Supplemental Data. Li et al. (2009). Nuclear activity of ROXY1, a glutaredoxin interacting with TGA factors, is required for petal development in *Arabidopsis thaliana*.

Supplemental Table 1. ROXY1 interacts with TGA factors in the nuclei of transiently transformed *Nicotiana benthamiana* leaves when the C-terminus of YFP (YC) was in-frame fused C-terminally to TGA2, TGA3, TGA7 and PAN, respectively.

	PAN-YC	TGA2-YC	TGA3-YC	TGA7-YC
YN-ROXY1	✔[a]	✔	✔	✔

[a]Ticks indicate YFP fluorescence reconstituted by an interaction.

Supplemental Data. Li et al. (2009). Nuclear activity of ROXY1, a glutaredoxin interacting with TGA factors, is required for petal development in *Arabidopsis thaliana*.

Supplemental Table 2. Floral phenotypes induced by expression of the chimeric *PAN* repressor *PANSRDX* under the control of the endogeneous regulatory sequence of *PAN* (*PANpro:PANSRDX*).

	Percentage of floral phenotypes of *PANpro:PANSRDX* T_1 transgenic plants[a]
Tetramerous wild type flowers developing four normal sepals, petals and stamens	14.6%
Pentamerous flowers with five normal sepals, petals and stamens	18.2%
Tetramerous flowers forming four petals with normal, small or folded shapes and four normal sepals and stamens	18.6%
Pentamerous flowers with five normal, small or folded petals and five normal sepals and stamens	48.6%

[a]500 flowers from 50 T_1 transgenic plants and 10 flowers per T_1 trangenic plant examined.

Supplemental Data. Li et al. (2009). Nuclear activity of ROXY1, a glutaredoxin interacting with TGA factors, is required for petal development in Arabidopsis thaliana.

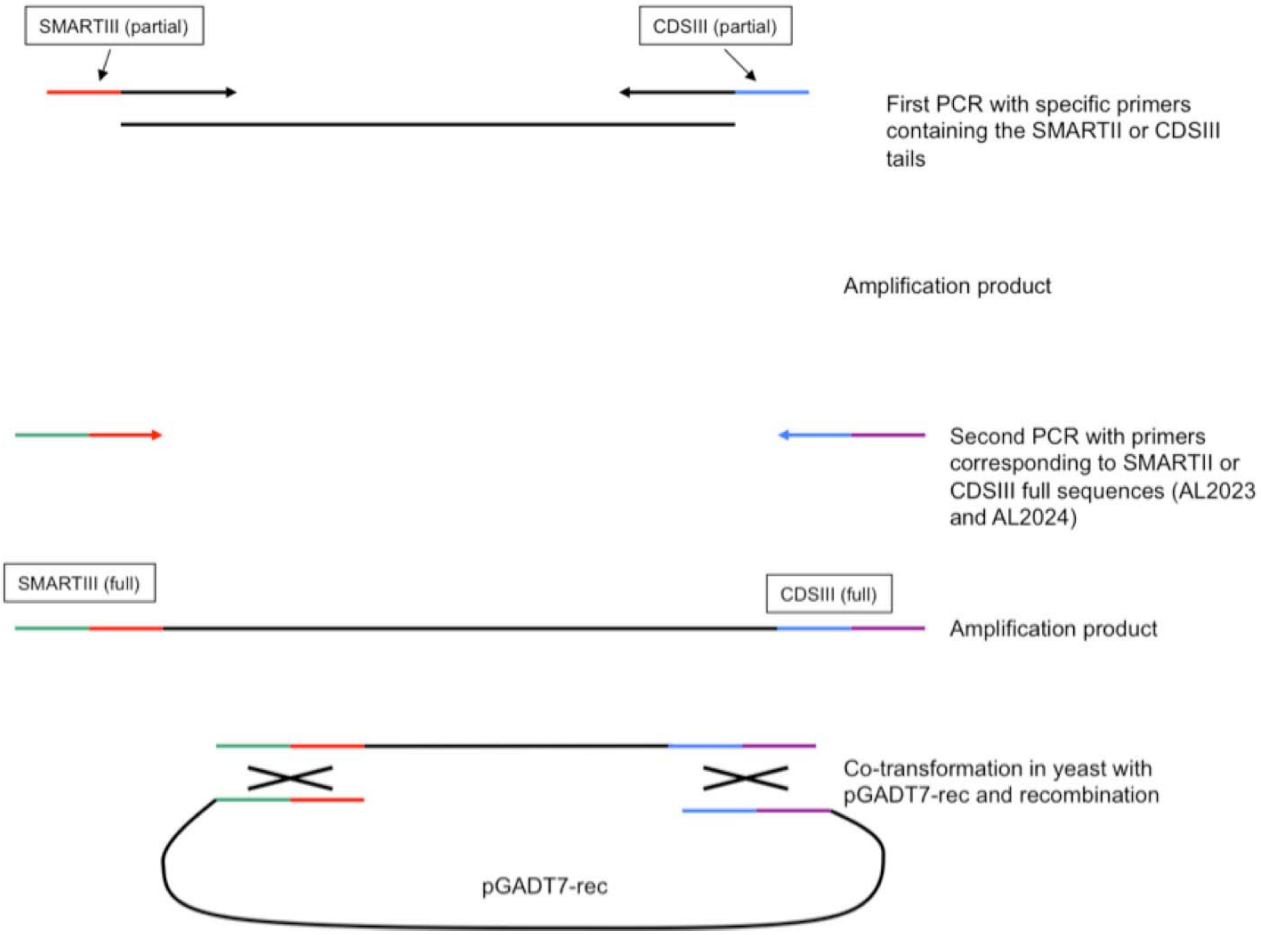
Supplemental Table 3. Primers Used in This Study.

Name	Sequence	Purpose
Primers for yeast-two hybrid screen		
AL2015	CAGAGTGGCCATTATGGCCC ATGATGAGTTCTTCTTCTCCAACACAAC	At1g77920 (TGA7) PCR+Rec in pGADT7-Rec first step-fwd
AL2016	CCGCCGAGGCGGCCGACATG CTTCTTCAAGACTTTTGATTCTGC	At1g77920 (TGA7) PCR+Rec in pGADT7-Rec first step -rev
AL2017	CAGAGTGGCCATTATGGCCC ATGAATTCGACATCGACACATTTTGTG	At5G65210 (TGA1) PCR+Rec in pGADT7-Rec first step-fwd
AL2018	CCGCCGAGGCGGCCGACATG CTACGTTGGTTCACGATGTCGAGTTG	At5G65210 (TGA1) PCR+Rec in pGADT7-Rec first step-rev
AL2019	CAGAGTGGCCATTATGGCCC ATGAATACAACCTCGACACATTTTGTTC	At5G10030 (TGA4) PCR+Rec in pGADT7-Rec first step-fwd
AL2020	CCGCCGAGGCGGCCGACATG TTACGTTGGTTCACGTTGCCTAGCC	At5G10030 (TGA4) PCR+Rec in pGADT7-Rec first step-rev
AL2021	CAGAGTGGCCATTATGGCCC ATGGAGATGATGAGCTCTTCTTCTTC	At1G22070 (TGA3) PCR+Rec in pGADT7-Rec first step-fwd
