

Continuum of Mechanisms for the Nucleophilic Substitution of Cyclic Acetals

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Supporting Information

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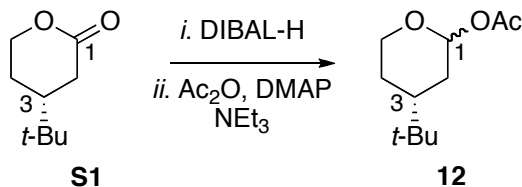
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I. General Experimental Considerations

^1H NMR and ^{13}C NMR spectra were recorded at ambient temperature at 400 or 500 MHz and 100 or 125 MHz, respectively, using a Bruker DRX 400 or DRX 500 spectrometer. Chemical shift data are reported in ppm downfield from tetramethylsilane and are referenced to the proton or carbon peaks for this standard. NMR resonance multiplicities are reported with the following abbreviations: br = broad, s = singlet, d = doublet, t = triplet, q = quartet, and m = multiplet, and coupling constants are reported in Hz. Infrared (IR) spectra were obtained using a MIDAC Prospect FT-IR spectrometer, a Perkin-Elmer Paragon 1000PC FT-IR, or a React-IR AS-1000 spectrometer. High resolution mass spectra were acquired on a VG Analytical 7070E, Fisons Autospec, or Waters LCT Premier spectrometer, and were obtained by peak matching. Microanalyses were performed by Atlantic Microlab Inc, Norcross, GA. Analytical gas-liquid

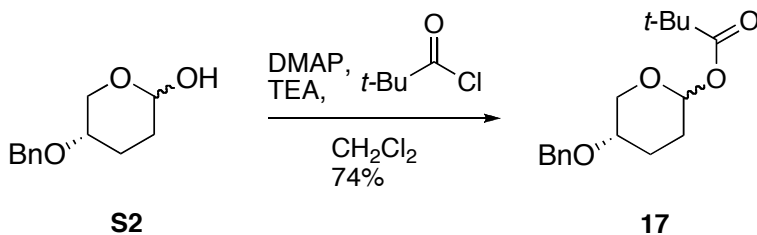
chromatography (GC) analyses were performed on an Agilent 6850 series chromatograph, equipped with an Agilent 6850 auto-sampler and a flame-ionization detector. A fused silica capillary column (30 m x 0.32 mm x 0.25 μ m) wall-coated with DB-1 (J & W Scientific) was used with helium as the carrier gas (25 psi column head pressure). Method of chromatography is as follows: Start temperature = 50 $^{\circ}$ C; ramp = 10 $^{\circ}$ C/min; final temperature = 250 $^{\circ}$ C. Reported melting points are uncorrected. Liquid chromatography was performed using forced flow (flash chromatography) of the indicated solvent system on silica gel (SiO₂) 60 (230–400 mesh). All reactions were performed under nitrogen atmosphere in dry glassware. Glassware was dried in one of the following methods: oven-dried for at least eight hours at 150 $^{\circ}$ C, or flame-dried under dynamic vacuum, or flame-dried under a stream of nitrogen. Unless otherwise noted, all reagents were obtained from commercial suppliers and, where appropriate, purified prior to use. Acetal **1**,¹ lactone **S1**,² lactol **S2**,¹ enoxy silane **8**,^{3,4} and silyl-ketene acetals **9**,^{4,5} **10**,^{4,6} and **11**^{4,7} were prepared by reported methods. THF, Et₂O, and CH₂Cl₂ were dried by filtration through alumina according to the procedure of Grubbs.⁸

II. Substrate Preparation



C3 *t*-Bu Acetal 12. To a cooled (-78 $^{\circ}$ C) solution of lactone **S1**² (1.00 g, 6.40) in CH₂Cl₂ (90 mL) was added diisobutylaluminum hydride (4.48 mL of a 1.5 M solution in toluene, 6.72 mmol) over 40 minutes. After stirring at -78 $^{\circ}$ C for 3 h, acetic anhydride

(2.66 mL, 28.2 mmol) and pyridine (1.81 mL, 22.4 mmol) were added to the reaction mixture dropwise followed by dimethylaminopyridine (0.820 g, 6.72 mmol) in one portion. The reaction mixture was allowed to slowly warm to room temperature and stir overnight. After this time, saturated aqueous ammonium chloride (50 mL) and methyl *tert*-butyl ether (300 mL) were added to the reaction mixture. The resulting emulsion was filtered through Celite and the layers were separated. The organic layer was washed with saturated aqueous Na₂HPO₄ (2 x 40 mL), saturated aqueous NaH₂PO₄ (2 x 40 mL) and saturated aqueous CuSO₄ (2 x 40 mL). The organic layer was dried over Na₂SO₄, filtered, and concentrated by rotary evaporation to yield a clear yellow oil (3.40 g). The residue was purified by flash chromatography (80:20 hexane/EtOAc) to afford **12** as a clear colorless oil (0.857 g, 67%): ¹H NMR (400 MHz, CDCl₃) δ 5.59 (dd, *J* = 9.5, 2.5, 1.0H), 4.09 (ddd, *J* = 11.6, 4.4, 1.7, 1.0H), 3.55 (dt, *J* = 11.6, 2.5, 1.1H), 2.11 (s, 3.1H), 1.85 (m, 1.0H), 1.54 (m, 1.1H), 1.40–1.23 (m, 3.4H), 0.87 (s, 9.5H); ¹³C NMR (125 MHz, CDCl₃) δ 169.6, 121.5, 95.3, 66.6, 44.4, 32.2, 31.8, 27.2, 26.2, 21.3; IR (thin film) 1754, 1232, 1060 cm⁻¹; HRMS (TOF MS ES+) *m/z* calcd for C₁₁H₂₀O₃Na (M + Na)⁺ 223.1310, found 223.1304. Anal. Calcd for C₁₁H₂₀O₃: C, 65.97; H, 10.07. Found: C, 65.69; H, 10.23.



Pivaloate Acetal 17. To a cooled (0 °C) solution of dimethylaminopyridine (DMAP, 0.368 g, 3.01 mmol), triethylamine (4.2 mL, 30 mmol), 4-benzyloxytetrahydropyran-1-ol¹ (**S2**, 1.57 g, 7.53 mmol) in CH₂Cl₂ (35 mL) was added

pivaloyl chloride (1.5 mL, 11 mmol) dropwise by syringe. The reaction mixture was then warmed to room temperature and stirred for 2 h. After this time, the reaction mixture was poured onto 160 mL of a saturated aqueous ammonium chloride solution. The layers were separated, and the aqueous phase was extracted with CH₂Cl₂ (4 x 200 mL). The combined organic extracts were dried (MgSO₄), filtered, and concentrated by rotary evaporation to yield a colorless oil, which formed white crystals of **17** (4.2 g) upon standing. The crystals were washed with pentane (2 x 10 mL), then further purified by flash chromatography (95:5 → 90:10 hexane/EtOAc) to afford **17** (as a 1: 1.5 mixture of anomers) as a white solid (1.6 g, 74%): mp 51–58 °C ¹H NMR (500 MHz, CDCl₃) δ 7.39–7.29 (m, 13.2H), 6.05 (t, *J* = 2.9, 1H), 6.00 (br, 1.5H), 4.60 (m, 5.2H), 3.93 (dd, *J* = 12.4, 2.4, 1H), 3.87–3.78 (m, 2.6H), 3.68–3.56 (m, 4.2H), 2.22 (m, 1H), 2.06–1.97 (m, 2.7H), 1.94–1.89 (m, 5.7H), 1.64–1.59 (m, 2.3H), 1.25 (s, 22.6H); ¹³C NMR (125 MHz, CDCl₃) δ 138.4, 138.3, 128.6, 128.5, 127.9, 127.74, 127.71, 127.66, 92.0, 90.6, 72.0, 70.9, 70.5, 70.4, 64.5, 64.1, 39.15, 39.13, 27.7, 27.1, 24.9, 24.4, 22.7; IR (thin film) 1736, 1155, 1128, 1097, 1027 cm⁻¹; HRMS (TOF MS ES+) *m/z* calcd for C₁₅H₂₀O₂Na (M + Na)⁺ 315.1572, found 315.1566. Anal. Calcd for C₁₇H₂₄O₄: C, 69.84; H, 8.27. Found: C, 69.50; H, 8.24.

III. Nucleophilic Substitution

General Procedure for Nucleophilic Substitution Reactions of Acetal 1 and 12: To a cooled (–78 °C) 0.1 M solution of acetal **1** (0.40–0.61 mmol, 1.0 equiv) and nucleophile (2.0–4.0 equiv) in CH₂Cl₂ was added the Lewis acid (1.4–1.6 equiv) dropwise. After stirring at –78 °C for 1–5 min, the reaction mixture was maintained at –

45 °C for 1–2 h. A saturated aqueous solution of NaHCO₃ (4.0 mL) was added and the solution was warmed to 22 °C. The resulting biphasic mixture was diluted with CH₂Cl₂ (2 mL) and H₂O (2 mL) and the layers were separated. The aqueous layer was extracted with CH₂Cl₂ (3 x 2 mL) and the combined organic layers were dried over Na₂SO₄, filtered, and concentrated *in vacuo*. Diastereomeric ratios were determined by GC and confirmed by GC/MS and ¹H NMR spectroscopy. Reported yields refer to purified material, and examples with each nucleophile are described in the text; those tables are reproduced here (Table 1 and Table 2).

Table 1. Nucleophile Screen with Me₃SiOTf.

Nu–SiMe₃ =

entry	Nu–SiMe ₃	<i>N</i> ^a	product	<i>cis:trans</i> ^{b,c}	yield (%) ^d
1	7	1.8	2	6:94	96
2	8	6.2	3	10:90	95
3	9	8.2	4	71:29	83
4	10	9.0	5	85:15	93
5	11	10.2	6	89:11	96

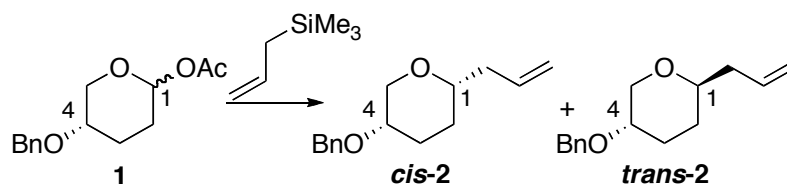
^a *N* = nucleophilicity parameter; see ref 9. ^b Determined by GC and ¹H NMR spectra the unpurified reaction mixtures. ^c Diastereoselectivities were independent of starting anomer ratio. ^d Isolated yield.

Table 2. Nucleophile screen with BF₃•OEt₂.

entry	Nu–SiMe ₃	<i>N</i> ^a	product	<i>cis:trans</i> ^{b,c}	yield (%) ^d
1	7	1.8	2	8:92	82
2	8	6.2	3	8:92	87

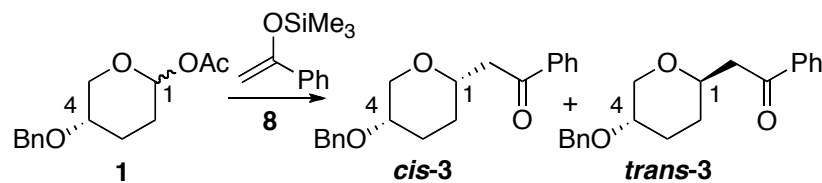
3	9	8.2	4	50:50	88
4	10	9.0	5	58:42	80
5	11	10.2	6	60:40	86

^a *N* = nucleophilicity parameter; see ref 9. ^b Determined by GC and ¹H NMR spectra of the unpurified reaction mixtures. ^c Diastereoselectivities were independent of starting anomer ratio. ^d Isolated yield.



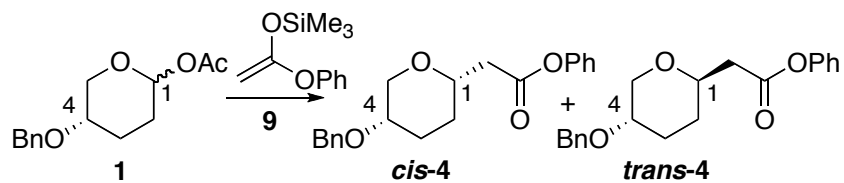
Allyl tetrahydropyran 2: (Table 1, Entry 1 and Table 2, Entry 1).¹ The standard procedure for nucleophilic substitution was followed with acetal **1** (1.0 equiv), allyltrimethylsilane (4.0 equiv), and Lewis acid ($\text{BF}_3 \cdot \text{OEt}_2$ or Me_3SiOTf ; 1.6 equiv in each case). Purification by flash chromatography (97.5:2.5 hexanes/EtOAc) afforded the product as a colorless, clear oil. Product yields and diastereoselectivities are reported in Table 1 and Table 2. The major isomer *trans*-**2** was isolated as a pure sample. Characterization data (¹H NMR, ¹³C NMR) matches reported values.¹

trans-**2**. ¹H NMR (500 MHz, CDCl_3) δ 7.35–7.24 (m, 5H), 5.80 (ddt, *J* = 17.2, 10.2, 7.0, 1H), 5.10–5.01 (m, 2H), 4.58 (d, *J* = 11.9, 1H), 4.52 (d, *J* = 11.9, 1H), 4.09 (ddd, *J* = 10.8, 4.7, 2.3, 1H), 3.43 (tt, *J* = 10.3, 4.6, 1H), 3.33–3.25 (m, 1H), 3.18 (t, *J* = 10.5, 1H), 2.30–2.22 (m, 1H), 2.22–2.12 (m, 2H), 1.78–1.70 (m, 1H), 1.49–1.38 (m, 1H), 1.36–1.25 (m, 1H); ¹³C NMR (125 MHz, CDCl_3) δ 138.7, 135.0, 128.6, 127.82, 127.78, 117.0, 77.1, 73.2, 70.94, 70.92, 40.4, 30.3, 30.2; HRMS (TOF MS ES+) *m/z* calcd for $\text{C}_{15}\text{H}_{20}\text{O}_2\text{Na}$ (*M* + Na)⁺ 255.1361, found 255.1369.



Ketone 3 (Table 1, Entry 2 and Table 2, Entry 2).¹⁰⁻¹² The standard procedure for nucleophilic substitution was followed with acetal **1** (1.0 equiv), enoxy silane **8** (4.0–4.1 equiv), and $\text{BF}_3 \cdot \text{OEt}_2$ (1.6 equiv) or Me_3SiOTf (1.4 equiv). Purification by flash chromatography (90:10 hexanes/EtOAc) afforded the product as an oily white solid. Product yields and diastereoselectivities are reported in Table 1 and Table 2. The major isomer *trans*-**3** was isolated as a pure sample.

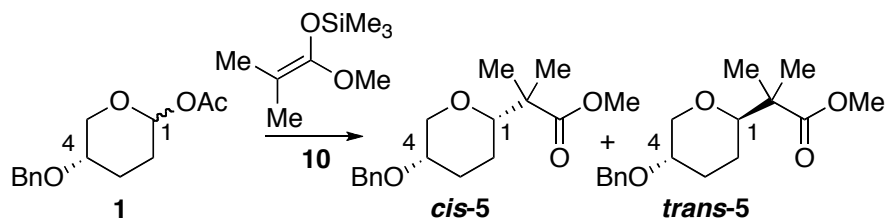
trans-**3**. mp 59–60.5 °C; ^1H NMR (500 MHz, CDCl_3) δ 7.95 (d, $J = 7.9$, 2H), 7.57 (t, $J = 7.3$, 1H), 7.46 (t, $J = 7.7$, 2H), 7.37–7.27 (m, 5H), 4.59 (d, $J = 11.9$, 1H), 4.53 (d, $J = 11.9$, 1H), 4.06 (ddd, $J = 10.8$, 4.6, 2.2, 1H), 3.98–3.88 (m, 1H), 3.48 (tt, $J = 10.0$, 4.5, 1H), 3.29 (dd, $J = 16.3$, 6.7, 1H), 3.24 (t, $J = 10.6$, 1H), 2.93 (dd, $J = 16.3$, 5.8, 1H), 2.27–2.18 (m, 1H), 1.97–1.87 (m, 1H), 1.60–1.48 (m, 1H), 1.46–1.35 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 198.2, 138.7, 137.3, 133.4, 128.8, 128.6, 128.4, 127.9, 127.8, 74.2, 73.0, 71.0, 70.9, 44.7, 30.8, 30.2; IR (thin film) 3061, 2860, 2352, 1681, 1596 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{20}\text{H}_{22}\text{O}_3\text{Na}$ ($\text{M} + \text{Na}$)⁺ 333.1467, found 333.1460. Anal. Calcd for $\text{C}_{20}\text{H}_{22}\text{O}_3$: C, 77.39; H, 7.14. Found: C, 77.14; H, 7.07.



Ester 4 (Table 1, Entry 3 and Table 2, Entry 3). The standard procedure for nucleophilic substitution was followed with acetal **1**, silyl ketene acetal **9** (2.0 equiv for the $\text{BF}_3 \cdot \text{OEt}_2$ -mediated reaction, 4.0 equiv for the Me_3SiOTf -mediated reaction), and Lewis acid ($\text{BF}_3 \cdot \text{OEt}_2$ or Me_3SiOTf ; 1.6 equiv in each case). Purification by flash chromatography (80:20 hexanes/EtOAc) afforded the product as a colorless, clear oil. Product yields and diastereoselectivities are reported in Table 1 and Table 2. Isomers *cis*-**4** and *trans*-**4** were isolated as pure samples for analytical purposes.

cis-**4**. ^1H NMR (500 MHz, CDCl_3) δ 7.40–7.17 (m, 8H), 7.07 (d, $J = 7.9$, 2H), 4.62–4.54 (m, 2H), 4.11 (d, $J = 12.5$, 1H), 3.92 (dddd, $J = 10.4$, 7.5, 5.3, 2.0, 1H), 3.54 (dd, $J = 12.5$, 1.2, 1H), 3.42 (br s, 1H), 2.86 (dd, $J = 15.5$, 7.9, 1H), 2.67 (dd, $J = 15.5$, 5.2, 1H), 2.12–2.06 (m, 1H), 1.86 (m, 1H), 1.71–1.63 (m, 1H), 1.62–1.55 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.0, 150.8, 138.8, 129.5, 128.5, 127.8, 127.7, 125.9, 121.8, 74.1, 70.4, 70.2, 69.7, 41.4, 27.0, 26.4; IR (thin film) 3066, 3031, 2944, 2858, 1756, 1594 cm^{-1} .

trans-**4**. ^1H NMR (500 MHz, CDCl_3) δ 7.40–7.19 (m, 8H), 7.08 (d, $J = 7.7$, 2H), 4.60 (d, $J = 11.9$, 1H), 4.54 (d, $J = 11.9$, 1H), 4.12 (ddd, $J = 10.8$, 4.7, 2.2, 1H), 3.84 (dddd, $J = 10.6$, 7.6, 5.2, 2.1, 1H), 3.49 (tt, $J = 10.1$, 4.5, 1H), 3.26 (t, $J = 10.5$, 1H), 2.74 (dd, $J = 15.2$, 7.9, 1H), 2.65 (dd, $J = 15.2$, 5.2, 1H), 2.27–2.19 (m, 1H), 1.92–1.84 (m, 1H), 1.57–1.40 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 169.9, 150.8, 138.6, 129.6, 128.7, 127.9, 127.8, 126.1, 121.8, 74.2, 72.8, 71.0, 70.9, 41.1, 30.5, 30.1; IR (thin film) 3064, 3033, 2939, 2863, 1756, 1594 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{20}\text{H}_{22}\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$)⁺ 349.1416, found 349.1420. Anal. Calcd for $\text{C}_{20}\text{H}_{22}\text{O}_4$: C, 73.60; H, 6.79. Found: C, 73.22; H, 6.80.

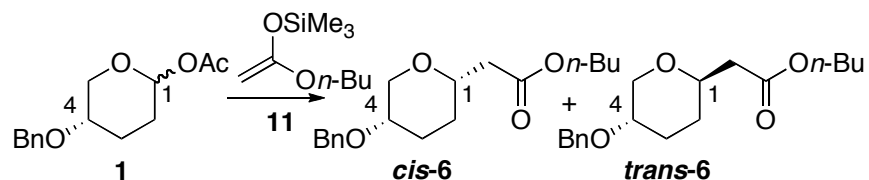


Ester 5 (Table 1, Entry 4 and Table 2, Entry 4). The standard procedure for nucleophilic substitution was followed with acetal **1**, silyl ketene acetal **10** (5.0 equiv for the $\text{BF}_3 \cdot \text{OEt}_2$ -mediated reaction, 4.3 equiv for the Me_3SiOTf -mediated reaction), and Lewis acid ($\text{BF}_3 \cdot \text{OEt}_2$ or Me_3SiOTf ; 1.6 equiv in each case). Purification by flash chromatography (90:10 hexanes/EtOAc) afforded the product as a colorless, clear oil. Product yields and diastereoselectivities are reported in Table 1 and Table 2. Isomers *cis*-**5** and *trans*-**5** were isolated as pure samples for analytical purposes.

cis-**5**. ^1H NMR (500 MHz, CDCl_3) δ 7.38–7.24 (m, 5H), 4.59 (d, $J = 12.3$, 1H), 4.51 (d, $J = 12.3$, 1H), 4.12 (d, $J = 12.5$, 1H), 3.68 (s, 3H), 3.55 (dd, $J = 11.4$, 1.7, 1H), 3.45 (dd, $J = 12.5$, 1.3, 1H), 3.38 (br s, 1H), 2.10 (dt, $J = 13.9$, 3.1, 1H), 1.81 (qd, $J = 13.1$, 3.7, 1H), 1.67–1.57 (m, 1H), 1.31 (br d, $J = 12.8$, 1H), 1.23 (s, 3H), 1.14 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 177.7, 139.1, 128.5, 127.64, 127.58, 82.4, 70.8, 69.9, 69.8, 52.0, 46.9, 27.8, 21.3, 20.64, 20.56; IR (neat) 2950, 2858, 1730, 1454, 1143, 1115 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 315.1572, found 315.1569. Anal. Calcd for $\text{C}_{17}\text{H}_{24}\text{O}_4$: C, 69.84; H, 8.27. Found: C, 70.06; H, 8.32.

trans-**5**. ^1H NMR (500 MHz, CDCl_3) δ 7.38–7.24 (m, 5H), 4.57 (d, $J = 12.0$, 1H), 4.52 (d, $J = 12.0$, 1H), 4.09 (ddd, $J = 10.7$, 4.7, 2.3, 1H), 3.67 (s, 3H), 3.46–3.37 (m, 2H), 3.16 (t, $J = 10.5$, 1H), 2.26–2.18 (m, 1H), 1.66–1.60 (m, 1H), 1.50–1.34 (m, 2H), 1.16 (s, 3H), 1.11 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 177.4, 138.8, 128.6, 127.9, 127.8, 82.3, 73.2, 71.3, 71.0, 52.1, 46.4, 30.3, 24.7, 21.7, 20.7; IR (neat) 2948, 2864, 1734, 1454, 1275,

1093 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$)⁺ 315.1572, found 315.1577. Anal. Calcd for $\text{C}_{17}\text{H}_{24}\text{O}_4$: C, 69.84; H, 8.27. Found: C, 70.00; H, 8.31.

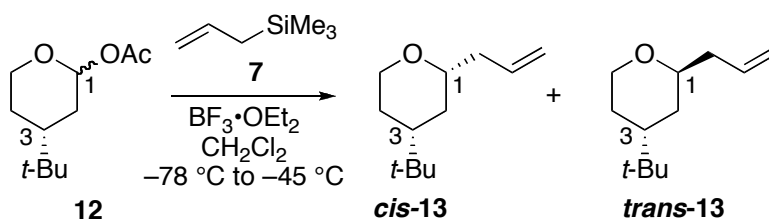


Ester 6 (Table 1, Entry 5 and Table 2, Entry 5). The standard procedure for nucleophilic substitution was followed with acetal **1**, silyl ketene acetal **11** (2.0 equiv for the $\text{BF}_3 \cdot \text{OEt}_2$ -mediated reaction, 3.8 equiv for the Me_3SiOTf -mediated reaction), and $\text{BF}_3 \cdot \text{OEt}_2$ (1.5 equiv) or Me_3SiOTf (1.6 equiv). Purification by flash chromatography (95:5 hexanes/EtOAc) afforded the product as a colorless, clear oil. Product yields and diastereoselectivities are reported in Table 1. Isomers *cis*-**6** and *trans*-**6** were isolated as pure samples for analytical purposes.

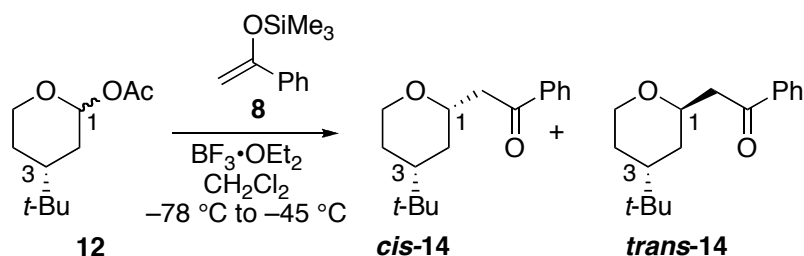
cis-**6**. ^1H NMR (500 MHz, CDCl_3) δ 7.38–7.24 (m, 5H), 4.60–4.53 (m, 2H), 4.12–4.03 (m, 3H), 3.85–3.77 (m, 1H), 3.51 (dd, $J = 12.5, 1.1, 1\text{H}$), 3.40 (br s, 1H), 2.62 (dd, $J = 15.5, 7.7, 1\text{H}$), 2.41 (dd, $J = 15.5, 5.4, 1\text{H}$), 2.10–2.01 (m, 1H), 1.77 (qd, $J = 13.2, 3.6, 1\text{H}$), 1.69–1.57 (m, 3H), 1.51 (d, $J = 12.0, 1\text{H}$), 1.36 (sextet, $J = 7.6, 2\text{H}$), 0.92 (t, $J = 7.5, 3\text{H}$); ^{13}C NMR (125 MHz, CDCl_3) δ 171.6, 138.8, 128.5, 127.7, 127.6, 74.2, 70.4, 70.1, 69.6, 64.5, 41.4, 30.8, 27.0, 26.4, 19.2, 13.9; IR (thin film) 2958, 2871, 1733, 1455, 1115 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{18}\text{H}_{26}\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$)⁺ 329.1729, found 329.1718. Anal. Calcd for $\text{C}_{18}\text{H}_{26}\text{O}_4$: C, 70.56; H, 8.55. Found: C, 70.39; H, 8.48.

trans-**6**. ^1H NMR (500 MHz, CDCl_3) δ 7.35–7.25 (m, 5H), 4.58 (d, $J = 11.9, 1\text{H}$), 4.52 (d, $J = 11.8, 1\text{H}$), 4.14–4.04 (m, 3H), 3.75–3.68 (m, 1H), 3.45 (tt, $J = 10.0, 4.6, 1\text{H}$), 3.21

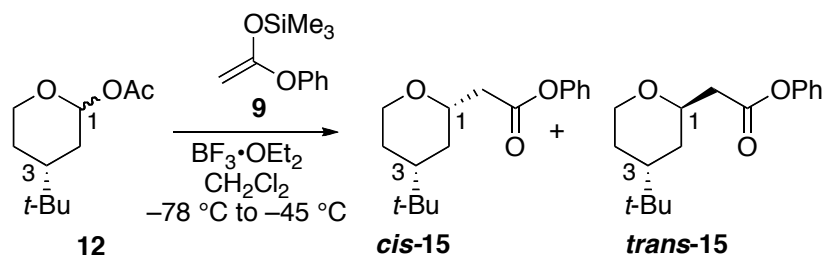
(t, $J = 10.5$, 1H), 2.49 (dd, $J = 15.1$, 7.8, 1H), 2.39 (dd, $J = 15.1$, 5.2, 1H), 2.19 (br d, $J = 12.4$, 1H), 1.79 (br d, $J = 12.9$, 1H), 1.61 (quintet, $J = 6.8$, 2H), 1.54–1.44 (m, 1H), 1.42–1.32 (m, 3H), 0.92 (t, $J = 7.5$, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 171.4, 138.6, 128.6, 127.9, 127.8, 74.2, 72.8, 71.0, 70.9, 64.6, 41.2, 30.8, 30.4, 30.1, 19.3, 13.9; IR (thin film) 2960, 2867, 1733, 1455, 1090 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{18}\text{H}_{26}\text{O}_4\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 329.1729, found 329.1720. Anal. Calcd for $\text{C}_{18}\text{H}_{26}\text{O}_4$: C, 70.56; H, 8.55. Found: C, 70.49; H, 8.70.



