

<u>Supplemental Figure S1</u> -

Fig. S1. Distribution of AChRs in the absence of the γ **-subunit**. (**A**) Whole-mount diaphragm muscles from E15.5 wild-type and $\gamma^{-/-}$ embryos were incubated with Texas Red-conjugated a-bungarotoxin (a-bgt). Images were collected with a low-power objective (10×) so the entire length of muscle fibers could be viewed under the same field. The a-bgt labeled AChRs were clustered in the central region of the wild-type muscle, but remained diffuse in the $\gamma^{-/-}$ muscle. Despite the diffused pattern, the central region (marked by the broken lines) of the $\gamma^{-/-}$ muscle was still more intensely labeled, compared with its flanking region. (**B**) The difference in fluorescence intensity was presented in a line-scan. The entire muscle fiber was manually traced from one end to the other (~800 µm) using NIH ImageJ software. The gray levels of each pixel were measured and plotted against its position along the muscle fiber (wild-type, black line; $\gamma^{-/-}$, grey line). The central region of the $\gamma^{-/-}$ muscle appeared more intensely labeled than its flanking region.

• Supplemental Figure S2 -

Fig. S2. Innervation pattern of hind limb muscles in wild-type and $\gamma < b > - / < b > - mice. ($ **A**,**B** $) Cross-section of E18.5 hind limb, stained with Hematoxylin and Eosin (H&E); note the apparent atrophy of the <math>\gamma^{-/-}$ muscles (B). (**C**,**D**) Low-magnification view of hind limb sections immunostained with synaptotagmin 2 (Syt2) antibody. Note a dramatic increase of innervation in all muscles in the $\gamma^{-/-}$ embryos (D), when compared with the wild-type muscles (C). (**E-H**) Higher magnification views of EDL and soleus are shown in the bottom row: nerve terminals in the wild-type muscles localized to small regions of the muscle (arrowheads in E,F), whereas nerve terminals in the $\gamma^{-/-}$ muscles were broadly distributed (arrows in G,H). TA, tibias anterior muscle; EDL, extensor digitorum longus muscle; MG, medial gastrocnemius muscle; LG, lateral gastrocnemius muscle; Sol, soleus; F, fibula. Scale bars: 200 µm in A-D; 100µm in E-H.

• Supplemental Figure S3 -

Fig. S3. Differentiation of the presynaptic nerve terminals at E16.5. Whole-mount diaphragm muscles labeled with synaptotagmin 2 (Syt2) antibody (A,D) and Texas-Red conjugated a-bungarotoxin (B,E). AChR clusters in the $\gamma^{-/-}$ muscle (arrowheads in E) were markedly reduced, compared with those

in the wild-type muscles (E). However, individual nerve terminals were intensely labeled by Syt2 antibody in both wild-type (arrowheads in A) and in $\gamma^{-/-}$ muscles (arrowheads in D), although the nerve terminals in $\gamma^{-/-}$ muscles were distributed across a much broader region. The majority of the nerve terminals in the wild-type muscle were closely apposed by AChR clusters (arrowheads in C), whereas the majority of the nerve terminals in the $\gamma^{-/-}$ muscles in the $\gamma^{-/-}$ muscles were not apposed by AChR clusters (arrows in F). Note, however, that aneural AChR clusters were still present in the wild-type muscles (arrows in B,C). Scale bar: 30 µm

• Supplemental Figure S4 -

Fig. S4. Low magnification views of whole-mount diaphragm muscles immunolabeled by synaptotagmin 2 antibody. In wild-type muscle, nerve terminals (arrowhead in A) were localized along the central region of the muscle; occasionally, some nerve sprouts (arrows in A) extended beyond the central region. In the $\gamma^{-/-}$ muscle, nerve terminals (arrowheads in B) were distributed across a broad region of the muscle, especially at the dorsal quadrant of the diaphragm. Excessive nerve sprouts were seen across the entire region (arrows in B). Scale bar: 500 µm.

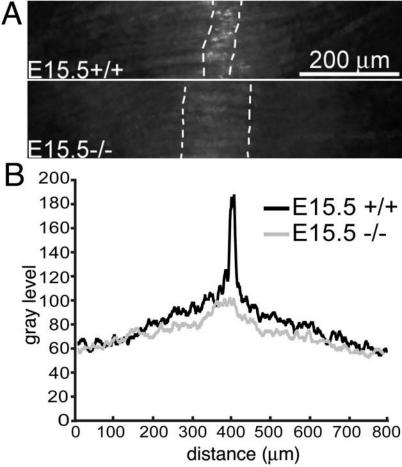
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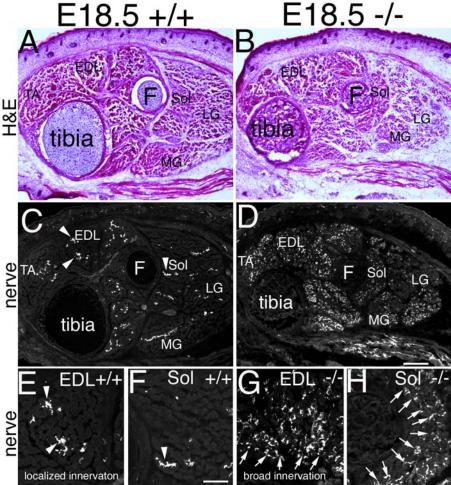
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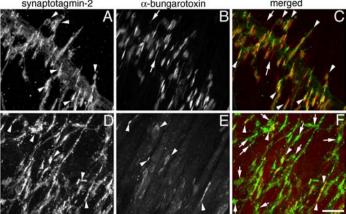
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