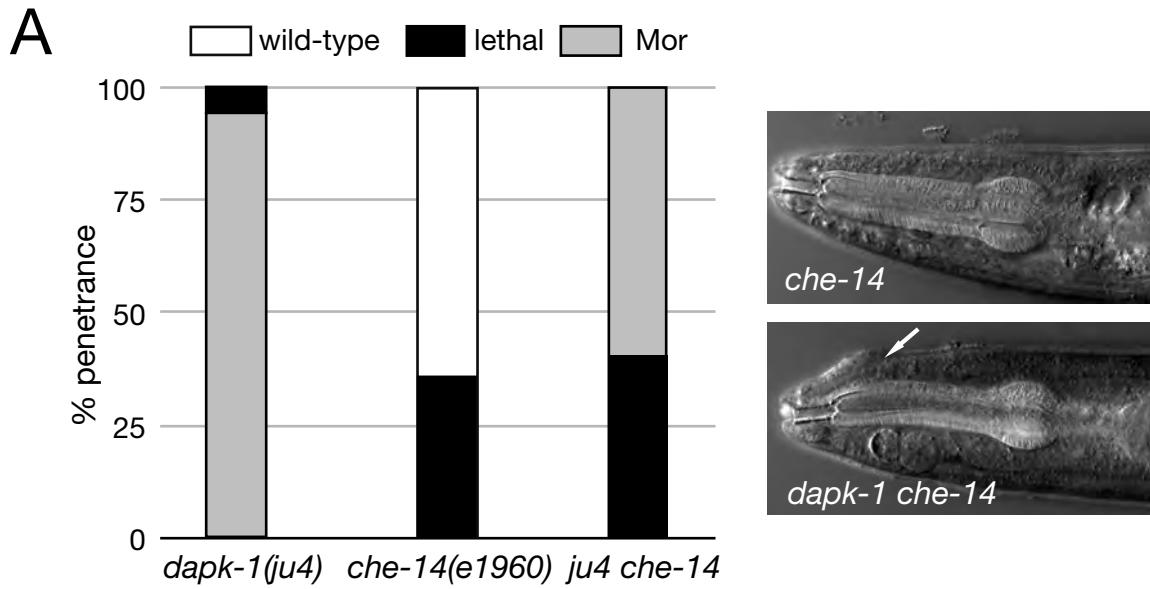


Fig. S2. The *dapk-1* phenotypes are not phenocopied, enhanced, or suppressed by loss of function in epithelial endocytosis or secretion pathways. (A) The *dapk-1(ju4)* *che-14(e1960)* double mutants display additive epidermal and lethal phenotypes; *che-14* encodes a Ptc family protein involved in apical secretion by the epidermis (3) ($n > 500$, animals for each genotype). Arrow indicates the head morphology (Mor) defect of *dapk-1* mutants. (B) Photomicrographs of head morphology in RNAi experiments inhibiting genes in the secretory or endocytic pathways. The effects of postembryonic feeding RNAi of the indicated gene are shown in *rrf-3* and in *dapk-1(ju469) rrf-3* backgrounds. We observed no detectable suppression or enhancement of the *dapk-1* phenotypes.