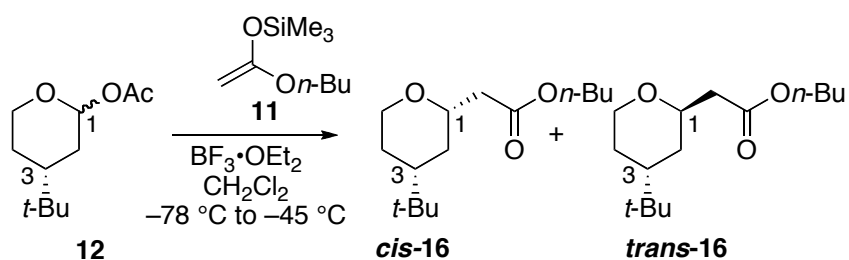
Allyl tetrahydropyran 13: The standard procedure for nucleophilic substitution was followed with acetal **12** (0.041 g, 0.21 mmol), allyltrimethylsilane (0.130 mL, 0.82 mmol), and $\text{BF}_3\cdot\text{OEt}_2$ (0.040 mL, 0.32 mmol). Analysis of the crude mixture by GC revealed a 99:1 *trans-13* to *cis-13* ratio. Purification by flash chromatography (98:2 pentane/ Et_2O) afforded *trans-13* as a colorless, clear oil (0.024 g, 67%): ^1H NMR (500 MHz, CDCl_3) δ 5.80 (ddt, $J = 17.1$, 10.1, 7.0, 1H), 5.12–5.04 (m, 2H), 4.04–3.98 (m, 1H), 3.73 (ddd, $J = 11.7$, 4.9, 2.0, 1H), 3.63 (td, $J = 11.9$, 2.7, 1H), 2.60–2.52 (m, 1H), 2.29 (dt, $J = 14.3$, 7.2, 1H), 1.58–1.46 (m, 3H), 1.45–1.33 (m, 2H), 0.84 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 135.8, 116.7, 72.9, 61.4, 39.5, 35.2, 32.3, 29.2, 27.6, 27.2; IR (thin film) 3078, 2944, 2860, 1642, 1364, 1125 cm^{-1} ; HRMS (TOF MS APCI) m/z calcd for $\text{C}_{12}\text{H}_{23}\text{O}$ ($\text{M} + \text{H}$) $^+$ 183.1749, found 183.1741. Anal. Calcd for $\text{C}_{12}\text{H}_{22}\text{O}$: C, 79.06; H, 12.16. Found: C, 79.02; H, 12.09.



Ketone 14: The standard procedure for nucleophilic substitution was followed with acetal **12** (0.040 g, 0.20 mmol), enoxy silane **8** (0.160 g, 0.83 mmol), and $\text{BF}_3 \cdot \text{OEt}_2$ (0.040 mL, 0.32 mmol). Analysis of the crude mixture by GC revealed a 98:2 *trans-14* to *cis-14* ratio. Purification by flash chromatography (95:5 hexane/EtOAc) afforded *trans-14* as a white solid (0.043 g, 83%): mp 74.5–75.0 °C; ^1H NMR (500 MHz, CDCl_3) δ 7.99–7.96 (m, 2H), 7.59–7.54 (m, 1H), 7.49–7.45 (m, 2H), 4.70–4.64 (m, 1H), 3.79 (ddd, $J = 11.7, 4.9, 2.1$, 1H), 3.68 (td, $J = 11.9, 2.7$, 1H), 3.44 (dd, $J = 15.3, 6.5$, 1H), 3.22 (dd, $J = 15.4, 7.4$, 1H), 1.67–1.56 (m, 3H), 1.49–1.33 (m, 2H), 0.84 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 198.8, 137.2, 133.3, 128.8, 128.4, 70.1, 62.2, 40.2, 39.8, 32.3, 30.0, 27.4, 27.2; IR (thin film) 2956, 2865, 1683 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{O}_2\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 283.1674, found 283.1680. Anal. Calcd for $\text{C}_{17}\text{H}_{24}\text{O}_2$: C, 78.42; H, 9.29. Found: C, 78.24; H, 9.31.



Ester 15: The standard procedure for nucleophilic substitution was followed with acetal **12** (0.042 g, 0.21 mmol), silyl ketene acetal **9** (0.160 g, 0.77 mmol), and $\text{BF}_3 \cdot \text{OEt}_2$ (0.040 mL, 0.32 mmol). Analysis of the crude mixture by GC revealed a 83:17 *trans*-**15** to *cis*-**15** ratio. Purification by flash chromatography (95:5 hexane/EtOAc) afforded a mixture of *trans*-**15** and *cis*-**15** as a colorless, clear oil (0.051 g, 93%). The *trans*-**15** isomer was isolated as a pure sample for analytical purposes: ^1H NMR (500 MHz, CDCl_3) δ 7.37 (t, $J = 7.6$, 2H), 7.22 (t, $J = 7.5$, 1H), 7.09 (d, $J = 7.6$, 2H), 4.67–4.60 (m, 1H), 3.86–3.80 (m, 1H), 3.73 (td, $J = 11.9$, 2.4, 1H), 3.11 (dd, $J = 14.3$, 8.8, 1H), 2.73 (dd, $J = 14.3$, 6.4, 1H), 1.70–1.58 (m, 3H), 1.49–1.34 (m, 2H), 0.87 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.3, 150.9, 129.6, 126.0, 121.8, 70.5, 62.0, 39.8, 36.7, 32.3, 29.9, 27.4, 27.2; IR (thin film) 2954, 2867, 1756, 1594, 1493, 1196 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{17}\text{H}_{24}\text{O}_3\text{Na}$ ($\text{M} + \text{Na}$) $^+$ 299.1623, found 299.1621. Anal. Calcd for $\text{C}_{17}\text{H}_{24}\text{O}_3$: C, 73.88; H, 8.75. Found: C, 74.14; H, 8.87.

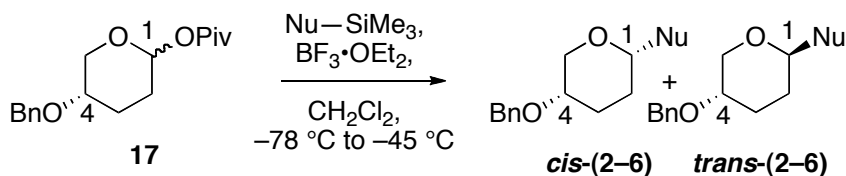


Ester 16: The standard procedure for nucleophilic substitution was followed with acetal **12** (0.040 g, 0.20 mmol), silyl ketene acetal **11** (0.154 g, 0.818 mmol), and $\text{BF}_3 \cdot \text{OEt}_2$ (0.040 mL, 0.32 mmol). Analysis of the crude mixture by GC revealed a 66:34 *trans*-**16** to *cis*-**16** ratio. Purification by flash chromatography (97.5:2.5 hexane/EtOAc)

afforded a mixture of *trans*-**16** and *cis*-**16** as a colorless, clear oil (0.035 g, 69%). Isomers *cis*-**16** and *trans*-**16** were isolated as pure samples for analytical purposes.

cis-**16**. ¹H NMR (500 MHz, CDCl₃) δ 4.10 (t, *J* = 6.7, 2H), 4.05–4.01 (m, 1H), 3.71 (dddd, *J* = 10.3, 7.4, 5.2, 1.9, 1H), 3.41 (td, *J* = 11.5, 2.0, 1H), 2.53 (dd, *J* = 15.0, 7.9, 1H), 2.40 (dd, *J* = 15.0, 5.2, 1H), 1.67–1.58 (m, 3H), 1.53 (dd, *J* = 9.3, 2.0, 1H), 1.43–1.35 (m, 2H), 1.33–1.24 (m, 2H), 1.08–0.99 (m, 1H), 0.93 (t, *J* = 7.4, 3H), 0.85 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 171.7, 74.8, 68.7, 64.5, 45.8, 42.2, 33.0, 32.3, 30.8, 27.3, 27.1, 19.3, 13.9; IR (thin film) 2958, 2869, 1737, 1468, 1364 cm⁻¹; HRMS (TOF MS ES+) *m/z* calcd for C₁₅H₂₉O₃ (M + H)⁺ 257.2117, found 257.2116. Anal. Calcd for C₁₅H₂₈O₃: C, 70.27; H, 11.01. Found: C, 70.44; H, 11.25.

trans-**16**. ¹H NMR (500 MHz, CDCl₃) δ 4.52–4.45 (m, 1H), 4.14–4.04 (m, 2H), 3.78–3.73 (m, 1H), 3.63 (td, *J* = 11.7, 2.4, 1H), 2.85 (dd, *J* = 14.4, 8.5, 1H), 2.49 (dd, *J* = 14.4, 6.5, 1H), 1.64–1.50 (m, 5H), 1.42–1.31 (m, 4H), 0.93 (t, *J* = 7.4, 3H), 0.84 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 171.9, 70.4, 64.6, 61.8, 39.7, 36.6, 32.2, 30.9, 29.9, 27.4, 27.1, 19.3, 13.9; IR (thin film) 2958, 2869, 1737, 1468, 1366 cm⁻¹; HRMS (TOF MS ES+) *m/z* calcd for C₁₅H₂₉O₃ (M + H)⁺ 257.2117, found 257.2125. Anal. Calcd for C₁₅H₂₈O₃: C, 70.27; H, 11.01. Found: C, 70.08; H, 11.21.



General Procedure for Nucleophilic Substitution Reactions of Pivaloate

Acetal 17: To a cooled (−78 °C) 0.1 M solution of **17** (0.40–0.69 mmol, 1.0 equiv) and

nucleophile (3.5 – 4.0 equiv) in CH₂Cl₂ was added BF₃•OEt₂ (1.6–3.5 equiv) dropwise. After stirring at –78 °C for 1–5 min, the reaction mixture was maintained at –45 °C for 1–2 h. Upon warming to 22 °C, a saturated aqueous solution of NaHCO₃ (4.0 mL) was added. The resulting biphasic mixture was diluted with CH₂Cl₂ (5 mL) and H₂O (2 mL), and the layers were separated. The aqueous layer was extracted with CH₂Cl₂ (3 x 8 mL) and the combined organic layers were dried over MgSO₄, filtered, and concentrated *in vacuo*. Diastereomeric ratios (Table S1) were determined by GC and confirmed by ¹H NMR spectroscopy. Reported yields (Table S1) refer to purified material.

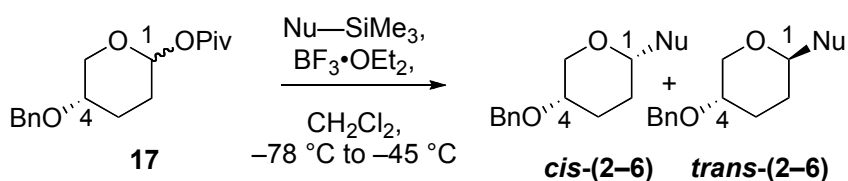


Table S1. Nucleophilic Substitution Reactions with Acetal **17**.

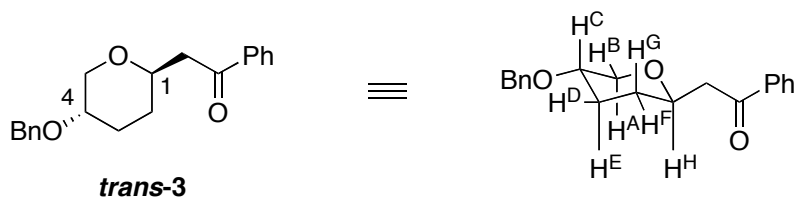
entry	Nu—SiMe ₃	product	<i>cis</i> : <i>trans</i> ^a	yield (%) ^b
1		2	12 : 88	78
2		3	10 : 90	67
3		4	49 : 51	47
4		6	52 : 48	18

^aMeasured by GC analysis of unpurified reaction mixture.

^bBased on purified products.

IV. Stereochemical Proofs of Nucleophilic Substitution Products

Figure S1. Diagnostic ^1H - ^1H coupling constants for *trans*-**3**.



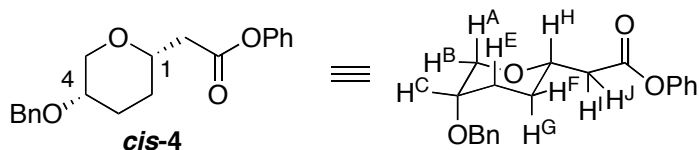
H^{A} : t, $J = 10.6$ Hz (gem $\text{ax}^{\text{A}}\text{-eq}^{\text{B}}$, $\text{ax}^{\text{A}}\text{-ax}^{\text{C}}$)
 H^{C} : tt, $J = 10.0$ Hz ($\text{ax}^{\text{C}}\text{-ax}^{\text{A}}$, $\text{ax}^{\text{C}}\text{-ax}^{\text{E}}$,
 4.5 Hz ($\text{ax}^{\text{C}}\text{-eq}^{\text{B}}$, $\text{ax}^{\text{C}}\text{-eq}^{\text{D}}$)

Figure S2. Relevant DPGFSE-NOE data (mixing time 2.0 s) for *trans*-**3**.



H^{C} irradiated: H^{G} (3.7%)
 H^{E} irradiated: H^{A} (3.8%), H^{H} (8.3%)
 H^{G} irradiated: H^{C} (8.2%)
 H^{H} irradiated: H^{A} (9.3%), H^{E} (4.1%)

Figure S3. Diagnostic ^1H - ^1H coupling constants for *cis*-**4**.



H^{A} : dd, $J = 12.5$ Hz (gem $\text{ax}^{\text{A}}\text{-eq}^{\text{B}}$), 1.2 Hz ($\text{ax}^{\text{A}}\text{-eq}^{\text{C}}$)
 H^{C} : br s
 H^{H} : dddd, $J = 10.4$ Hz ($\text{ax}^{\text{H}}\text{-ax}^{\text{G}}$), 7.5 Hz ($\text{ax}^{\text{H}}\text{-H}^{\text{I}}$),
 5.3 Hz ($\text{ax}^{\text{H}}\text{-H}^{\text{J}}$), 2.0 Hz ($\text{ax}^{\text{H}}\text{-eq}^{\text{F}}$)

Figure S4. Relevant DPGSE-NOE data (mixing time 2.0 s) for *cis-4*.

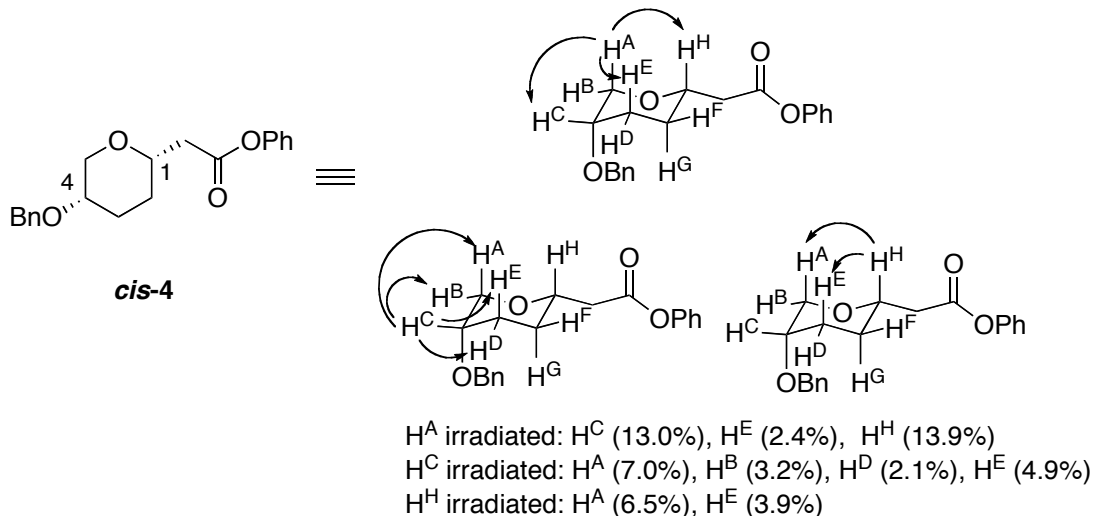


Figure S5. Diagnostic 1H - 1H coupling constants for *trans-4*.

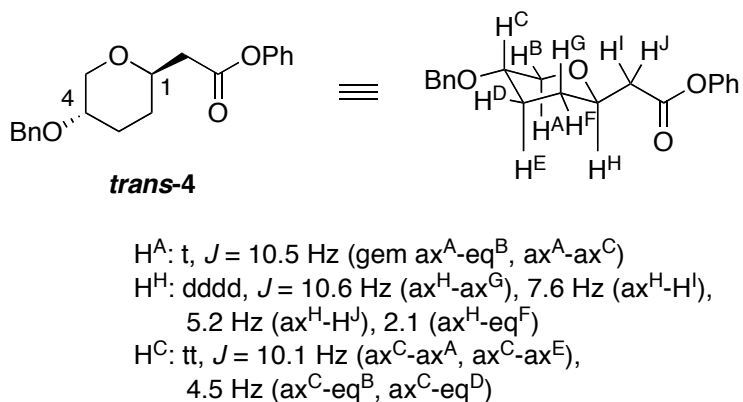


Figure S6. Relevant DPGSE-NOE data (mixing time 2.0 s) for *trans-4*.

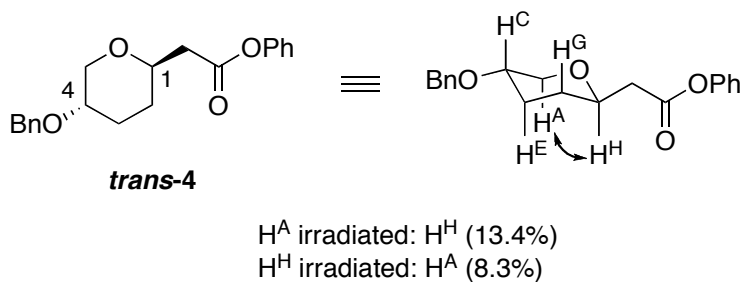
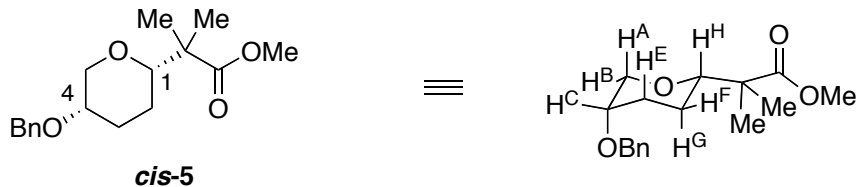


Figure S7. Diagnostic ^1H - ^1H coupling constants for *cis*-**5**.

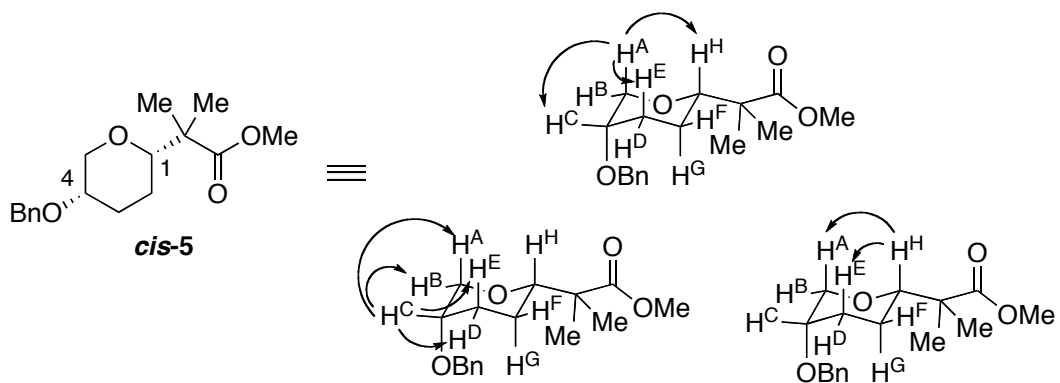


H^{A} : dd, $J = 12.5$ Hz (gem $\text{ax}^{\text{A}}\text{-eq}^{\text{B}}$), 1.3 Hz ($\text{ax}^{\text{A}}\text{-eq}^{\text{C}}$)

H^{C} : br s

H^{H} : dd, $J = 11.4$ Hz ($\text{ax}^{\text{H}}\text{-ax}^{\text{G}}$), 1.7 Hz ($\text{ax}^{\text{H}}\text{-eq}^{\text{F}}$)

Figure S8. Relevant DPGSE-NOE data (mixing time 2.0 s) for *cis*-**5**.

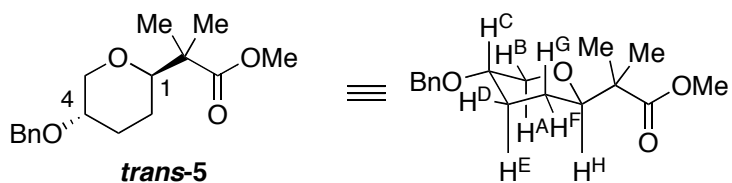


H^{A} irradiated: H^{C} (7.7%), H^{E} (2.9%), H^{H} (12.1%)

H^{C} irradiated: H^{A} (4.1%), H^{B} (3.2%), H^{D} (2.1%), H^{E} (4.7%)

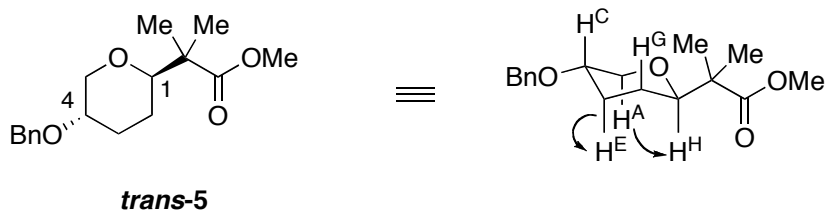
H^{H} irradiated: H^{A} (6.3%), H^{E} (3.8%)

Figure S9. Diagnostic ^1H - ^1H coupling constants for *trans*-**5**.



H^{A} : t, $J = 10.5$ Hz (gem $\text{ax}^{\text{A}}\text{-eq}^{\text{B}}$, $\text{ax}^{\text{A}}\text{-ax}^{\text{C}}$)

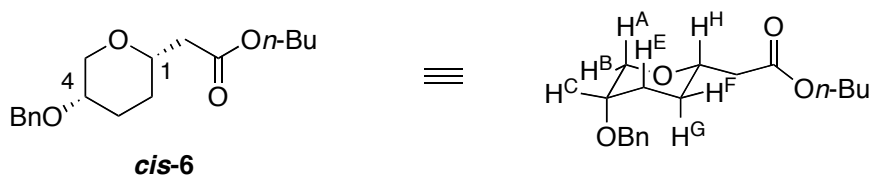
Figure S10. Relevant DPGSE-NOE data (mixing time 2.0 s) for *trans-5*.



trans-5

H^A irradiated: H^H (14.0%), H^E (3.6%)

Figure S11. Diagnostic ¹H-¹H coupling constants for *cis-6*.

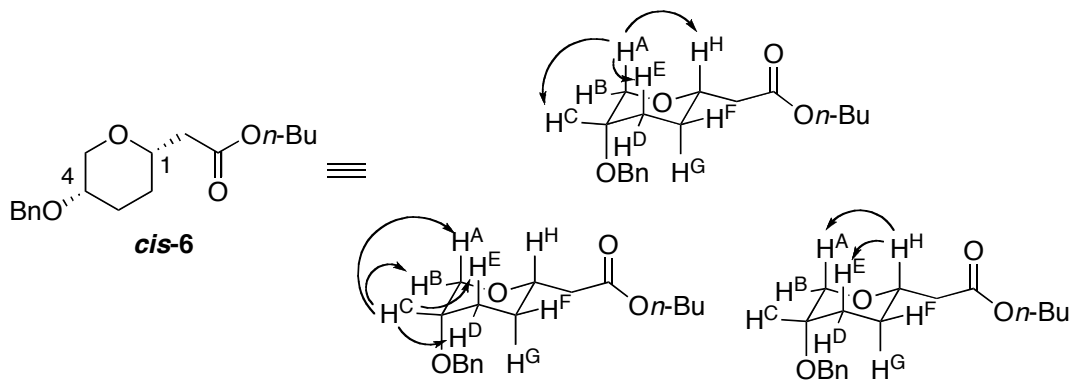


cis-6

H^A: dd, $J = 12.5$ Hz (gem ax^A-eq^B), 1.1 Hz (ax^A-eq^C)

H^C: br s

Figure S12. Relevant DPGSE-NOE data (mixing time 2.0 s) for *cis-6*.



cis-6

H^A irradiated: H^C (11.8%), H^E (2.4%), H^H (13.1%)

H^C irradiated: H^A (6.4%), H^B (3.1%), H^D (2.0%), H^E (4.6%)

H^H irradiated: H^A (6.5%), H^E (3.8%)

Figure S13. Diagnostic ^1H - ^1H coupling constants for *trans*-6.

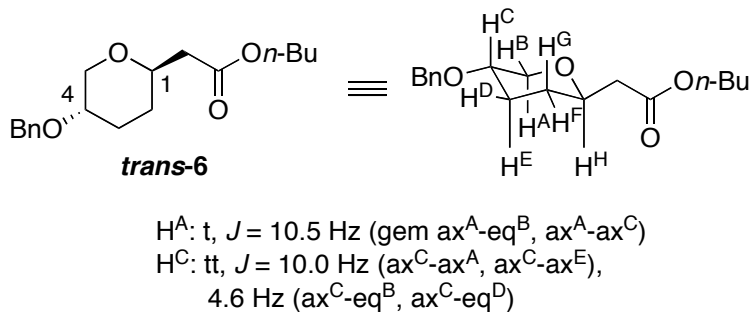


Figure S14. Relevant DPGSE-NOE data (mixing time 2.0 s) for *trans*-6.

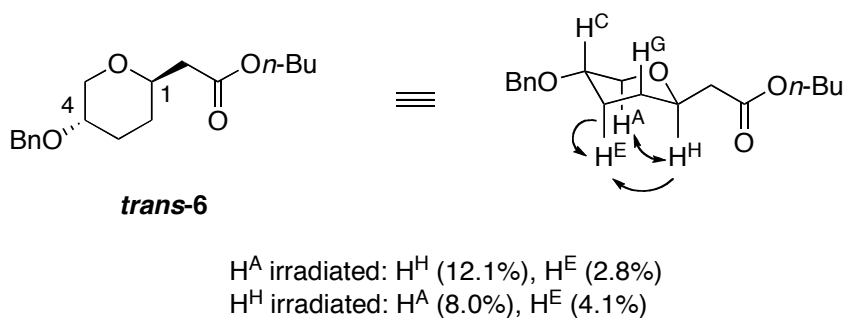


Figure S15. Relevant DPGSE-NOE data (mixing time 2.0 s) for *trans*-13 (analysis assumes an equatorial *tert*-butyl group).

