

Research Paper

Anxa4 Genes are Expressed in Distinct Organ Systems in *Xenopus laevis* and *tropicalis* But are Functionally Conserved

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KEY WORDS

annexin 4, kidney, liver, evolution, paralog and ortholog, *Xenopus laevis*, *Xenopus tropicalis*, morpholino, organogenesis

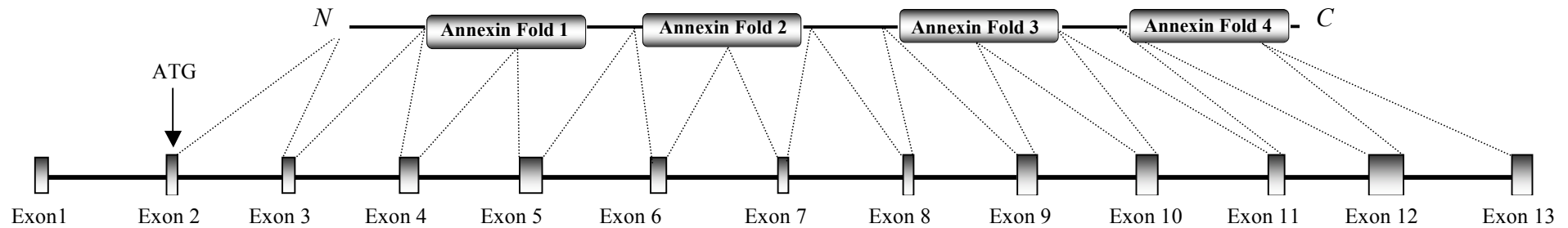
ABBREVIATIONS

Anx	Annexin
Anx4	Annexin4
anxa4a	<i>X. laevis</i> Annexin4
Xt anx4	<i>X. tropicalis</i> anx4
anxa4b	<i>X. laevis</i> annexin4 pseudoallele
sd	standard deviation

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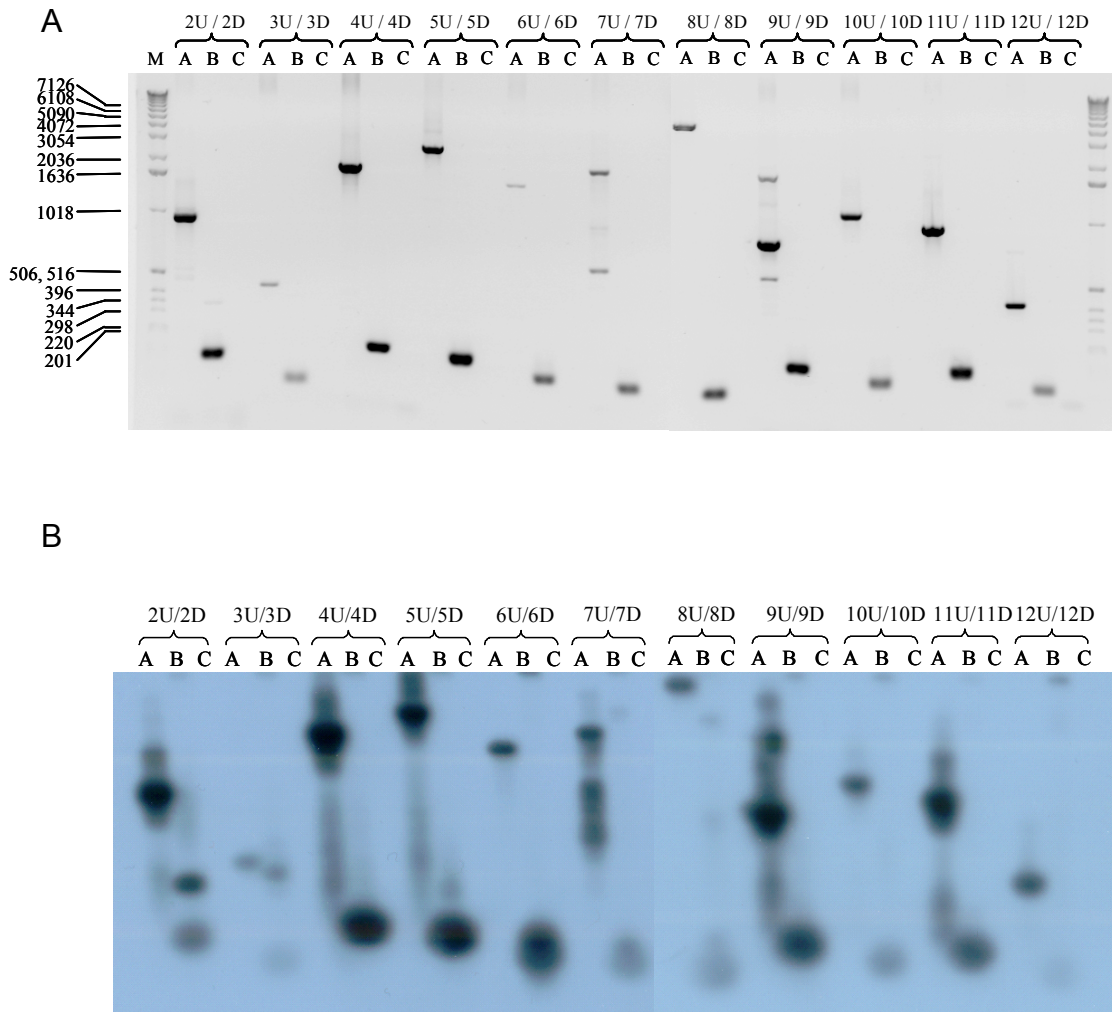
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SUPPLEMENTARY MATERIALS



