

Supporting Information

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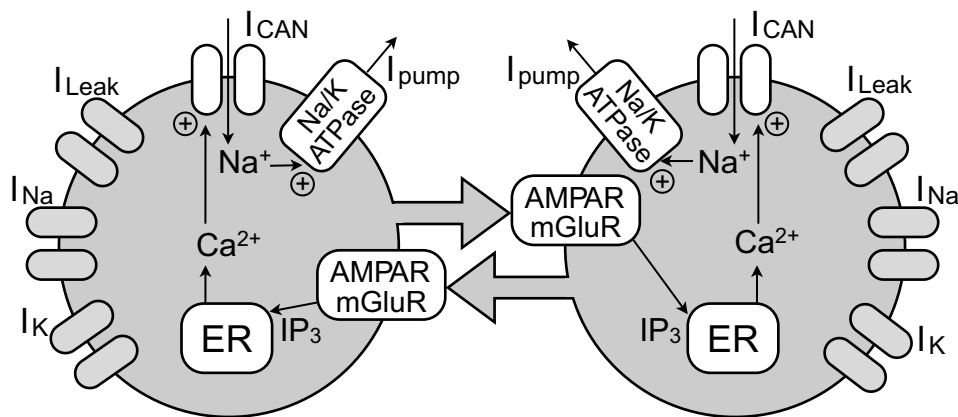


Fig. S1. Schematic diagram of the model. Reciprocally coupled neurons motivated by respiratory neurons of the mammalian preBötC were equipped with Hodgkin-Huxley-style “fast” Na^+ and delayed-rectifier K^+ currents (I_{Na} , I_{K}), and a K^+ -dominated leakage current (I_{Leak}). I_{Na} , I_{K} , and I_{Leak} are shaded gray to indicate that their role is mostly in setting resting potential and generating spikes. As discussed in the text, the bursting dynamics in the model depend on a Ca^{2+} -activated nonspecific cationic current (I_{CAN}), material balance equations that determine the intracellular concentrations of Ca^{2+} and Na^+ , and some type of activity-dependent net outward current (Figs. 3 and 4 and Figs. S3 and S4). In the basic model illustrated, the net outward current is mediated by an Na/K ATPase pump (I_{pump}), as analyzed in Figs. 3, S3A, and S4A. Other net outward currents included a voltage-dependent M-like K^+ current (I_{M} , Fig. 4A and Figs. S3B and S4B), Ca^{2+} -dependent K^+ current ($I_{\text{K-Ca}}$, Fig. 4B and Figs. S3C and S4C), and voltage-dependent inactivation of inward persistent Na^+ current (I_{NaP} , Fig. 4C and Figs. S3D and S4D).