AL2022	CCGCCGAGGCGGCCGACATG GAAAATATCAGCCAACACTTTCTG	At1G22070 (TGA3) PCR+Rec in pGADT7-Rec first step-rev
AL2023	TCCCACCCAAGCAGTGGTATCAACG CAGAGTGGCCATTATGGCCC	PCR+Rec in pGADT7-Rec univ primers fwd
AL2024	GTATCGATGCCCACCCTCTAGAGG CCGCCGAGGCGGCCGACATG	PCR+Rec in pGADT7-Rec univ primers rev
MZ2032	CAGAGTGGCCATTATGGCCC ATGGGAGATACTAGTCCAAGAACATC	At5g06960(TGA5) PCR+Rec in pGADT7-Rec first step-fwd
MZ2033	CCGCCGAGGCGGCCGACATG CCTTCGTTGAAGTATAATAGAGTCATGG	At5g06960(TGA5) PCR+Rec in pGADT7-Rec first step-rev
MZ2034	CAGAGTGGCCATTATGGCCC ATGGCTGATACCAGTCCGAGAACTG	At5g06950(TGA2) PCR+Rec in pGADT7-Rec first step-fwd
MZ2035	CCGCCGAGGCGGCCGACATG GTACAGCTGACATGTGACCAAAATACAG	At5g06950(TGA2) PCR+Rec in pGADT7-Rec first step-rev
MZ2036	CAGAGTGGCCATTATGGCCC ATGCAGAGCAGCTTCAAAACCGTTCC	At1g68640(PAN) PCR+Rec in pGADT7-Rec first step-fwd
MZ2037	CCGCCGAGGCGGCCGACATG TTAGTCTCTAGGTCTGGCTAACC	At1g68640(PAN) PCR+Rec in pGADT7-Rec first step-rev
MZ2038	CAGAGTGGCCATTATGGCCC ATGGCTGATACCAGTTCAAGGACTG	At3g12250(TGA6) PCR+Rec in pGADT7-Rec first step-fwd
MZ2039	CCGCCGAGGCGGCCGACATG CACACAACACTCGAGTCTTGCGGACC	At3g12250(TGA6) PCR+Rec in pGADT7-Rec first step-rev
AL1377	CCGgaattcATGCAATACCAGACAGAATCCGTG	At3g02000(ROXY1) forward primer to clone ROXY1 in Y2H vectros, contains EcoR I site
AL1378	CGGggatccTCAGAGCCAGAGAGCGCCGGCGTC	At3t02000(ROXY1) reverse primer to clone ROXY1 in Y2H vectros. contains BamH I site
ROXY1AN-F	CCGgaattcATGAGCGCGGTGGTGATCTTCAGCGTG	At3t02000(ROXY1) forward primer for N-terminal deletion of ROXY1
ROXY1AC-R	CGGggatccTCAGAGAAGAGGAACGAGAGAGCCGTTTATG	At3g02000(ROXY1) reverse primer for C-terminal deletion of ROXY1
ROXY1AI-m-F	GACATCCAGGCAGCCCTCATTCTCCGCGTCGTCTCATC	At3g02000(ROXY1) forward middle primer for deletion of ROXY1intervening region
ROXY1AI-m-R	GATGAAGACGACCGGGAGAATGAGGGCTCGCTGGATGTC	At3g02000(ROXY1) reverse middle primer for deletion of ROXY1intervening region
Primers for BiFC experiments		
PAN-pSYC-F	*GGGGACAAGTTTGTACAAAAAAGCAGGCTACCATGCAGAGCAGCTTCAAAACCGTTC*	amplifivine the codine seauence of *PAN* for SPLIT YFP analvsis
PAN-pSYC-R	GGGGACCACTTTGTACAAGAAAGCTGGGTCGTCTCTAGGTCTGGCTAACC	amplifivine the codine seauence of *PAN* for SPLIT YFP analvsis
PAN-pE-SPYCE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGCAGAGCAGCTTCAAAACCGTTC	amplifivine the codine seauence of *PAN* for SPLIT YFP analvsis
PAN-pE-SPYCE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTTAGTCTCTAGGTCTGGCTAACC	amplifivine the codine seauence of *PAN* for SPLIT YFP analvsis