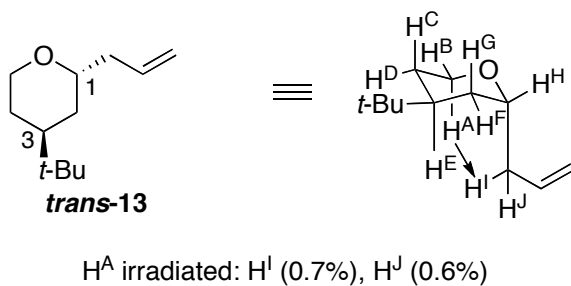


Figure S16. Relevant DPGSE-NOE data (mixing time 2.0 s) for *trans*-**14** (analysis assumes an equatorial *tert*-butyl group).

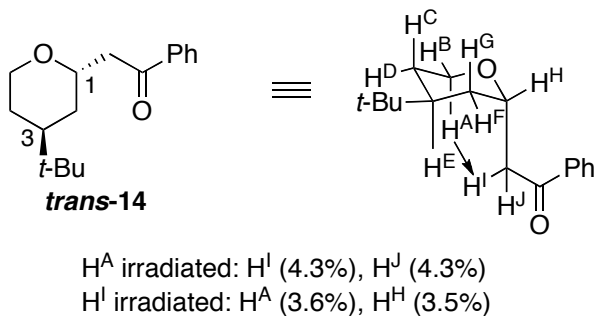


Figure S17. Relevant DPGSE-NOE data (mixing time 2.0 s) for *trans*-**15** (analysis assumes an equatorial *tert*-butyl group).

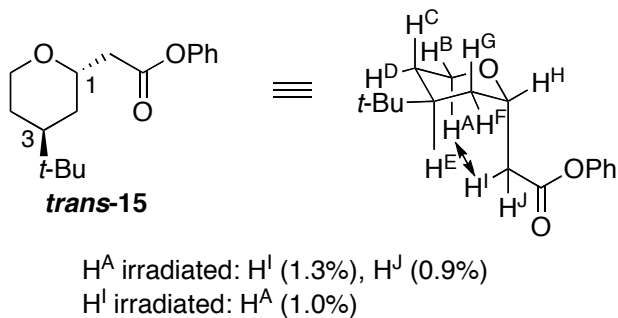


Figure S18. Relevant DPGSE-NOE data (mixing time 2.0 s) for *trans*-**16** (analysis assumes an equatorial *tert*-butyl group).

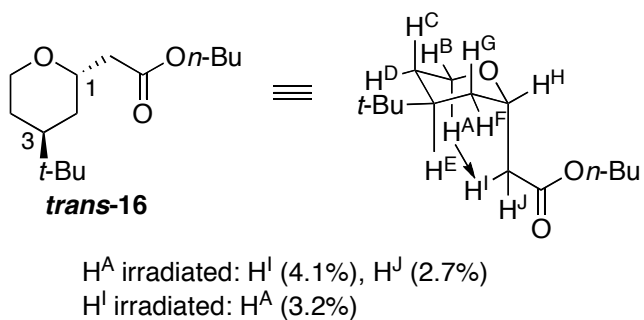
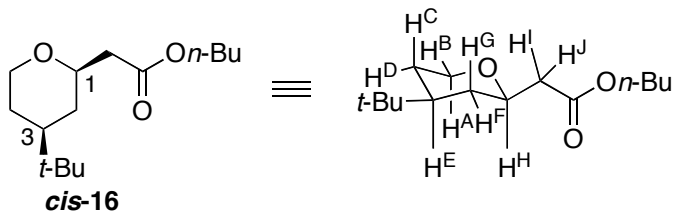
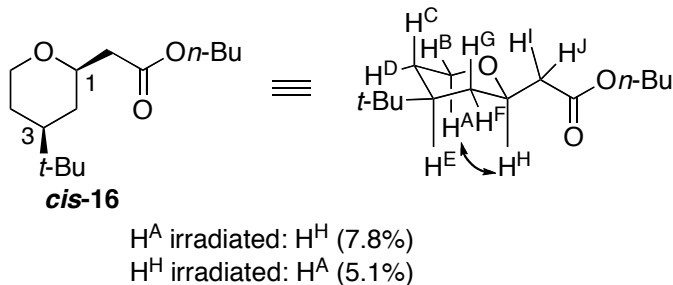


Figure S19. Diagnostic ^1H - ^1H coupling constants for *cis*-**16** (analysis assumes an equatorial *tert*-butyl group).



H^{H} : dddd, $J = 10.3$ Hz ($\text{ax}^{\text{H}}\text{-ax}^{\text{G}}$), 7.4 Hz ($\text{ax}^{\text{H}}\text{-H}^{\text{I}}$),
 5.2 Hz ($\text{ax}^{\text{H}}\text{-H}^{\text{J}}$), 1.9 Hz ($\text{ax}^{\text{H}}\text{-eq}^{\text{F}}$)
 H^{I} : dd, $J = 15.0$ Hz (gem $\text{H}^{\text{I}}\text{-H}^{\text{J}}$), 7.9 Hz ($\text{ax}^{\text{H}}\text{-H}^{\text{I}}$)
 H^{J} : dd, $J = 15.0$ Hz (gem $\text{H}^{\text{I}}\text{-H}^{\text{J}}$), 5.2 Hz ($\text{ax}^{\text{H}}\text{-H}^{\text{I}}$)

Figure S20. Relevant DPGSE-NOE data (mixing time 2.0 s) for *cis*-**16** (analysis assumes an equatorial *tert*-butyl group).



V. Competition Experiments

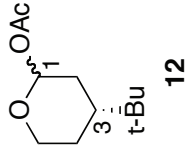
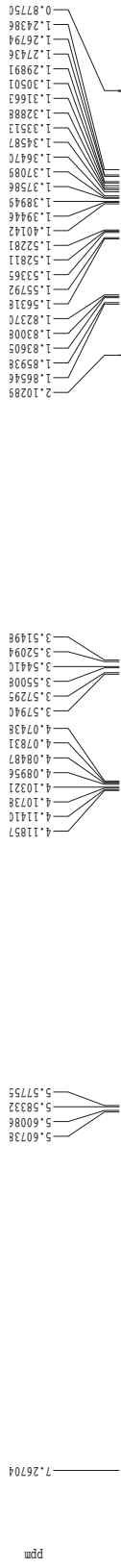
Standard Competition Experiment Conditions. A solution of acetal (0.20 mmol, 1.0 equiv.) and nucleophiles (1.0 mmol each, 5.0 equiv. each) in anhydrous CH_2Cl_2 (2.0 mL) was prepared. An aliquot (0.050 mL) was removed, diluted with CDCl_3 and analyzed by ^1H NMR spectroscopy to confirm the molar ratio of the acetal and nucleophiles prior to addition of Lewis acid. Additional quantities of nucleophiles or

acetal were added to adjust the stoichiometry as needed. The reaction mixture was then cooled to $-78\text{ }^{\circ}\text{C}$, and Lewis acid (0.32 mmol, 1.6 equiv) was added dropwise. After stirring at $-78\text{ }^{\circ}\text{C}$ for 5 min, the reaction mixture was warmed to $-45\text{ }^{\circ}\text{C}$ for 1–2 h before quenching. A saturated aqueous solution of NaHCO_3 (4.0 mL) was added, and the solution was warmed to $22\text{ }^{\circ}\text{C}$, diluted with CH_2Cl_2 (2 mL) and H_2O (2 mL), and the layers were separated. The aqueous layer was extracted with CH_2Cl_2 (3 x 2 mL) and the combined organic layers were dried over Na_2SO_4 , filtered, and concentrated *in vacuo*. Chemoselectivity values were determined by GC after addition of dodecane standard. Chemoselectivity data reported are based on response factor calibration curves generated from authentic samples of pure products with dodecane as internal standards. Diastereoselectivities are reported as uncorrected values.

VI. Bibliography

- (1) Romero, J. A. C.; Tabacco, S. A.; Woerpel, K. A. *J. Am. Chem. Soc.* 2000, 122, 168-169.
- (2) Bergdahl, M.; Eriksson, M.; Nilsson, M.; Olsson, T. *J. Org. Chem.* 1993, 58, 7238-7244.
- (3) House, H. O.; Czuba, L. J.; Gall, M.; Olmstead, H. D. *J. Org. Chem.* 1969, 34, 2324-2336.
- (4) Colvin, E. W. *Silicon Reagents in Organic Synthesis*; Academic Press Inc.: London, 1988.
- (5) Slougui, N.; Rousseau, G. *Synth. Commun.* 1987, 17, 1-11.
- (6) Collins, D. J.; Jacobs, H. A. *Australian Journal of Chemistry* 1986, 39, 2095-2110.
- (7) Miura, K.; Takeyama, Y.; Oshima, K.; Utimoto, K. *Bull. Chem. Soc. Jpn.* 1991, 64, 1542-1553.
- (8) Pangborn, A. B.; Giardello, M. A.; Grubbs, R. H.; Rosen, R. K.; Timmers, F. J. *Organometallics* 1996, 15, 1518-1520.
- (9) Mayr, H.; Kempf, B.; Ofial, A. R. *Acc. Chem. Res.* 2003, 36, 66-77.
- (10) Compound 3 has been synthesized previously, however the reported analytical data was in error. The correct analytical data is presented here.
- (11) Ayala, L.; Lucero, C. G.; Romero, J. A. C.; Tabacco, S. A.; Woerpel, K. A. *J. Am. Chem. Soc.* 2003, 125, 15521-15528.
- (12) Shenoy, S.; Woerpel, K. A. *Org. Lett.* 2005, 7, 1157-1160.

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 D1 0.10000000 sec
 ACRESF 0.00000000 sec
 ACPRK 0.01500000 sec

==== CHANNEL f1 =====
 NU1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SFO1 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300152 MHz
 WDW EN
 SSB 0
 LB 0.30 Hz
 GB 2.00
 PC 2.00

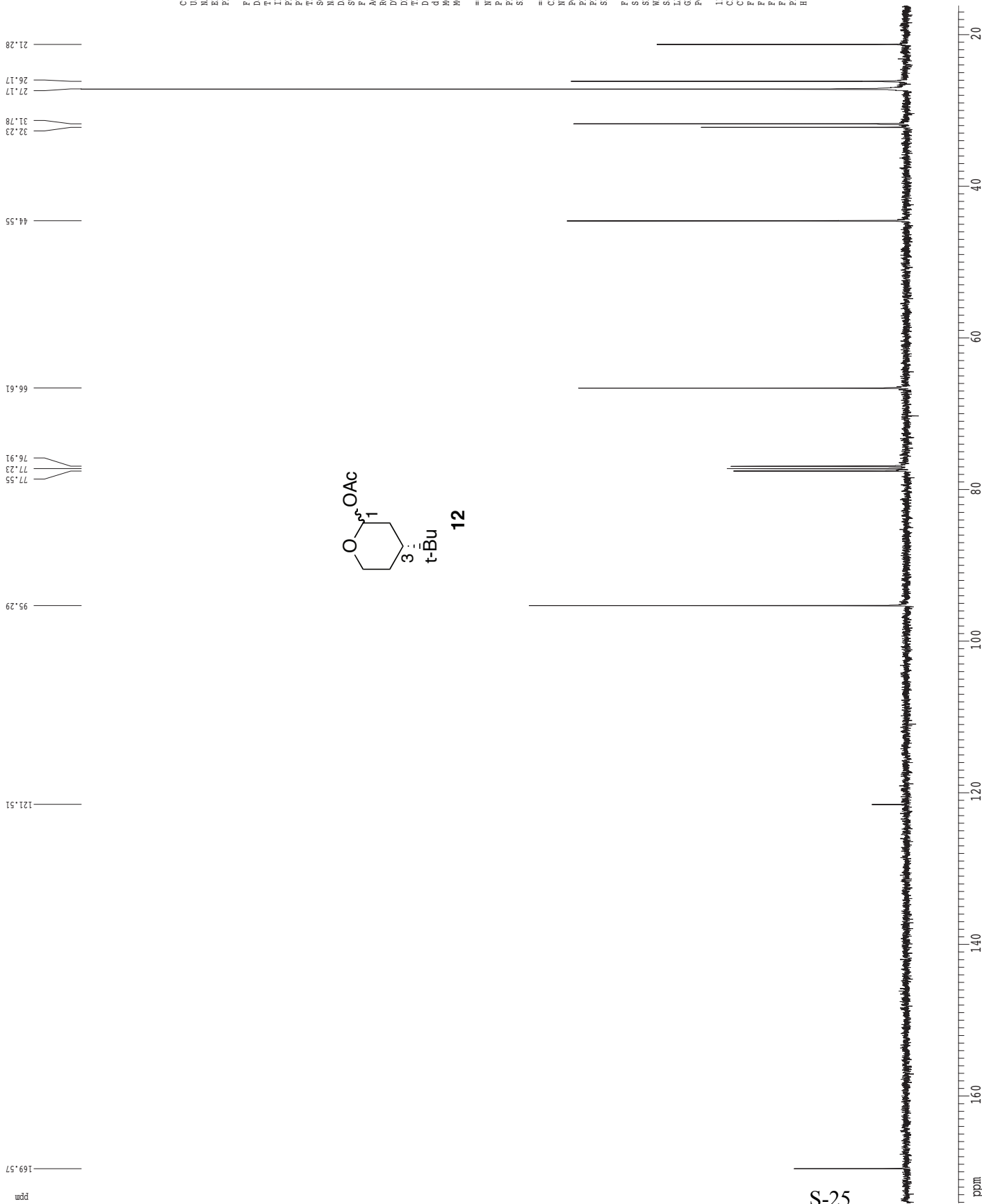
1D NMR plot parameters
 CX 22.80 cm
 CY 70.00 cm
 F1P 7.695 ppm
 F1 3078.83 Hz
 F2P 0.020 ppm
 F2 8.13 Hz
 PPMON 0.33659 ppm/cm
 PPMCN 134.67969 Hz/cm

S-24

Integral

ppm

Jrk-2-71 purified C₃-tBu acetate
 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      krump
NAME      jrk-2-71
EXPNO     14
PROCNO    1

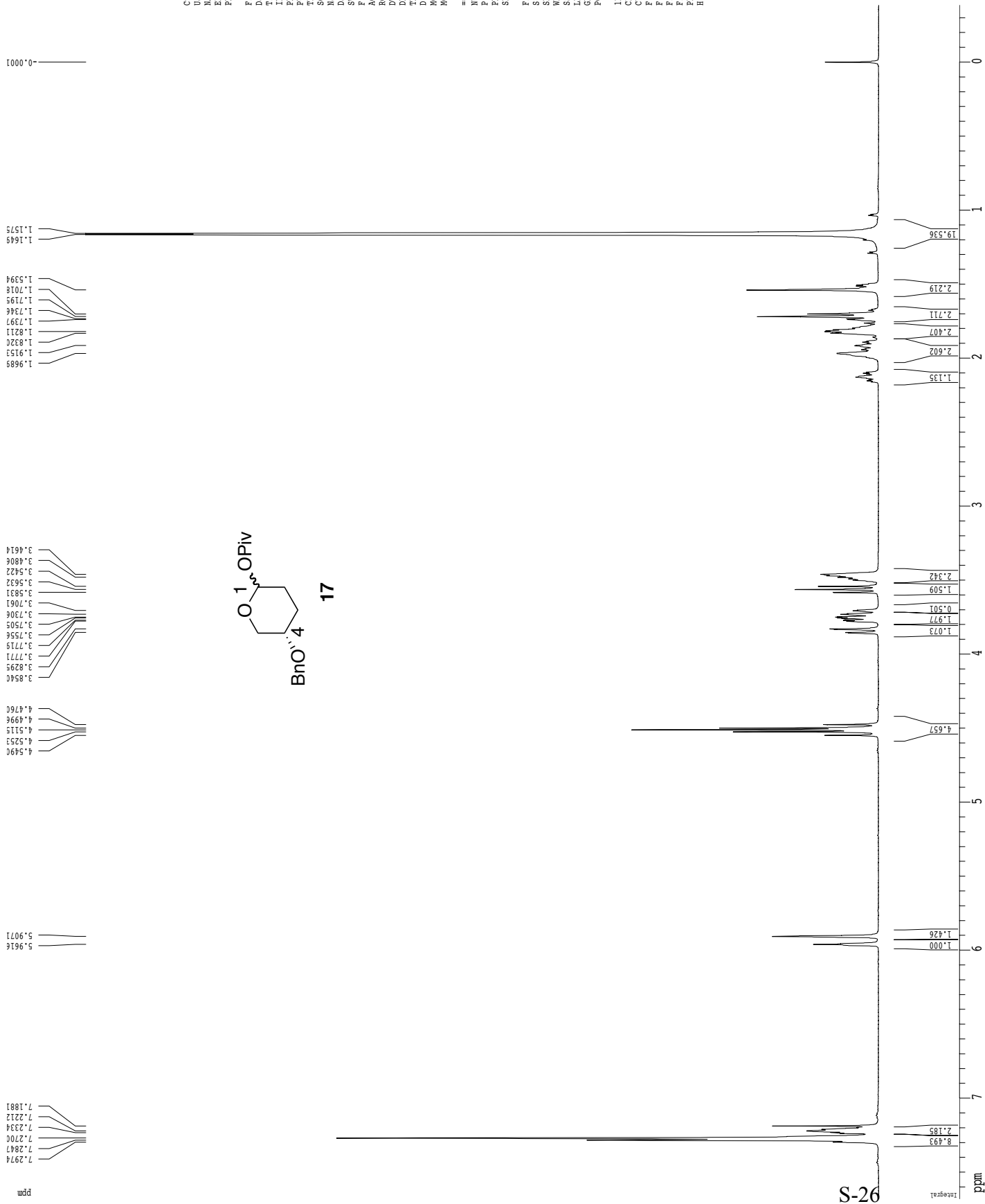
F2 - Acquisition Parameters
Date_     20080820
Time      10.26
INSTRUM   dx400
PROBHD    5 mm QNP H/F/P
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         122
DS         4
SWH        24154.50 Hz
FIDRES     0.1368570 Hz
AQ         1.3566452 sec
RG         9195.2
DW         20.700 usec
DE         20.39 usec
TE         298.0 K
D1         0.10000000 sec
d11        0.03000000 sec
MCREST     0.00000000 sec
MORPK      0.01500000 sec

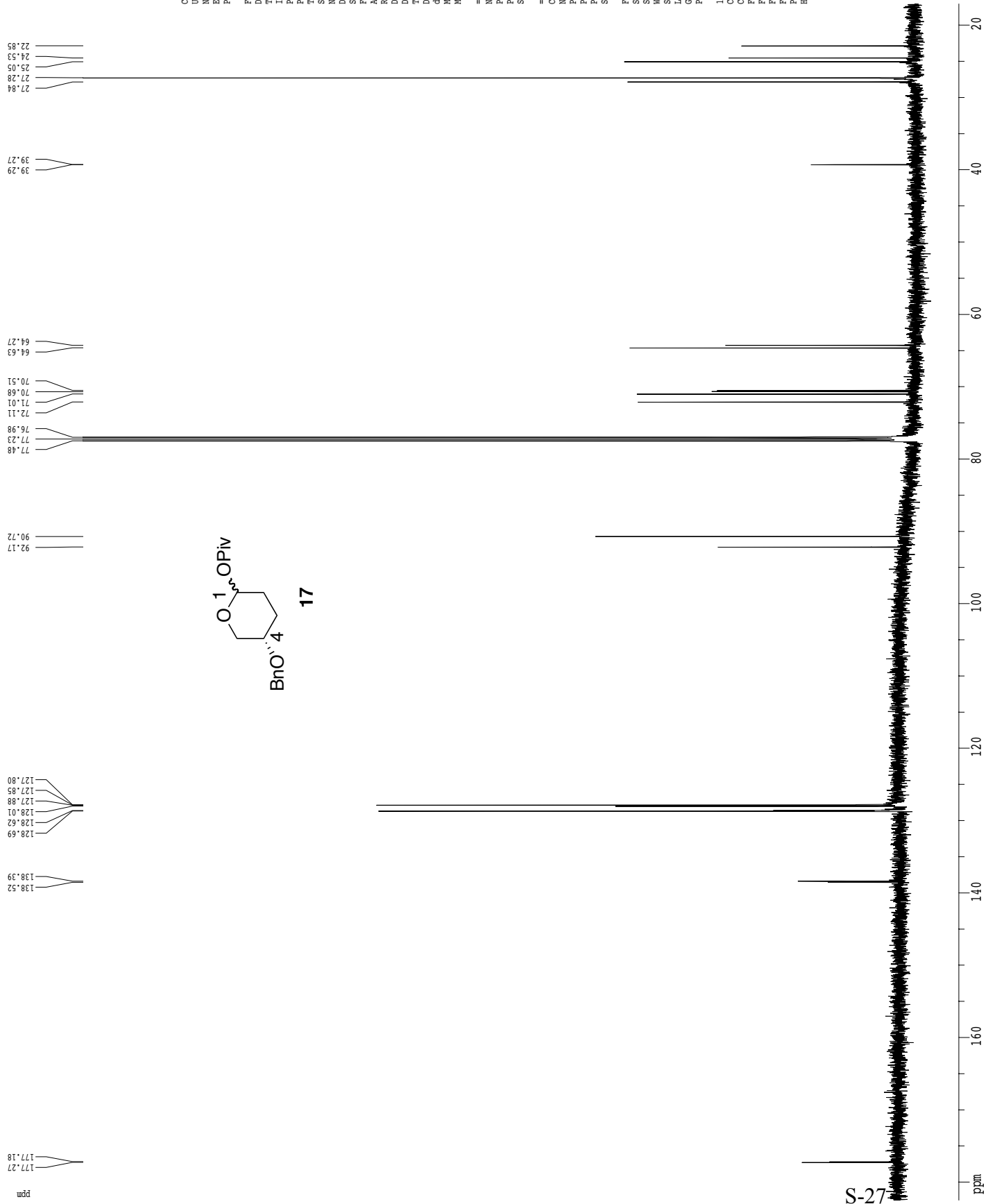
===== CHANNEL f1 =====
NUC1       13C
P1         10.30 usec
PL1        0.00 dB
SFO1       100.627964 MHz

===== CHANNEL f2 =====
CPDPRG2    mlev16
NUC2       1H
P2         90.00 usec
PL2        0.00 dB
PL12       17.70 dB
SFO2       400.1328009 MHz

F2 - Processing parameters
SI         65536
SF         100.6127576 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.00