	E1	I1	E2	I2	E3	I3	E4	I4	E5	I5	E6	I6	E7	I7	E8	I8	E9	I9	E10	I10	E11	I11	E12	I12	E13
<i>X. laevis</i>	37	>1400	55(<u>9</u>)	833	<u>88</u>	379	<u>95</u>	~1700	<u>114</u>	~2350	<u>91</u>	~1350	<u>80</u>	~1950	<u>57</u>	~4100 (5500)	<u>94</u>	~700	<u>96</u>	~1100	<u>59</u>	~900	<u>123</u>	338	112(<u>60</u>)
<i>X. tropicalis</i>	45	8373	57(<u>9</u>)	744	<u>88</u>	375	<u>95</u>	1414	<u>114</u>	1273	<u>91</u>	784	<u>80</u>	396	<u>57</u>	2757	<u>94</u>	558	<u>96</u>	1328	<u>59</u>	2840	<u>123</u>	398	114(<u>60</u>)
<i>H. sapiens</i>	26	ND	59(<u>9</u>)	6479	<u>88</u>	16391	<u>95</u>	1757	<u>114</u>	1417	<u>91</u>	2597	<u>80</u>	1979	<u>57</u>	3391	<u>94</u>	2411	<u>96</u>	583	<u>59</u>	1359	<u>123</u>	4633	943(<u>60</u>)
<i>M. musculus</i>	47	27640	49(<u>3</u>)	5004	<u>88</u>	2760	<u>95</u>	2850	<u>114</u>	959	<u>91</u>	1570	<u>80</u>	973	<u>57</u>	2080	<u>94</u>	4126	<u>96</u>	1447	<u>59</u>	1158	<u>123</u>	4011	911(<u>60</u>)
<i>D. rerio</i>	257	10490	58(<u>9</u>)	2941	<u>88</u>	2869	<u>95</u>	5479	<u>114</u>	3480	<u>91</u>	99	<u>80</u>	446	<u>57</u>	1645	<u>94</u>	2203	<u>96</u>	80	<u>59</u>	76	<u>123</u>	2360	219(<u>60</u>)

Supplementary Figure 1. Exon / intron organization of *anxa4* genes. Exons are represented by shaded boxes and introns by a horizontal line. The total size in base pairs is given in the corresponding column. Coding sequences are underlined. Exon 2 contains the start codon (atg) and exon 13 contains the stop codon (taa). Sizes of exons 1 and 13 were determined by reference to the cDNA sequences (See Table 1 for Accession numbers). Each exon is positioned relative to the *anxa4* protein domains. Figures in *italics* are generated from size estimates obtained by reference to PCR amplified bands to size markers; these introns have not been fully sequenced. *X.laevis* intron 1 was never amplified by PCR but was located in the 5'UTR using the Universal Genome Walker kit (CLONTECH laboratories, Inc). Its exact size could not be determined. Genomic sequences for the human, mouse and zebrafish *anxa4* genes were identified by BLAST on the NCBI website. The Genbank accession numbers for these sequences are: *H.sapiens*, NT_022184; *M.musculus*, NT_039353 and *D.rerio*, NW_001513125. *X.tropicalis anxa4* genomic sequences, located on the scaffold 143, were identified by BLAST on the JGI *Xenopus* website (<http://genome.jgi-psf.org/Xentr4/Xentr4.home.html>). ND, not determined.



