TSG-6 TRANSFERS PROTEINS BETWEEN GLYCOSAMINOGLYCANS VIA A SER₂₈-MEDIATED COVALENT CATALYTIC MECHANISM

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SUPPLEMENTAL DATA

Fig. S1. Purification of HC•TSG-6 cross-links using strong cation exchange-HPLC. Purified IαI and TSG-6 were incubated to generate HC•TSG-6 complexes. Subsequently the proteins were denatured and digested with trypsin, and the resulting peptides were applied to a strong cation exchange column. The column was developed by a gradient of increasing KCl concentration and the eluting peptides monitored at 220 nm. The figure shows the strong cation exchange-HPLC trace. The fraction numbers are indicated above the x-axis. Pool 1 and pool 2 were used for subsequently purification of HC1•TSG-6 and HC2•TSG-6, respectively.



<u>Fig. S2.</u> MS analyses of strong cation exchange-HPLC fractions. Fractions 22-44 (+/- NaOH treatment) from the strong cation exchange-HPLC were analyzed by MALDI-TOF MS. The fractions sensitive to NaOH treatment are shown. (*A*) Fraction 31, (*B*) fraction 31 after NaOH treatment, (*C*) fraction 34, (*D*) fraction 34 after NaOH treatment, (*E*) fraction 35, and (*F*) fraction 35 after NaOH treatment. The observed MH⁺'s correspond to the presence of HC1•TSG-6 cross-link in fraction 31 (m/z 2174.05) and the presence of HC2•TSG-6 cross-link in fraction 34-35 (m/z 1943.92).





<u>Fig. S3.</u> Tandem MS analyses of the tryptic TSG-6 peptide that participates in the cross-links. The tryptic TSG-6 peptide released after NaOH treatment of the cross-links was purified using RP-HPLC, subsequently the peptide was modified using the CAF derivatization protocol, and finally analyzed by tandem MS. (*A*) MS/MS analysis of the TSG-6 peptide (Fig. 2B, peak β) released from the HC1•TSG-6 complex. (*B*) MS/MS analysis of the TSG-6 peptide (Fig. 3B, peak λ) released from the HC2•TSG-6 complex. The two spectra ((*A*) and (*B*)) are very similar and the TSG-6 sequence D₂₂GIFHNSIWLER₃₃ was easily derived from both spectra. In contrast to the MS/MS analysis of the cross-link (Fig. 4) the Ser in the NaOH-released TSG-6 peptide did not lose water upon fragmentation. It supports that Ser₂₈ is involved in the cross-link.

$\mathsf{TABLE}\,S1$

Analysis of fragment ions from MS/MS of the CAF modified HC1 •TSG-6 cross-link

The purified HC1•TSG-6 cross-link (Fig. 2A, peak α) was modified using the CAF derivatization protocol and subjected to MS/MS analysis (Fig. 4A-B). The resulting fragment ions were subsequently examined. The CAF protocol favors the formation of Y-ions and only "weak" B-ions were detected. The detected fragment ion series correlate with an ester bond between the internal Ser residue in the TSG-6 peptide and the terminal Asp residue of HC1.

Y-ions:

Sequence: DGIFHNSIWLER, where Ser is dehydrated.

	INIH+		NIH+	
Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	175,13	-	-	R
Y2"	304,16	304,18	0,02	ER
Y3"	417,25	417,26	0,01	LER
Y4"	603,33	603,34	0,01	WLER
Y5"	716,41	716,42	0,01	IWLER
Y6"	785,42	785,45	0,03	S(-H ₂ O)IWLER
Y7"	899,46	899,49	0,03	NS(-H ₂ O)IWLER
Y8"	1036,52	1036,56	0,04	HNS(-H ₂ O)IWLER
Y9"	1183,59	1183,62	0,03	FHNS(-H ₂ O)IWLER
Y10"	1296,68	-	-	IFHNS(-H ₂ O)IWLER
Y11"	1353,70	1353,73	0,03	GIFHNS(-H ₂ O)IWLER
Y12"	1468,72	1468,76	0,04	DGIFHNS(-H ₂ O)IWLER
Intact	1604,72	1604,73	0,01	"CAF"DGIFHNS(-H ₂ O)IWLER