1D NMR plot parameters
XZ        22.80 cm
Y1         174.107 ppm
Y2         175.1735 Hz
F1         16.177 ppm
F2         1627.57 Hz
PRNCHN    6.92675 ppm/cm
HZCN      696.91992 Hz/cm
  
```





Current Data Parameters
 USER krumpe
 NAME Jrk-2-54
 EXPNO 114
 PROCNO 1

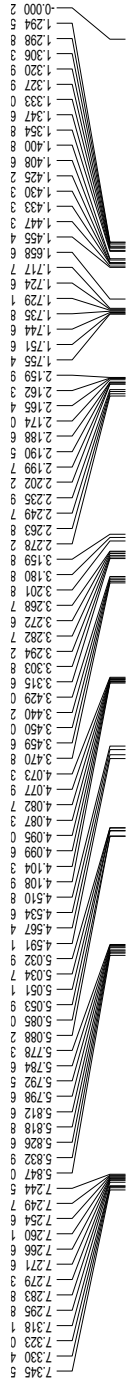
F2 - Acquisition Parameters
 Date_ 20080429
 Time 16.06
 INSTRUM cryo500
 PROBD 5 mm CPTCI LH-
 PULPROG zgpg30
 TD 6536
 SOLVENT CDCl3
 DS 114
 SFO1 125.7604011 MHz
 SF 125.7604011 MHz
 FIDRES 0.462388 Hz
 AQ 1.0813940 sec
 RG 13004
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MCHRG 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 15.00 usec
 PL1 23.00 dB
 SFO1 125.7604011 MHz

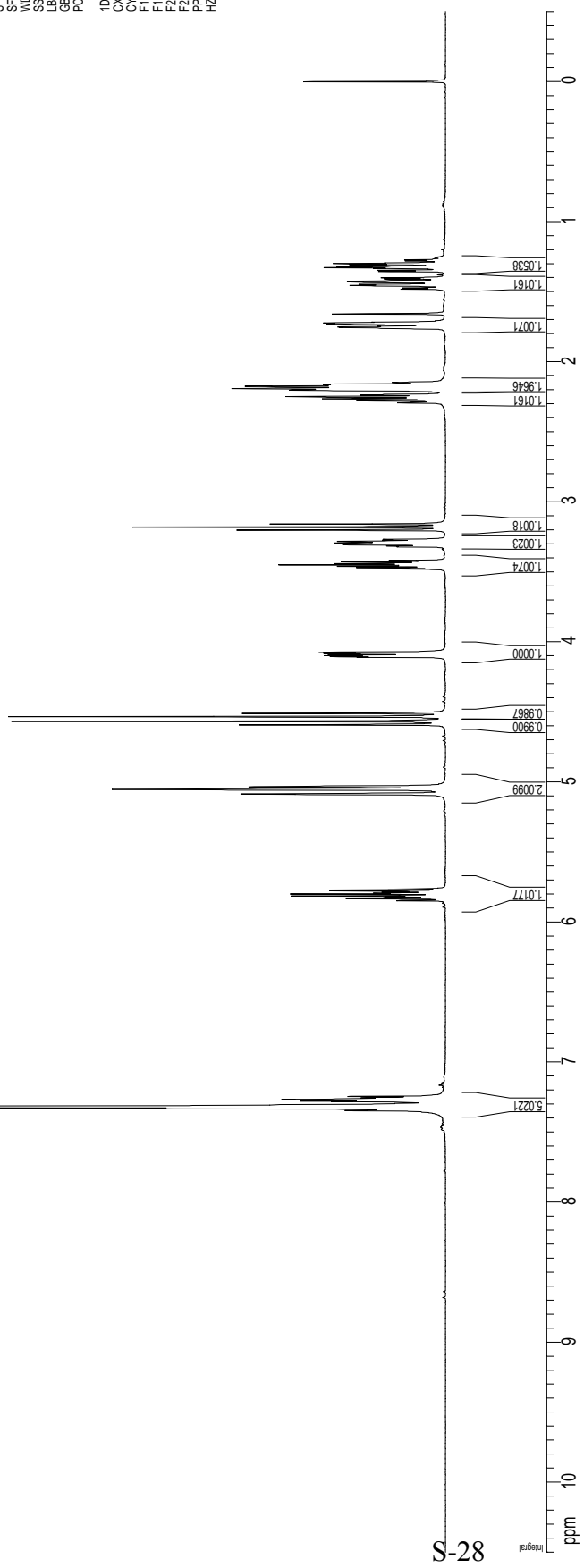
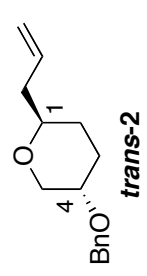
==== CHANNEL f2 =====
 CPDPRG2 wahtz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 23.54 dB
 SFO2 500.225011 MHz

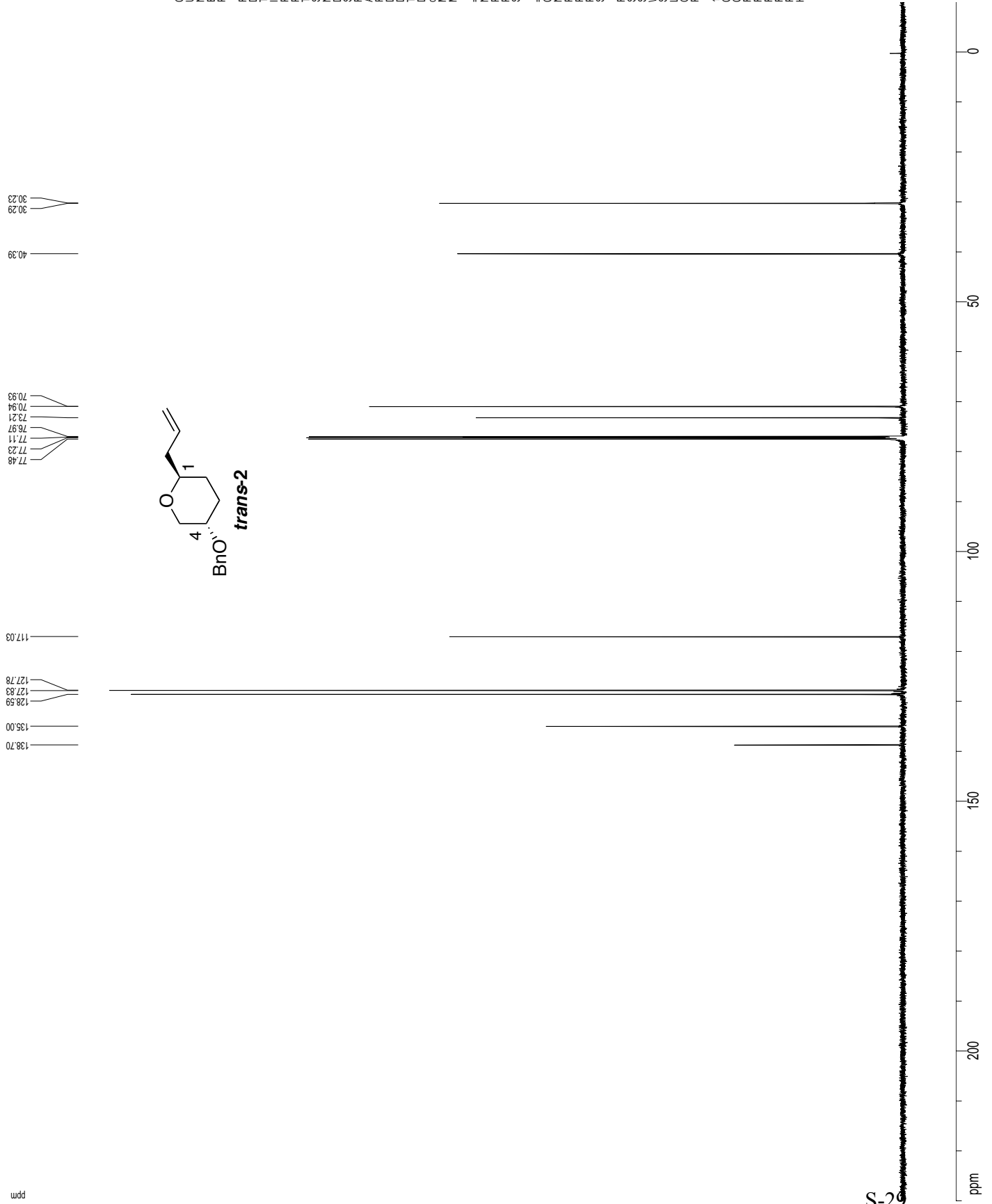
F2 - Processing parameters
 SI 65536
 SF 125.7604011 MHz
 WDW EN
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 35.00 cm
 FIP 182.472 ppm
 F1 22951.37 Hz
 F2P 17.072 ppm
 F2 2147.31 Hz
 PPMCH 7.25438 ppm/cm
 HZCH 912.45911 Hz/cm



Current Data Parameters
 USER walter
 NAME WAS-III-177-23
 EXPNO 1
 PROCNO 1
 F2-Acquisition Parameters
 Date_ 20071030
 Time 17:58
 INSTRUM cryo500
 PROBHD 5 mm CPTCI-1H-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3T
 NS 1
 DS 0
 SVH 8072.820 Hz
 FIDRES 0.086043 Hz
 AQ 5.0993986 sec
 SFO1 500.136099 MHz
 DQ 62.400 usec
 DE 288.0 K
 TE 6.00 usec
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCVRK 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 1.80 dB
 SFO1 500.2250015 MHz
 F2-Processing parameters
 SI 32768
 SF 500.220386 MHz
 SSF 65.535 usec
 WDW EM
 EN 0
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00
 1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 FIP 10.500 ppm
 F2P 9262.31 Hz
 F3P 9.000 ppm
 F2 -250.11 Hz
 PPMCM 0.48246 ppm/cm
 HZCM 241.33423 Hz/cm





Current Data Parameters
USER walter
NAME WAS-III-177-23
EXPNO 3
PROCNO 1

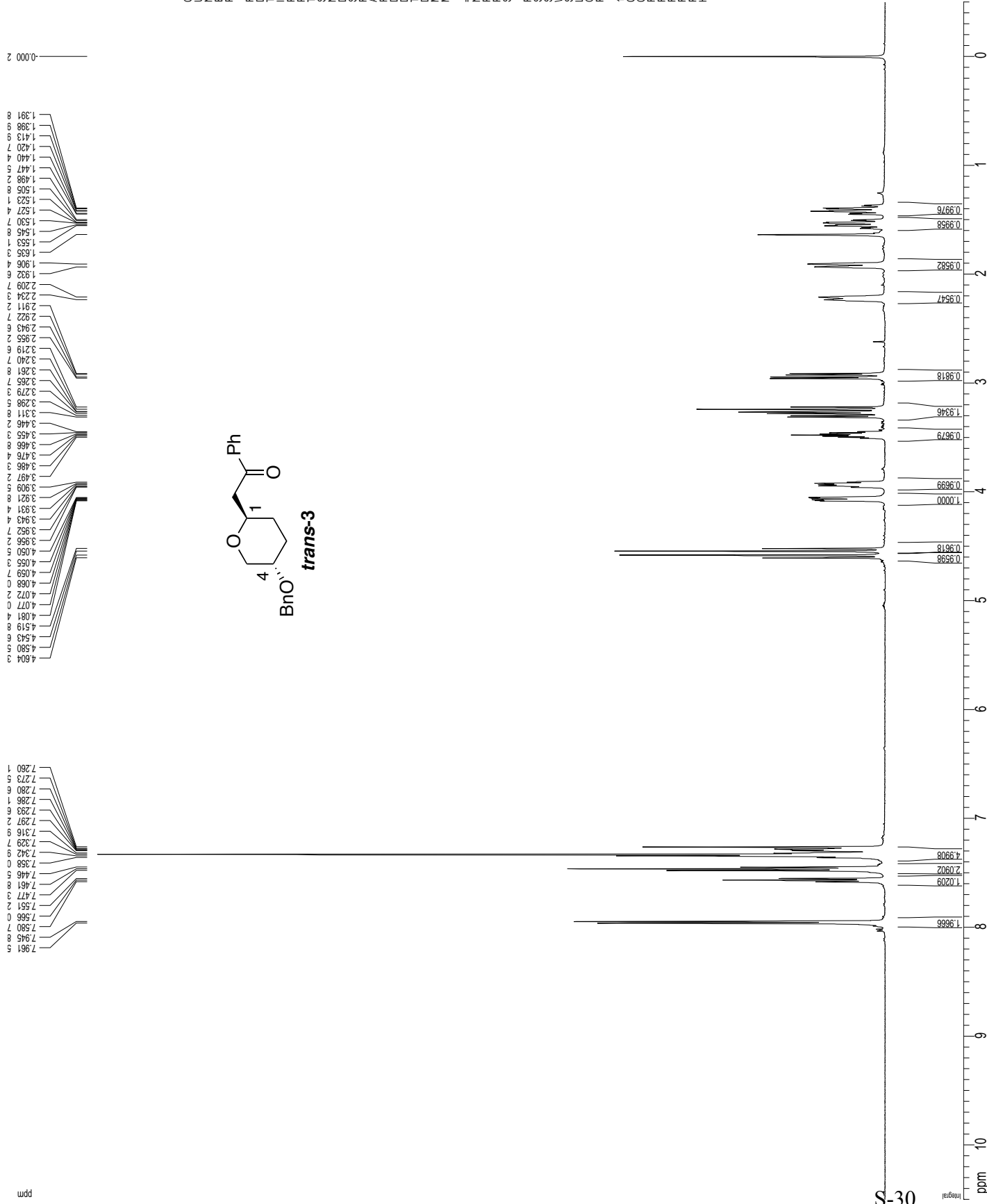
F2 - Acquisition Parameters
Date_ 20071030
Time 18.01
INSTRUM cryo500
PROBHD 5 mm CPTC 1H-
PULPROG zgpg30
TD 65472
SOLVENT CDCl3
NS 65
DS 4
SWH 30303.031 Hz
FIDRES 0.463222 Hz
AQ 1.0794635 sec
RG 8192
DW 16.500 usec
DE 6.00 usec
TE 288.0 K
D1 0.25000000 sec
d11 0.03000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

==== CHANNEL f1 =====
NUC1 13C
P1 15.00 usec
PL1 -1.00 dB
SFO1 125.760462 MHz

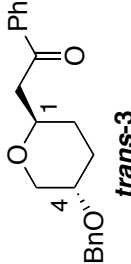
==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.60 dB
PL12 23.54 dB
SFO2 500.225011 MHz

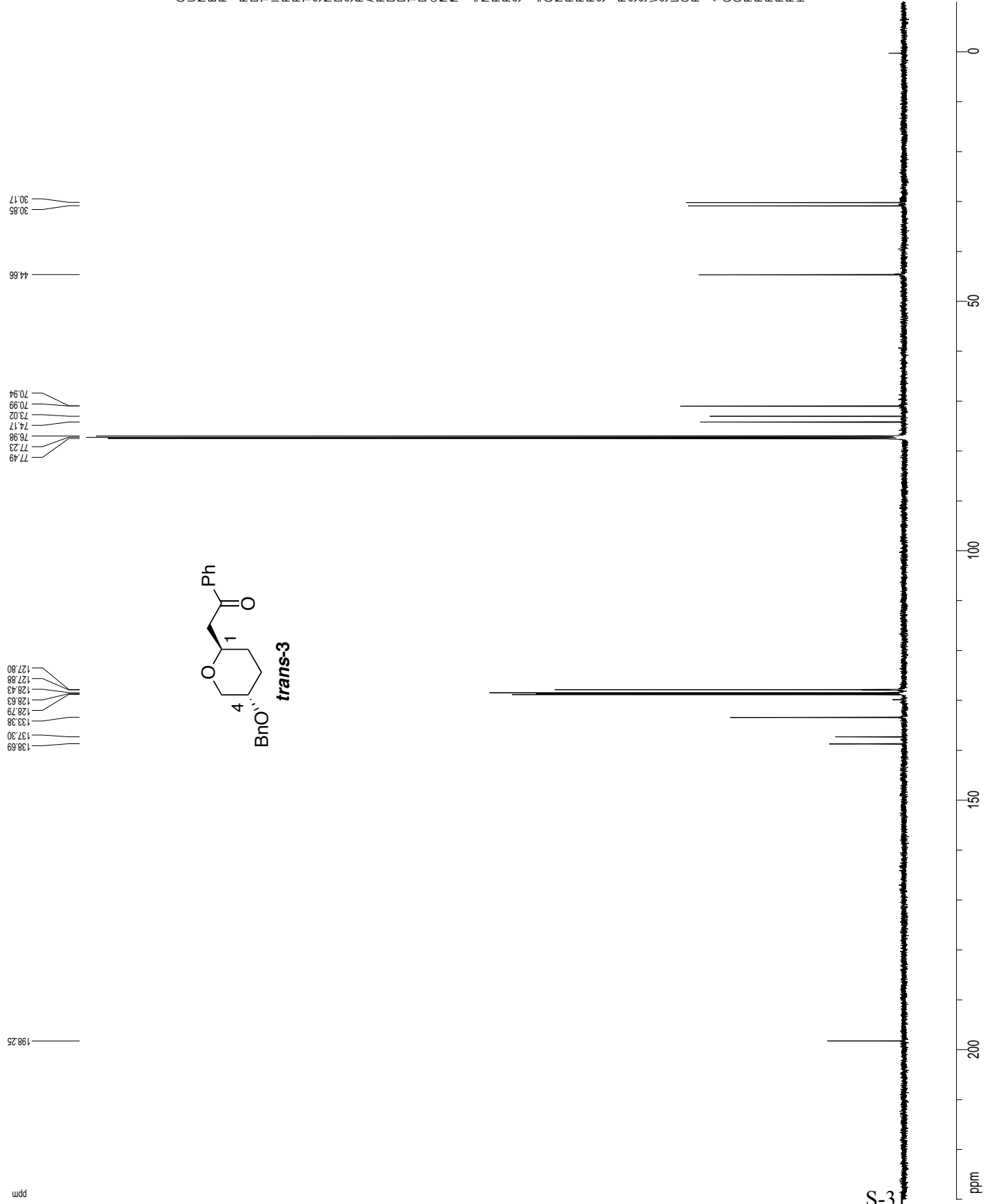
F2 - Processing parameters
SI 65536
SF 125.760462 MHz
WDW EM
SSB 0
GB 1.00 Hz
CB 2.00
PC

1D NMR parameters
CX 22.80 cm
CY 15.65 cm
F1P 230.000 ppm
F1 28929.49 Hz
F2P -10.000 ppm
F2 -1257.80 Hz
PPH1CM 10.52632 ppm/cm
HZCM 1324.00427 Hz/cm



Current Data Parameters
 USER walter
 NAME WAS-III-31-25
 EXPNO 31
 PROCNO 1
 F2-Acquisition Parameters
 Date_ 20070221
 Time 9:45
 INSTRUM cry500
 PROBHD 5 mm CPTCI-1H
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3T
 NS 8
 DS 0
 SWH 8072.820 Hz
 FIDRES 0.058043 Hz
 AQC 5.0899774 sec
 RG 4.5
 DW 6.400 usec
 DE 6.000 usec
 TE 290.1 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCVRK 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 8.00 usec
 PL1 1.60 dB
 SFO1 500.2250015 MHz
 F2-Processing parameters
 SI 32768
 SF 500.220319 MHz
 SD 6500 Hz
 WDW EM
 EN 0
 LB 0.30 Hz
 GB 0
 PC 4.00
 1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 10.500 ppm
 F2P 5252.31 Hz
 F3P 0.000 ppm
 F2 -250.11 Hz
 PPMCM 0.48246 ppm/cm
 HZCM 241.33423 Hz/cm





Current Data Parameters
 USER: walter
 NAME: WAS-III-31-25
 EXPNO: 33
 PROCNO: 1

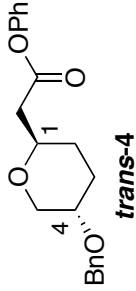
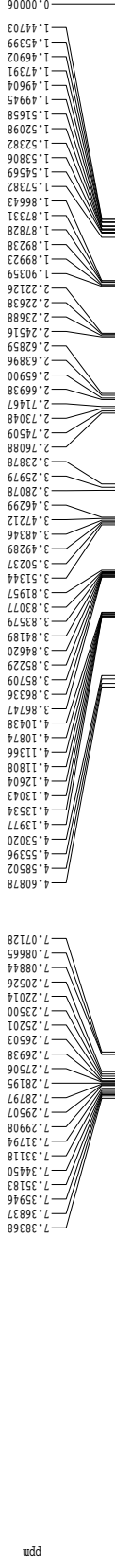
F2 - Acquisition Parameters
 Date_: 20070621
 Time: 9:53
 INSTRUM: cryo500
 PULPROG: zgpg30
 TD: 65472
 SOLVENT: CDCl3
 NS: 225
 DS: 4
 SWH: 30303.031 Hz
 FIDRES: 0.463222 Hz
 AQ: 1.0794470 sec
 RG: 11585.2
 DW: 16.500 usec
 DE: 6.00 usec
 TE: 288.0 K
 d1: 0.25000000 sec
 d11: 0.03000000 sec
 MCREST: 0.00000000 sec
 MCWRR: 0.01500000 sec

==== CHANNEL f1 =====
 NUC1: ¹³C
 P1: 15.00 usec
 PL1: -1.00 dB
 SFO1: 125.762548 MHz

==== CHANNEL f2 =====
 CPDPRG2: waltz16
 NUC2: ¹H
 PCPD2: 100.00 usec
 PL2: 1.60 dB
 PL12: 23.54 dB
 SFO2: 500.225011 MHz

F2 - Processing parameters
 SI: 65536
 SF: 125.7604015 MHz
 WDW: EM
 SSB: 0
 GB: 1.00 Hz
 GC: 0
 PC: 2.00

1D NMR parameters
 CX: 22.80 cm
 CY: 15.65 cm
 F1P: 230.000 ppm
 F1: 28929.49 Hz
 F2P: -10.000 ppm
 F2: -1257.80 Hz
 PPMICM: 10.52632 ppm/cm
 HZCM: 1324.00427 Hz/cm



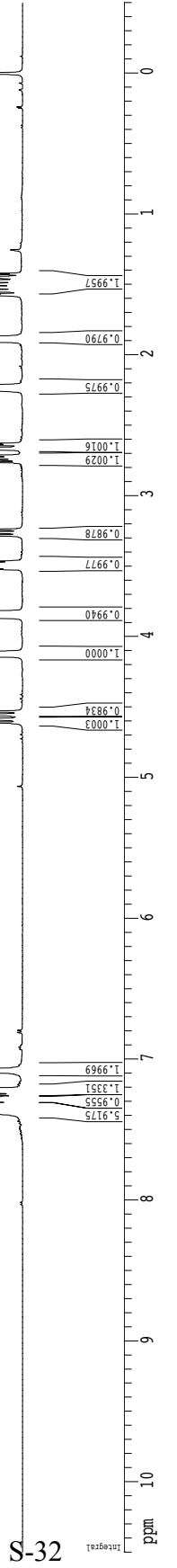
Current Data Parameters
USER: walter
NAME: WAS-III-154-28
EXPNO: 3
PROCNO: 1

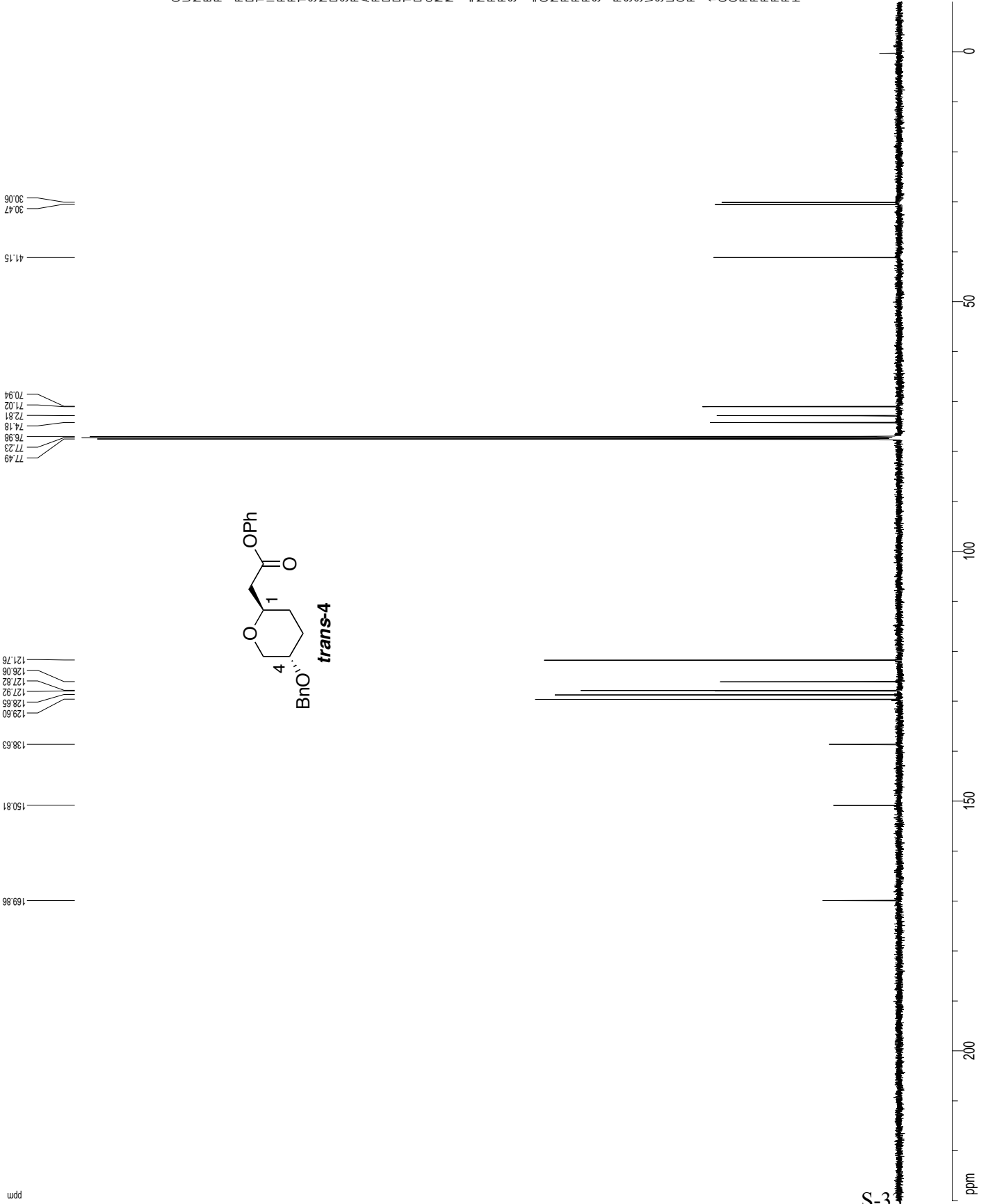
F2 - Acquisition Parameters
Date_ : 20080403
Time : 23.43
INSTRUM: cryso00
PROBHD: 5 mm CPXI 1H-
PULPROG: zg30
TD: 81728
SOLVENT: CDCl3T
NS: 8
DS: 0
SWH: 8012.920 Hz
FIDRES: 0.090043 Hz
AQ: 5.099999 sec
RG: 3
SFO1: 500.136099 MHz
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 4.00

1D NMR plot parameters
CX: 22.80 cm
CY: 15.00 cm
FLP: 10.500 ppm
F1: 52.8231 Hz
F2: -2.0311 ppm
FPCW: 0.88246 ppm/cm
HZCN: 241.33823 Hz/cm

==== CHANNEL f1 =====
NUC1: 1H
P1: 8.00 usec
PL1: 1.60 dB
SFO1: 500.225015 MHz

F2 - Processing parameters
SI: 32768
SF: 500.220351 MHz
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 4.00





Current Data Parameters
 USER walter
 NAME WAS-III-154-28
 EXPNO 2
 PROCNO 1

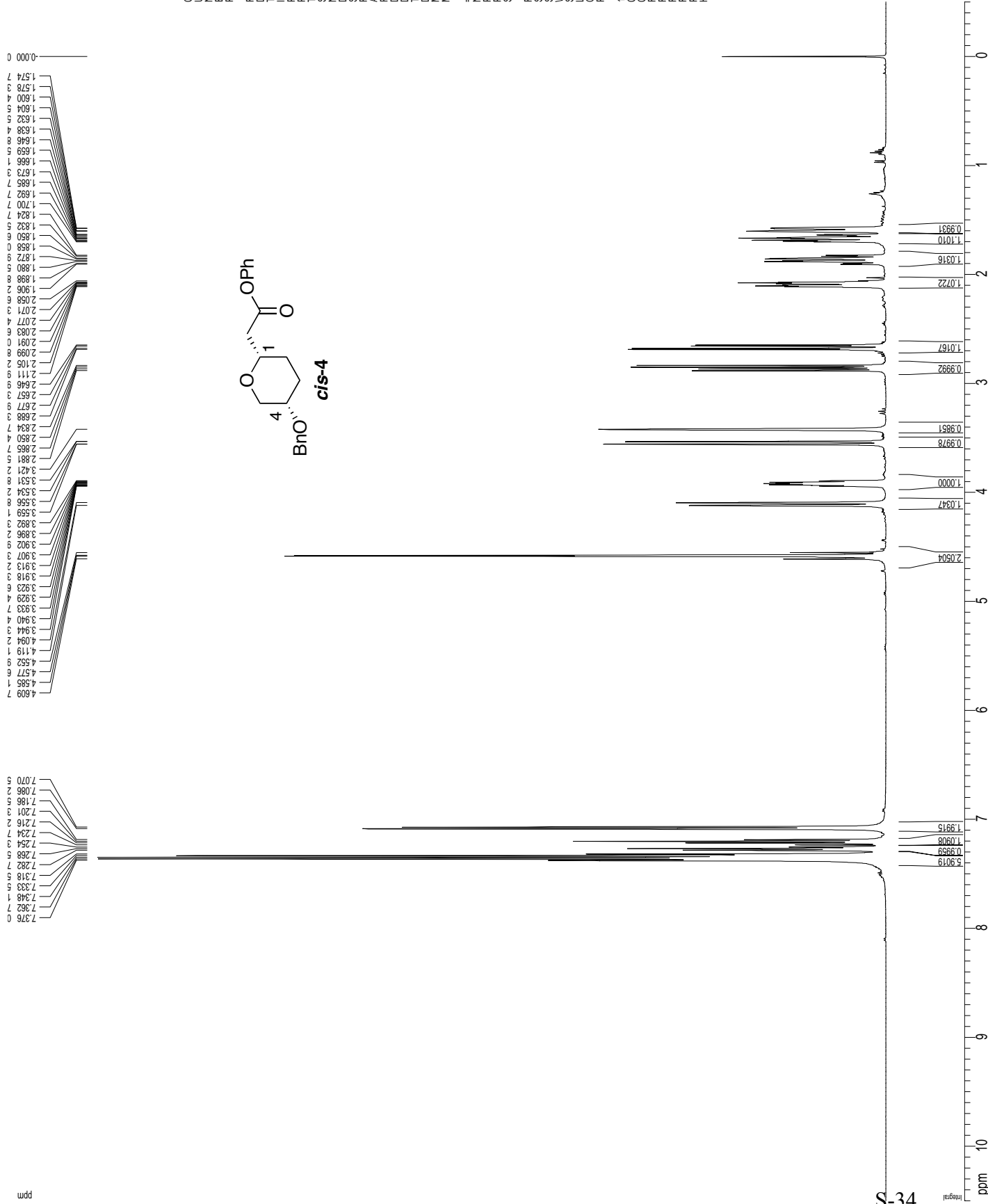
F2 - Acquisition Parameters
 Date_ 20071002
 Time 16.37
 INSTRUM cryo500
 PULPROG zgpg30
 TD 65472
 SOLVENT CDCl3
 NS 112
 DS 0
 SWH 30303.031 Hz
 FIDRES 0.463222 Hz
 AQ 1.0794635 sec
 RG 10321.3
 DW 16.500 usec
 DE 6.00 usec
 TE 288.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 15.00 usec
 PL1 -1.00 dB
 SFO1 125.762548 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 23.54 dB
 SFO2 500.225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7604015 MHz
 WDW EM
 SSB 0
 GB 1.00 Hz
 PC 2.00

1D NMR parameters
 CX 22.80 cm
 CY 15.65 cm
 F1 230.000 ppm
 F2 28929.49 Hz
 F2P -10.000 ppm
 F2 -1257.80 Hz
 PPM1CM 10.52632 ppm/cm
 HZCM 1324.00427 Hz/cm



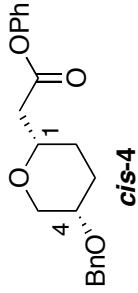
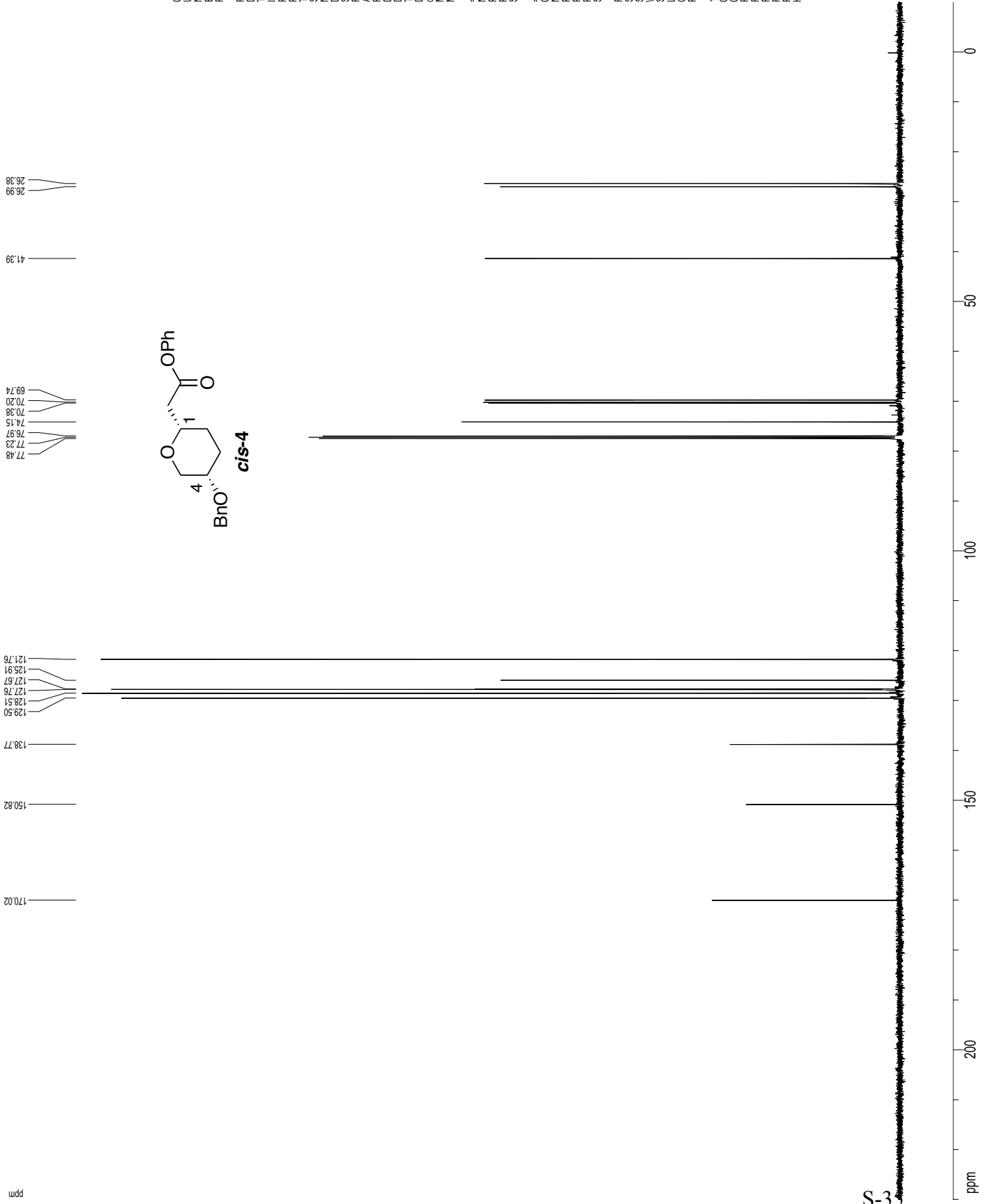
Current Data Parameters
 USER: walter
 NAME: WAS-III-154-30
 EXPNO: 3
 PROCNO: 1

F2 - Acquisition Parameters
 Date_ 200702
 Time 23.48
 INSTRUM: cryo500
 PROBHD: 5 mm CPTCI-1H-
 PULPROG: zg30
 TD: 81728
 SOLVENT: CDCl3T
 NS: 8
 DS: 0
 SWH: 8072.820 Hz
 FIDRES: 0.086043 Hz
 AQ: 5.095986 sec
 RG: 37.700
 DW: 6.400 usec
 DE: 6.000 usec
 TE: 298.0 K
 D1: 0.4000000 sec
 MCREST: 0.0000000 sec
 MCVRK: 0.0150000 sec

==== CHANNEL f1 =====
 NUC1: 1H
 P1: 8.00 usec
 PL1: -1.80 dB
 SFO1: 500.2250015 MHz

F2 - Processing parameters
 SF: 500.220436 MHz
 DSF: 65.000000 MHz
 WDW: EM
 SSB: 0
 LB: 0.30 Hz
 GB: 0
 PC: 4.00

1D NMR plot parameters
 CX: 22.80 cm
 CY: 15.00 cm
 F1P: 10.500 ppm
 F2P: 5252.31 Hz
 F3P: 0.000000 ppm
 F2: -250.11 Hz
 PPMCM: 0.48246 ppm/cm
 HZCM: 241.33423 Hz/cm



Current Data Parameters
 USER walter
 NAME WAS-III-154-30
 EXPNO 2
 PROCNO 1

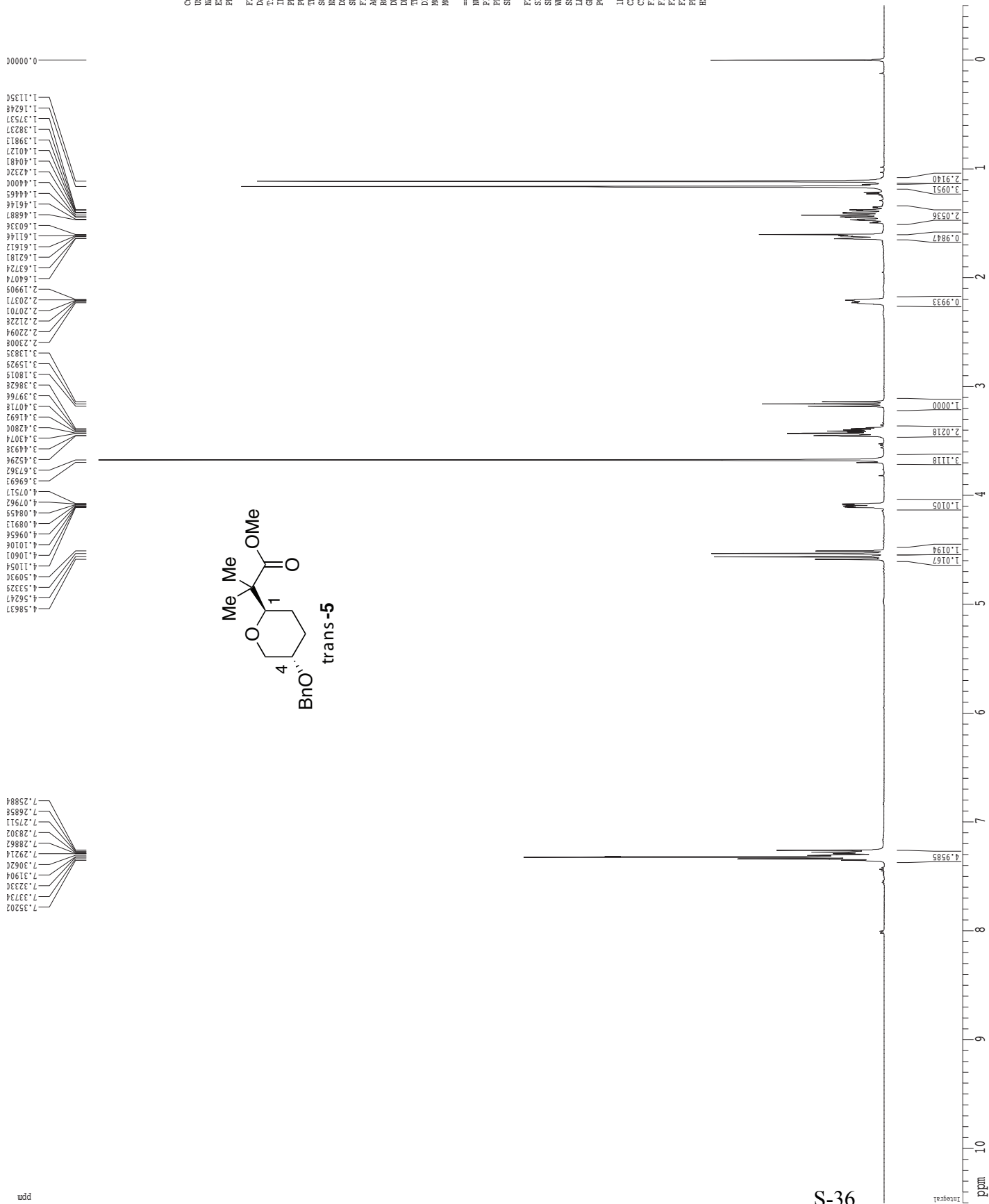
F2 - Acquisition Parameters
 Date_ 20071002
 Time 16.47
 INSTRUM cryo500
 PROBO 5 mm CPTC 1H-
 PULPROG zgpg30
 TD 65428
 SOLVENT CDCl3
 NS 76
 DS 0
 SWH 30003.031 Hz
 FIDRES 0.463222 Hz
 AQ 1.0794635 sec
 RG 13004
 DW 16.500 usec
 DE 6.00 usec
 TE 288.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

==== CHANNEL f1 =====
 NUC1 13C
 P1 15.00 usec
 PL1 -1.00 dB
 SFO1 125.762548 MHz

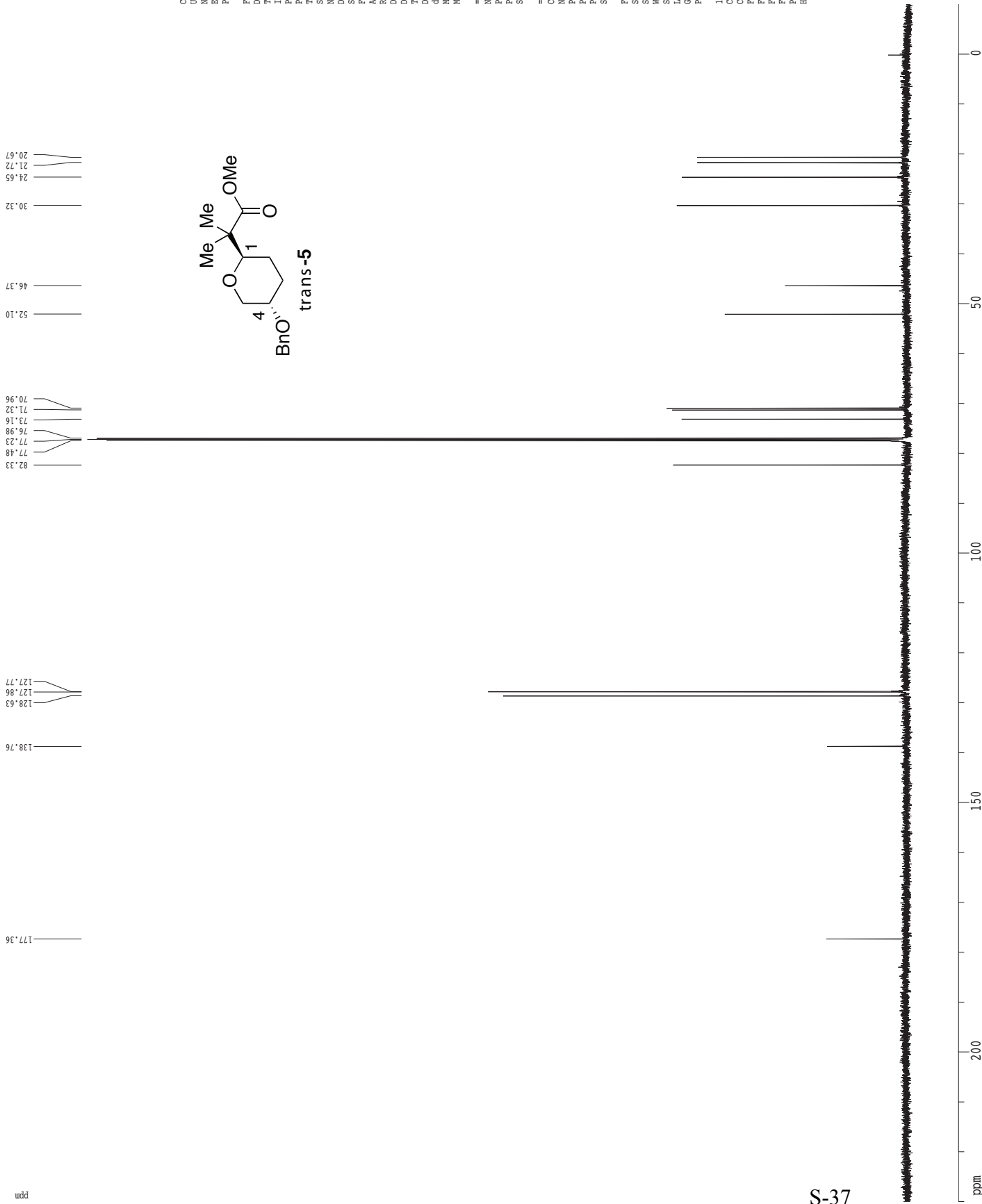
==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 23.54 dB
 SFO2 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7604112 MHz
 WDW EM
 SSB 0
 GB 1.00 Hz
 PC 2.00

1D NMR ph2 parameters
 CX 22.80 cm
 CY 15.65 cm
 F1P 230.000 ppm
 F1 28929.49 Hz
 F2P -10.000 ppm
 F2 -1257.80 Hz
 PPM1CM 10.52632 ppm/cm
 HZCM 1324.00439 Hz/cm



WAS-IV_17-35
 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      walter
NAME      WAS-IV-17-35
EXPNO     2
PROCNO    1

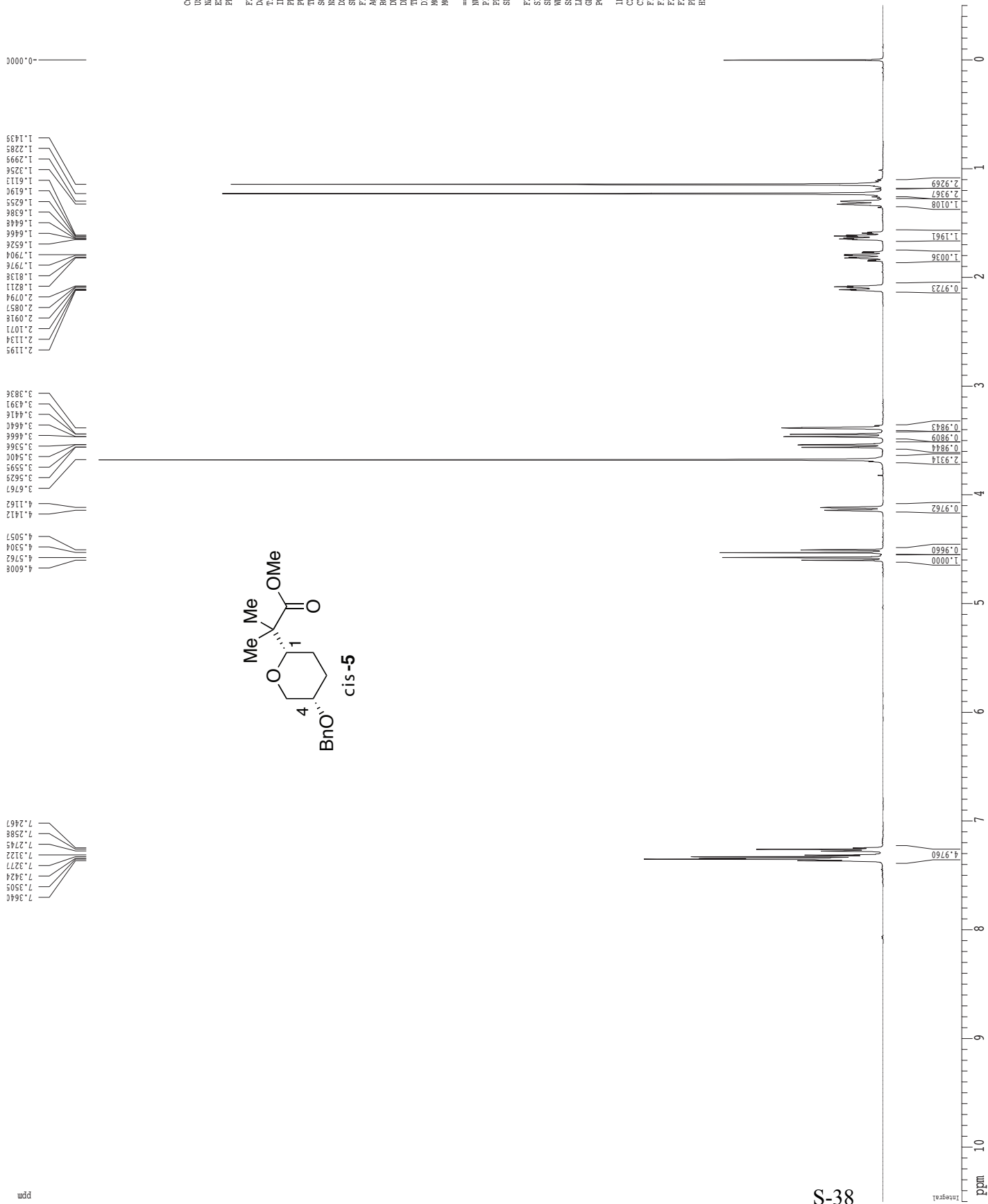
F2 - Acquisition Parameters
Date_     20080408
Time      11.07
INSTRUM   cryo500
PROBHD    5 mm CPXI.H-
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         106
DS         3003.03 Hz
SWH        0.246238 Hz
FIDRES     1.6814105 sec
AQ         1.0004
RG         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
MCREST     0.00000000 sec
MORPK      0.01500000 sec

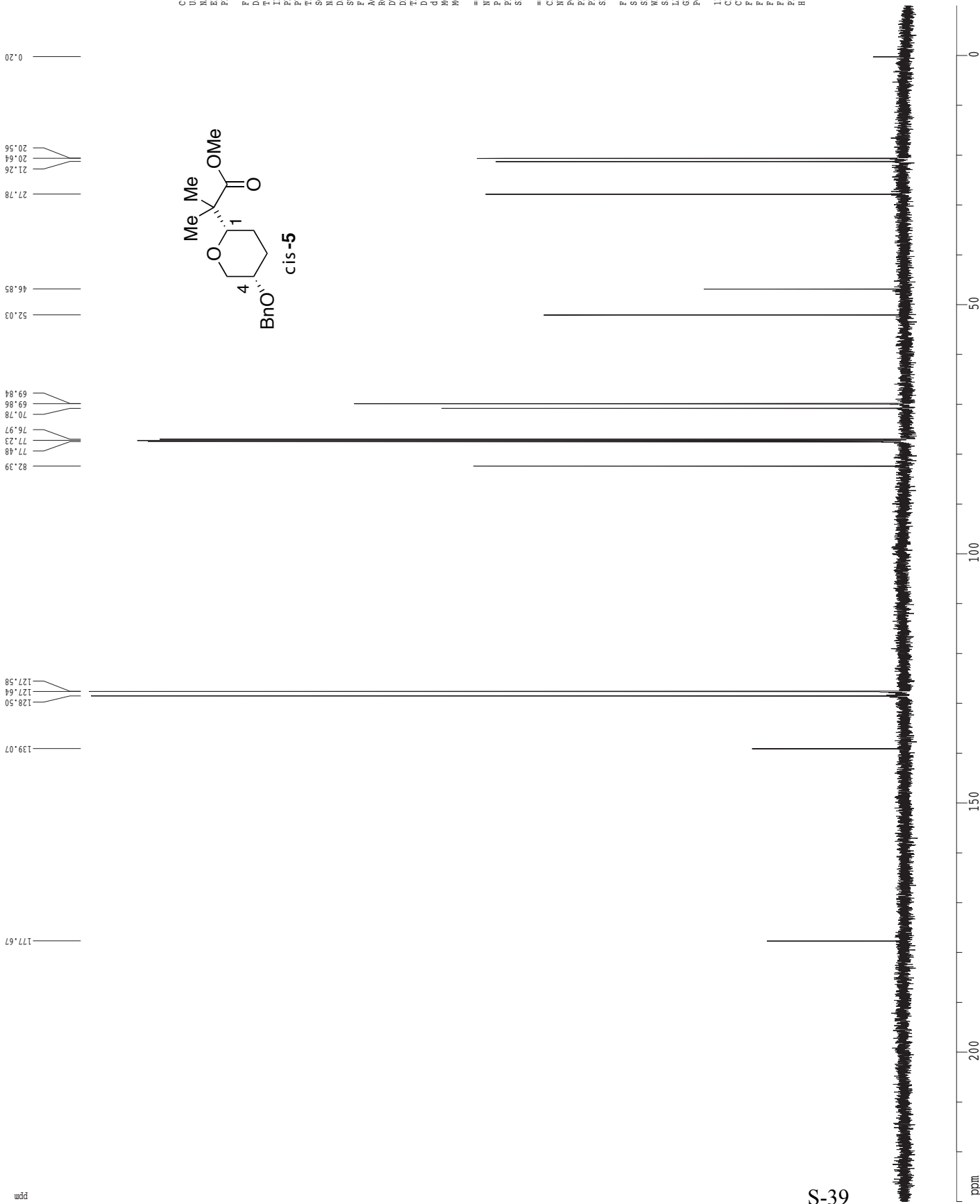
===== CHANNEL f1 =====
NUC1       13C
P1         14.75 usec
PL1        -1.00 dB
SFO1       125.7942348 MHz

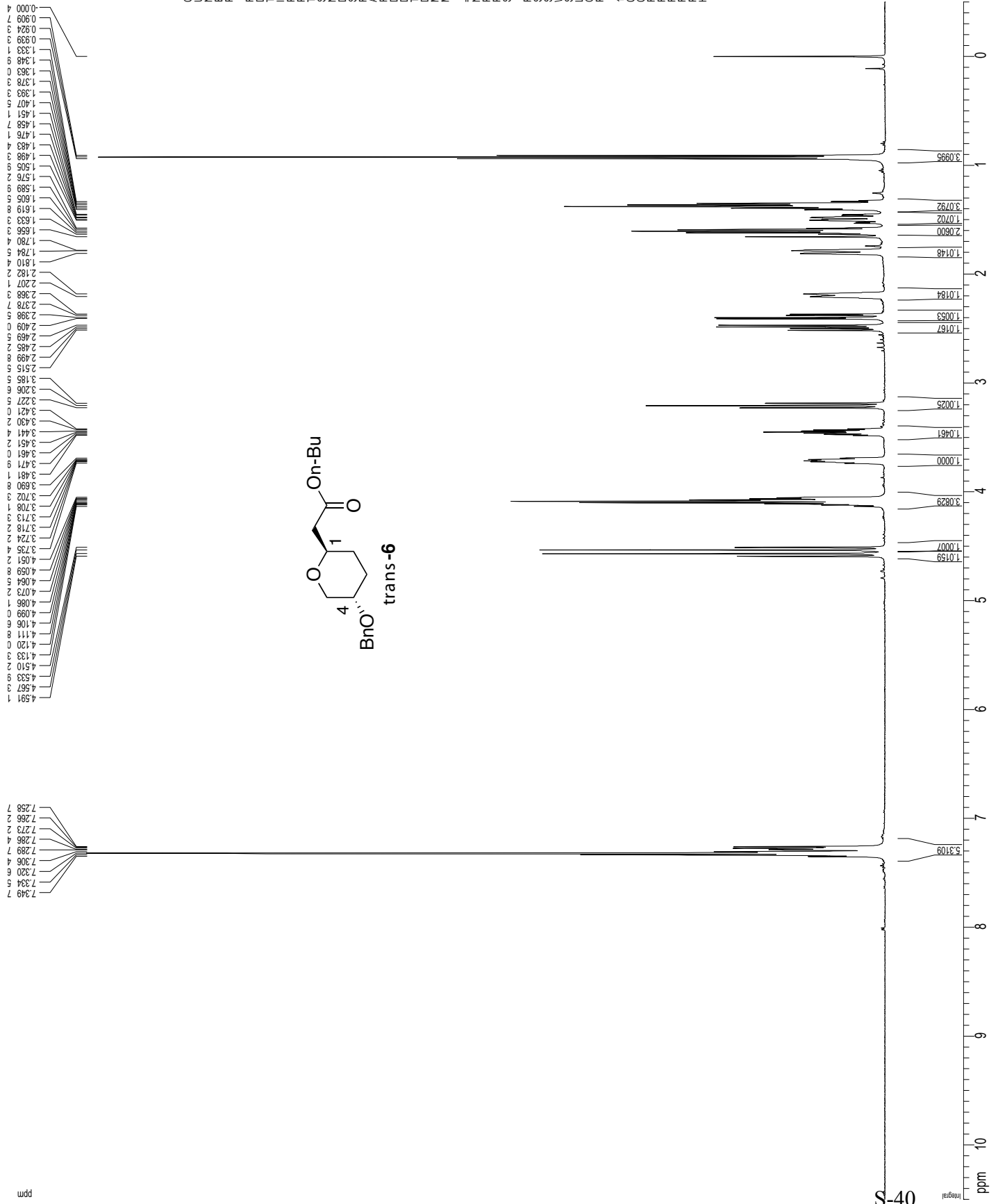
===== CHANNEL f2 =====
CPDPRG2    waltz16
NUC2
P2         100.00 usec
PL2        -1.60 dB
PL12       24.80 dB
SFO2       500.225011 MHz

F2 - Processing parameters
SI         65536
SF         125.7844006 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00

ID NMR plot parameters
X         72.80 cm
Y         72.80 cm
F1        230.000 ppm
F2        28929.49 Hz
F3        -10.000 ppm
F4        -1257.80 Hz
PRNCM     10.52632 ppm/cm
HZCM      1324.00477 Hz/cm
  
