

Table S4. Bacterial strains and plasmids used in this study

Strains	Genotype or markers; characteristics and uses	Reference or source
<i>Shewanella oneidensis</i>		
MR-1	Dissimilatory metal-reducing strain (Lake Oneida, NY)	Myers and Nealson (1988) <i>Science</i> 240:1319-1321.
$\Delta ldhA$	SO0968 (<i>ldhA</i>) deletion derivative of <i>S. oneidensis</i> MR-1	This study
$\Delta dld-II$	SO1521 (<i>lldD</i>) deletion derivative of <i>S. oneidensis</i> MR-1	This study
$\Delta lldE$	SO1520 (<i>lldE</i>) deletion derivative of <i>S. oneidensis</i> MR-1	This study
$\Delta lldF$	SO1519 (<i>lldF</i>) deletion derivative of <i>S. oneidensis</i> MR-1	This study
$\Delta lldG$	SO1518 (<i>lldG</i>) deletion derivative of <i>S. oneidensis</i> MR-1	This study
$\Delta dld-II\Delta lldF$	SO1519 (<i>lldF</i>)/SO1521 (<i>lldD</i>) double deletion derivative of <i>S. oneidensis</i> MR-1	This study
<i>Escherichia coli</i>		
BL21/DE3	Host used for purification of <i>S. oneidensis</i> MR-1 LdhA (SO0968)	Stratagene, La Jolla, CA
DH10B (GeneHog)	Host used for overexpression of <i>E. coli</i> K12 <i>ykgEFG</i> and <i>S. oneidensis</i> MR-1 <i>dld-II</i> and <i>lldEFG</i> genes	Invitrogen, Carlsbad, CA
Top10	Host used for construction of pBBR1MCS-5 clones. Invitrogen F- <i>mcrA</i> $\Delta(mrr-hsdRMS-mcrBC)$ $\phi 80lacZ\Delta M15$ $\Delta lacX74$ <i>recA1</i> <i>araD139</i> $\Delta(ara-leu)7697$ <i>galU</i> <i>galK</i> <i>rpsL</i> (<i>Str^r</i>) <i>endA1</i> <i>nupG</i>	Invitrogen, Carlsbad, CA
EC100D <i>pir</i> -116	Host used for construction of pDS3.1 clones; F ⁻ <i>mcrA</i> $\Delta(mrr-hsdRMS-mcrBC)$ $\phi 80lacZ\Delta M15$ $\Delta lacX74$ <i>recA1</i> <i>endA1</i> <i>araD139</i> $\Delta(ara, leu)7697$ <i>galU</i> <i>galK</i> λ^{-} <i>rpsL</i> <i>nupG</i> <i>pir</i> -116(DHFR)	Epicenter Biotechnologies, Madison, WI
B2155 Ipir	Donor strain used for conjugation with <i>S. oneidensis</i> MR-1; <i>thrB1004 pro thi strA hsdS lacZ</i> $\Delta M15$ (F9 <i>lacZ</i> $\Delta M15$ <i>lacIq</i> <i>traD36 proA1 proB1</i>) $\Delta dapA::erm$ (Erm ^r) <i>pir</i> ::RP4 (::kan (Km ^r) from SM10)	Dehio and Meyer (1997) <i>J Bacteriol</i> 179:538-540.
WM3064	Donor strain used for conjugation with <i>S. oneidensis</i> MR-1; <i>thrB1004 pro thi rpsL hsdS lacZ</i> $\Delta M15$ RP4-1360 (<i>araBAD</i>)567 <i>dapA1341::[erm pir(wt)]</i>	Saltikov, <i>et al.</i> (2003) <i>Appl Environ Microbiol</i> 69: 2800-2809.
$\Delta lldD$	Deletion derivative of <i>E. coli</i> K-12; Db3605 (<i>lldD</i>), used for complementation analysis	Baba and Mori (2008) <i>Methods Mol Biol</i> 416:171-81.
Δdld	Deletion derivative of <i>E. coli</i> K-12; $\Delta b2133$ (Δdld), used for complementation analysis	Baba and Mori (2008) <i>Methods Mol Biol</i> 416:171-81.
<i>Bacillus subtilis</i>		
168 Marburg	Type strain, <i>trpC2</i>	Culture collection of Japanese and European <i>B. subtilis</i> Functional Analysis Programs