ROXY1-pE-SPYNE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGCAATACCAGACAGAATCGTGG	amplifivine the codine seauence of *ROXY1* for SPLIT YFP analvsis
ROXY1-pE-SPYNE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCAGAGCCAGAGAGCGCCGGCGTC	amplifivine the codine seauence of *ROXY1* for SPLIT YFP analvsis
ROXY1CBd-pE-SPYNE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCAGCAGAAGAGGAACGAGAGAGCCGTTTATG	amplifivine the codine seauence of *ROXY1Cbd* for SPLIT YFP analvsis
TGA3-pSYC-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTACCATGGAGATGATGAGCTCTTCTTCTTC	amplifivine the codine seauence of *TGA3* for SPLIT YFP analvsis
TGA3-pSYC-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCAGTGTGTTCTCGTGGACGAGC	amplifivine the codine seauence of *TGA3* for SPLIT YFP analvsis
TGA3-pE-SPYCE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGGAGATGATGAGCTCTTCTTCTTC	amplifivine the codine seauence of *TGA3* for SPLIT YFP analvsis
TGA3-pE-SPYCE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCAAGTGTGTTCTCGTGGACGAGC	amplifivine the codine seauence of *TGA3* for SPLIT YFP analvsis
TGA1-pE-SPYCE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGAATTCGACATCGACACATTTTGTGCC	amplifivine the codine seauence of *TGA1*for SPLIT YFP analvsis
TGA1-pE-SPYCE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCACTACGTTGGTTCACGATGTCGAGTTGCCCAAC	amplifivine the codine seauence of *TGA1* for SPLIT YFP analvsis
TGA2-pE-SPYCE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGGCTGATACCAGTCCGAGAACTGATG	amplifivine the codine seauence of *TGA2* for SPLIT YFP analvsis
TGA2-pE-SPYCE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCACTCTCTGGGTCGAGCAAGCCATAAG	amplifivine the codine seauence of *TGA2* for SPLIT YFP analvsis
TGA4-pE-SPYCE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGAATACAACCTCGACACATTTTGTTCCACCG	amplifivine the codine seauence of *TGA4* for SPLIT YFP analvsis
TGA4-pE-SPYCE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCAATACGTTGGTTCACGTTGCCTAGC	amplifivine the codine seauence of *TGA4* for SPLIT YFP analvsis
TGA5-pE-SPYCE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGGGAGATACTAGTCCAAGAACATC	amplifivine the codine seauence of *TGA5* for SPLIT YFP analvsis
TGA5-pE-SPYCE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCACTCTCTTGGTCTGGCAAGC	amplifivine the codine seauence of *TGA5* for SPLIT YFP analvsis
TGA6-pE-SPYCE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGGCTGATACCAGTTCAAGGACTG	amplifivine the codine seauence of *TGA6* for SPLIT YFP analvsis