```







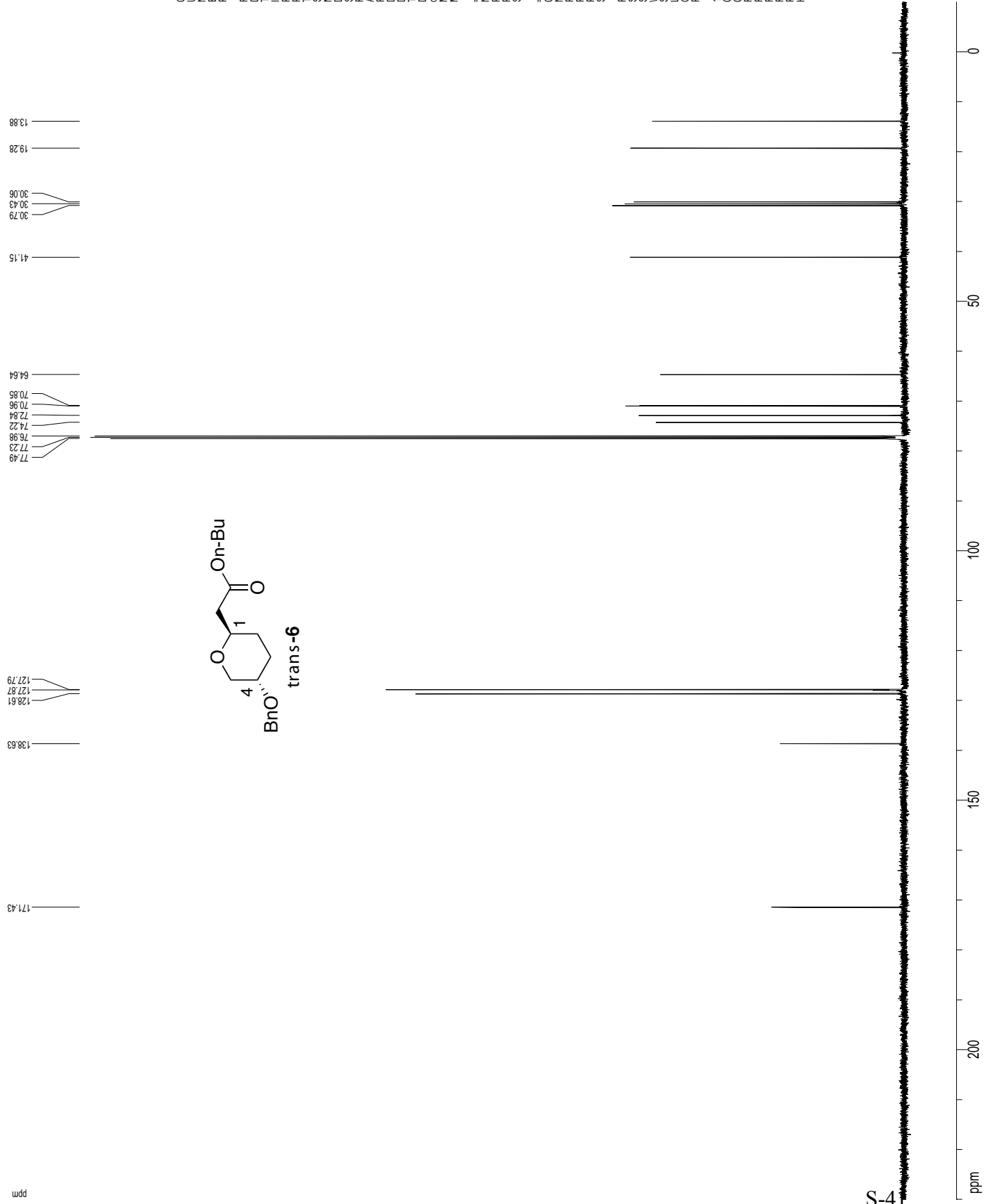
Current Data Parameters
 USER: walter
 NAME: WAS-III-97-25
 EXPNO: 5
 PROCNO: 1