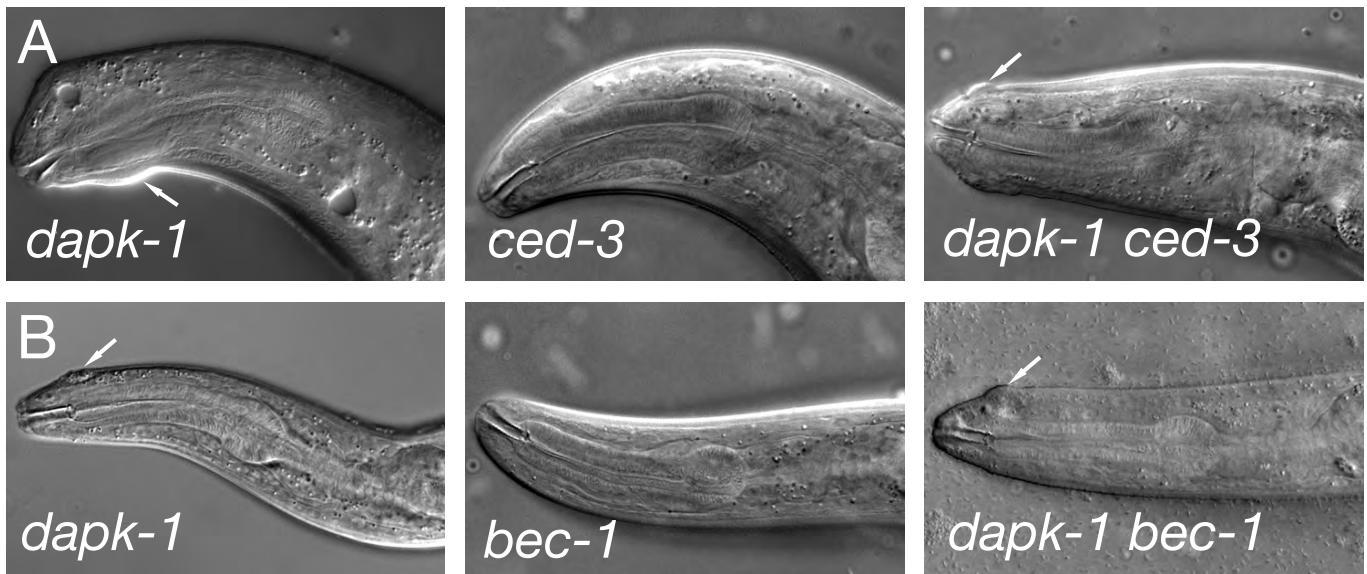


Fig. S3. Lack of genetic interaction of *dapk-1* with apoptosis or autophagy mutants in head morphogenesis. (A) Nomarski DIC micrographs of head morphology of adult *dapk-1(ju4)*, *ced-3(n717)*, and *ju4; n717* double mutants, showing *ced-3* neither phenocopies nor suppresses *dapk-1* epidermal morphology defects (arrows). (B) The *bec-1* mutants neither phenocopy nor suppress the *dapk-1* epidermal defects (arrows). As most *ok700* animals arrest in larval development, images are of L4 stage head region of *bec-1(ok700)* and *dapk-1; bec-1* double mutants. See Table S2 for quantitation.