Supplementary Figure 2. Identification of the *X. laevis anxa4* introns.

(A) PCR amplification of introns 2 – 12 of *anxa4* gene.

The putative intron boundaries of the *Xenopus laevis anxa4* gene were predicted by reference to the organization of mammalian *anxa4* genes. Primers were designed complementary to exon sequence adjacent to the putative intron boundaries (Supplementary Table 1). Each intron was amplified by PCR from *X. laevis* genomic DNA extracted from adult blood. Each amplification was optimised by gradient PCR by modifying the MgCl₂ concentration and the annealing temperature. The optimized conditions used for each intronic amplification, are indicated in Supplementary Table 1. PCR was carried out using the Taq polymerase from Invitrogen and Expand Long Template PCR polymerase (intron 7). For each primer pair, lane A shows the amplified genomic fragment, lane B shows the amplified cDNA fragment and lane C shows the no DNA input control. M, 1Kb DNA marker (Invitrogen).

(B) Southern blot analysis of the PCR amplified intronic sequences using *anxa4a* cDNA labelled with α^{32} P-dGTP. For each primer pair, lane A shows the amplified genomic fragment, lane B shows the amplified cDNA fragment and lane C shows the no DNA input control.

Intron	Sequence at exon-intron junction						Amino acid interrupted
	5' splice donor			3' splice acceptor			
2	<i>Xl</i>	ATG	GCA	GCA	gt aagatataaacct.....ttcctgttctat ag	CTC GGA ACT AAG	
	<i>Xt</i>	ATG	GCA	ACA	gt aagtatgaacct.....tttctgttctct ag	CTA GGA ACT AAG	
	<i>Hs</i>	ATG	GCC	ATG	gt aagttgtgaaat.....tcttgtgtgctc ag	GCA ACC AAA GGA	
	<i>Mm</i>			ATG	gt gagtgccctcgcc.....ttctgtgccttc ag	GAA GCC AAA GGA	
	<i>Dr</i>	ATG	GCA	CGG	gt aagtgaataaac.....tttttttactgt ag	TTG GGA AAC CGT	
3	<i>Xl</i>	ATG	AAA	GGA GCA G	gt gagtactagtagc.....cccctgtttct ag	GC ACC GAT GAA	Gly-33
	<i>Xt</i>	ATG	AAA	GGA GCA G	gt gagtactgatat.....gtgcccgtttct ag	GC ACC GAT GAA	Gly-33
	<i>Hs</i>	ATG	AAA	GGG CTC G	gt atgtgtcctgtct.....ccttcttcccc ag	GC ACC GAT GAA	Gly-33
	<i>Mm</i>	ATG	AAA	GGC CTC G	gt atgtactcttct.....catcttgtctcc ag	GT ACT GAT GAA	Gly-30
	<i>Dr</i>	ATG	AAA	GGG GCA G	gt gagagagacct.....aagctacttttc ag	GT ACC AAC GAG	Gly-33
4	<i>Xl</i>	ACT	GTT	GGA AAA	gt aagtcraagccaa.....attatcattttt ag	GAT CTT GAT GAT	
	<i>Xt</i>	ACT	ATT	GGA AAA	gt aagtcraaaccaa.....gttcttattttt ag	GAT CTG GAG GAT	
	<i>Hs</i>	ACC	ATC	GCC AGG	gt aggccaacagctct.....ttccttgcaattgc ag	GAC TTG ATA GAC	
	<i>Mm</i>	ACC	ATT	GCC AGG	gt agggcccagctct.....ctcgtttgtttgc ag	GAC CTG ATT GAG	
	<i>Dr</i>	AGC	GTG	GGA AAG	gt aaaacgtaaac.....aaacgctctttc ag	GAA TTG ATG GAC	
5	<i>Xl</i>	AAA	GCC	ATG AAG	gt gggtgttctgtg.....gctatggttttt ag	GGT GCA GGG ACA	
	<i>Xt</i>	AAA	GCC	ATG AAG	gt gggtgttctgaa.....gtttgtttttc ag	GGT GCA GGG ACA	
	<i>Hs</i>	AGG	GCC	ATG AAG	gt ctgtgtctcttcc.....ggtttcttgttt ag	GGA GCC GGC ACT	
	<i>Mm</i>	AGG	GCC	ATG AAG	gt ctgtgtgtccct.....gcctcaccattt ag	GGA GCT GGC ACG	
	<i>Dr</i>	AAC	GCC	ATC AAG	gt tagtgaggatag.....tattttgtgtct ag	GGA GCA GGG ACG	