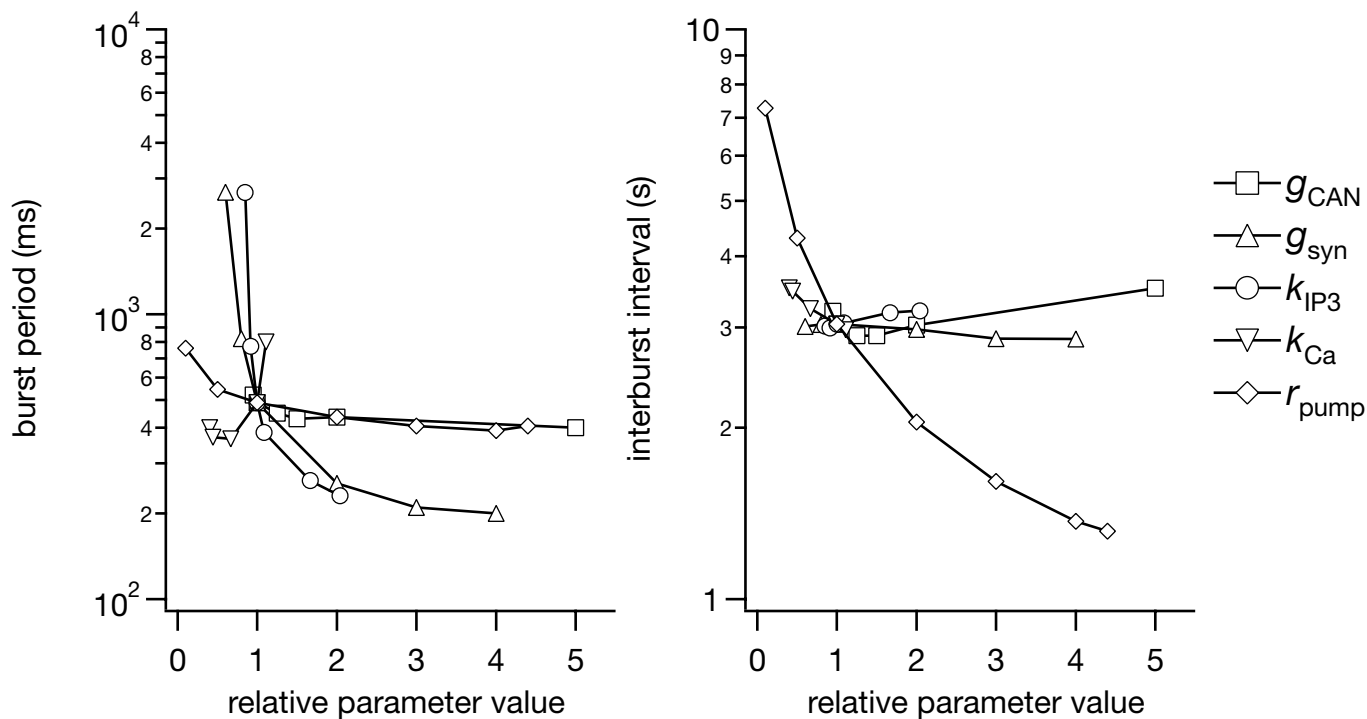


Fig. 52. Robustness parameter study in the self-coupled model with I_{pump} . Burst period (Left) and interburst interval (Right), plotted over the range of each parameter for which the self-coupled neuron generated rhythmic bursts. Such bursts were defined as trajectories analogous to Fig. 3D, with a pair of AH crossings followed by a pair of SNIC crossings in the $(Ca^{2+}-Na^+)$ plane in each period. Each parameter value was normalized relative to its default value (see Derivation of the model, SI Appendix). The parameters varied include: g_{CAN} (squares), g_{syn} (upward triangles), k_{IP3} (circles), k_{Ca} (downward triangles), and r_{pump} (diamonds).