Sequence: DGIFHNSIWLER, with intact Ser. $$\rm MH$\ensuremath{+}$$

Ion	Theoretical	Observed	Δ MH+	Peptide
Y2"	304,16	304,18	0,02	ER
Y3"	417,25	417,26	0,01	LER
Y4"	603,33	603,34	0,01	WLER
Y5"	716,41	716,42	0,01	IWLER
Y6"	803,44	-	-	SIWLER
Y7"	917,48	-	-	NSIWLER
Y8"	1054,54	1054,56	0,02	HNSIWLER
Y9"	1201,61	1201,62	0,01	FHNSIWLER
Y10"	1314,70	-	-	IFHNSIWLER
Y11"	1371,72	1371,76	0,04	GIFHNSIWLER
Y12"	1486,74	1486,79	0,05	DGIFHNSIWLER
Intact	1622,74	1622,73	-0,01	"CAF"DGIFHNSIWLER

Sequence: VTGVDTD (cross-linked to DGIFHNSIWLER)

_	MH+			
Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	1737,77*	1737,80	0,03	D
Y2"	1838,82	1838,81	-0,01	TD
Y3"	1953,85	1953,86	0,01	DTD
Y4"	2052,91	2052,91	0,00	VDTD
Y5"	2109,94	2109,93	-0,01	GVDTD
Y6"	2210,98	2210,98	0,00	TGVDTD
Y7"	2310,05	2310,07	0,02	VTGVDTD
Intact	2446,05	2446,05	0,00	"CAF"VTGVDTD

* = TSG-6 peptide (1485,74) + CAF (136) + Asp (133,04) - H₂0 (18,02) + H+ (1,01)

Sequence: VTGVDTD (cross-linked to GIFHNSIWLER)

	MH+			
Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	1486,74*	1486,79	0,05	D
Y2"	1587,79	1587,85	0,06	TD
Y3"	1702,82	1702,84	0,02	DTD
Y4"	1801,88	1801,95	0,07	VDTD
Y5"	1858,91	1858,95	0,04	GVDTD
Y6"	1959,95	1960,00	0,05	TGVDTD
Y7"	2059,02	-	0,00	VTGVDTD
Intact	2195,02	2195,01	-0,01	"CAF"VTGVDTD

* = TSG-6 peptide (1370.71) + Asp (133,04) - H_{20} (18,02) + H+ (1,01)

Sequence: VTGVDTD (cross-linked to FHNSIWLER)

MH+				
Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	1316,63*	1316,68	0,05	D
Y2"	1417,68	1417,73	0,05	TD
Y3"	1532,70	1532,73	0,02	DTD
y4"	1631,77	-	-	VDTD
Y5"	1688,79	1688,86	0,07	GVDTD
Y6"	1789,83	1789,89	0,06	TGVDTD
Y7"	1888,91	-	-	VTGVDTD
Intact	2024,91	2024,93	0,02	"CAF"VTGVDTD
*	(1000 (0))		21)	

* = TSG-6 peptide (1200,60) + Asp (133,04) - $H_20(18,02) + H+(1,01)$

Sequence: VTGVDTD (cross-linked to HNSIWLER)

	MH+		MH+		
Ion	Theoretical	Observed	Δ MH+	Peptide	
Y1"	1169,57*	1169,61	0,04	D	
Y2"	1270,62	1270,65	0,03	TD	
Y3"	1385,64	1385,68	0,04	DTD	
Y4"	1484,71	-	-	VDTD	
Y5"	1541,73	1541,76	0,03	GVDTD	
Y6"	1642,78	1642,85	0,07	TGVDTD	
Y7"	1741,85	-	-	VTGVDTD	
Intact	1877,85	1877,86	0,01	"CAF"VTGVDTD	

* = TSG-6 peptide (1053,54) + Asp (133,04) - $H_20(18,02) + H_1(1,01)$

Sequence: VTGVDTD (cross-linked to NSIWLER) MH+

Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	1032,51*	1032,53	0,02	D
Y2"	1133,56	1133,59	0,03	TD
Y3"	1248,59	1248,61	0,02	DTD
Y4"	1347,66	-	-	VDTD
Y5"	1404,68	1404,70	0,02	GVDTD
Y6"	1505,73	1505,74	0,01	TGVDTD
Y7"	1604,80	-	-	VTGVDTD
Intact	1740,80	1740,83	0,03	"CAF"VTGVDTD

* = TSG-6 peptide (916,48) + Asp (133,04) - H₂0 (18,02) + H+ (1,01)

Sequence: VTGVDTD (cross-linked to SIWLER)

	MH	+		
Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	918,46*	918,49	0,03	D
Y2"	1019,52	1019,53	0,01	TD
Y3"	1134,55	1134,54	-0,01	DTD
Y4"	1233,62	-	-	VDTD
Y5"	1290,64	1290,67	0,03	GVDTD
Y6"	1391,69	1391,74	0,05	TGVDTD
Y7"	1490,76	-	-	VTGVDTD
Intact	1626,76	1626,77	0,01	"CAF"VTGVDTD
* = TSG-6	6 peptide (802,43) + Asp (1	33,04) - H20 (18,02) + H+ (1,4	01)	

B-ions:

Sequence: DGIFHNSIWLER, where Ser is dehydrated.