6	<i>Xl</i>	ACC	TAC	AAA ATA A	gt aagtattataaa.....tttctgctttgc ag	AA TAT GGT AAA	Lys-133
	<i>Xt</i>	ACC	TAC	AGA ATA A	gt aagttagaaaaag.....tttttgcaattac ag	AA TAT GGT AAA	Lys-133
	<i>Hs</i>	ACC	TAC	CAG CAG C	gt acgtgacatccg.....cactctcttgc ag	AA TAT GGA CGG	Gln-133
	<i>Mm</i>	ACA	TAC	CAG CAG C	gt aagtagcagccc.....caattttttggt ag	AA TAT GGA AGG	Gln-131
	<i>Dr</i>	GCT	TAC	AAG AAA G	gt gcatgctagtta.....ccctcattactc ag	AA CAT GCA AAG	Glu-133
7	<i>Xl</i>	TCT	TTG	GCT GCT	gt aagtaccacaac.....ttgttcatttgc ag	GGT GGG AGA GAC	
	<i>Xt</i>	TCT	TTG	GCT GCT	gt aagtaccacaac.....gcattcctttgc ag	GGT GGG AGA GAC	
	<i>Hs</i>	TCT	CTG	TCA GCT	gt gagtgactgtct.....tttctcactcct ag	GGT GGG AGG GAT	
	<i>Mm</i>	TTC	CTG	TCG GCG	gt gctgtgtcttct.....ccatcttcccac ag	GCT GGC AGG GAT	
	<i>Dr</i>	TCA	TTG	CTC ACG	gt aaataaacaat.....tgctttgtttac ag	GCT GGG CGT GAC	
8	<i>Xl</i>	CAA	GAT	GCC AAT	gt aagtcagtctat.....cactttcttctc ag	GAC CTA TAT GAA	
	<i>Xt</i>	CAG	GAT	GCC AAT	gt aagtcagtctat.....tactttcttctc ag	GAA CTA TAT GAA	
	<i>Hs</i>	CAG	GAT	GCC CAG	gt tggtagaactgc.....ttgtttggttcc ag	GAC CTG TAT GAG	
	<i>Mm</i>	CAG	GAT	GCC CAG	gt ttgtggaccgca.....ttgtttgcatc ag	GAA CTG TAT GAG	
	<i>Dr</i>	CAG	GAC	GCA AAG	gt gaggattttgac.....ttatgctttatc ag	GAC ATC TAT GAG	
9	<i>Xl</i>	CAC	TTA	CTG AAA G	gt gagcatgtgaga.....gcttttttctc ag	TT TTT GAA GAG	Val-210
	<i>Xt</i>	CAC	TTG	CTG AAG G	gt gagcttatctcc.....ggttttattctc ag	TT TTT GAT GAA	Val-210
	<i>Hs</i>	CAC	CTG	TTG CAT G	gt aaggcacttact.....gttattgtctgc ag	TG TTT GAT GAA	Val-210
	<i>Mm</i>	CAT	CTG	CTC CAC G	gt gggtagcccacct.....gttattgtctac ag	TG TTT GAT GAA	Val-208
	<i>Dr</i>	GCC	CTT	CTG CGA G	gt aaacacagtaaa.....ctgtttttccc ag	TG TTT CAG GAG	Val-210
10	<i>Xl</i>	CTT	TTG	GCA ATC G	gt aagtatgtcagg.....tctttgccttgc ag	TG AAA TGC ATA	Val-242
	<i>Xt</i>	CTT	TTG	GCA GTC G	gt gagtagcttagg.....tctttgccttgc ag	TG AAA TGC TTG	Val-242
	<i>Hs</i>	CTG	CTG	GCT ATA G	gt aagctggttaggg.....ttttctttgac ag	TA AAG TGC ATG	Val-242
	<i>Mm</i>	CTG	TTG	GCT ATA G	gt aagctggtgagc.....cgttttctccac ag	TG AAG TGC ATG	Val-240
	<i>Dr</i>	TTT	CTG	GCA ATA G	gt cagcgttagtgc.....gtttgccttttc ag	TG AAA GTC ATA	Val-242
11	<i>Xl</i>	AAA	TCT	ATG AAG	gt gagtgctttttt.....cttctgttttgc ag	GGT CTG GGG ACA	
	<i>Xt</i>	AAA	TCT	ATG AAG	gt gagccctttttt.....cttctgttttgc ag	GGT CTG GGG ACA	
	<i>Hs</i>	AAA	TCG	ATG AAG	gt aatggcctttat.....gttttcttctt ag	GGC TTG GGC ACC	
	<i>Mm</i>	AAA	TCC	ATG AAG	gt aagcagctgtgc.....actccttctct ag	GGC TTA GGC ACT	
	<i>Dr</i>	AAG	TCA	ATG AAG	gt acacaaaatcac.....atatcatttctt ag	GGT TTA GGG ACT	
12	<i>Xl</i>	TCA	TTT	ATT AAG	gt gagagaagaatt.....tttgtttccgc ag	GGT GAC TGC TCG	
	<i>Xt</i>	TCC	TTT	ATT AAG	gt aagagaggaatt.....ttttgtttctgc ag	GGT GAC TGC TCT	
	<i>Hs</i>	TCG	TTC	ATC AAG	gt aggtcacagcag.....tccctatcgaa ag	GGT GAC ACA TCT	
	<i>Mm</i>	TCT	TTC	ATC AAG	gt aggaacatcatc.....ctcctttctcc ag	GGT GAC ACT TCC	
	<i>Dr</i>	TCC	TTT	ATA AAG	gt gagacattacac.....ttgtctttttc ag	GGC GAC ACG TCG	