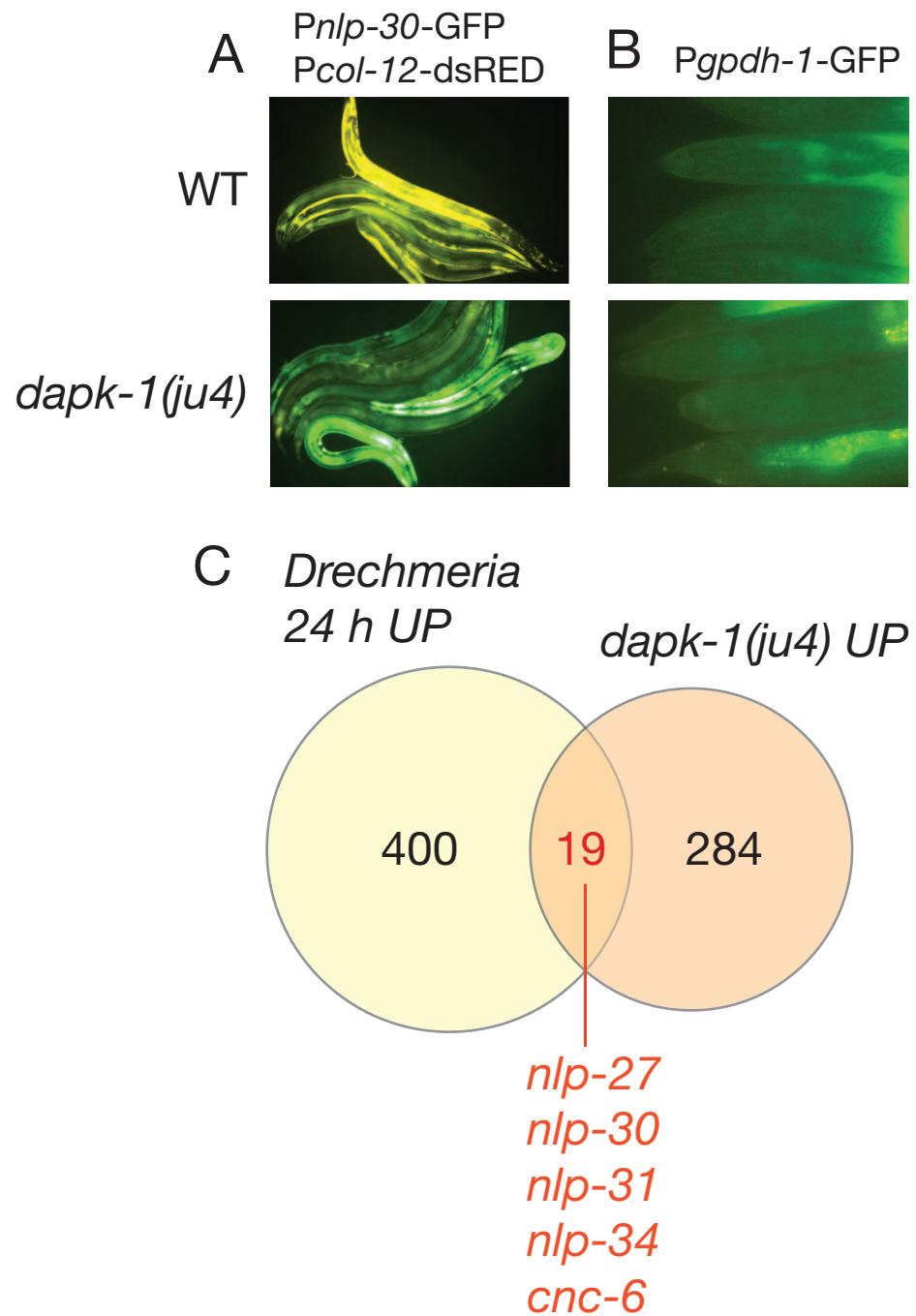


Fig. S4. Up-regulation of cutaneous innate immune response but not osmotic stress response in *dapk-1* animals. (A and B) Expression of *Pnlp-30-GFP/Pcol-12-dsRed* (A) in day 1 adults and (B) *Pgpdh-1-GFP* in L4 stage wild type and *dapk-1(ju4)*. Animals were photographed under identical camera settings. Residual fluorescence in the *Pgpdh-1-GFP* panels is due to gut autofluorescence. (C) Venn diagram of overlap between top-quartile *dapk-1(ju4)* up-regulated genes (303 genes) and *Drechmeria* infected worms (24 h, 419 genes). The 19 genes in common, listed in Table S4, include the AMPs *nlp-27*, *nlp-30*, *nlp-31*, *nlp-34*, and *cnc-6*. None of the 303 genes up-regulated in *dapk-1* are in the 254 genes of the bacterial “common response” (4).

