

Functional Characterization of *tImK* Unveiling Unstable Carbinolamide Intermediates in the Tallysomyacin Biosynthetic Pathway*

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EXPERIMENTAL PROCEDURES

Breaklight assay—Breaklight assay of the DNA cleavage activity of TLM A, TLM K-1(Cu-free) and TLM K-2 was carried out according to the literature procedure (1). A labeled oligonucleotide (i.e. breaklight), 5'-6-FAM-GGGTTAAGGGTTTTCCCTTAACCC-3'BHQ1 with a 5'-6-carboxyfluorescein (5'-6-FAM) and a 3'-Black Hole Quencher 1 (BHQ1), was purchased from Integrated DNA Technologies (Coralville, IA) and used as the DNA substrate. The assay mixture contains TLM A (200 nM), K-1 (Cu-free) (0.2 μ M) or K-2 (0.2 μ M), respectively, and 3.2 nM of the labeled oligonucleotide in 25 mM Tris-HCl buffer (pH7.5). The assay reaction was initiated by addition of $(\text{NH}_4)_2\text{Fe}(\text{SO}_4)_2$. All samples were filtered before analysis. The reactions were monitored over 5 min with a FluoroMax-3 spectrofluorometer equipped with DataMax for Windows (Instruments SA, Edison, NJ), and analyzed via a time base scan ($\lambda_{\text{ex}}=485$ nm, $\lambda_{\text{em}}=517$ nm) in a Suprasil quartz cuvette (10-mm path) fitted with a magnetic stirring bar in a total volume of 2 mL. The temperature was controlled by a Haake Circulator DC10 set to 37 °C. Total cleavage of the breaklight was defined as the fluorescence emission under saturated cleaving conditions, 2 mg/mL DNase I, 37 °C, 4 hr. Emission units were converted to the amount of labeled oligonucleotide used within a procedure, thereby equating labeled oligonucleotide degradation as a function of the emission of fluorescence.

TABLE S1. Plasmids used in this study.

Plasmids	Description	Source/reference
SuperCos1	Cosmid vector containing a neomycin/kanamycin resistance gene (<i>neo</i>)	Stratagene
pIJ780	λ RED-mediated PCR-targeting vector containing a viomycin resistance gene (<i>vph</i>)	(2)
pBS8004	Φ C31-derived integrative vector containing an <i>aac(3)IV-tsr-oriT</i> cassette	(3)
pBS8008	Cosmid containing downstream part of the <i>tlm</i> cluster and an <i>aac(3)IV-tsr-oriT</i> cassette	(3)
pBS8010	λ RED-mediated PCR-targeting vector, derived from pBS8008, containing a neomycin-kanamycin resistance gene (<i>neo</i>)	This study
pBS8011	Cosmid containing downstream part of the <i>tlm</i> cluster, with <i>tlmK</i> replaced by <i>neo</i>	This study
pBS8012	Cosmid carrying the Δ <i>tlmK</i> in-frame deletion	This study
pBS8013	Integrative expression vector carrying <i>ErmE</i> * promoter and Φ C31 integration function	This study
pBS8014	pBS8013-derived construct for Δ <i>tlmK</i> complementation in which the expression of <i>tlmK</i> is under <i>ErmE</i> *	This study

TABLE S2. Bacterial strains used in this study

Strains	Description	Source/reference
<i>Escherichia coli</i>		
DH5 α		(4)
ET12567		(5)
BW25113/pIJ790		(2)
BT340		(2)
<i>Streptoalloteichus hindustanus</i>		
E465-94 (ATCC31158)	Wild type TLM producer	ATCC, Rockville, MD, USA
SB8003	Δ tlmK	This study
SB8004	Δ tlmK /pBS8014	This study

TABLE S3. Oligonucleotides as PCR primers used in this study

Oligonucleotides	Sequence
For replacement of <i>vph</i> in pIJ780 with <i>neo</i> from SuperCos1	
neo-FRT1	5'-G TTCCTATTCTCTAGAAAGTATAGGAACTTCGAAGTTCCCACGCTG CCGCAAGCACTCAG-3'
neo-FRT2	5'-CTATACTTTCTAGAGAATAGGAACTTCGGAATAGGAACTTGCTAGC TTGGTCGGTCATTTCTGAACC-3'
For replacement of <i>tImK</i> with FRT- <i>neo</i>	
tImK-frt1	5'-ccctgatcccgcccccaacgaaggaagccgcatgATTCCGGGGATCCGTCGAC C-3'
tImK-frt2	5'-ccggagcgggatggcgctttctaccactcccggtcaTGTAGGCTGGAGCTGCTTC-3' (Low case letters represent DNA sequence originating from <i>S. hindustanus</i> E465-94 and upper case letter represent DNA sequence flanking the FRT- <i>neo</i> cassette from pBS8101)
For PCR and Southern blot verification of mutant strain	
tImK-up	5'-CGCTGACAGCGCCGTCTGGG-3'
tImK-down	5'-GGCGTTGAGCATCGGGGTGG-3'
For cloning of the <i>ErmE</i> * promoter	
PermEI-f	5'-GGAATTCGTGATGCTAGTCGCGGTTGATC-3'
PermEI-r	5'-GGAATTCGTAATCATGCATTATCTCCTTCTCGCTGGATCCTACCAA CCGG-3'
For cloning of <i>tImK</i>	
tImK-NsiI	5'-TCCAATGCATGGGCCAGTCCTGGTGGTC-3'
tImK-XbaI	5'-GCTCTAGATCAGGCCCGGCGGGGAGAT-3'

REFERENCES

1. Biggins, J. B., Prudent, J. R., Marshall, D. J., Ruppen, M., and Thorson, J. S. (2000) *Proc. Natl. Acad. Sci. U. S. A.* **97**, 13537-13542
2. Gust, B., Challis, L., Fowler, K., Kieser, T., and Chater, F. (2003) *Proc. Natl. Acad. Sci. U. S. A.* **100**, 1541-1546
3. Tao, M., Wang, L., Wendt-Pienkowski, E., George, N. P., Galm, U., Zhang, G., Coughlin, J. M., and Shen, B. (2007) *Mol. BioSyst.* **3**, 60-74
4. Sambrook, J., and Russell, W. (2001) *Molecular Cloning: a Laboratory Manual*. 3rd Ed., Cold Spring Harbor Laboratory Press: New York
5. MacNeil, J., Gewain, M., Ruby, L., Dezeny, G. Gibbons, H., and MacNeil, T. (1992) *Gene*, **111**, 61-68