•	MH+				
Ion	Theoretical	Observed	Δ MH+	Peptide	
B1"	252,04	-	-	"CAF"D	
B2"	309,06	-	-	"CAF"DG	
B3"	422,14	-	-	"CAF"DGI	
B4"	569,21	-	-	"CAF"DGIF	
B5"	706,27	-	-	"CAF"DGIFH	
B6"	820,31	830,30	-0,01	"CAF"DGIFHN	
B7"	889,32	889,34	0,02	"CAF"DGIFHNS(-H ₂ O)	
B8"	1002,41	1002,41	0,00	"CAF"DGIFHNS(-H ₂ O)I	
B9"	1188,49	-	-	"CAF"DGIFHNS(-H2O)IW	
B10"	1301,57	-	-	"CAF"DGIFHNS(-H2O)IWL	
B11"	1430,61	-	-	"CAF"DGIFHNS(-H2O)IWLE	
Intact	1604,72	1604,73	0,01	"CAF"DGIFHNS(-H2O)IWLER	

$\mathsf{TABLE}\,S2$

Analysis of fragment ions from MS/MS of CAF modified HC2•TSG-6 cross-link

The purified HC2•TSG-6 cross-link (Fig. 3A, peak γ) was modified using the CAF derivatization protocol and subjected to MS/MS analysis (Fig. 4C-D). The resulting fragment ions were subsequently examined. The CAF protocol favors Y-ions and only a single "weak" B-ion series was detected. The detected fragment ion series correlate with an ester bond between the internal Ser residue in the TSG-6 peptide and the terminal Asp residue of HC2.

<u>Y-ions:</u>

Sequence: DGIFHNSIWLER, where Ser is dehydrated. MH+

Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	175,13	175,14	0,01	R
Y2"	304,16	304,17	0,01	ER
Y3"	417,25	417,24	-0,01	LER
Y4"	603,33	603,32	-0,01	WLER
Y5"	716,41	716,40	-0,01	IWLER
Y6"	785,42	785,42	0,00	S(-H ₂ O)IWLER
Y7"	899,46	899,47	0,01	NS(-H ₂ O)IWLER
Y8"	1036,52	1036,51	-0,01	HNS(-H ₂ O)IWLER
Y9"	1183,59	1183,58	-0,01	FHNS(-H ₂ O)IWLER
Y10"	1296,68	-	-	IFHNS(-H ₂ O)IWLER
Y11"	1353,70	1353,70	0,00	GIFHNS(-H ₂ O)IWLER
Y12"	1468,72	1468,70	-0,02	DGIFHNS(-H ₂ O)IWLER
Intact	1604,72	1604,69	-0,03	"CAF"DGIFHNS(-H2O)IWLER

Sequence: DGIFHNSIWLER, with intact Ser. $$\rm MH$\ensuremath{+}$$

Ion	Theoretical	Observed	Δ MH+	Peptide
Y2"	304,16	304,17	0,01	ER
Y3"	417,25	417,24	-0,01	LER
Y4"	603,33	603,32	-0,01	WLER
Y5"	716,41	716,40	-0,01	IWLER
Y6"	803,44	803,43	-0,01	SIWLER
Y7"	917,48	-	-	NSIWLER
Y8"	1054,54	1054,53	-0,01	HNSIWLER
Y9"	1201,61	1201,59	-0,02	FHNSIWLER
Y10"	1314,70	-	-	IFHNSIWLER
Y11"	1371,72	1371,70	-0,02	GIFHNSIWLER
Y12"	1486,74	1486,73	-0,01	DGIFHNSIWLER
Intact	1622,74	1622,70	-0,04	"CAF"DGIFHNSIWLER

Sequence: VEND (cross-linked to DGIFHNSIWLER)

