

Supplementary Figures

FIGURE S1. LPA and S1P activate ERK in adherent microvessel endothelial cells. Serum starved MVECs were treated with 20 μ M LPA (A) or 1 μ M S1P (B) for the times indicated. Cells were treated with increasing concentrations of LPA (C) or S1P (D) for 6 min. Cell lysates were prepared and immunoblotted for changes in ERK activation. Membranes were stripped and reprobed with antibodies directed against ERK2 to demonstrate equal loading.

FIGURE S2. Inhibition of ERK activation blocks lysophospholipid-mediated MVEC proliferation. Serum starved MVECs were treated with 5 μ M U0126 for 30 min prior to addition of 20 μ M LPA. [³H]-thymidine incorporation (A) as an indicator of cell proliferation was determined as described in Materials and Methods. Statistical differences were measured by comparison with cells in the absence of the inhibitor. ** p <0.01; *** p <0.001. In panel B, serum starved cells were treated with U0126 as indicated for 30 min prior to stimulation with either 20 μ M LPA or 1 μ M S1P for 6 min. Cell lysates were subjected to SDS-PAGE and immunoblotted for changes in phospho-ERK. Stripped membranes were reprobed with an anti-ERK2 antibody for loading control.

FIGURE S3. Lysophospholipid-mediated ERK activation and proliferation in MVECs is G_i-dependent. Serum starved MVECs were treated with increasing concentrations of pertussis toxin (PTx) as indicated for 4 h. Cells were then incubated with 20 μ M LPA or 1 μ M S1P for 6 min, lysed and immunoblotted for changes in phospho-ERK (A). Membranes were stripped and reprobed with antibodies directed against ERK2 to demonstrate equal loading. Cells treated with

25 ng/ml PTx overnight were assayed for LPA (20 μ M) mediated [3 H]-thymidine incorporation (B). Statistical differences were measured by comparison with cells in the absence of PTx.

*** p <0.001.

FIGURE S4. Effect of growth factor receptor inhibition on lysophospholipid-mediated

ERK activation in MVECs. Serum starved cells were treated with either 5 μ M AG1478, an

EGF receptor inhibitor (A, B), or 5 μ M SU5416, a VEGF receptor inhibitor (C, D), for 30 min

prior to stimulation with either 10 ng/ml EGF, 10 ng/ml VEGF, 20 μ M LPA, or 1 μ M S1P for 6

min. Cell extracts were subjected to immunodetection using phospho-specific antibodies to ERK

(A, C). Membranes were stripped and reprobed with antibodies directed against ERK2 as

loading control. Alternatively, immunoblots were probed with phospho-tyrosine-specific (pTyr)

antibodies (B, D), stripped and reprobed with antibodies against the EGF receptor (B) or the

VEGF receptor (D).

FIGURE S5. LPA and S1P activate Akt through a G_i-PI3K-dependent pathway. Serum

starved MVECs were treated with 20 μ M LPA (A) or 1 μ M S1P (B) for various times as

indicated. Proteins separated on SDS-PAGE were transferred to nitrocellulose and probed with

antibodies directed against phosphorylated Ser473 of Akt and phospho-specific antibodies to

ERK. Membranes were stripped and reprobed for ERK2 as loading control. Alternatively, cells

were treated with 10 μ M LY294002, an inhibitor of PI3K, for 30 min (C) or 1 ng/ml PTx for 20

h (D) prior to stimulation with 20 μ M LPA or 1 μ M S1P for 6 min. Immunoblots were probed

with anti-pS473Akt and anti-pERK antibodies. Membranes were stripped and reprobed with an

anti-ERK2 antibody as loading control.

