

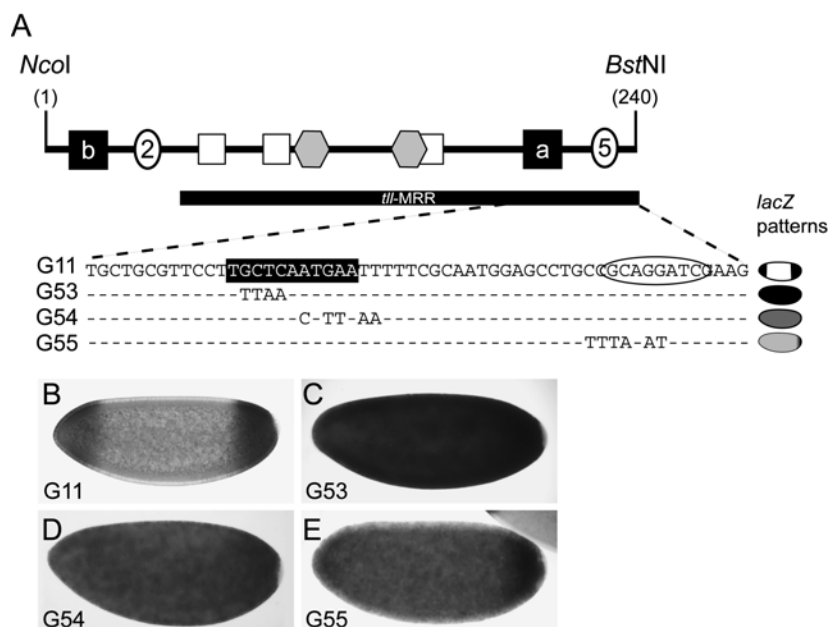
## Supplementary figures

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mel TGCTCAATGAAtttttcgcaatggcagc
sim TGCTCAATGAAtttttcgcaatggcagc
yak TGCTCAATGAAtttttcgcaatggcagc
ana TGCTCAATGAA-ttttcgcaatggcagc
ere TGCTCAATGAA-ttttcgcaatggcagc
pse TGCTCAATGAA-ttttcgcaatggcagc
vir TGCTCAATGAA-ttttcgcaatggcagc
moj TGCTCAATGAAtttttcgcaatggcagc
gri TGCTCAATGAA-ttttcgcaatggcagc
Hsf BS      GAA-nnTTCnnGAA
  
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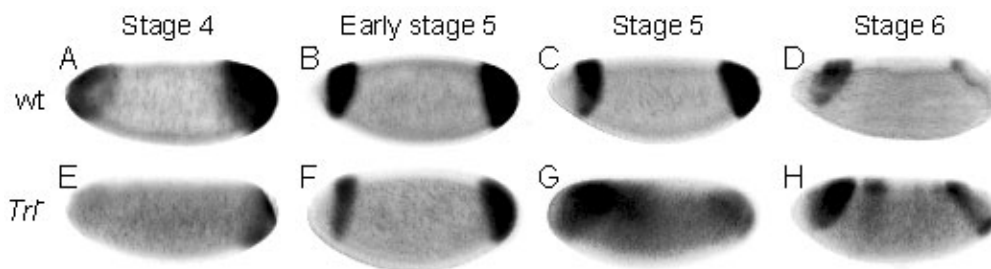
**Figure S1. DNA sequences in the *tor*-RE and downstream region are highly conserved among nine *Drosophila* species.**

DNA sequences of the *tor*-RE (upper case) and downstream region (lower case) in *Drosophila melanogaster* (*mel*), *simulans* (*sim*), *yakuba* (*yak*), *ananassae* (*ana*), *erecta* (*ere*), *pseudoobscura* (*pse*), *virilis* (*vir*), *mojavensis* (*moj*) and *grimshawi* (*gri*) were retrieved from the UCSC Genome Bioinformatics (<http://genome.ucsc.edu/>). The consensus sequence bound by Hsf trimer (Hsf BS) is shown below the multiple-aligned sequences. The TGAG sequences in the reverse strand of the *tor*-RE are shaded. Two GAAs in an inverted-repeat fashion at the 3' end and flanking region of the *tor*-RE are boxed. The spacing between the GAA repeats is 3 bp in *D. mel*, *sim*, *yak* and *moj*, and is 2 bp in the remaining five species.