	MH	+		
Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	1737,77*	1737,72	-0,05	D
Y2"	1851,81	1851,78	-0,03	ND
Y3"	1980,86	1980,80	-0,06	END
Y4"	2079,92	2079,88	-0,04	VEND
Intact	2215,92	2215,86	-0,06	"CAF"VEND
* = TSG-	6 peptide (1485,74) + CAF	(136) + Asp (133,04) - H ₂ 0 (1	8,02) + H+ (1,01)	

Sequence: VEND (cross-linked to GIFHNSIWLER)

	MH	+			
Ion	Theoretical	Observed	Δ MH+	Peptide	
Y1"	1486,74*	1486,73	-0,01	D	
Y2"	1600,78	1600,75	-0,03	ND	
Y3"	1729,83	1729,82	-0,01	END	
Y4"	1828,89	1828,89	0,00	VEND	
Intact	1964,89	1964,87	-0,02	"CAF"VEND	
* = TSG-	6 peptide (1370,71) + Asp	$(133,04) - H_20(18,02) + H + (1)$,01)		

Sequence: VEND (cross-linked to FHNSIWLER) MH+

Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	1316,63*	1316,62	-0,01	D
Y2"	1430,67	1430,65	-0,02	ND
Y3"	1559,72	1559,71	-0,01	END
y4"	1658,78	1658,76	-0,02	VEND
Intact	1794,78	1794,75	-0,03	"CAF"VEND

* = TSG-6 peptide (1200,60) + Asp (133,04) - H_{20} (18,02) + H+(1,01)

Sequence: VEND (cross-linked to HNSIWLER)

	MH	+			
Ion	Theoretical	Observed	Δ MH+	Peptide	
Y1"	1169,57*	1169,56	-0,02	D	
Y2"	1283,61	1283,60	-0,01	ND	
Y3"	1412,66	1412,63	-0,03	END	
Y4"	1511,72	1511,71	-0,01	VEND	
Intact	1647,72	1647,68	-0,04	"CAF"VEND	
Y4" Intact	1511,72 1647,72	1511,71 1647,68	-0,01 -0,04	VENL "CAF"VENL	

* = TSG-6 peptide (1053,54) + Asp (133,04) - H_20 (18,02) + H+(1,01)

Sequence: VEND (cross-linked to NSIWLER)

	MH	+		
Ion	Theoretical	Observed	Δ MH+	Peptide
Y1"	1032,51*	1032,49	-0,02	D
Y2"	1146,55	1146,54	-0,01	ND
Y3"	1275,60	1275,58	-0,02	END
Y4"	1374,66	1374,63	-0,03	VEND
Intact	1510,66	1510,63	-0,03	"CAF"VEND

* = TSG-6 peptide (916,48) + Asp (133,04) - $H_20(18,02) + H_1(1,01)$

Sequence: VEND (cross-linked to SIWLER)

_	MH	+		
Ion Theoretical		Observed	Δ MH+	Peptide
Y1"	918,46*	918,45	-0,01	D
Y2"	1032,50	1032,49	-0,01	ND
Y3"	1161,55	1161,55	0,00	END
Y4"	1260,61	1260,61	0,00	VEND
Intact	1396,61	-	-	"CAF"VEND
* = TSG-6	6 peptide (802,43) + Asp (1	33,04) - H20 (18,02) + H+ (1,	01)	

B-ions:

Sequence: DGIFHNSIWLER, where Ser is dehydrated.

	MH	+				
Ion	Theoretical	Observed	Δ MH+	Peptide		
B1"	252,04	-	-	"CAF"D		
B2"	309,06	-	-	"CAF"DG		
B3"	422,14	-	-	"CAF"DGI		
B4"	569,21	-	-	"CAF"DGIF		
B5"	706,27	-	-	"CAF"DGIFH		
B6"	820,31	820,28	-0,03	"CAF"DGIFHN		
B7"	889,32	889,31	-0,01	"CAF"DGIFHNS(-H ₂ O)		
B8"	1002,41	1002,39	-0,02	"CAF"DGIFHNS(-H ₂ O)I		
B9"	1188,49	-	-	"CAF"DGIFHNS(-H2O)IW		
B10"	1301,57			"CAF"DGIFHNS(-H2O)IWL		
B11"	1430,61	0,61 1430,65 0,04		"CAF"DGIFHNS(-H2O)IWLE		
Intact	1604,72	1604,69		"CAF"DGIFHNS(-H2O)IWLER		