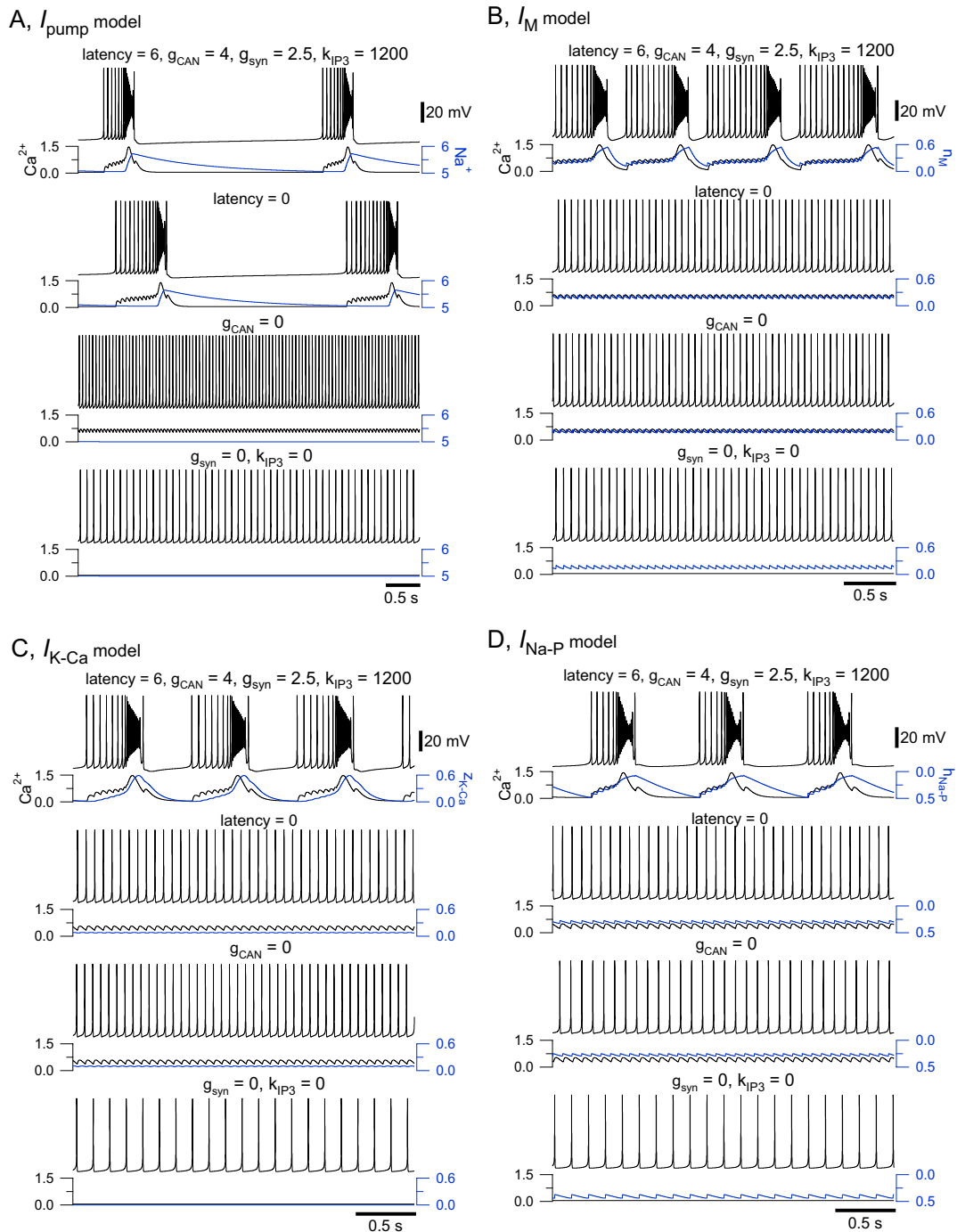


Fig. S3. Simulations of the self-coupled model using different net outward currents, showing that the parameters g_{CAN} , g_{syn} , and k_{IP3} and the currents they influence are critical, regardless of which activity-dependent net outward current is used in the model. (A–D) Sets of 4 time courses of model outputs. Each sweep consists of 3 traces: membrane potential (*Upper*, black), Ca^{2+} (*Lower*, black), and the model-specific slow variable (*Lower*, blue, superimposed with Ca^{2+} dynamics). The upper sweep in each panel serves as control and lists default standard parameter values for latency, g_{CAN} , g_{syn} , and k_{IP3} . Subsequent (*Lower*) sweeps in each panel each show results given by varying one or more of the parameters from the control condition. (A) I_{pump} model with Na^{+} as the slow variable. (B) I_{M} model with n_{M} as the slow variable. (C) $I_{\text{K-Ca}}$ model with $z_{\text{K-Ca}}$ as the slow variable. (D) $I_{\text{Na-P}}$ model with $h_{\text{Na-P}}$ as the slow variable.