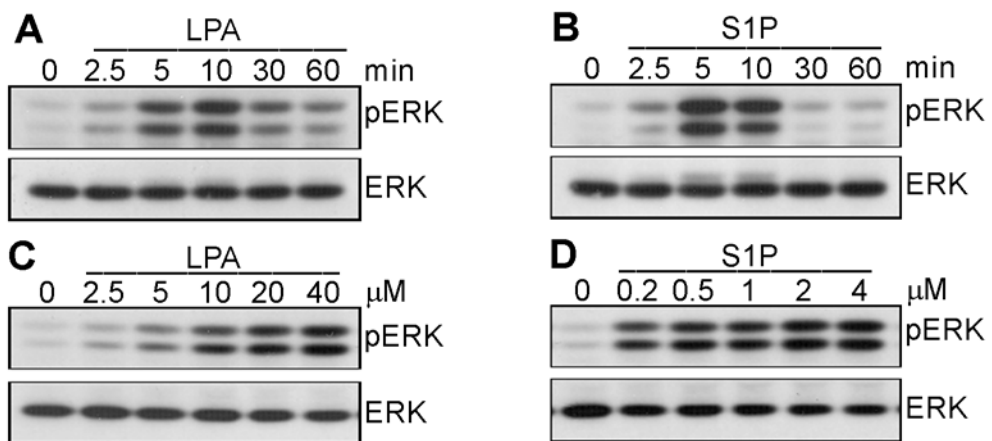


Figure S1

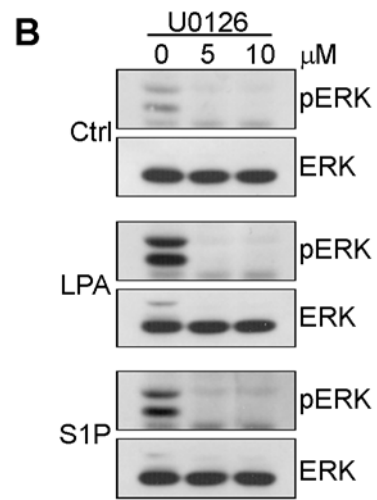
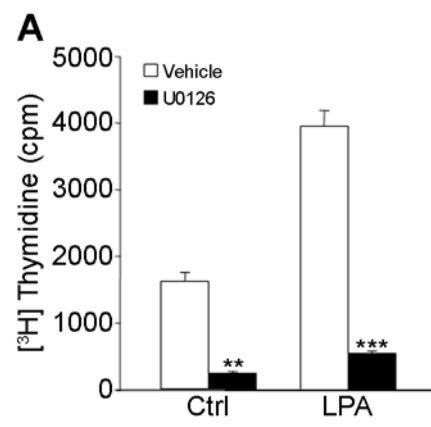


Figure S2

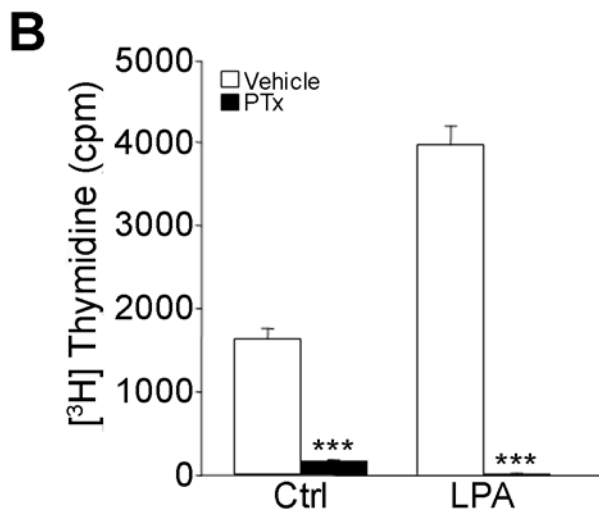
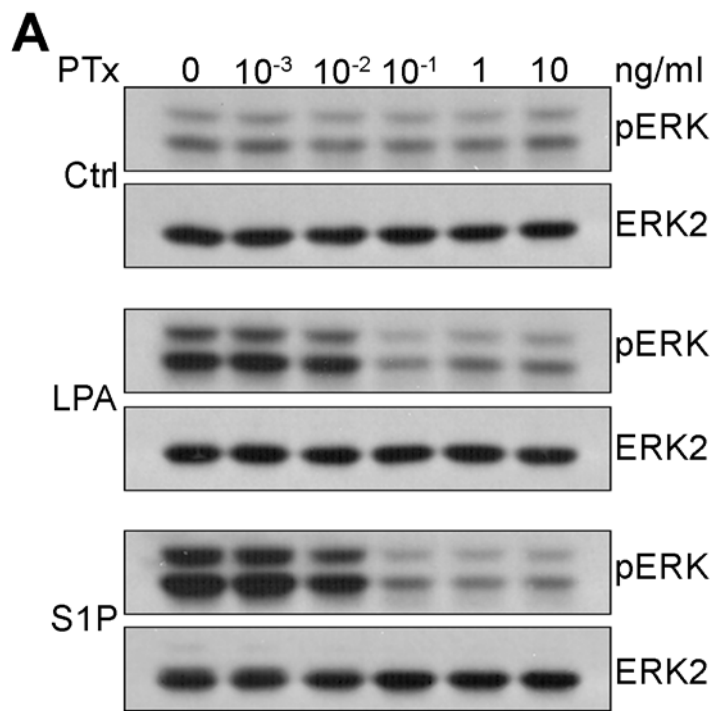


