## **Supporting Information**

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**Fig. S1.** The loss of caspase-2 results in reduced apoptosis in nontransformed and E1A/Ras-transformed MEFs. Primary nontransformed *caspase-2<sup>+/+</sup>* and *caspase-2<sup>-/-</sup>* MEFs, and E1A/Ras-transformed *caspase-2<sup>+/+</sup>* and *caspase-2<sup>-/-</sup>* MEFs were exposed to  $\gamma$ -radiation (10 Gy), vincristine (50  $\mu$ g/ml), or cisplatin (30  $\mu$ M) for the indicated times and apoptosis analyzed by Annexin V staining. Results are mean  $\pm$  SEM from 3 experiments (\**P* < 0.05).



**Fig. 52.** Caspase-2 contributes to apoptosis induced by IR. (*A*) Apoptosis level of nontransformed and E1A/Ras-transformed *caspase-2<sup>-/-</sup>* MEFs following restoration of caspase-2 expression via transfection of caspase-2-GFP. Cell were  $\gamma$ -irradiated (IR) and then cultured for 48 or 120 h before apoptosis analysis. Results are expressed as mean  $\pm$  SEM from 3 experiments (\**P* < 0.05). (*B*) Immunoblots showing levels of caspase-2 protein following transfection of *caspase-2<sup>-/-</sup>* MEFs and E1A/Ras-transformed *caspase-2<sup>-/-</sup>* MEFs with the caspase-2-GFP expression construct. Data from 2–3 separate transfection experiments are shown. Asterisk indicates a band that presumably represents caspase-2 from which GFP has been cleaved off. The relative positions of caspase-2-GFP (78 kDa), caspase-2 (51 kDa), and  $\beta$ -actin (42 kDa) are indicated by arrowheads. Same blots were sequentially probed with a caspase-2 antibody and a  $\beta$ -actin antibody.



**Fig. S3.** Reduced levels of *p21* expression in late-passage *caspase-2<sup>-/-</sup>* MEFs. Wild-type (*casp2<sup>+/+</sup>*) and *caspase-2<sup>-/-</sup>* (*casp2<sup>-/-</sup>*) MEFs were grown in high-glucose DMEM and RNA extracted at passages P1, P4, P6, and P8, as indicated. Levels of *p21* transcript were measured in triplicate reactions using real-time qPCR and expressed relative to the internal control gene  $\beta$ -actin. The data shown are averages from a single experiment. Similar data were obtained in 2 independent experiments.

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**Fig. S4.**  $E_{\mu}$ -**Myc** and caspase- $2^{-/-}/E_{\mu}$ -Myc mice show lymphocytic infiltrates characteristic of lymphoma. Representative histology of lymph nodes, liver, and lung from  $E_{\mu}$ -Myc (LHS) and caspase- $2^{-/-}/E_{\mu}$ -Myc (RHS) mice showing (arrows) increased lymphocytic infiltrates and prominent lymphoblasts in blood smear. Sections were stained with H&E.

DNA



**Fig. S5.** The loss of caspase-2 results in reduced apoptosis in lymphoma cells.  $E\mu$ -Myc and  $caspase-2^{-/-}/E\mu$ -Myc lymphoma cells were treated with (A) zoledronic acid for 24 h, (B) cytochalasin D for 24 h, or (C) vincristine for 24 h, or (D)  $\gamma$ -irradiated and cultured for 48 h. Apoptosis was determined by Annexin V staining using flow cytometry. Results are expressed as mean  $\pm$  SEM from 3–4 separate experiments (\*P < 0.05).



**Fig. S6.** VDVADase activity in cells undergoing IR-induced apoptosis following Chk1 inhibition. Caspase activity in *caspase*- $2^{+/+}$  and *caspase*- $2^{-/-}$  (A) and E1A/Ras-transformed *caspase*- $2^{+/+}$  and *caspase*- $2^{-/-}$  MEFs (B) was assessed by cleavage of VDVAD-AMC substrate by cell lysates. Results from 3 experiments are expressed as mean  $\pm$  SEM relative fluorescence units (RFU). \*P < 0.05.