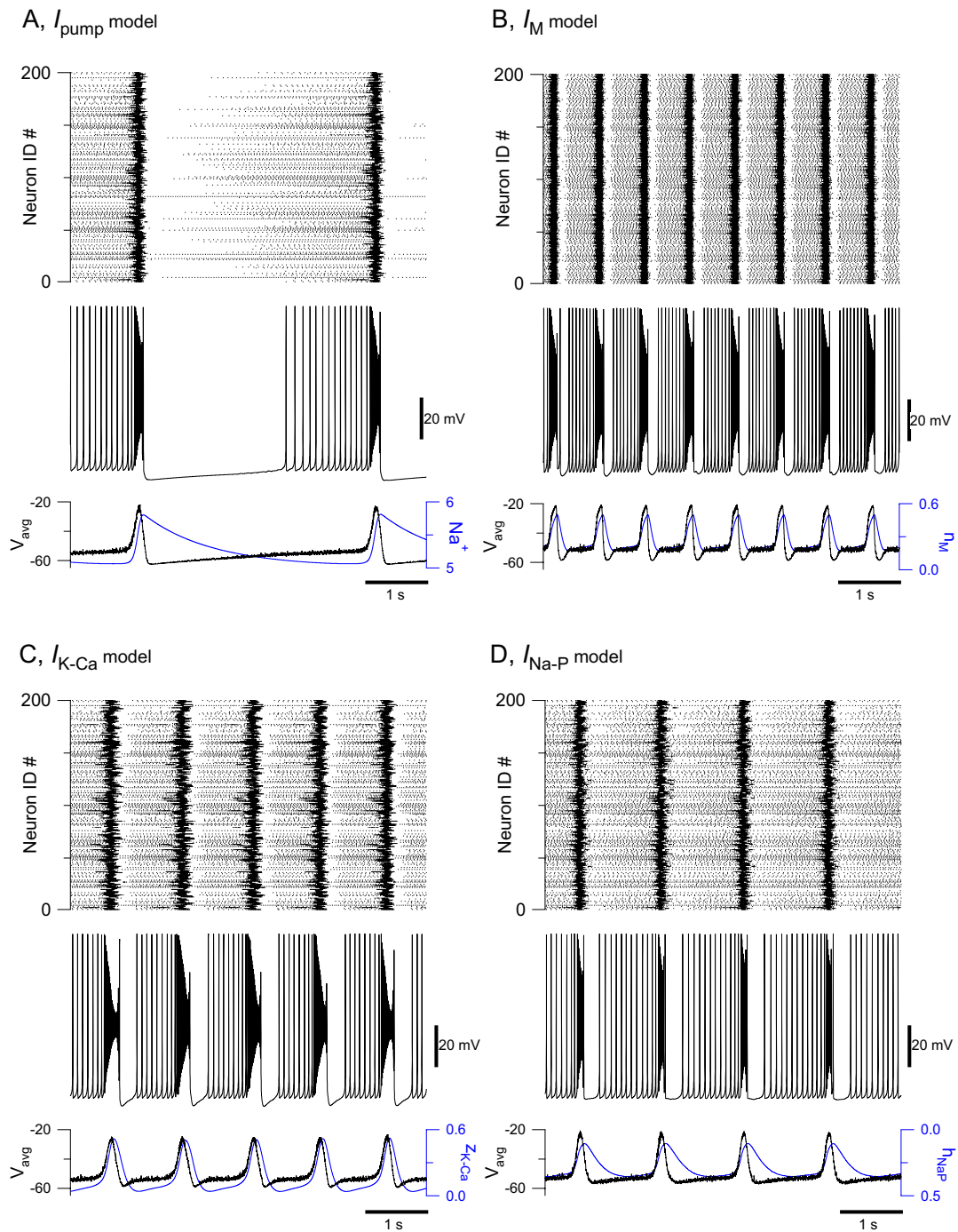


Fig. S4. Rhythmic burst activity in large-scale network models. Each panel consists of a raster diagram (*Top*) showing the spike times of 200 simulated neurons, the voltage trajectory of a representative neuron (*Middle*), the average membrane voltage for all neurons in the network (*Bottom*, black) along with the average slow variable (blue). (*A–D*) Network activity with different activity-dependent outward currents including I_{pump} , where average Na^+ concentration is the slow variable (*A*), M -like K^+ current (I_M), where average n_M is the slow variable (*B*), Ca^{2+} -dependent K^+ current ($I_{\text{K-Ca}}$), where average $z_{\text{K-Ca}}$ is the slow variable (*C*), and persistent Na^+ current ($I_{\text{Na-P}}$), where average $h_{\text{Na-P}}$ is the slow variable (*D*).

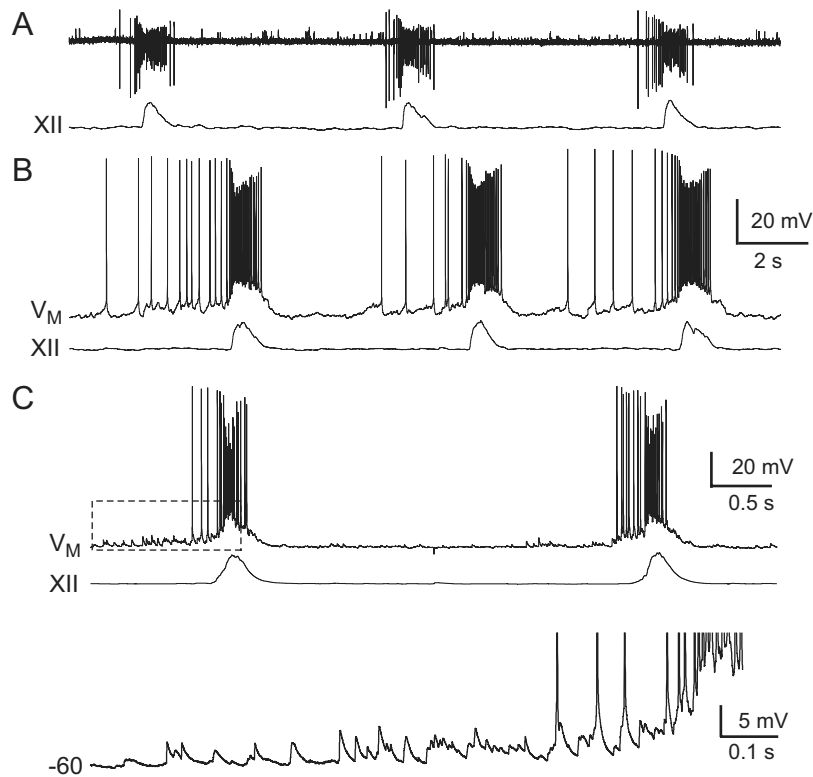


Fig. S5. Respiratory rhythm generation in vitro. (A) On-cell patch recording of a preBötC neuron (*Upper*) with inspiratory motor discharge recorded from the hypoglossal (XII) motor nerve root (*Lower*). (B) Whole-cell recording conditions in the same cell as A (*Upper*) with XII motor output (*Lower*). Baseline membrane potential was -60 mV. (C) Two consecutive cycles in a representative preBötC neuron (top trace) with XII motor output (lower trace). Ramp-like depolarization and EPSPs occur 300–1000 ms before the inspiratory phase; spiking occurs in the final 300 ms before the inspiratory phase. Voltage-dependent spike inactivation, i.e., depolarization block, is visible during the inspiratory bursts. Temporal summation of EPSPs and low-rate (≈ 5 – 10 Hz) spiking activity in the preBötC shows evidence for recurrent synaptic excitation during respiratory rhythmogenesis. Broken lines indicate the region of the recording that is expanded in the *Inset*. (*Inset*) Illustrates temporal summation of EPSPs and preinspiratory spiking in greater detail.

Other Supporting Information Files

[SI Appendix](#)