Figure S3

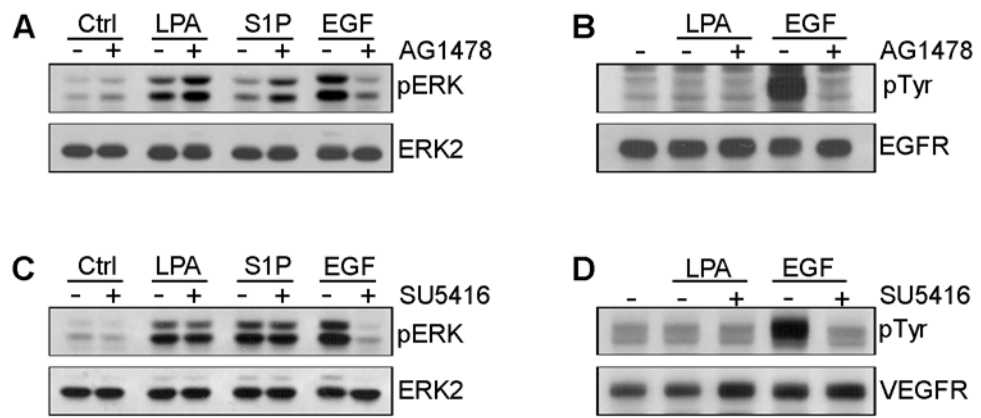


Figure S4

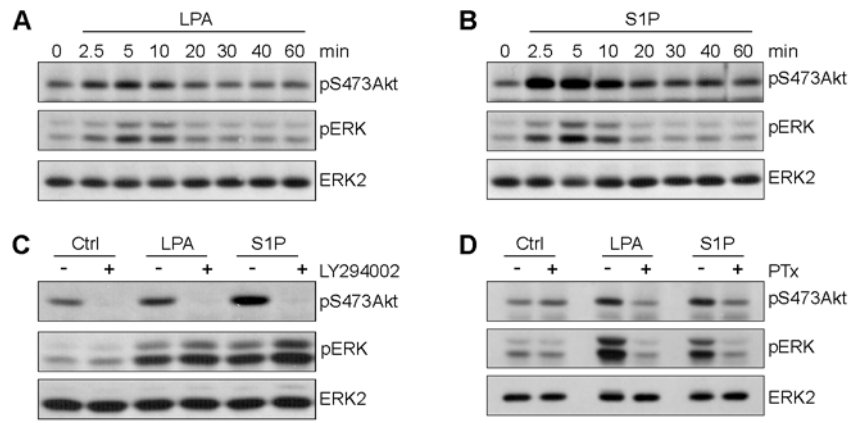


Figure S5