TGA6-pE-SPYCE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCACTCTCTTGGCCGGGCAAGCCACAAGGAAC	amplifivine the codine seauence of *TGA6* for SPLIT YFP analvsis
TGA7-pE-SPYCE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGATGAGTTCTTCTTCTCCAACAC	amplifivine the codine seauence of *TGA7* for SPLIT YFP analvsis
TGA7-pE-SPYCE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCATTAAGTTGGTTCTTGTGGACGAGC	amplifivine the codine seauence of *TGA7* for SPLIT YFP analvsis
ROXY1AN-pE-SPYNE-F	GGGGACAAGTTTGTACAAAAAAGCAGGCTATGAGCGCGGTGGTGATCTTCAGCGTG	amplifivine the codine seauence of *ROXY1AN* for SPLIT YFP analvsis
ROXY1AN-pE-SPYNE-R	GGGGACCACTTTGTACAAGAAAGCTGGGTTCAGAGCCAGAGAGCGCCGGCGTCTTTG	amplifivine the codine seauence of *ROXY1AN* for SPLIT YFP analvsis
Primers for in situ hybridization		
ROXY1-245 bp-T3-F	GCGGAATTAACCCTCACTAAAGGGCCAACAAACTTTAGCCAATC	Amplifiving a 245 bp fragment from the *ROXY1* 5' end for mRNA in situ hybridization
ROXY1-245 bp-T7-R	GCTCGTATACGACTCACTATAGGGCACGTGCTCACGCTGAAGATC	Amplifiving a 245 bp fragment from the ROXY1 5' end for mRNA in situ hybridization
ROXY1-225 bp-T3-F	GCGGAATTAACCCTCACTAAAGGGCTCGATCCCTTCCTCTCCTAGCTTTC	Amplifiving a 225 bp fragment from the *ROXY1* 3' end mRNA in situ hybridization
ROXY1-225 bp-T7-R	GCGCGTAATACGACTCACTATAGGGCTGTTTACTATTATGATTTAATG	Amplifiving a 225 bp fragment from the ROXY1 3' end mRNA in situ hybridization
PAN-588 bp-F	GATGAATCCTCCAGCCGTACTGATTC	Amplifiving a 588 bp fragment from the *PAN* 5' end for mRNA in situ hybridization
PAN-588 bp-T7-R	GCTCGTATACGACTCACTATAGGGCTGTGGAAGTGTCAGTATGTTG	Amplifiving a 588 bp fragment from the *PAN* 5' end for mRNA in situ hybridization
TGA2-371 bp-F	CTGTATTTTGGTCACATGTCAGCTG	Amplifiving a 371 bp fragment from the *TGA2* 3' end for mRNA in situ hybridization
TGA2-371 bp-T7-R	GCTCGTATACGACTCACTATAGGGCTCAAATAGAACATCTTAATATTGTTAAATGATTACAGTACAG	Amplifiving a 371 bp fragment from the *TGA2* 3' end for mRNA in situ hybridization
Primers for genotvpine *roxv1 pan* double mutants		
ST2712	TCCATGTTGTGTCCCATTAAAC	3' primer for *pan* genotvping, PCR with ST13 for an about 900 bp wild type band
ST2713	CGTTCCTTTCACTCCTGATTTC	5' primer for *pan* genotvping, start from -145 bp from ATG ROXY1
ST2711	GCGTGGACCGCTTGCTGCAACT	LBb of pBIN-pROK2 for SALK lines, 110bp from LB of T-DNA
SX896	CAGAGTTAGACTCAGAGGTGTAGTAGG	5' primer for *roxv1* genotvping, start from -145 bp from ATG ROXY1
SX897	GAATAGATACGGCGTCAGTTAATACCG	3' primer for *roxv1* genotvpine. PCR with SX896 for an about 864 bp wild tvne band
SX451	GATAAAGGAAAGGCCATCGTTG	T-DNA right border primer used by GABI-KAT ko mutants for sequencing, pAC161, start from 3007 bp in pAC161 for sequencing for right border
