Supporting Information

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Fig. S1. Characterization of thiocillins by NMR. ¹H-NMR spectra of thiocillin I (compound **3**) (*A*) and YM-266183 [compound **6**, also known as QN3323A (1)] (*B*) are shown. Compounds were dissolved in DMSO- d_6 , and data were collected on a Varian 400-MHz NMR spectrometer. Spectrum *A* contains minor peaks that are consistent with the presence of QN3323Y1 (1), which differs from 3 only in the olefin geometry at Dhb4, as a minor product.

^{1.} Kamigiri K, et al. (2006) US Patent 7030085 (April 18, 2006).

Table S1. Primers used in this study

Primer	Sequence	Comments
LCW001	5'-gtacggatcccatactcccagattatttgcagtcgtaac-3'	BamHI (LF-5')
LCW002	5'-gtacgtcgacgagaggagtatacatatgcttactg-3'	Sall (LF-3')
LCW010	5'-gtacggatccgaaataaactctatttcagaaattgag-3'	BamHI (cam1–5')
LCW011	5'-gtac <u>ctcgag</u> gtccgattgttcggacacacctaagc-3'	Xhol (cam1–3')
LCW012	5'-gtacggatccgccaactcccttcagttcatttacagg-3'	BamHI (cam2–5')
LCW013	5'-gtac <u>ctcgag</u> cttactcttcaataaatcagggaacttg-3'	XhoI (cam2–3')
LCW014	5'-gtacggatccggacgattatgaaagtcctttcgcag-3'	BamHI (cam3–5')
LCW015	5'-gtac <u>ctcgag</u> ccatcgtccgtacaagtgaataattc-3'	Xhol (cam3–3')
LCW028	5'-gctgaattcataagcttatactagtgtc-3'	IMX_check1_3′
LCW029	5'-gtgtagcatgtctcattcaattttgagg-3'	IMX_check2_5′
LCW032	5'-gtgatataacatgaaaccaataaactatg-3'	IM2_check1_5′
LCW033	5'-ctacaaggtgtgagaccgatttttcaac-3'	IM2_check2_3′
LCW034	5'-catgaataactataccgttatgaaatcc-3'	IM3_check1_5′
LCW035	5'-cacctacaagaaatttttggaaactaac-3'	IM3_check2_3′
LCW040	5'-gttagaagactatgatgtaccgtggatc-3'	IM4_check1_5′
LCW041	5'-ccgcggtttgatcgactacgatgatatc-3'	IM4_check2_3′
LCW042	5'-caaagcacgctacatctgatgagagaaac-3'	IM1_check1a_5′
LCW043	5'-gtgcttacctaatcctggccctatctta-3'	IM1_check2a_3′

Restriction sites are underlined.

Table S2. Plasmids used in this study

Plasmid	Comments	Source
pKM082	Carries MLS resistance gene.	David Rudner, Harvard Medical School
pLW106	Derived from pKM082. Transformed into <i>B. cereus</i> ATCC14579 to generate IM1. (pKM082 + cam2)	This study
pLW105	Derived from pKM082. Transformed into <i>B. cereus</i> ATCC14579 to generate IM2. (pKM082 + cam1)	This study
pLW111	Derived from pKM082. Transformed into <i>B. cereus</i> ATCC14579 to generate IM3. (pKM082 + LF)	This study
pLW107	Derived from pKM082. Transformed into <i>B. cereus</i> ATCC14579 to generate IM4. (pKM082 + cam3)	This study

Table S3. Strains used in this study

Comments	Source	
Thiocillin producer	ATCC	
Derivative of <i>B. cereus</i> ATCC 14579 harboring pLW106 (MLS resistance)	This study	
Derivative of <i>B. cereus</i> ATCC 14579 harboring pLW105 (MLS resistance)	This study	
Derivative of <i>B. cereus</i> ATCC 14579 harboring pLW111 (MLS resistance)	This study	
Derivative of <i>B. cereus</i> ATCC 14579 harboring pLW107 (MLS resistance)	This study	
	Comments Thiocillin producer Derivative of <i>B. cereus</i> ATCC 14579 harboring pLW106 (MLS resistance) Derivative of <i>B. cereus</i> ATCC 14579 harboring pLW105 (MLS resistance) Derivative of <i>B. cereus</i> ATCC 14579 harboring pLW111 (MLS resistance) Derivative of <i>B. cereus</i> ATCC 14579 harboring pLW107 (MLS resistance)	

δ , ppm	Description	Integration	J, Hz
9.65	S	1	
9.52	S	1	
8.58	S	1	
8.46-8.30	m	7	
8.25	S	1	
7.99	S	1	
7.92-7.90	m	1	
7.55	d	1	7.8
6.52-6.47	m	2	
5.48	d	1	10.2
5.19	S	1	
5.05-4.97	m	2	
4.72-4.58	m	3	
4.50-4.48	m	1	
3.98-3.90	m	1	
3.75-3.65	m	1	
3.07-3.03	m	2	
2.07	S	3	
1.73–1.71	m	6	
1.37	m	3	
1.24–1.20	m	6	
1.01	d	3	6.0

Table S4. 400-MHz	¹ H NMR data	of 3 in	d ₆ -DMSO	at 25	°C
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s, singlet; d, doublet; t, triplet; q, quartet; br, broad; m, multiplet.

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δ, ppm	Description	Integration	J, Hz
9.65	S	1	
9.60	S	1	
8.58	S	1	
8.46-8.32	m	8	
8.25	S	1	
7.99	S	1	
7.55	d	1	8.0
6.58–6.55	m	1	
6.50-6.47	m	1	
5.48-5.44	m	1	
5.19	S	1	
5.04-4.98	m	2	
4.70-4.67	m	2	
4.51–4.98	m	1	
3.96–3.91	m	1	
3.88–3.86	m	2	
2.07	S	3	
1.73–1.71	m	6	
1.37	m	3	
1.24–1.20	m	6	
1.01	d	3	6.0

Table S5. 400-MHz ¹H NMR data of 6 in d_{6} -DMSO at 25 °C

s, singlet; d, doublet; t, triplet; q, quartet; br, broad; m, multiplet.