Supplementary Figure 3: Exon / intron splice junctions of *anxa4* gene sequences.

Each intronic amplification was ligated into pGEMt-easy as recommended by the manufacturer (Promega, UK) and confirmed as *anxa4* genomic sequences by in-house sequencing. Intronic boundaries were identified by alignment between the genomic and cDNA sequences and according to the intron/exon boundary consensus sequences (Breathnach and Chambon, 1981). Sequences at the exon/intron boundaries from different species were compared by alignment. Shading represents conservation between species at this base in two or more of the presented intronic sequences. Intronic boundaries **gt...ag** are shown in bold. *Xl*, *Xenopus laevis*; *Xt*, *Xenopus tropicalis*; *Hs*, *Homo sapiens*; *Mm*, *Mus musculus*; *Dr*, *Danio rerio*.

Breathnach, R. and Chambon, P. Organization and expression of eucaryotic split genes coding for proteins. *Annu Rev Biochem* 1981; 50:349-83.

Supplementary Table 1. Primers designed to amplify up *X. laevis anxa4* introns

Intron	Sequence	Optimum [MgCl₂]	Optimum Temperature (°C)	Cycles	
2	U-CAGCCGACATCTGCTTAGAA D-CCTTTCATGGCGTTCCTCAG	3mM	60	30	This work
3	U-CTGAGGAACGCCATGAAAGG D-GCAATGACGTCAATGAC	7.5mM	60	30	This work
4	U-GTCATTGACGTCATTGC D-GTTCCTCCACATCATAGAGA	5mM	60	30	This work
5	U-CCGAACAGCAGGGAACCTT D-TCTCCTCTGCGCTGCGAGAT	3mM	60	30	This work
6	U-TCTCGCAGCGCAGAGGAGAT D-CTCTCTGGAACATGAAAGAC	5mM	60	30	This work
7	U-GGAGGACGATATTTGCTCAG D-TCATTCACGGTGCTGCTCTG	1.5mM	60	30	This work Seville et al. (2002)
9	U-CAGAGCAGCACCGTGAATGA D-CCACTTCTTCTCACC GGCTT	4mM	60	30	This work
10	U-AAGCCGGTGAGAAGAAGTGG D-GTGTCCCAGACATTTCAAGATT	3mM	60	30	This work
11	U-AGATCTTGAGGCCAGTATAA D-CAATCGTTCTGCAAAGTAGG	5mM	60	30	This work
12	U-AAGAGCAGGCCAGCCTACTT D-TCTTGAACTCGCAGCGGATT	1.5mM	60	30	This work
13	U-TCCGCTGCGAGTTCAAGAAG D-GAGCACCTTCCTGTAATCTC	3mM	60	30	This work