Primers for veneratine PAN-SRDX reoressors		
35Spro-PAN-F	CCCCCGGGACCATGCAGAGCAGCTTCAAAACCGTTC	5' primer for amplification of PAN coding region
35Spro-PAN-SRDX-R	GCTCTAGATCAAGCGAAACCCAACCTAAGCTCCAAGTCAAGGTCCAAGTCTCTAGGTCTGGCTAACC	3' primer for amplification of PAN coding region, incorporating the SRDX-coding sequence
PANpro-PAN-SRDX-F	CCCCCGGGCTTCGGAAAAGAGAAACACTCAAGACTCTAAACC	5' primer for amplification of the genomic region of *PAN*, 2.5 kb upstream of the start codon
PANpro-PAN-SRDX-R	GCTCTAGATCAACCTTGTAGAAGCGTAATAATTG	3' primer for amplification of the genomic region of *PAN*, 1.7 kb downstream of the stop codon
PANpro-PAN-SRDX-m-F	CTCTATGGTTAGCCAGACCTAGAGACTTGGACCTTGACTTGGAGCTTAGGTTGGGTTTCGCTTTGA	5' primer for fusing the SRDX-coding region to PAN coding sequence
PANpro-PAN-SRDX-m-R	CCTTTGGTCATCATGACTCATGCAGGCTTTTTATTCAAAGCGAAACCCAACCTAAGCTCCAAGTCAAGGTCCAA	3' primer for fusing the SRDX-coding region to PAN coding sequence
Primers for mutagenesis of PAN Cysteines and complementation analysis		
35Spro-PAN-R	GCTCTAGATCAGTCTCTAGGTCTGGCTAACCATAG	3' primer for amplification of wild type PAN coding region with 5' primer 35Spro-PAN-F
35Spro-PAN-C27S-m-F	GAGGAGATAGTTCTCTTGAGGAGTTTC	5' primer for mutagenization of PAN C27S
35Spro-PAN-C27S-m-R	GAAACTCCTCAAGAGAACTATCTCCTC	3' primer for mutagenization of PAN C27S
35Spro-PAN-C68S-m-F	CGGTTATGGATTCTGGTGGTGGTGGTG	5' primer for mutagenization of PAN C68S
35Spro-PAN-C68S-m-R	CACCACCACCACCAGAATCCATAACCG	3' primer for mutagenization of PAN C68S
35Spro-PAN-C87S-m-F	GGAGAAGAGGAGTCTTTGGACACAG	5' primer for mutagenization of PAN C87S
35Spro-PAN-C87S-m-R	CTGTGTCCAAAGACTCCTCTTCTCC	3' primer for mutagenization of PAN C87S
35Spro-PAN-C114S-m-F	CAGCAGTTGGTCTGATTCGGTTTC	5' primer for mutagenization of PAN C114S
35Spro-PAN-C114S-m-R	GAAACCGAATCAGACCAACTGCTG	3' primer for mutagenization of PAN C114S
35Spro-PAN-C145S-m-F	GGCTACAAATTCTTCAGATCAATC	5' primer for mutagenization of PAN C145S
35Spro-PAN-C145S-m-R	GATTGATCTGAAGAATTTGTAGCC	3' primer for mutagenization of PAN C145S
35Spro-PAN-C340S-m-F	GATAGGCATTAGCAACCTTCAGC	5' primer for mutagenization of PAN C340S
35Spro-PAN-C340S-m-R	GCTGAAGGTTGCTAATGCCTATC	3' primer for mutagenization of PAN C340S
Primers for studies on intracellular localication of ROXY1		
35Spro-YFP1-F	CCCCCGGGACCATGGTGAGCAAGGGCGAGGAGCTGTTC	5' primer for amplifying the coding region of YFP
35Spro-YFP1(without TCA)-m-R	GAGTTCAGCAAGCTTGTACAAACTCTCGAGGGGCTGCAGCTTGTACAGCTCGTCCATG	3' primer for amplifying the coding sequence of YFP and constructing YFP-ROXY1