TABLE S3

NH2-terminal protein sequencing of the purified HC•TSG-6 cross-links and unreacted TSG-6

	The close mines field and is a speptices (peak of).											
Position	22	23	24	25	26	27	28	29	30	31	32	33
TSG-6, residue:	D	G	Ι	F	Н	Ν	S	Ι	W	L	Е	R
Amount (pmol):	25.1	14.6	7.0	9.0	3.8	5.9	-	1.6	-	1.3	0.9	-
% of initial yield	100	58.2	27.9	35.9	15.1	23.5	-	6.4	-	5.2	3.6	-

T 1.	1 11101			``
The cross-li	nked HCT and	1 ISG-6 per	ptides (peak)	α):

Position	632	633	634	635	636	637	638
HC1, residue:	V	Т	G	V	D	Т	D
Amount (pmol):	25.5	12.0	18.8	8.5	11.7	3.2	-
% of initial yield	100	47.1	73.7	33.3	45.9	12.5	-

The cross-linked HC2 and TSG-6 peptides (peak γ):

									• ·			
Position	22	23	24	25	26	27	28	29	30	31	32	33
TSG-6, residue:	D	G	Ι	F	Н	Ν	S	Ι	W	L	Е	R
Amount (pmol):	15.9	17.1	9.5	9.4	3.5	6.2	-	1.7	-	1.2	1.1	-
% of initial yield	100	108	59.7	59.1	22.0	39.0	-	10.7	-	7.5	6.9	-

Position	645	646	647	648
HC2, residue:	V	Е	Ν	D
Amount (pmol):	34.5	23.6	16.1	-
% of initial yield	100	68.4	46.7	-

	150-0 (unfeacted):											
Position	18	19	20	21	22	23	24	25	26	27	28	29
Residue:	W	G	F	K	D	G	Ι	F	Н	Ν	S	Ι
Amount (pmol):	-	36.0	31.8	7.7	26.5	23.1	15.7	17.3	4.2	7.8	7.2	8.1
% of initial yield	-	100	88.3	21.4	73.6	64.2	43.6	48.1	11.7	21.7	20.0	22.5

Position	30	31	32	33	34	35	36
Residue:	W	L	Е	R	Α	Α	G
Amount (pmol):	-	7.1	5.0	-	9.6	10.8	9.2
% of initial yield	-	19.7	13.9	-	26.7	30.0	25.6

TSG-6 (unreacted):

TABLE S4

LC-MS/MS analyses of TSG-6 expressed in mammalian cells.

Concentrated medium from mammalian cells transfected with respectively wild type TSG-6 cDNA, S28A mutant TSG-6 cDNA, and extracellular superoxide dismutase cDNA (control) were resolved by SDS-PAGE. Bands of interest were excised, digested with trypsin and analyzed by LC-MS/MS. Several proteins were positively identified by the Mascot search engine in each of the three analyzed media samples. TSG-6 was identified in medium from both the transfection with wild type cDNA and from the transfection with mutant cDNA, but not in the medium from the control transfection. The identification of TSG-6 is shown below. The data demonstrates that both wild type and mutant TSG-6 have been expressed. Mr (*Expt*), the experimental derived relative molecular mass. Mr (*Calc*), calculated relative molecular mass.

Expression of wild type TSG-6: Protein score: 260 MS/MS spectrum number: 45 Matched MS/MS spectrum: 6

Sequence coverage: 23 %								
Peptides	Mr (Expt)	Mr (Calc)	ΔMr	Peptide score				
LTYAEAK	794.33	794.42	-0.09	25				
TGIIDYGIR	1006.42	1006.54	-0.12	55				
ECGGVFTDPK	1122.37	1122.50	-0.13	38				
WDAYCYNPHAK	1437.42	1437.61	-0.19	24				
DGIFHNSIWLER	1485.56	1485.74	-0.18	48				
FLSDASVTAGGFQIK	1539.60	1539.79	-0.20	70				

Expression of mutant S28A TSG-6:

Protein score: 103 MS/MS spectrum number: 38 Matched MS/MS spectrum: 3 Sequence coverage: 10 %

Peptides	Mr (Expt)	Mr (Calc)	ΔMr	Peptide score
TGIIDYGIR	1006.44	1006.54	-0.10	54
YVAMDPVSK	1008.39	1008.49	-0.10	17
ECGGVFTDPK	1122.39	1122.50	-0.12	32