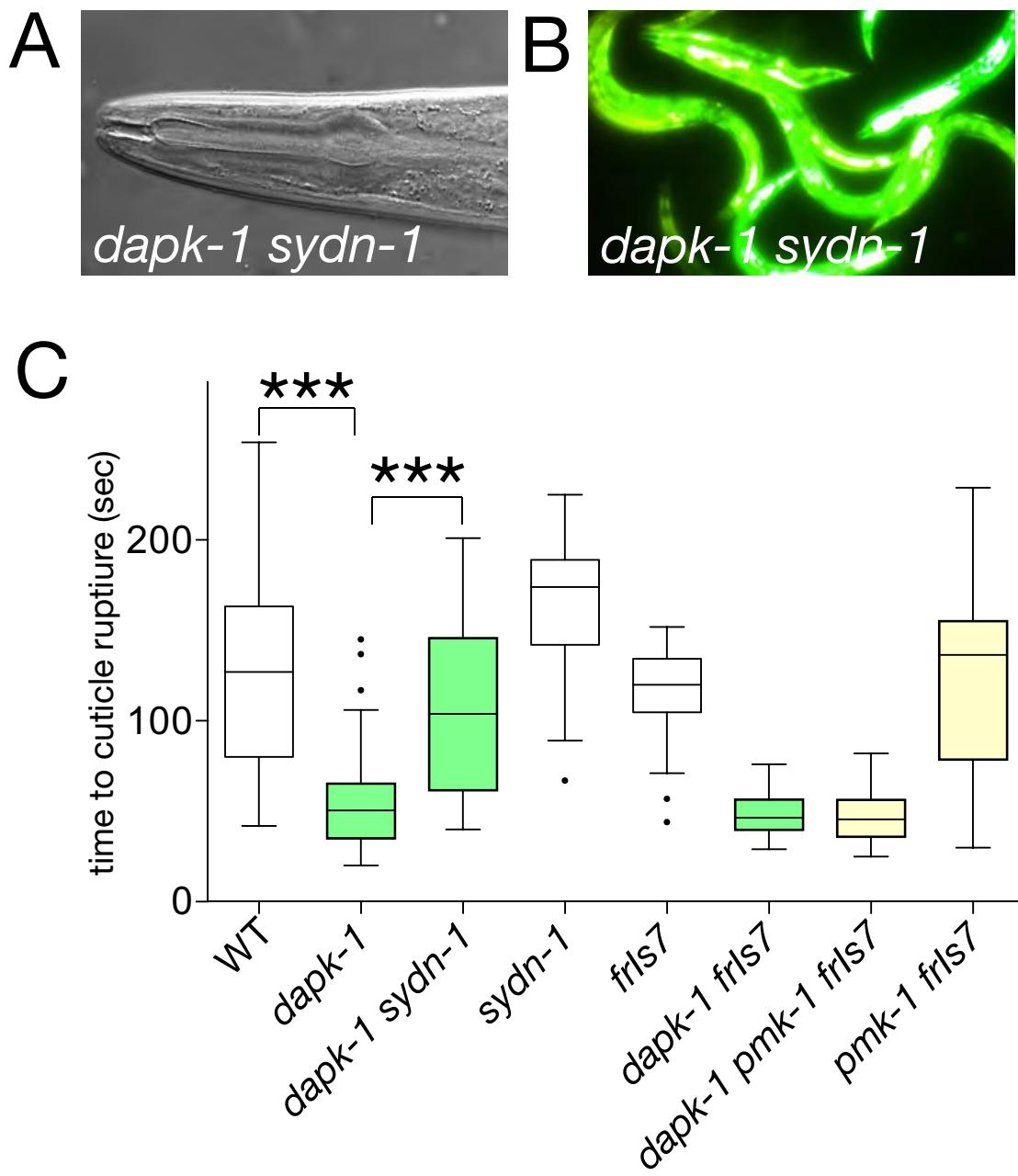


Fig. S5. Genetic suppression of *dapk-1* indicates up-regulation of AMPs is independent of morphological defects. (A and B) The *dapk-1(ju4)* *sydn-1(ju541)* double mutants have normal head morphology (A) and up-regulated *Pnlp-29-GFP* (B). (C) The *dapk-1* cuticle fragility defects are suppressed by *sydn-1* but not by *pmk-1*; Tukey boxplots ($n = 50$, for each genotype); tests of significance use *t* test, $P < 0.001$.

Table S1. Quantitation of *dapk-1* morphogenetic phenotypes

Genotype, temperature	Morphological defect, %	<i>n</i>
<i>dapk-1(ju4)</i> 25 °C	100	538
<i>dapk-1(ju4)</i> 20 °C	100	309
<i>dapk-1(ju4)</i> 15 °C	98	454
<i>dapk-1(ju469)</i> 25 °C	42	632
<i>dapk-1(ju469)</i> 20 °C	16	280
<i>dapk-1(ju469)</i> 15 °C	12	253
<i>dapk-1(ju557)</i> 25 °C	79	507
<i>dapk-1(ju557)</i> 20 °C	33	183
<i>dapk-1(ju557)</i> 15 °C	23	262
<i>dapk-1(gk219)</i> 25 °C	65	743
<i>dapk-1(gk219)</i> 20 °C	19	281
<i>dapk-1(gk219)</i> 15 °C	12	222

The *dapk-1* alleles form an allelic series, in order of phenotypic severity: *ju4* > *ju557* > *gk219* > *ju469*. All 4 alleles are temperature-sensitive, suggesting loss of *dapk-1* function reveals an underlying temperature-sensitive process.