35Spro-ROXY1-m-F	CTGCAGCCCTCGAGAGTTTGTACAAGCTTGCTGAACTCATGCAATACCAGACAGAATCGTGG	5' primer for amplifying the coding sequence of ROXY1 and constructine YFP-ROXY1
35Spro-ROXY1-R (with TCA)	GCTCTAGATCAGAGCCAGAGAGCGCCGGCGTCTTTG	3' primer for amplifying the coding sequence of ROXY1
35Spro-NLS-YFP1-F	CACCCGGGAGATGGAGCCAAAGAAGAAGAAAGGTTGAAGACCCAAAG GTGAGCAAGGGCGAGGAGCTG	5' primer for amplifying the coding sequence of YFP, incoporating a NLS-coding sequence at the 5'
35Spro-YFP1(without TCA)-R	GCTCTAGAGTCCTTGTACAGCTCGTCCATGCCGAG	3' primer for amplifying the coding sequence of YFP and introducing the first YFP fragment
35Spro-Spe I-Nhe I-YFP2/3- F	GGACTAGTGCTAGCATGGTGAGCAAGGGCGAGGAGCTGTTC	5' primer for amplifying the coding sequence of YFP and introducing the second and third YFP fragments
35Spro-Spe I-Nhe I-Xba I-YFP2(without TCA)-R	GGACTAGTGCTAGCTCTAGACTTGTACAGCTCGTCCATGCCGAG	3' primer for amplifying the coding sequence of YFP (without the stop codon) and introducing the second YFP fragment
Primers for complementation analvsis of wild tvne and mutated versions of ROXY1 and the other selected 12 CC-tvne GRXs		
ROXY1pro-ROXY1-F	GCTCTAGAATGCAATACCAGACAGAATCGTG	5' primer for amplifvine the codine region of ROXY1
ROXY1pro-ROXY1-R	GCTCTAGATCAGAGCCAGAGAGCGCCGGCGTC	3' primer for amplifying the coding region of ROXY1
ROXY1pro-ROXY1AN-F	GCTCTAGAATGAGCGCGGTGGTGATCTTCAGCGTG	5' primer for amplifying the coding region of ROXY1AN
ROXY1pro-ROXY1AC-R	GCTCTAGATCAGAGAAGAGGAACGAGAGAGCCGTTTATG	3' primer for amplifying the coding region of ROXY1AC
ROXY2pro-ROXY2-F	GCTCTAGAATGCAATACAAAACAGAAACTCGAGGG	5' primer for amplifvine the codine region of ROXY2
ROXY2pro-ROXY2-R	GCTCTAGATCAGAGCCAAAGAGCGCCAGCATCTTTGAG	5' primer for amplifying the coding region of ROXY2
ROXY3pro-ROXY3-F	GCTCTAGAATGGATGTGGTAGCAAGATTAGCG	5' primer for amplifying the coding region of ROXY3
ROXY3pro-ROXY3-R	GCTCTAGACTAAAGCCACAAAGCACCAG	5' primer for amplifying the coding region of ROXY3
ROXY4pro-ROXY4-F	GCTCTAGAATGGAGAAATAAGAGATTTGTCGTCG	5' primer for amplifying the coding region of ROXY4
ROXY4pro-ROXY4-R	GCTCTAGACTATAACCAAATGGCCTTAGCG	5' primer for amplifying the coding region of ROXY4
ROXY5pro-ROXY5-F	GCTCTAGAATGGAAACGAGTAAGAAGATTTGGCATCG	5' primer for amplifying the coding region of ROXY5
ROXY5pro-ROXY5-R	GCTCTAGATCACAACCATATGGCGTTAG	5' primer for amplifying the coding region of ROXY5
ROXY6pro-ROXY6-F	GCTCTAGAATGGACAAAGTTATGAGAATGTCGTCCG	5' primer for amplifvine the codine region of ROXY6