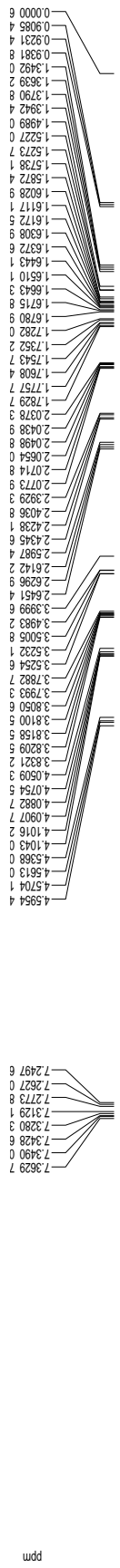
F2 - Acquisition Parameters
 Date_: 20070516
 Time: 14.43
 INSTRUM: crys500
 PROBHID: 5 mm CPTCI-1H-
 PULPROG: zg30
 TD: 81728
 SOLVENT: CDCl3T
 NS: 1
 DS: 0
 SWH: 8072.820 Hz
 FIDRES: 0.086043 Hz
 AQ: 5.0889774 sec
 SFO1: 500.136099 MHz
 DQ: 6.300 usec
 DE: 6.000 usec
 TE: 298.0 K
 D1: 0.10000000 sec
 MCREST: 0.00000000 sec
 MCVRK: 0.01500000 sec

==== CHANNEL f1 =====
 NUC1: 1H
 P1: 8.00 usec
 PL1: 1.60 dB
 SFO1: 500.2250015 MHz

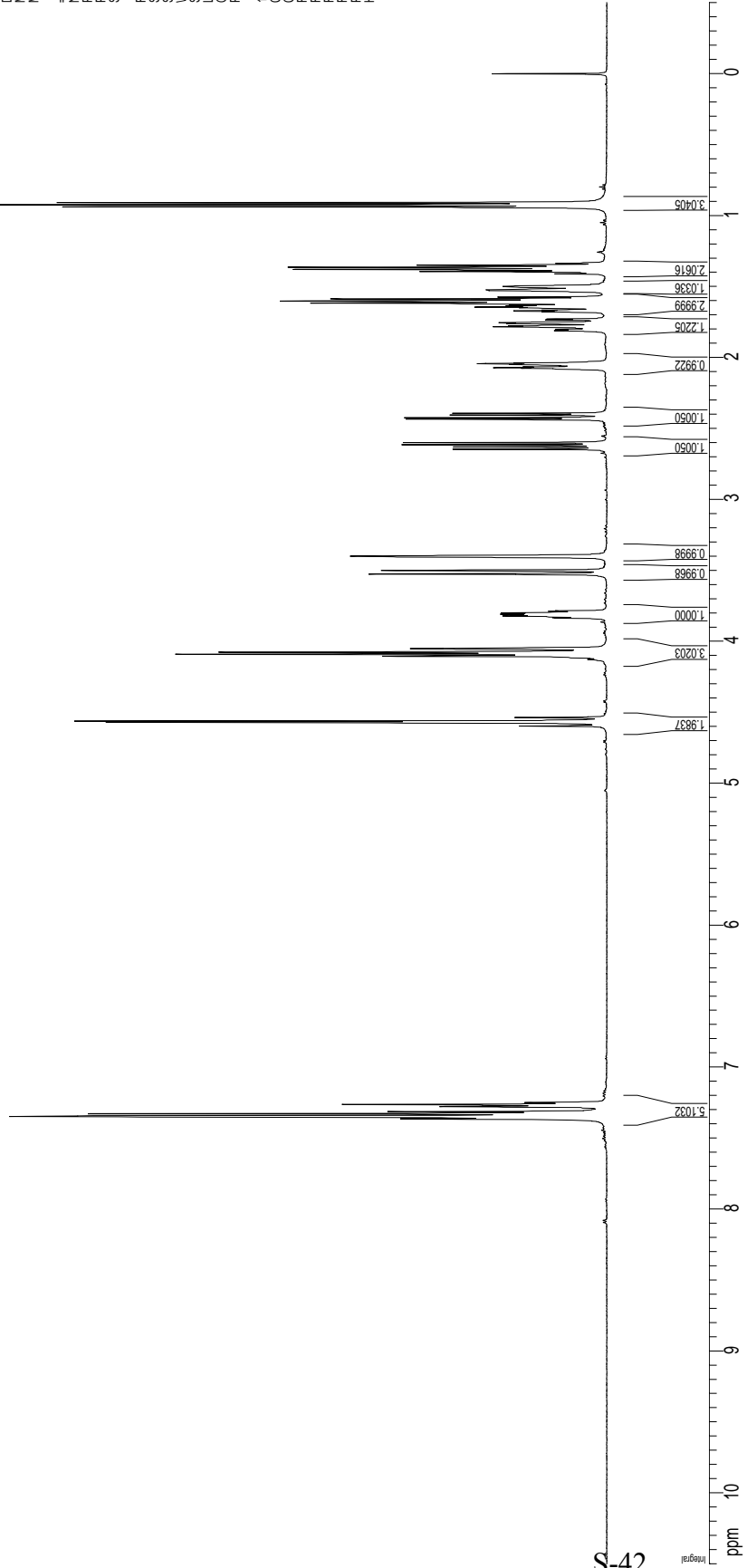
F2 - Processing parameters
 SI: 32768
 SF: 500.220324 MHz
 DSF: 65536
 WDW: EM
 SSB: 0
 LB: 0.30 Hz
 GB: 0
 PC: 4.00

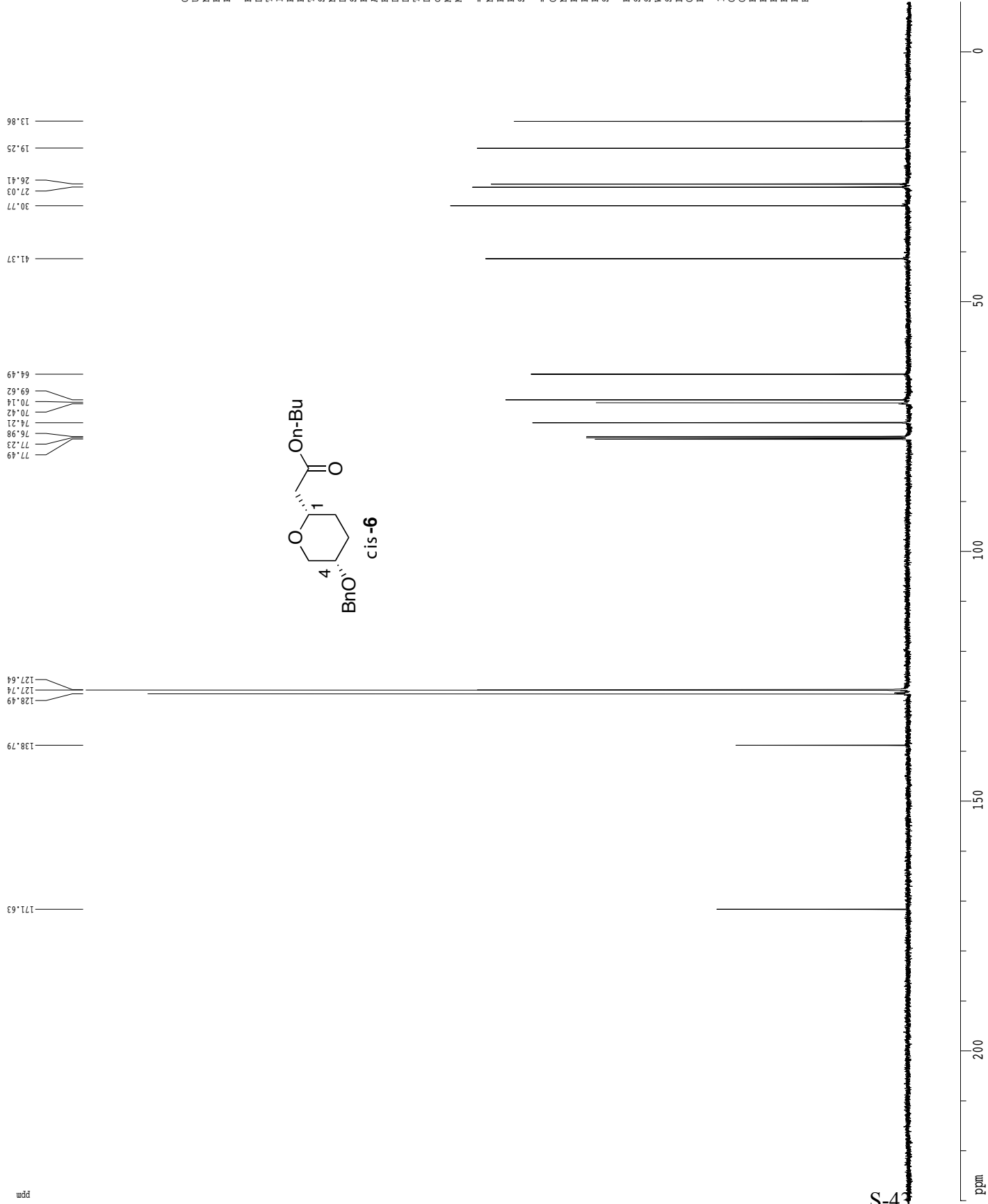
1D NMR plot parameters
 CX: 22.80 cm
 CY: 15.00 cm
 F1P: 10.500 ppm
 F2P: 5252.31 Hz
 F3P: 0.000 ppm
 FZ: -250.1 Hz
 PPM1CM: 0.48246 ppm/cm
 HZCM: 241.33423 Hz/cm

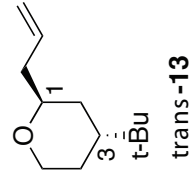




Current Data Parameters
 USER: walter
 NAME: WAS-III-97-27
 EXPNO: 5
 PROCNO: 1
 F2-Acquisition Parameters
 Date_ 20070516
 Time 14.36
 INSTRUM: cryo500
 PROBHD: 5 mm CPTCI 1H-
 PULPROG: zg30
 TD: 81728
 SOLVENT: CDCl3T
 NS: 1
 DS: 0
 SWH: 8072.820 Hz
 FIDRES: 0.086043 Hz
 AQ: 5.0899774 sec
 RG: 5
 DV: 6240.000 sec
 DE: 6.000 sec
 TE: 298.0 K
 D1: 0.40000000 sec
 MCREST: 0.00000000 sec
 MCVRK: 0.01500000 sec
 ===== CHANNEL f1 =====
 NUC1: 1H
 P1: 8.00 usec
 PL1: 1.60 dB
 SFO1: 500.2250015 MHz
 F2-Processing parameters
 SF: 500.220305 MHz
 DSF: 65.00000000
 WDW: EM
 SSB: 0
 LB: 0.30 Hz
 GB: 0
 PC: 4.00
 1D MMR plot parameters
 CX: 22.80 cm
 CY: 15.00 cm
 F1P: 10.500 ppm
 F2P: 52.231 Hz
 F3P: 0.00000000 ppm
 F2: -250.11 Hz
 PPM1CM: 0.48246 ppm/cm
 HZCM: 241.33423 Hz/cm







```

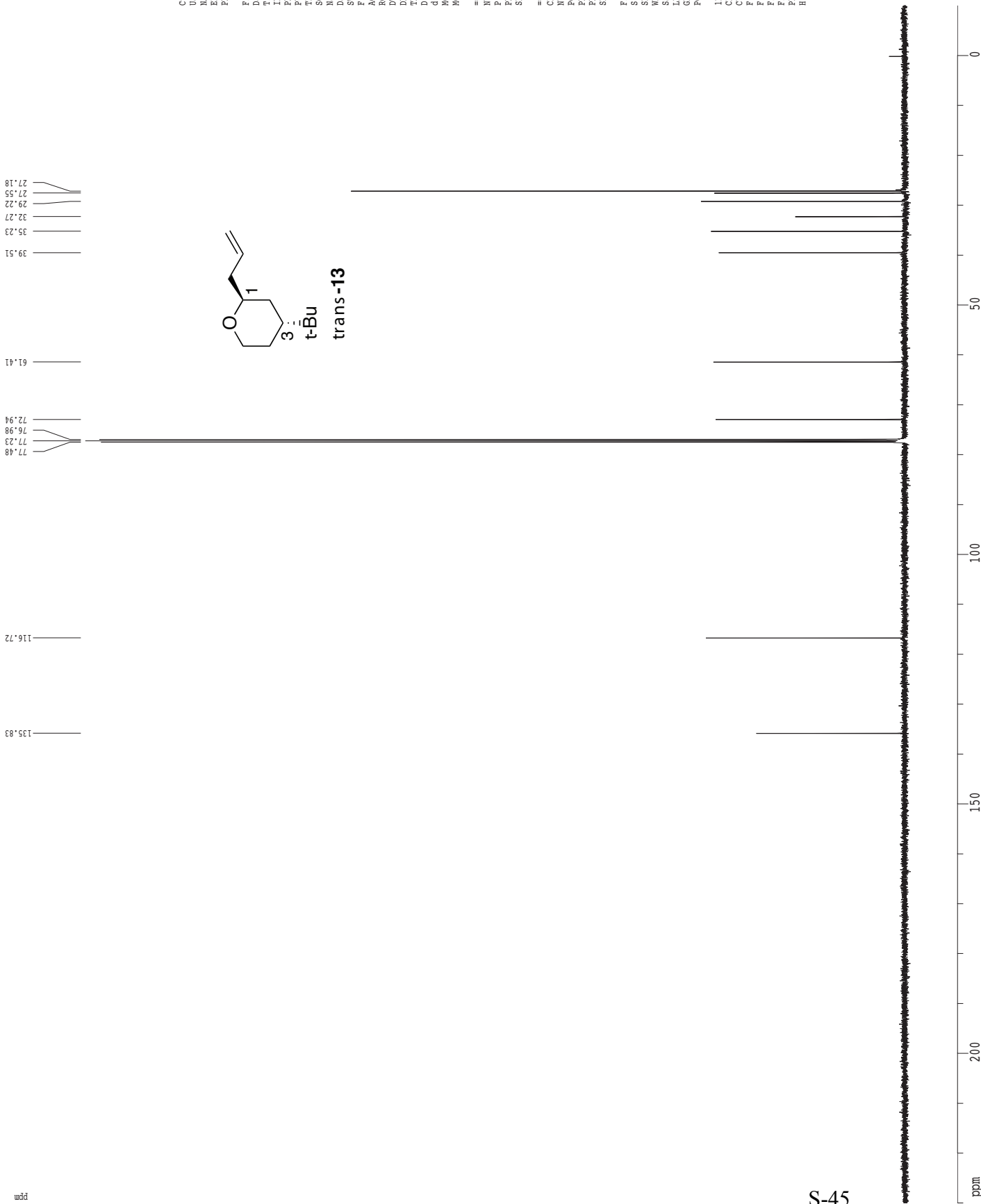
Current Data Parameters
USER      walter
NAME      WAS-III-58-27
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20070212
Time      17.18
INSTRUM   cryo500
PROBHD    5 mm CPXI IH-
PULPROG   zgpg30
NUC1       13
SOLVENT   CDCl3
NS         8
DS         0
SWH        8012.820 Hz
FIDRES     0.098043 Hz
AQ         5.0998774 sec
RG         5
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
ACRESF     0.00000000 sec
ACQRW      0.01500000 sec

===== CHANNEL f1 =====
NUC1       13
P1         8.00 usec
PL1        1.60 dB
SFO1       500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200305 MHz
WDW        EM
SSB        0
LB         0.20 Hz
GB         0
PC         4.00

ID NMR plot parameters
CX         22.80 cm
CY         15.00 cm
F1P        8.000 ppm
F1         4001.76 Hz
F2P        -0.500 ppm
F2         -250.11 Hz
PPMCH     0.37281 ppm/cm
HZCH      186.49555 Hz/cm
    
```



Current Data Parameters
USER walter
NAME MS-111-0427
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20070212
Time 17:22
INSTRUM cryo500
PROBHD 5 mm CPXI 1H-
PULPROG zgpg30
TD 65418
SOLVENT CDCl3
NS 150
DS 4
SWH 30303.03 Hz
FIDRES 0.246222 Hz
AQ 1.0794470 sec
RG 91895.2
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.2500000 sec
d11 0.0300000 sec
MCREST 0.0000000 sec
MORPK 0.0150000 sec

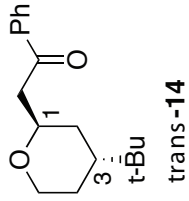
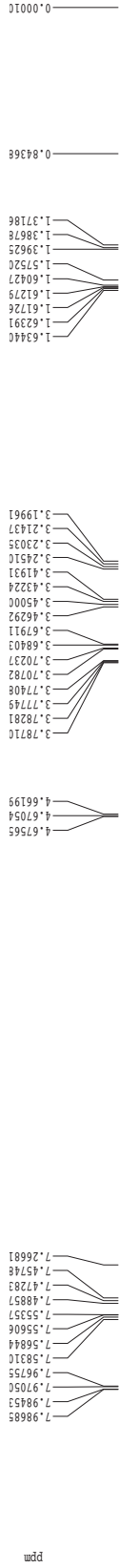
==== CHANNEL f1 =====
NUC1 13C
P1 15.00 usec
PL1 -1.00 dB
SFO1 125.7942348 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.60 dB
PL12 23.54 dB
SFO2 500.2225011 MHz

F2 - Processing parameters
SI 65536
SF 125.7803987 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters
X 72.94 cm
Y 23.54 cm
F1 28929.49 ppm
F2 -10.000 ppm
F2 -1257.80 Hz
PRCM 10.52632 ppm/cm
HZCM 1324.00427 Hz/cm

WAS-III-62-26
1H spectrum



Current Data Parameters
 USER: walter
 NAME: WAS-III-62-26
 EXPNO: 1
 PROCNO: 1

F2 - Acquisition Parameters
 Date_: 20070222
 Time: 17:38
 INSTRUM: cryo500
 PROBD: 5 mm CPXI IH-
 PULPROG: zgpg30
 SOLVENT: CDCl3
 NS: 8
 DS: 0
 SWH: 8012.820 Hz
 FIDRES: 0.098043 Hz
 AQ: 5.0998774 sec
 RG: 6.3
 DW: 62.400 usec
 DE: 6.00 usec
 TE: 298.0 K
 D1: 0.10000000 sec
 ACRESF: 0.00000000 sec
 ACPRR: 0.01500000 sec

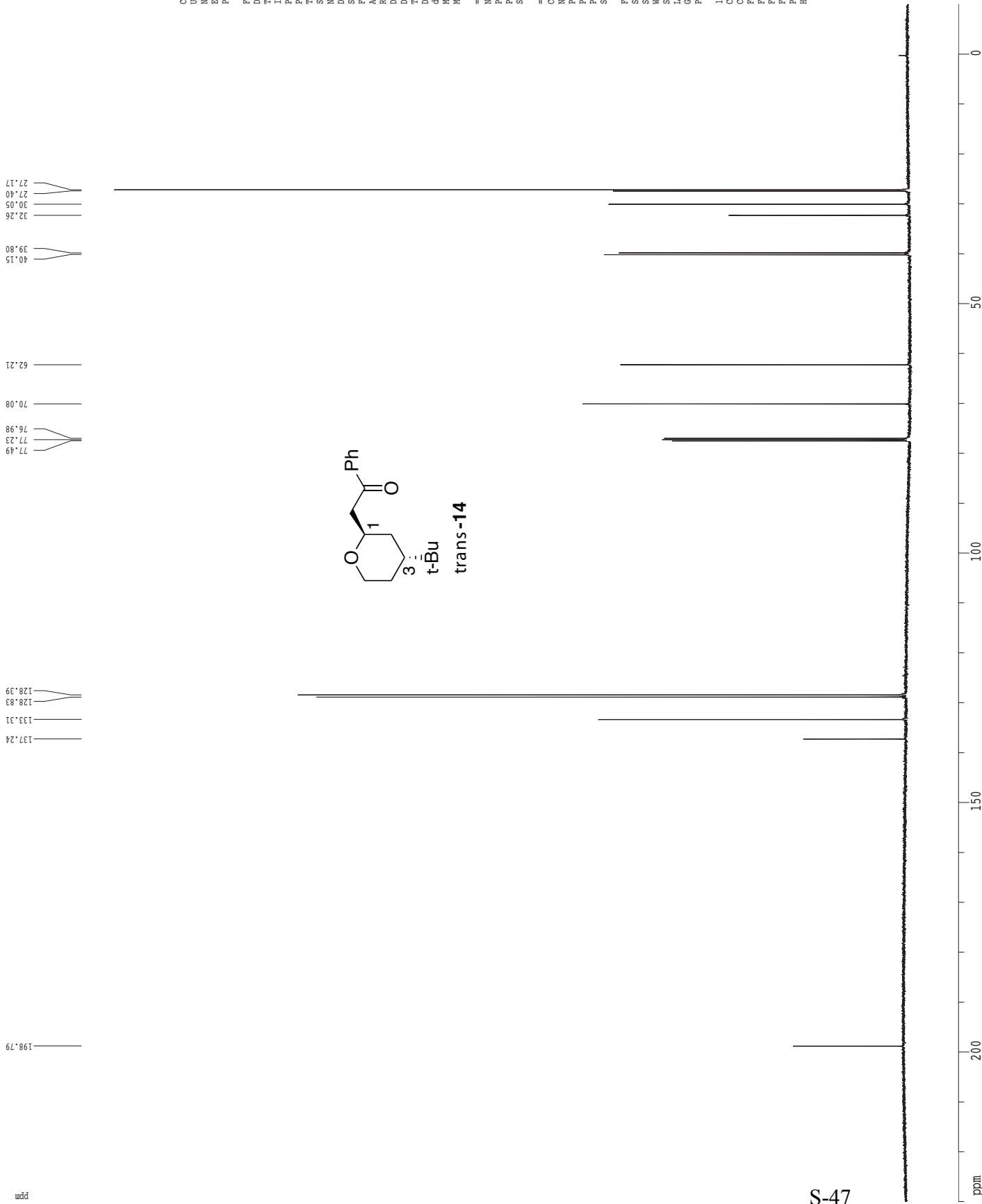
==== CHANNEL f1 =====
 NU1: 1H
 P1: 8.00 usec
 PL1: 1.60 dB
 SFO1: 500.2235015 MHz

F2 - Processing parameters
 SI: 65536
 SF: 500.2200278 MHz
 WDW: EN
 SSB: 0
 LB: 0.30 Hz
 GB: 0
 PC: 4.00

ID NMR plot parameters
 CX: 22.80 cm
 CY: 15.00 cm
 F1P: 9.000 ppm
 F1: 4501.98 Hz
 F2P: -0.500 ppm
 F2: -250.11 Hz
 PPMCM: 0.41667 ppm/cm
 HZCM: 204.42502 Hz/cm