Table S2. *dapk-1* double mutants

Genotype	Morphological defect, %	n
<i>dapk-1(ju4)</i>	100	538
<i>dapk-1(ju4) unc-54(e190)*</i>	100	112
<i>dapk-1(ju4); ced-3(n717)</i>	100	606
<i>dapk-1(ju4); ced-2(e1752)</i>	100	538
<i>dapk-1(ju469)</i>	42	632
<i>dapk-1(ju469); ced-3(n717)</i>	89	294
<i>dapk-1(ju469); ced-2(e1752)</i>	90	476
<i>dapk-1(ju4); bec-1(ok700)*</i>	100	53
<i>dapk-1(ju4); atgr-18(gk378)*</i>	100	231
<i>dapk-1(gk219); unc-51(e369)</i>	93	268
<i>dapk-1(ju4); pmk-1(km25)*</i>	100	666
<i>dapk-1(ju4); sek-1(km4)*</i>	100	271
<i>dapk-1(ju4); nsy-1(ky397)</i>	98	333
<i>dapk-1(ju4); tir-1(tm3036), 20 °C</i>	100	656
<i>dapk-1(ju4); sydn-1(ju541)</i>	0	513

Double mutants were scored at 25 °C or at room temperature (22–23 °C) (indicated by asterisks). Non-*dapk-1* single mutants have normal epidermal morphology. Because *dapk-1* partial loss of function mutants (*ju469*, *gk219*) are mildly enhanced by mutations that either prevent apoptosis, autophagy, or phagocytosis, these enhancement effects may reflect a nonspecific effect of overall sickness, or may reflect a cryptic role for all three processes in epidermal morphogenesis.

Table S3. Transgenic rescue of *dapk-1*

Genotype	Morphological defect, %	<i>n</i>
<i>dapk-1(ju4)</i>	100	538
<i>dapk-1(ju4); juEx886 [Pdapk-1-DAPK-1]</i>	10	62
<i>dapk-1(gk219)</i>	65	743
<i>dapk-1(gk219); juEx890 [Pdapk-1-DAPK-1]</i>	2	125
<i>dapk-1(gk219); juEx891 [Pdapk-1-DAPK-1]</i>	1	207
<i>dapk-1(ju4); juEx1681 [Pdpy-7-DAPK-1]</i>	19	317
<i>dapk-1(ju4); juEx1682 [Pdpy-7-DAPK-1]</i>	3	260
<i>dapk-1(ju4); juEx921 [Pdapk-1-GFP::DAPK-1]</i>	8	107
<i>dapk-1(gk219); juEx924[Pdapk-1-GFP::DAPK-1]</i>	0	160

All transgenic lines were scored at 25 °C. The transgenic rescue of *ju4* or *gk219* by all full-length DAPK-1 transgenes is highly significant by the Fisher exact test (data not shown).

Table S4. Genes upregulated in *dapk-1* and in *Drechmeria* infected worms

Gene	Median fold <i>dapk-1</i>	Median fold infected	Function
B0213.17 <i>nlp-34</i>	2.55	3.34	AMP
B0213.2 <i>nlp-27</i>	1.34	1.53	AMP
B0213.5 <i>nlp-30</i>	1.68	2.78	AMP
B0213.6 <i>nlp-31</i>	1.28	2.51	AMP
C06A8.3	1.39	1.18	—
C06G4.2a <i>clp-1</i>	1.48	1.20	Calpain
C13A10.2	1.26	1.23	—
C45B2.8	1.39	1.65	—
F19B6.1b	1.34	1.25	plakoglobin-like
F36H12.5	1.33	1.26	—
F55H12.4	1.36	2.25	—
K02F6.6	1.25	1.20	BTB/POZ domain
R13A5.6 <i>ttr-8</i>	1.28	1.24	Transthyretin
T19B10.2 <i>phi-59</i>	1.24	1.84	—
Y46E12A.1 <i>cnc-6</i>	1.73	1.85	AMP
Y53G8AR.6	1.35	1.25	SAP domain
Y54G11A.9	1.49	1.25	—
Y71H2AM.18	1.29	1.18	DEAD box
ZK637.14	1.44	1.33	RING finger

Common genes from the top quartiles of up-regulated genes. Data from *Drechmeria* infected worms are 24 h post infection (1).