ROXY6pro-ROXY6-R	GCTCTAGATCAACATAAAATATGGCTTCACTAGGGGAACG	5' primer for amplifying the coding region of ROXY6
ROXY7pro-ROXY7-F	GCTCTAGAATGGACAAAGTTGTGAGAATGTCGTC	5' primer for amplifvine the codine region of ROXY7
ROXY7pro-ROXY7-R	GCTCTAGATCAACATAGATTGGCTTGAAACGGC	5' primer for amplifying the coding region of ROXY7
ROXY8pro-ROXY8-F	GCTCTAGAATGGACAAGGTTATGAGAATGTCATCGG	5' primer for amplifvine the codine region of ROXY8
ROXY8pro-ROXY8-R	GCTCTAGACTAGTTATGAAATGACTGATACGGC	5' primer for amplifying the coding region of ROXY8
ROXY9pro-ROXY9-F	GCTCTAGAATGGACAAAGTGATGAGAATGTCTTCAGAG	5' primer for amplifvine the codine region of ROXY9
ROXY9pro-ROXY9-R	GCTCTAGACTAGTAAAGGATGGACTGATAGGG	5' primer for amplifying the coding region of ROXY9
ROXY10pro-ROXY10-F	CTGCTAGCATGGATATGATAACGAAGATGGTGATGG	5' primer for amplifvine the codine region of ROXY10
ROXY10pro-ROXY10-R	CTGCTAGCTCAAACCCACAATGCACCAG	5' primer for amplifying the coding region of ROXY10
ROXY11pro-ROXY11-F	GCTCTAGAATGGAGAATCTACAAAAGATGATCTCCG	5' primer for amplifvine the codine region of ROXY11
ROXY11pro-ROXY11-R	GCTCTAGATCAAAGCCATAAAGCTCCAGC	5' primer for amplifying the coding region of ROXY11
ROXY12pro-ROXY12-F	GCTCTAGAATGGAGAACCTACAGAAGATGATCTC	5' primer for amplifvine the codine region of ROXY12
ROXY12pro-ROXY12-R	GCTCTAGATTAAAGCCATAATGCTCCAGCACG	5' primer for amplifying the coding region of ROXY12
ROXY13pro-ROXY13-F	GCTCTAGAATGGATAAGCTACAGAAGATGATCTCC	5' primer for amplifvine the codine region of ROXY13
ROXY13pro-ROX13-R	GCTCTAGATCAAAGCCATAGTGCTCCAACC	5' primer for amplifying the coding region of ROXY13
ROXY14pro-ROXY14-F	GCTCTAGAATGGAGAAGCTACAGAAGATGACC	5' primer for amplifvine the codine region of ROXY14
ROXY14pro-ROXY14-R	GCTCTAGATCAAAGCCATAACGCCCCAAC	5' primer for amplifying the coding region of ROXY14
ROXY15pro-ROXY15-F	GCTCTAGAATGGAAGATACAAAAGATGATCTCCG	5' primer for amplifvine the codine region of ROXY15
ROXY15pro-ROXY15-R	GCTCTAGATCAAAGCCATAAAGCCCCAAAGCG	5' primer for amplifying the coding region of ROXY15
ROXY16pro-ROXY16-F	GCTCTAGAATGGAGAAGATATCAAATTTGTTAGAAGACAAGC	5' primer for amplifvine the codine region of ROXY16
ROXY16pro-ROXY16-R	GCTCTAGATCAAATCCATATGGCTCCAGCTCTTC	5' primer for amplifying the coding region of ROXY16
ROXY17pro-ROXY17-F	GCTCTAGAATGGAAAGCGTTAGAAGTTTAGTTGAAG	5' primer for amplifvine the codine region of ROXY17
ROXY17pro-ROXY17-R	GCTCTAGATTACACCCATATGGCTCCG	5' primer for amplifying the coding region of ROXY17
ROXY18pro-ROXY18-F	GCTCTAGAATGCAAAAAGCAATTCGACCATACG	5' primer for amplifvine the codine region of ROXY18