<i>yvfV</i> ::pMUTIN2	<i>trpC2</i> , pMUTIN2 insertion in <i>yvfV</i> (<i>Bsu3402</i>), <i>Em^r</i>	Culture collection of Japanese and European <i>B. subtilis</i> Functional Analysis Programs
<i>yvfW</i> ::pMUTIN2	<i>trpC2</i> , pMUTIN2 insertion in <i>yvfW</i> (<i>Bsu3401</i>), <i>Em^r</i>	Culture collection of Japanese and European <i>B. subtilis</i> Functional Analysis Programs
<i>yvbY</i> ::pMUTIN2	<i>trpC2</i> , pMUTIN2 insertion in <i>yvbY</i> (<i>Bsu3401</i>), <i>Em^r</i>	Culture collection of Japanese and European <i>B. subtilis</i> Functional Analysis Programs
Plasmids		
pBAD-TOPO	4.1-kb vector used for gene expression in <i>E. coli</i> ; <i>Amp^r</i> , <i>araC</i>	Invitrogen, Carlsbad, CA
pDS3.0	Suicide vector derived from pCDV224; <i>Amp^r</i> , <i>Gm^r</i> , <i>sacB</i>	Wan, <i>et al.</i> (2004) <i>J Bacteriol</i> 186:8385-8400.
pBBR1MCS-5	5.1-kb broad-host range plasmid; <i>Gm^r</i> , <i>lacZ</i>	Kovach, <i>et al.</i> (1994) <i>BioTechniques</i> 16, 800–802.
pMCSG7	Vector used for cloning <i>ldhA</i>	Stols, <i>et al.</i> (2002) <i>Protein Expr Purif</i> 25, 8-15.
pBAD (SO1521)	SO1521 gene was cloned into pBAD-TOPO	This study
pBAD (SO1518-19-20)	2.7-kb PCR fragment containing <i>lldEFG</i> operon was cloned into pBAD-TOPO	This study
pBAD (SO1519-20)	2.1-kb PCR fragment containing <i>lldEF</i> genes was cloned into pBAD-TOPO	This study
pBAD (SO1520)	SO1520 gene was cloned into pBAD-TOPO	This study
pBAD (YkgEFG)	2.8-kb PCR fragment containing <i>E. coli</i> K-12 <i>ykgEFG</i> genes (b0306-8) was cloned into pBAD-TOPO	This study
pBBR <i>lldEFG</i>	<i>lldEFG</i> 2.7 kb <i>Bam</i> HI- <i>Pme</i> I restriction fragment of pBAD- <i>lldEFG</i> cloned in the <i>Bam</i> HI- <i>Pvu</i> II site of pBBR1MCS-5	This study
pBBR <i>dld-II</i>	<i>dld-II</i> PCR product cloned into the x site of pBBR1MCS-5	This study
pDSD <i>ldhA</i>	2-kb fusion PCR fragment encoding DNA flanking <i>ldhA</i> cloned in pDS3.0; used to delete <i>ldhA</i> from the MR-1 genome	This study
pDSD <i>dld-II</i>	2-kb fusion PCR fragment encoding DNA flanking <i>dld-II</i> cloned in pDS3.0; used to delete <i>dld-II</i> from the MR-1 genome	This study
pDSD <i>lldE</i>	2-kb fusion PCR fragment encoding DNA flanking <i>lldE</i> cloned in pDS3.0; used to delete <i>lldE</i> from the MR-1 genome	This study
pDSD <i>lldF</i>	2-kb fusion PCR fragment encoding DNA flanking <i>lldF</i> cloned in pDS3.0; used to delete <i>lldF</i> from the MR-1 genome	This study
pDSD <i>lldG</i>	2-kb fusion PCR fragment encoding DNA flanking <i>lldG</i> cloned in pDS3.0; used to delete <i>lldG</i> from the MR-1 genome	This study