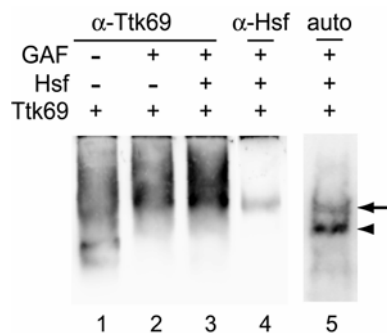
**Figure S2. Two different factors may bind to the *tor*-RE.**

(A) The diagram shows the locations of the *tor*-REs (solid squares), GAF (open squares) and Ttk69 (open ovals) binding sites in a 240-bp *tll* cis-regulatory region from *Nco*I to *Bst*NI. The DNA sequence of the 3' end of *tll* minimal regulatory region (*tll*-MRR; 186 bp in length) is shown (1). Shaded hexagons represent two putative Hsf binding sites in addition to the *tor*-RE. DNA sequence in 3' portion of the *tll*-MRR is shown below the diagram. Closed rectangle and open oval delimitate sequences for the *tor*-RE and the Ttk69 binding site, TC5, respectively. DNA sequences with or without base substitutions are indicated by letters and “-”, respectively. *lacZ* expression patterns were determined by in situ hybridization with a digoxigenin-labeled RNA as a probe. The cartoons at the right summarize the patterns driven by wild-type (G11, B) or three different mutated *tll*-MRRs, G53 (C), G54 (D) or G55 (E). Except for the pattern of G54, the other expression patterns have been published (2).



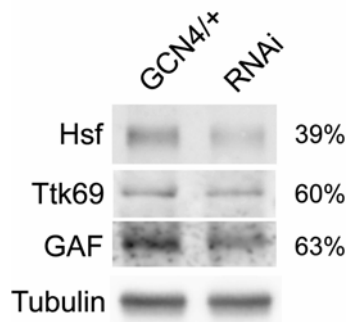
**Figure S3. The anterior stripe of *tll* expression in embryos with reduced *Trf* activity shifts posteriorly at late stage 5.**

Embryos from females obtained from GLC with *Trf*<sup>R85</sup> (E-H) were used to determine *tll* expression using in situ hybridization. *tll* expression patterns in embryos from GLC with wild type serve as controls (A-D). The embryonic stages are late stage 4 (stage 4; A and E), early (B and F) and late stage 5 (C and G), and stage 6 (D and H). Embryos are arranged in a sagittal view, with the anterior towards the left.



**Figure S4. Hsf and Ttk69 exist in the *tor*-RE-protein complex.**

Shift-western blotting of the complexes formed with GAF (2  $\mu$ l in lanes 2-5), Hsf (3  $\mu$ l in lanes 3-5), Ttk69 (4  $\mu$ l in lanes 1-5) and [<sup>32</sup>P]-labeled *tor*-RE, which are the same as those used in EMSA (Figure 4A). The binding reaction was carried out at room temperature. The DNA-protein complexes were subjected to be separated in a 6% of native polyacrylamide gel and transferred onto stacked nitrocellulose and nylon membranes. Hsf and Ttk69 on nitrocellulose membrane were detected by anti-Hsf and Ttk69 antibodies, respectively. “auto” represents an autoradiogram that shows position of the radiolabeled probe on nylon membrane.



**Figure S5. Reduced levels of GAF, Hsf and Ttk69 proteins in embryos with simultaneous knock-down of the three genes.**

Proteins extracted from embryos laid by females of *GCN4>hsf*, *Trl* and *ttk69* RNAi were separated in an 8% SDS polyacrylamide gel and detected by western blotting with anti-GAF, Hsf or Ttk69 antibodies and a chemiluminescent assay kit. Tubulin served as a loading control to normalize the percentage of protein reduction. The percentages of the remaining proteins in the embryos are shown at the right.

## References

1. Liaw, G.J., Rudolph, K.M., Huang, J.D., Dubnicoff, T., Courey, A.J. and Lengyel, J.A. (1995) The *torso* response element binds GAGA and NTF-1/Elf-1, and regulates *tailless* by relief of repression. *Genes Dev*, **9**, 3163-3176.
2. Chen, Y.J., Chiang, C.S., Weng, L.C., Lengyel, J.A. and Liaw, G.J. (2002) Tramtrack69 is required for the early repression of *tailless* expression. *Mech Dev*, **116**, 75-83.