ROXY18pro-ROXY18-R	GCTCTAGATCACCAGCGGATTCGGTCGAC	5' primer for amplifying the coding region of ROXY18
ROXY19pro-ROXY19/GRX480-F	GCTCTAGAATGCAAGGAACGATTTCTTGTGC	5' primer for amplifvine the codine region of ROXY19/GRX480
ROXY19pro-ROXY19/GRX480-R	GCTCTAGATCACAACCACAGAGCCCCAACTTCCTTAAG	5' primer for amplifvine the codine region of ROXY19/GRX480
ROXY20pro-ROXY20-F	GCTCTAGAATGAAGACGATGCGAGGTTTACG	5' primer for amplifvine the codine region of ROXY20
ROXY20pro-ROXY20-R	GCTCTAGATTATGCCCATAAGGCTCCAACC	5' primer for amplifying the coding region of ROXY20
ROXY21pro-ROXY21-F	GCTCTAGAATGATGCAAGAATTAGGCTTACAACG	5' primer for amplifvine the codine region of ROXY21
ROXY21pro-ROXY21-R	GCTCTAGATCATACCCAAAGAGCTCCAACTTGGAC	5' primer for amplifying the coding region of ROXY21

Supplemental Data. Li et al. (2009). Nuclear activity of ROXY1, a glutaredoxin interacting with TGA factors, is required for petal development in *Arabidopsis thaliana*.

Supplemental Methods. The recombination strategy used for cloning coding sequences of PAN and TGAs into the pGADT7-rec yeast expression vector.

Coding sequences of PAN and TGAs were cloned into the pGADT7-rec yeast expression vector following a recombination strategy. First, coding regions were amplified with primers that contain a gene specific sequence at the 3' end and part of the SMARTIII or CDSIII extension at the 5' end for the forward or reverse primers, respectively. The full length of the SMARTIII and CDSIII sequences were reconstructed by amplifying the PCR product with AL2023 and AL2024 primers. These primers correspond to the SMARTIII and CDSIII full length regions and mediate the in yeast recombination of the PCR fragment into the pGADT7-rec. The product of the second amplification was pooled with a SmaI-linearized pGADT7-rec vector and co-transformed into AH109 harbouring the bait to be tested. Transformed yeast was first plated on media lacking Trp and Leu to select for pGADT7-rec/PCR product successful recombination. Single colonies were then chosen to be challenged on SD/-Trp-His-Leu-Ade and 3.0 mM 3-AT. If needed, the pGADT7-rec insert was isolated from the chosen colonies by PCR with AL2023 and AL2024 primers and sequence


SMARTIII (partial)
CDSIII (partial)
First PCR with specific primers containing the SMARTII or CDSIII tails
Amplification product
Second PCR with primers corresponding to SMARTII or CDSIII full sequences (AL2023 and AL2024)
SMARTIII (full)
CDSIII (full)
Amplification product
Co-transformation in yeast with pGADT7-rec and recombination
pGADT7-rec