WAS-III-62-26
 13C spectrum with 1H decoupling



```

Current Data Parameters
USER      walter
NAME      WAS-III-62-26
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20070222
Time      17.42
INSTRUM   cryo500
PROBHD    5 mm CPTCI LH-
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         222
DS         4
SWH        30303.00 Hz
FIDRES     0.2462388 Hz
AQ         1.0819380 sec
RG         115851.2
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
MCREST    0.00000000 sec
MORPK     0.01500000 sec

===== CHANNEL f1 =====
NUC1       13C
P1         15.00 usec
PL1        -1.00 dB
SFO1       125.7942348 MHz

===== CHANNEL f2 =====
NUC2       13C
P2         15.00 usec
PL2        -1.00 dB
SFO2       125.7942348 MHz

===== CHANNEL f3 =====
NUC3       13C
P3         15.00 usec
PL3        -1.00 dB
SFO3       125.7942348 MHz

===== CHANNEL f4 =====
NUC4       13C
P4         15.00 usec
PL4        -1.00 dB
SFO4       125.7942348 MHz

===== CHANNEL f5 =====
NUC5       13C
P5         15.00 usec
PL5        -1.00 dB
SFO5       125.7942348 MHz

===== CHANNEL f6 =====
NUC6       13C
P6         15.00 usec
PL6        -1.00 dB
SFO6       125.7942348 MHz

===== CHANNEL f7 =====
NUC7       13C
P7         15.00 usec
PL7        -1.00 dB
SFO7       125.7942348 MHz

===== CHANNEL f8 =====
NUC8       13C
P8         15.00 usec
PL8        -1.00 dB
SFO8       125.7942348 MHz

===== CHANNEL f9 =====
NUC9       13C
P9         15.00 usec
PL9        -1.00 dB
SFO9       125.7942348 MHz

===== CHANNEL f10 =====
NUC10      13C
P10        15.00 usec
PL10       -1.00 dB
SFO10      125.7942348 MHz

===== CHANNEL f11 =====
NUC11      13C
P11        15.00 usec
PL11       -1.00 dB
SFO11      125.7942348 MHz

===== CHANNEL f12 =====
NUC12      13C
P12        15.00 usec
PL12       -1.00 dB
SFO12      125.7942348 MHz

===== CHANNEL f13 =====
NUC13      13C
P13        15.00 usec
PL13       -1.00 dB
SFO13      125.7942348 MHz

===== CHANNEL f14 =====
NUC14      13C
P14        15.00 usec
PL14       -1.00 dB
SFO14      125.7942348 MHz

===== CHANNEL f15 =====
NUC15      13C
P15        15.00 usec
PL15       -1.00 dB
SFO15      125.7942348 MHz

===== CHANNEL f16 =====
NUC16      13C
P16        15.00 usec
PL16       -1.00 dB
SFO16      125.7942348 MHz

===== CHANNEL f17 =====
NUC17      13C
P17        15.00 usec
PL17       -1.00 dB
SFO17      125.7942348 MHz

===== CHANNEL f18 =====
NUC18      13C
P18        15.00 usec
PL18       -1.00 dB
SFO18      125.7942348 MHz

===== CHANNEL f19 =====
NUC19      13C
P19        15.00 usec
PL19       -1.00 dB
SFO19      125.7942348 MHz

===== CHANNEL f20 =====
NUC20      13C
P20        15.00 usec
PL20       -1.00 dB
SFO20      125.7942348 MHz

===== CHANNEL f21 =====
NUC21      13C
P21        15.00 usec
PL21       -1.00 dB
SFO21      125.7942348 MHz

===== CHANNEL f22 =====
NUC22      13C
P22        15.00 usec
PL22       -1.00 dB
SFO22      125.7942348 MHz

===== CHANNEL f23 =====
NUC23      13C
P23        15.00 usec
PL23       -1.00 dB
SFO23      125.7942348 MHz

===== CHANNEL f24 =====
NUC24      13C
P24        15.00 usec
PL24       -1.00 dB
SFO24      125.7942348 MHz

===== CHANNEL f25 =====
NUC25      13C
P25        15.00 usec
PL25       -1.00 dB
SFO25      125.7942348 MHz

===== CHANNEL f26 =====
NUC26      13C
P26        15.00 usec
PL26       -1.00 dB
SFO26      125.7942348 MHz

===== CHANNEL f27 =====
NUC27      13C
P27        15.00 usec
PL27       -1.00 dB
SFO27      125.7942348 MHz

===== CHANNEL f28 =====
NUC28      13C
P28        15.00 usec
PL28       -1.00 dB
SFO28      125.7942348 MHz

===== CHANNEL f29 =====
NUC29      13C
P29        15.00 usec
PL29       -1.00 dB
SFO29      125.7942348 MHz

===== CHANNEL f30 =====
NUC30      13C
P30        15.00 usec
PL30       -1.00 dB
SFO30      125.7942348 MHz

===== CHANNEL f31 =====
NUC31      13C
P31        15.00 usec
PL31       -1.00 dB
SFO31      125.7942348 MHz

===== CHANNEL f32 =====
NUC32      13C
P32        15.00 usec
PL32       -1.00 dB
SFO32      125.7942348 MHz

===== CHANNEL f33 =====
NUC33      13C
P33        15.00 usec
PL33       -1.00 dB
SFO33      125.7942348 MHz

===== CHANNEL f34 =====
NUC34      13C
P34        15.00 usec
PL34       -1.00 dB
SFO34      125.7942348 MHz

===== CHANNEL f35 =====
NUC35      13C
P35        15.00 usec
PL35       -1.00 dB
SFO35      125.7942348 MHz

===== CHANNEL f36 =====
NUC36      13C
P36        15.00 usec
PL36       -1.00 dB
SFO36      125.7942348 MHz

===== CHANNEL f37 =====
NUC37      13C
P37        15.00 usec
PL37       -1.00 dB
SFO37      125.7942348 MHz

===== CHANNEL f38 =====
NUC38      13C
P38        15.00 usec
PL38       -1.00 dB
SFO38      125.7942348 MHz

===== CHANNEL f39 =====
NUC39      13C
P39        15.00 usec
PL39       -1.00 dB
SFO39      125.7942348 MHz

===== CHANNEL f40 =====
NUC40      13C
P40        15.00 usec
PL40       -1.00 dB
SFO40      125.7942348 MHz

===== CHANNEL f41 =====
NUC41      13C
P41        15.00 usec
PL41       -1.00 dB
SFO41      125.7942348 MHz

===== CHANNEL f42 =====
NUC42      13C
P42        15.00 usec
PL42       -1.00 dB
SFO42      125.7942348 MHz

===== CHANNEL f43 =====
NUC43      13C
P43        15.00 usec
PL43       -1.00 dB
SFO43      125.7942348 MHz

===== CHANNEL f44 =====
NUC44      13C
P44        15.00 usec
PL44       -1.00 dB
SFO44      125.7942348 MHz

===== CHANNEL f45 =====
NUC45      13C
P45        15.00 usec
PL45       -1.00 dB
SFO45      125.7942348 MHz

===== CHANNEL f46 =====
NUC46      13C
P46        15.00 usec
PL46       -1.00 dB
SFO46      125.7942348 MHz

===== CHANNEL f47 =====
NUC47      13C
P47        15.00 usec
PL47       -1.00 dB
SFO47      125.7942348 MHz

===== CHANNEL f48 =====
NUC48      13C
P48        15.00 usec
PL48       -1.00 dB
SFO48      125.7942348 MHz

===== CHANNEL f49 =====
NUC49      13C
P49        15.00 usec
PL49       -1.00 dB
SFO49      125.7942348 MHz

===== CHANNEL f50 =====
NUC50      13C
P50        15.00 usec
PL50       -1.00 dB
SFO50      125.7942348 MHz

===== CHANNEL f51 =====
NUC51      13C
P51        15.00 usec
PL51       -1.00 dB
SFO51      125.7942348 MHz

===== CHANNEL f52 =====
NUC52      13C
P52        15.00 usec
PL52       -1.00 dB
SFO52      125.7942348 MHz

===== CHANNEL f53 =====
NUC53      13C
P53        15.00 usec
PL53       -1.00 dB
SFO53      125.7942348 MHz

===== CHANNEL f54 =====
NUC54      13C
P54        15.00 usec
PL54       -1.00 dB
SFO54      125.7942348 MHz

===== CHANNEL f55 =====
NUC55      13C
P55        15.00 usec
PL55       -1.00 dB
SFO55      125.7942348 MHz

===== CHANNEL f56 =====
NUC56      13C
P56        15.00 usec
PL56       -1.00 dB
SFO56      125.7942348 MHz

===== CHANNEL f57 =====
NUC57      13C
P57        15.00 usec
PL57       -1.00 dB
SFO57      125.7942348 MHz

===== CHANNEL f58 =====
NUC58      13C
P58        15.00 usec
PL58       -1.00 dB
SFO58      125.7942348 MHz

===== CHANNEL f59 =====
NUC59      13C
P59        15.00 usec
PL59       -1.00 dB
SFO59      125.7942348 MHz

===== CHANNEL f60 =====
NUC60      13C
P60        15.00 usec
PL60       -1.00 dB
SFO60      125.7942348 MHz

===== CHANNEL f61 =====
NUC61      13C
P61        15.00 usec
PL61       -1.00 dB
SFO61      125.7942348 MHz

===== CHANNEL f62 =====
NUC62      13C
P62        15.00 usec
PL62       -1.00 dB
SFO62      125.7942348 MHz

===== CHANNEL f63 =====
NUC63      13C
P63        15.00 usec
PL63       -1.00 dB
SFO63      125.7942348 MHz

===== CHANNEL f64 =====
NUC64      13C
P64        15.00 usec
PL64       -1.00 dB
SFO64      125.7942348 MHz

===== CHANNEL f65 =====
NUC65      13C
P65        15.00 usec
PL65       -1.00 dB
SFO65      125.7942348 MHz

===== CHANNEL f66 =====
NUC66      13C
P66        15.00 usec
PL66       -1.00 dB
SFO66      125.7942348 MHz

===== CHANNEL f67 =====
NUC67      13C
P67        15.00 usec
PL67       -1.00 dB
SFO67      125.7942348 MHz

===== CHANNEL f68 =====
NUC68      13C
P68        15.00 usec
PL68       -1.00 dB
SFO68      125.7942348 MHz

===== CHANNEL f69 =====
NUC69      13C
P69        15.00 usec
PL69       -1.00 dB
SFO69      125.7942348 MHz

===== CHANNEL f70 =====
NUC70      13C
P70        15.00 usec
PL70       -1.00 dB
SFO70      125.7942348 MHz

===== CHANNEL f71 =====
NUC71      13C
P71        15.00 usec
PL71       -1.00 dB
SFO71      125.7942348 MHz

===== CHANNEL f72 =====
NUC72      13C
P72        15.00 usec
PL72       -1.00 dB
SFO72      125.7942348 MHz

===== CHANNEL f73 =====
NUC73      13C
P73        15.00 usec
PL73       -1.00 dB
SFO73      125.7942348 MHz

===== CHANNEL f74 =====
NUC74      13C
P74        15.00 usec
PL74       -1.00 dB
SFO74      125.7942348 MHz

===== CHANNEL f75 =====
NUC75      13C
P75        15.00 usec
PL75       -1.00 dB
SFO75      125.7942348 MHz

===== CHANNEL f76 =====
NUC76      13C
P76        15.00 usec
PL76       -1.00 dB
SFO76      125.7942348 MHz

===== CHANNEL f77 =====
NUC77      13C
P77        15.00 usec
PL77       -1.00 dB
SFO77      125.7942348 MHz

===== CHANNEL f78 =====
NUC78      13C
P78        15.00 usec
PL78       -1.00 dB
SFO78      125.7942348 MHz

===== CHANNEL f79 =====
NUC79      13C
P79        15.00 usec
PL79       -1.00 dB
SFO79      125.7942348 MHz

===== CHANNEL f80 =====
NUC80      13C
P80        15.00 usec
PL80       -1.00 dB
SFO80      125.7942348 MHz

===== CHANNEL f81 =====
NUC81      13C
P81        15.00 usec
PL81       -1.00 dB
SFO81      125.7942348 MHz

===== CHANNEL f82 =====
NUC82      13C
P82        15.00 usec
PL82       -1.00 dB
SFO82      125.7942348 MHz

===== CHANNEL f83 =====
NUC83      13C
P83        15.00 usec
PL83       -1.00 dB
SFO83      125.7942348 MHz

===== CHANNEL f84 =====
NUC84      13C
P84        15.00 usec
PL84       -1.00 dB
SFO84      125.7942348 MHz

===== CHANNEL f85 =====
NUC85      13C
P85        15.00 usec
PL85       -1.00 dB
SFO85      125.7942348 MHz

===== CHANNEL f86 =====
NUC86      13C
P86        15.00 usec
PL86       -1.00 dB
SFO86      125.7942348 MHz

===== CHANNEL f87 =====
NUC87      13C
P87        15.00 usec
PL87       -1.00 dB
SFO87      125.7942348 MHz

===== CHANNEL f88 =====
NUC88      13C
P88        15.00 usec
PL88       -1.00 dB
SFO88      125.7942348 MHz

===== CHANNEL f89 =====
NUC89      13C
P89        15.00 usec
PL89       -1.00 dB
SFO89      125.7942348 MHz

===== CHANNEL f90 =====
NUC90      13C
P90        15.00 usec
PL90       -1.00 dB
SFO90      125.7942348 MHz

===== CHANNEL f91 =====
NUC91      13C
P91        15.00 usec
PL91       -1.00 dB
SFO91      125.7942348 MHz

===== CHANNEL f92 =====
NUC92      13C
P92        15.00 usec
PL92       -1.00 dB
SFO92      125.7942348 MHz

===== CHANNEL f93 =====
NUC93      13C
P93        15.00 usec
PL93       -1.00 dB
SFO93      125.7942348 MHz

===== CHANNEL f94 =====
NUC94      13C
P94        15.00 usec
PL94       -1.00 dB
SFO94      125.7942348 MHz

===== CHANNEL f95 =====
NUC95      13C
P95        15.00 usec
PL95       -1.00 dB
SFO95      125.7942348 MHz

===== CHANNEL f96 =====
NUC96      13C
P96        15.00 usec
PL96       -1.00 dB
SFO96      125.7942348 MHz

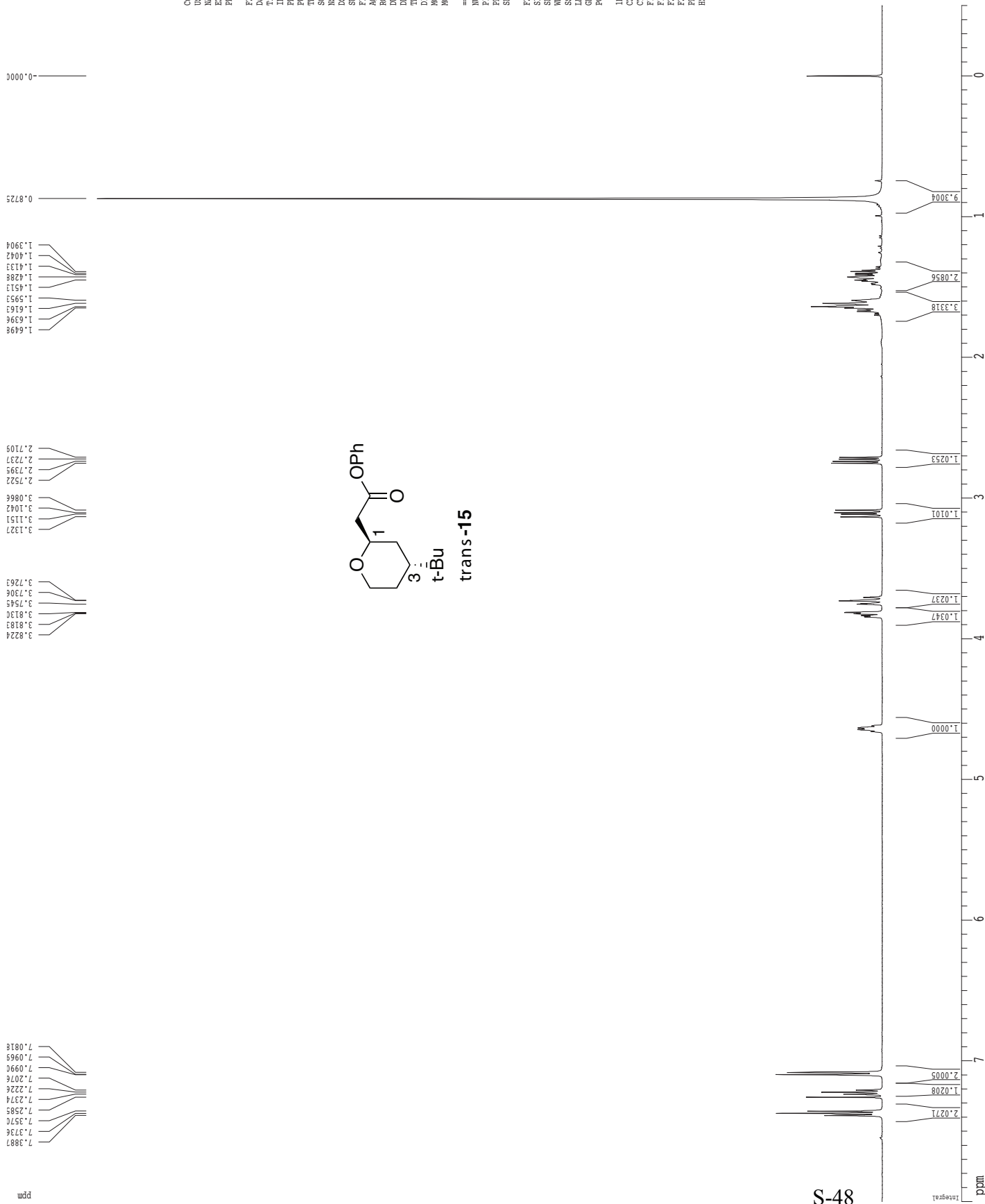
===== CHANNEL f97 =====
NUC97      13C
P97        15.00 usec
PL97       -1.00 dB
SFO97      125.7942348 MHz

===== CHANNEL f98 =====
NUC98      13C
P98        15.00 usec
PL98       -1.00 dB
SFO98      125.7942348 MHz

===== CHANNEL f99 =====
NUC99      13C
P99        15.00 usec
PL99       -1.00 dB
SFO99      125.7942348 MHz

===== CHANNEL f100 =====
NUC100     13C
P100       15.00 usec
PL100      -1.00 dB
SFO100     125.7942348 MHz
  
```

WAS-III-59-27
1H spectrum



Current Data Parameters
 USER walter
 NAME WAS-III-59-27
 EXPNO 3
 PROCNO 1

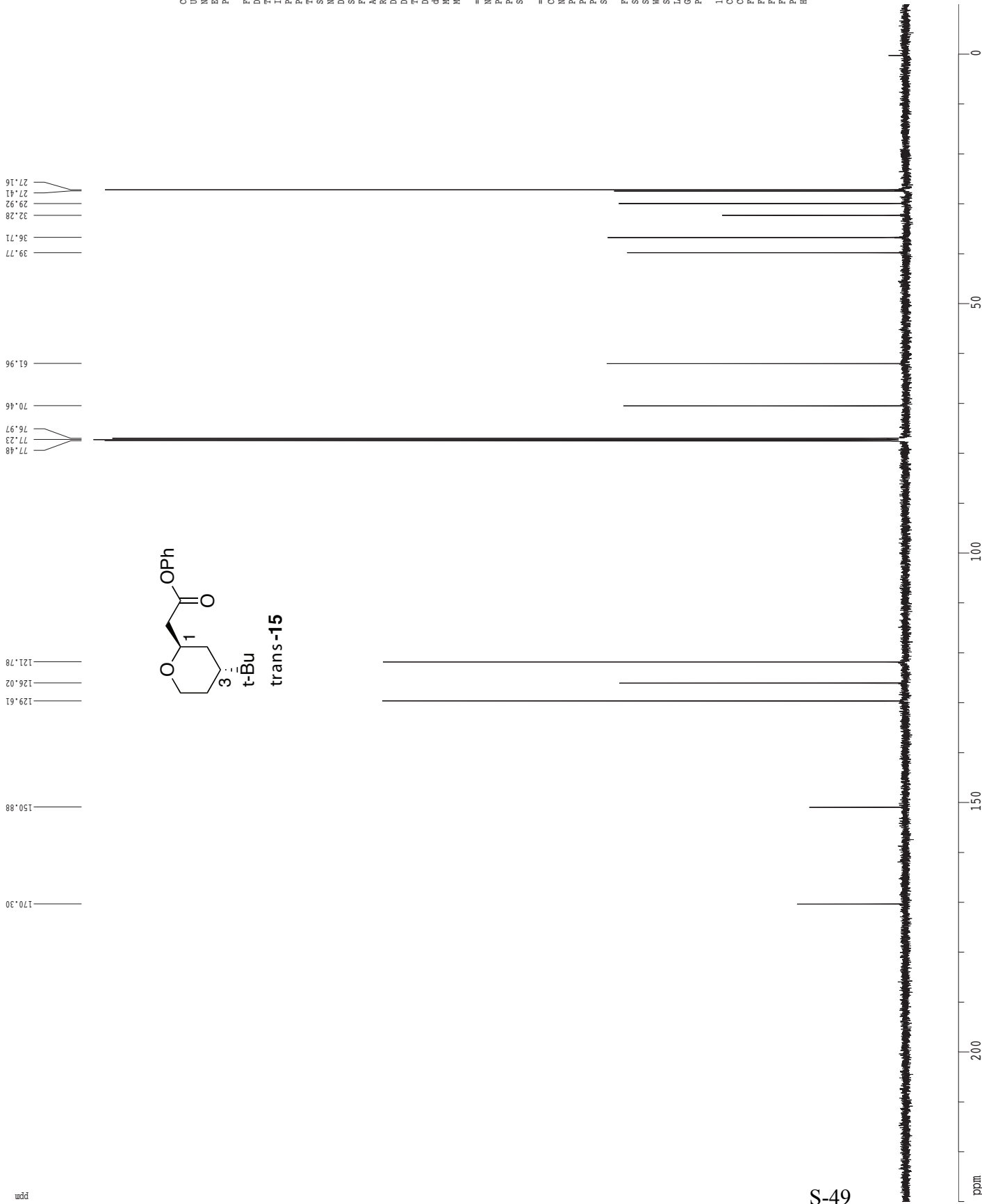
F2 - Acquisition Parameters
 Date_ 20070213
 Time 9.11
 INSTRUM cryo500
 PROBDH 5 mm CPCLP 1H-
 PULPROG zgpg30
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 4.5
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 ACRESF 0.0000000 sec
 ACPRR 0.0100000 sec

==== CHANNEL f1 =====
 NUCL1 1H
 P1 8.00 usec
 PL1 1.60 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200331 MHz
 WDW EN
 SSB 0
 LB 0.20 Hz
 GB 0
 PC 4.00

ID NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 FIP 8.000 ppm
 F1 4001.76 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.37281 ppm/cm
 HZCM 186.49555 Hz/cm

WAS-III-59-27
 13C spectrum with 1H decoupling



Current Data Parameters
 USER walter
 NAME WAS-III-59-27
 EXPNO 5
 PROCNO 1

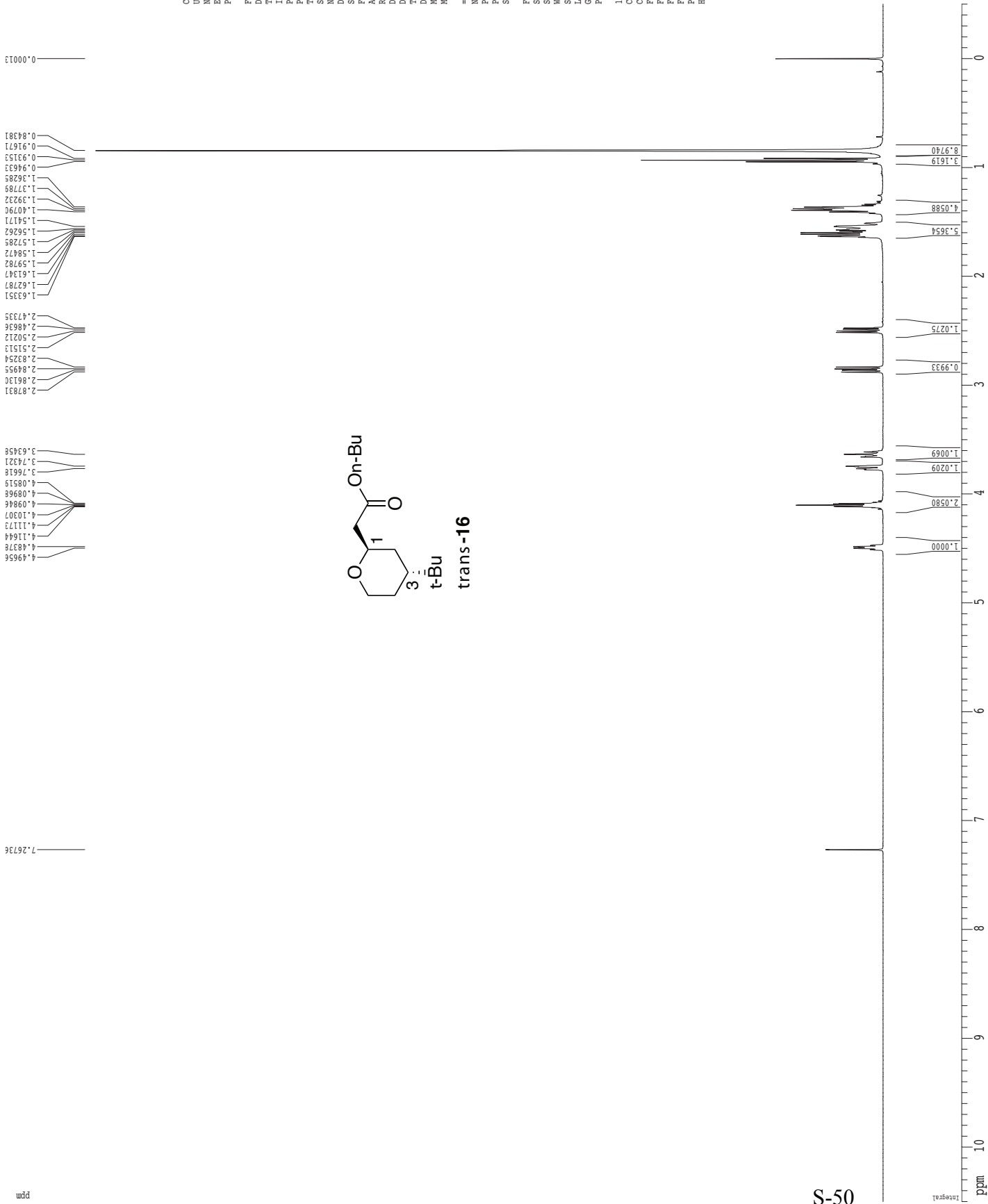
F2 - Acquisition Parameters
 Date_ 20070213
 Time 9.14
 INSTRUM cryo500
 PROBHD 5 mm CPTCI LH-
 PULPROG zgpg30
 TD 65418
 SOLVENT CDCl3
 NS 94
 DS 4
 SWH 30303.00 Hz
 FWHZ 0.64222 Hz
 AQ 1.0794470 sec
 RG 130004
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MONRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 15.00 usec
 PL1 -1.00 dB
 SFO1 125.7942348 MHz

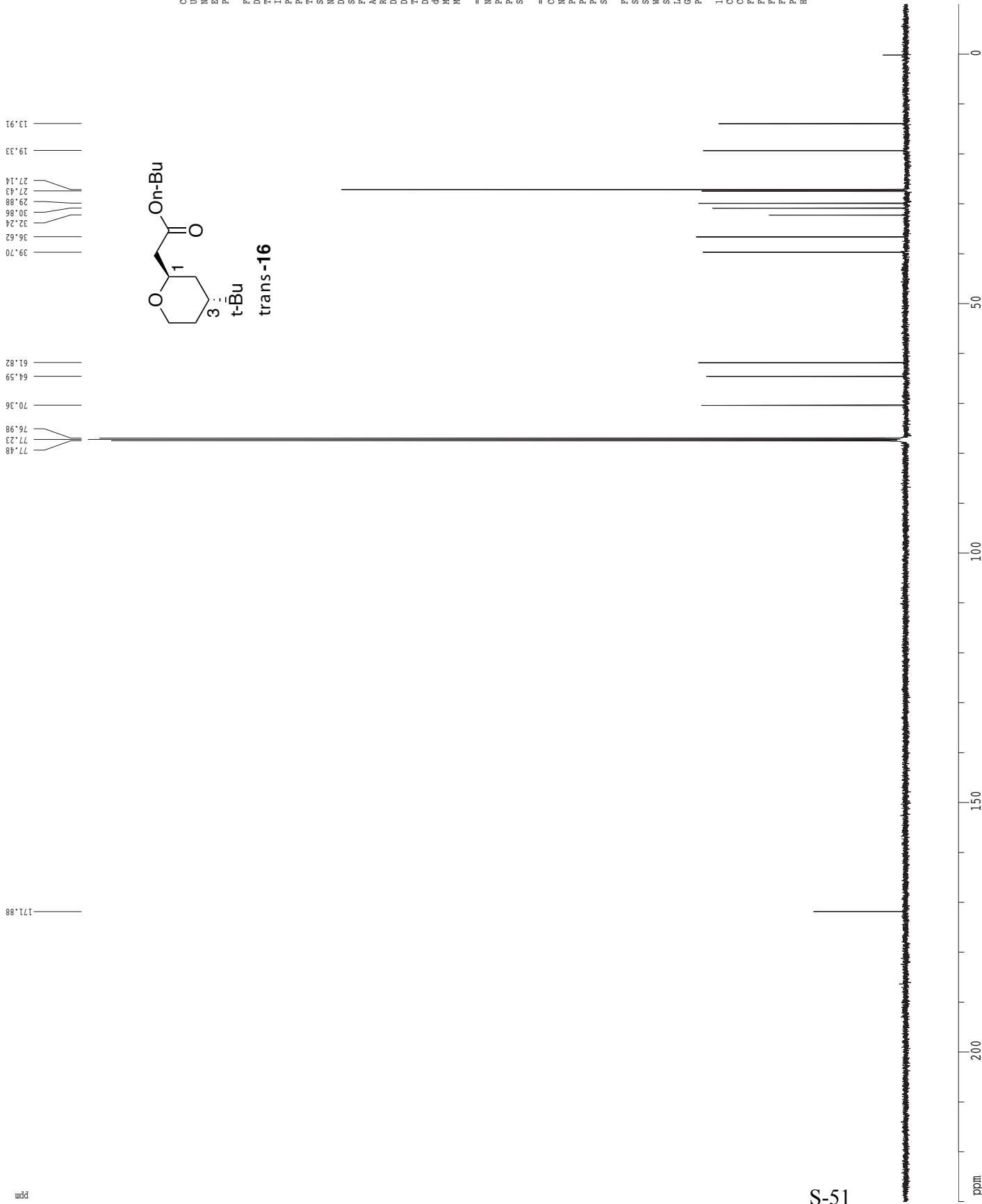
===== CHANNEL f2 =====
 CUPRG2 waltz16
 NUC2 1H
 P2 100.00 usec
 PL2 1.60 dB
 PL12 23.54 dB
 SFO2 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.784020 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

ID NMR plot parameters
 X 72.50 cm
 Y 11.00 cm
 F1 230.000 ppm
 F2 28929.49 Hz
 F3 -10.000 ppm
 F4 -1257.80 Hz
 PPMCM 10.52632 ppm/cm
 HZCM 1324.00427 Hz/cm



WAS-III-166-27
 13C spectrum with 1H decoupling



Current Data Parameters
 USER walter
 NAME WAS-III-166-27
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20071002
 Time 16.51
 INSTRUM cryo500
 PROBDH 5 mm CPTCI LH-
 PULPROG zgpg30
 TD 65418
 SOLVENT CDCl3T
 NS 179
 DS 30003.00 Hz
 SFO1 125.761322 Hz
 AQ 1.0794655 sec
 RG 130004
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MONRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 15.00 usec
 PL1 -1.00 dB
 SFO1 125.761322 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 -1.60 dB
 PL12 23.54 dB
 SFO2 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.761322 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 X 72.80 cm
 Y 1.00 cm
 Z 230.000 ppm
 F1 28929.49 Hz
 F2 -10.000 ppm
 F2 -1257.80 Hz
 PRCM 10.52632 ppm/cm
 HZCM 1324.00415 Hz/cm



Current Data Parameters
 USER: walter
 NAME: WAS-III-166-26
 EXPNO: 5
 PROCNO: 1

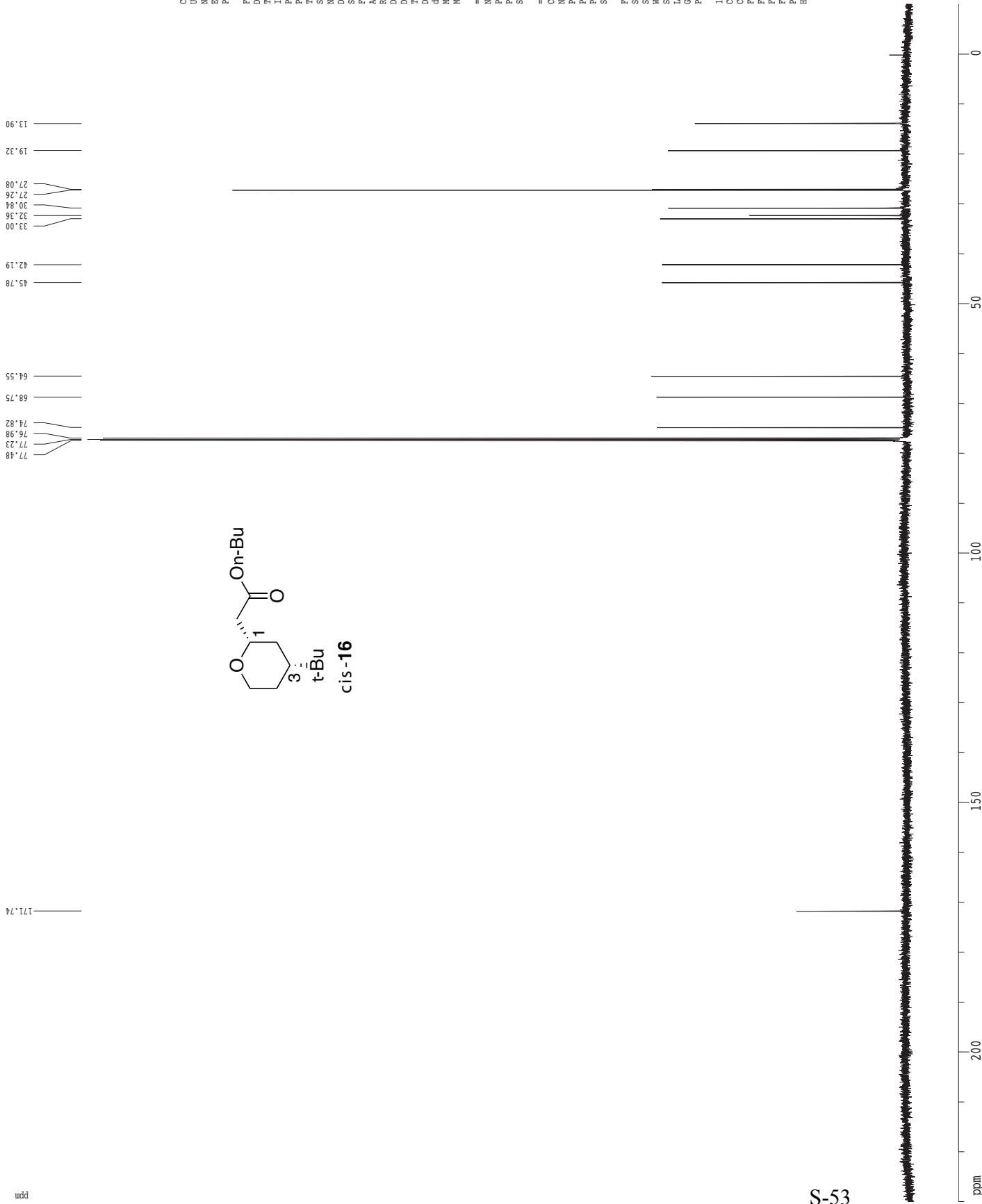
F2 - Acquisition Parameters
 Date_: 20071002
 Time: 23.07
 INSTRUM: cryo500
 PROBD: 5 mm CPXI IH-
 PULPROG: zgpg30
 SOLVENT: CDCl3
 NS: 8
 DS: 0
 SWH: 8012.820 Hz
 FIDRES: 0.098043 Hz
 AQ: 5.0999398 sec
 RG: 4.5
 DW: 62.400 usec
 DE: 6.00 usec
 TE: 298.0 K
 D1: 0.10000000 sec
 ACRESF: 0.00000000 sec
 ACPRR: 0.01500000 sec

==== CHANNEL f1 =====
 NU1: 1H
 P1: 8.00 usec
 PL1: 1.60 dB
 SFO1: 500.2235015 MHz

F2 - Processing parameters
 SI: 65536
 SF: 500.2200266 MHz
 WDW: EN
 SSB: 0
 LB: 0.20 Hz
 GB: 0
 PC: 4.00

ID NMR plot parameters
 CX: 22.80 cm
 CY: 15.00 cm
 F1P: 10.500 ppm
 F1: 5252.31 Hz
 F2P: -0.500 ppm
 F2: -250.11 Hz
 PPMCM: 0.48246 ppm/cm
 HZCM: 241.33423 Hz/cm

WAS-III-166-26
 13C spectrum with 1H decoupling



Current Data Parameters
 USER walter
 NAME WAS-III-166-26
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20071002
 Time 16.42
 INSTRUM cryo500
 PROBHD 5 mm CPTCI LH-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3T
 NS 77
 DS 30303.00 Hz
 SFO1 0.642388 Hz
 AC 1.0814105 sec
 RG 1.0004
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MONRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 15.00 usec
 PL1 -1.00 dB
 SFO1 125.7942348 MHz

===== CHANNEL f2 =====
 CPMPC2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 -1.60 dB
 PL12 23.54 dB
 SFO2 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7803987 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

LD NMR plot parameters
 X 72.80 cm
 Y 1.00 cm
 Z 20.000 ppm
 F1 28929.49 Hz
 F2 -10.000 ppm
 F2 -1257.80 Hz
 PPMCM 10.52632 ppm/cm
 HZCM 1324.00427 Hz/cm

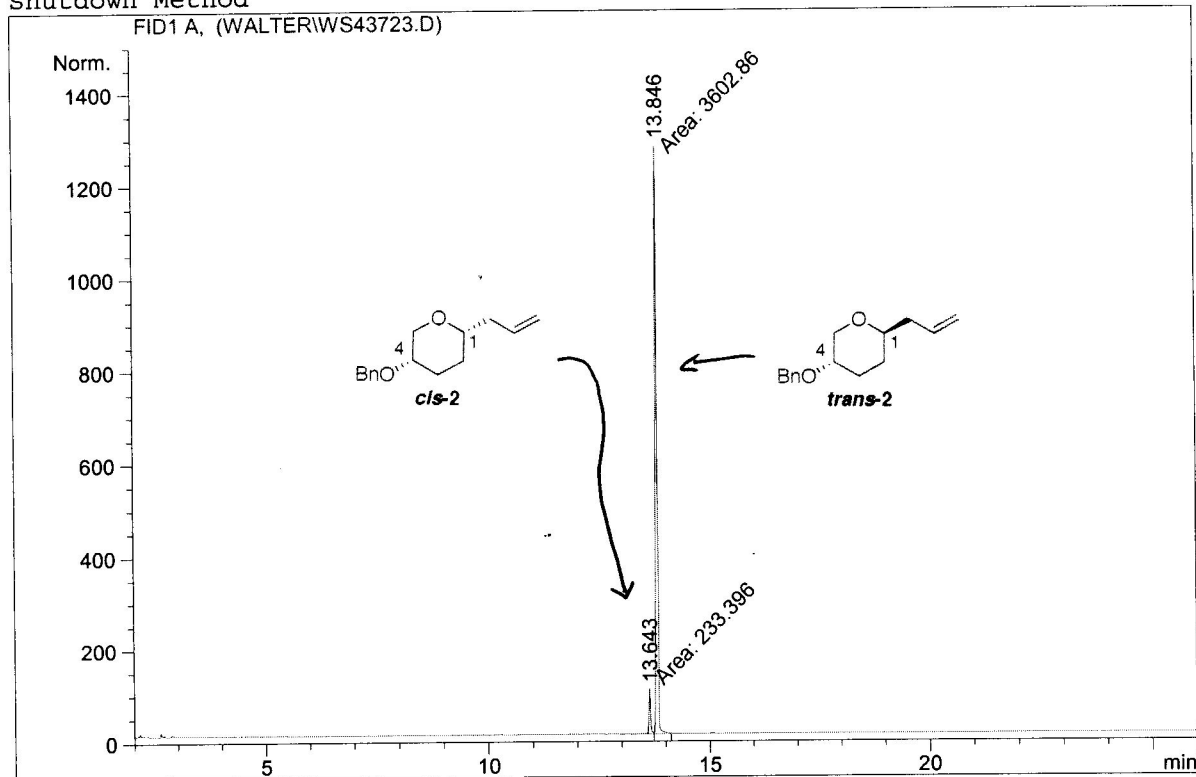
```

=====
Injection Date   : 5/2/2008 3:46:51 PM      Seq. Line   :    7
Sample Name     : WAS-IV-37-23             Location    : Vial 20
Acq. Operator   : jelena                   Inj         :    1
                                           Inj Volume  : 1 µl
                                           Actual Inj Volume : 5 µl
Different Inj Volume from Sequence !
Acq. Method     : C:\HPCHEM\1\METHODS\WALTER.M
Last changed    : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
Last changed    : 6/10/2008 9:37:46 AM by jelena
                (modified after loading)
=====

```

Table 1, Entry 1

Shutdown Method



```

=====
Area Percent Report
=====

```

```

Sorted By       :      Signal
Multiplier      :      1.0000
Dilution        :      1.0000

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	13.643	MM	0.0399	233.39619	97.60922	6.08395
2	13.846	MM	0.0473	3602.86206	1270.33093	93.91605

```
Totals :                3836.25826 1367.94016
```

Results obtained with enhanced integrator!

```

=====
*** End of Report ***
=====

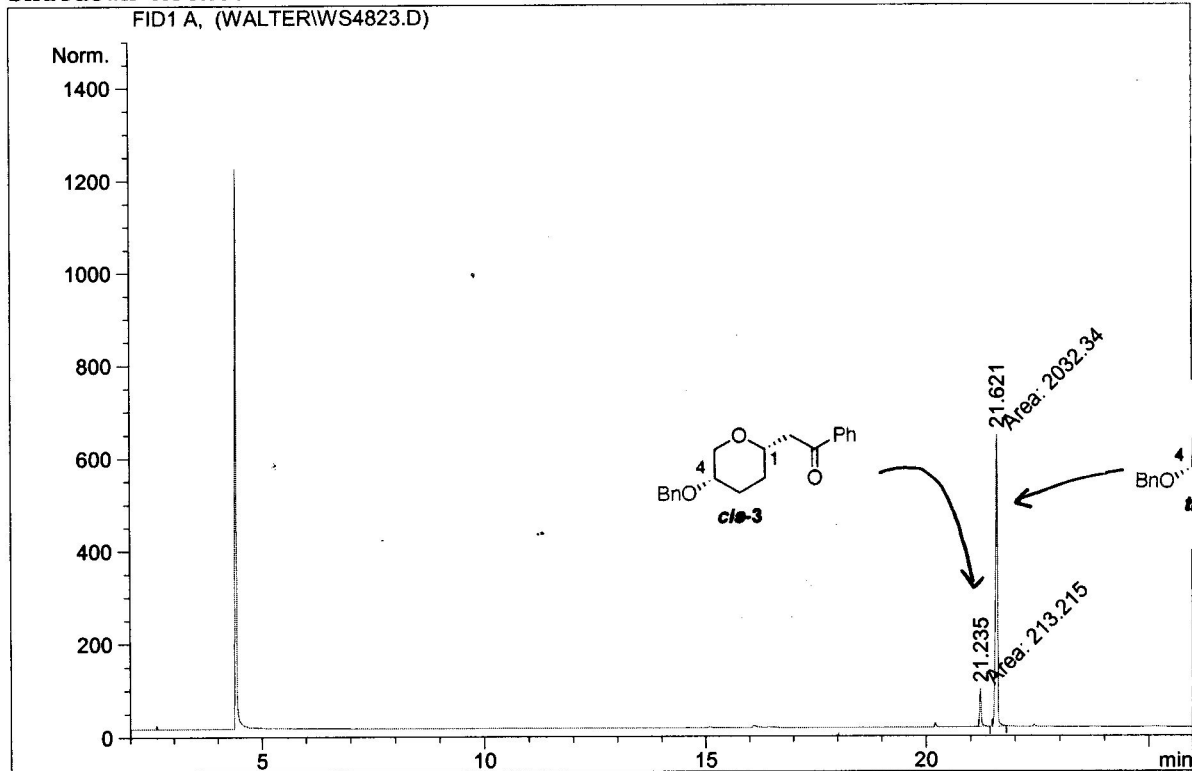
```

```

=====
Injection Date : 2/14/2008 7:31:59 PM      Seq. Line :    2
Sample Name    : WAS-IV-8-23                Location  : Vial 8
Acq. Operator  : jelena                      Inj       :    1
                                           Inj Volume: 1 µl
                                           Actual Inj Volume: 5 µl
Different Inj Volume from Sequence !
Acq. Method    : C:\HPCHEM\1\METHODS\WALTER.M
Last changed   : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
Last changed   : 6/10/2008 9:38:48 AM by jelena
                (modified after loading)
    
```

Table 1, Entry 2

Shutdown Method



Area Percent Report

```

Sorted By      :      Signal
Multiplier    :      1.0000
Dilution      :      1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	21.235	MM	0.0430	213.21509	82.68630	9.49497
2	21.621	MM	0.0537	2032.34253	630.24927	90.50503

Totals : 2245.55762 712.93557

Results obtained with enhanced integrator!

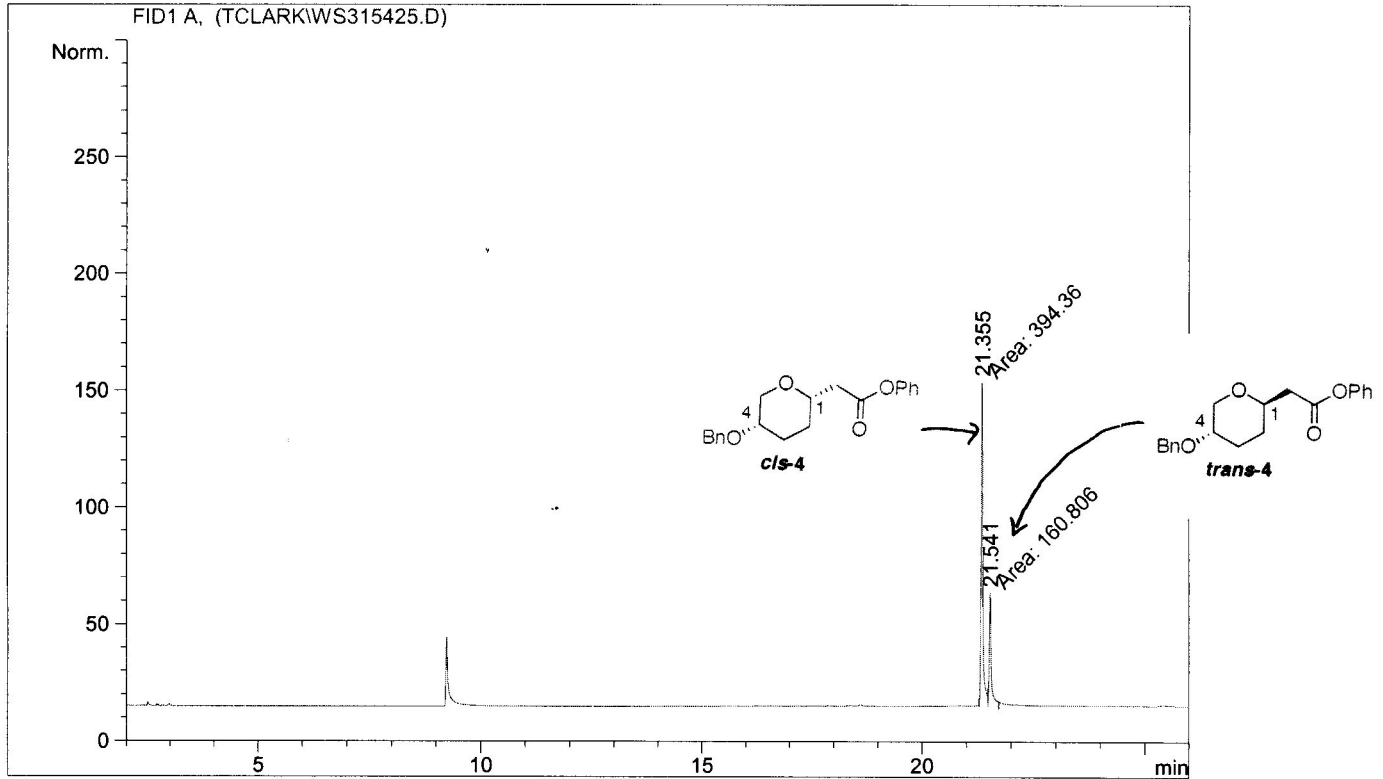
*** End of Report ***

```

=====
Injection Date   : 9/11/2007 1:57:12 PM      Seq. Line   :    1
Sample Name     : WAS-III-154-25            Location    : Vial 4
Acq. Operator   : jelena                    Inj         :    1
                                           Inj Volume  : 1 µl
                                           Actual Inj Volume : 5 µl
Different Inj Volume from Sequence !
Acq. Method    : C:\HPCHEM\1\METHODS\WALTER.M
Last changed   : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
Last changed   : 6/10/2008 9:47:47 AM by jelena
                (modified after loading)
Shutdown Method

```

Table 1, Entry 3



Area Percent Report

```

Sorted By      : Signal
Multiplier    : 1.0000
Dilution      : 1.0000

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	21.355	MM	0.0474	394.36014	138.73720	71.03465
2	21.541	MM	0.0551	160.80571	48.66586	28.96535

Totals : 555.16585 187.40305

Results obtained with enhanced integrator!

*** End of Report ***

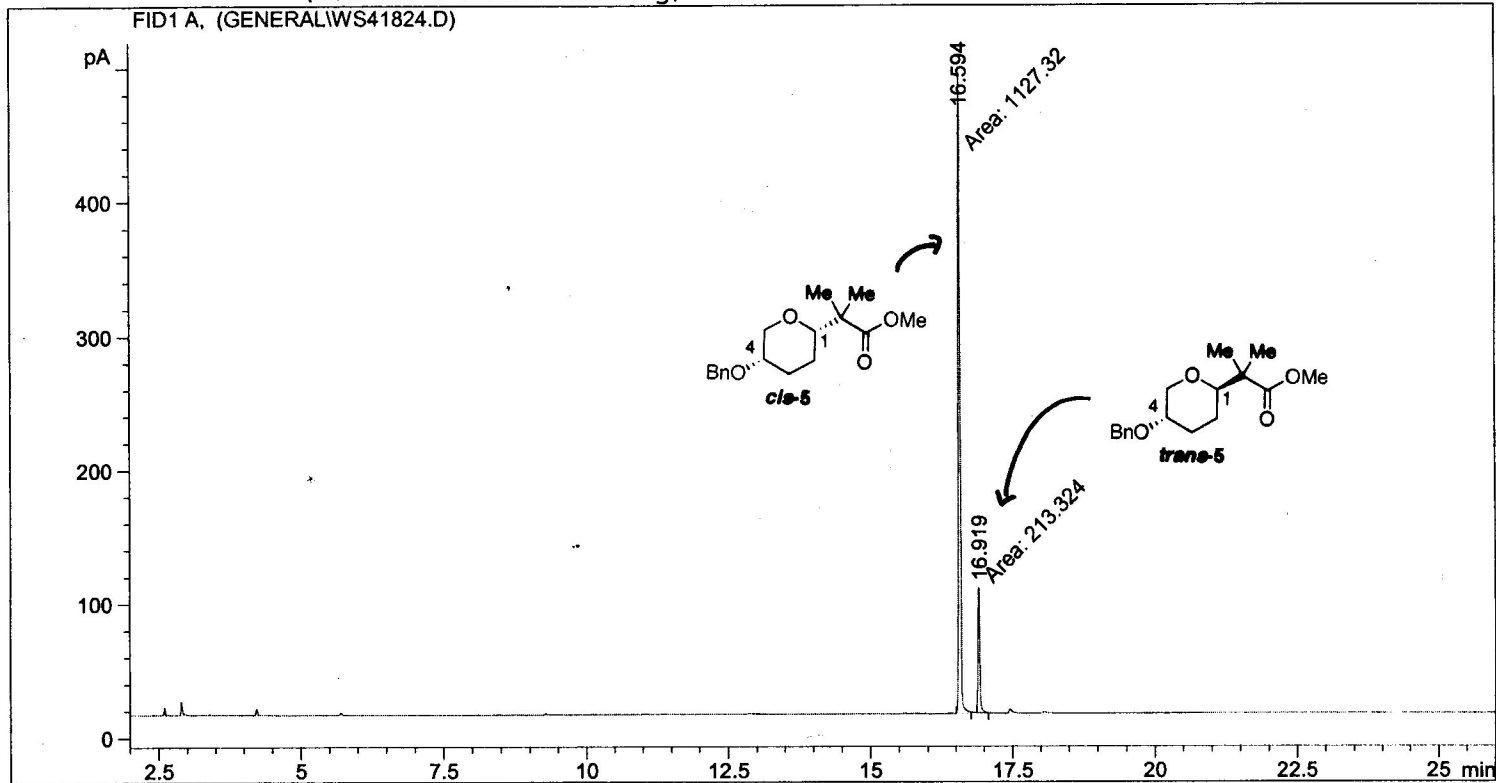

```

=====
Injection Date   : 3/3/2008 9:09:07 PM      Seq. Line   :    2
Sample Name     : WAS-IV-18-24              Location    : Vial 2
Acq. Operator   : jelena                    Inj         :    1
                                           Inj Volume  : 1 µl
                                           Actual Inj Volume : 5 µl

Different Inj Volume from Sequence !

Acq. Method     : C:\HPCHEM\1\METHODS\WALTER.M
Last changed    : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\OFF.M
Last changed    : 3/4/2008 9:18:21 AM by jelena
                 (modified after loading)
    
```

Table 1, Entry 4



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	16.594	MM	0.0392	1127.31763	478.89899	84.08791
2	16.919	MM	0.0377	213.32408	94.20467	15.91209

Totals : 1340.64171 573.10366

Results obtained with enhanced integrator!

*** End of Report ***

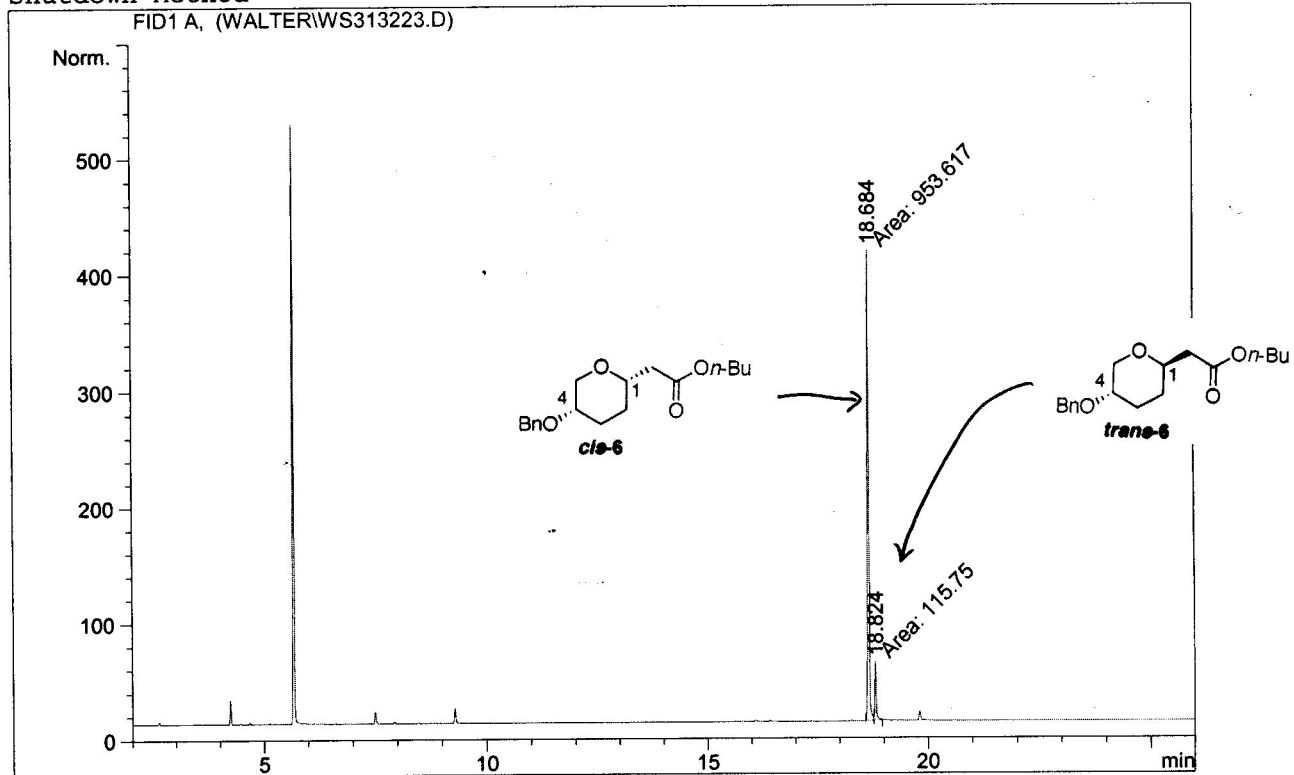
```

=====
Injection Date   : 7/19/2007 3:19:51 PM      Seq. Line   :    3
Sample Name     : WAS-III-132-23            Location    : Vial 17
Acq. Operator   : jelena                    Inj         :    1
                                           Inj Volume  : 1 µl
                                           Actual Inj Volume : 5 µl
Different Inj Volume from Sequence !
Acq. Method     : C:\HPCHEM\1\METHODS\WALTER.M
Last changed    : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
Last changed    : 6/10/2008 9:41:43 AM by jelena
                 (modified after loading)
=====

```

Table 1, Entry 5

Shutdown Method



Area Percent Report

```

=====
Sorted By      :      Signal
Multiplier     :      1.0000
Dilution      :      1.0000
=====

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	18.684	MM	0.0393	953.61658	404.30756	89.17585
2	18.824	MM	0.0384	115.74980	50.21415	10.82415

Totals : 1069.36638 454.52170

Results obtained with enhanced integrator!

*** End of Report ***

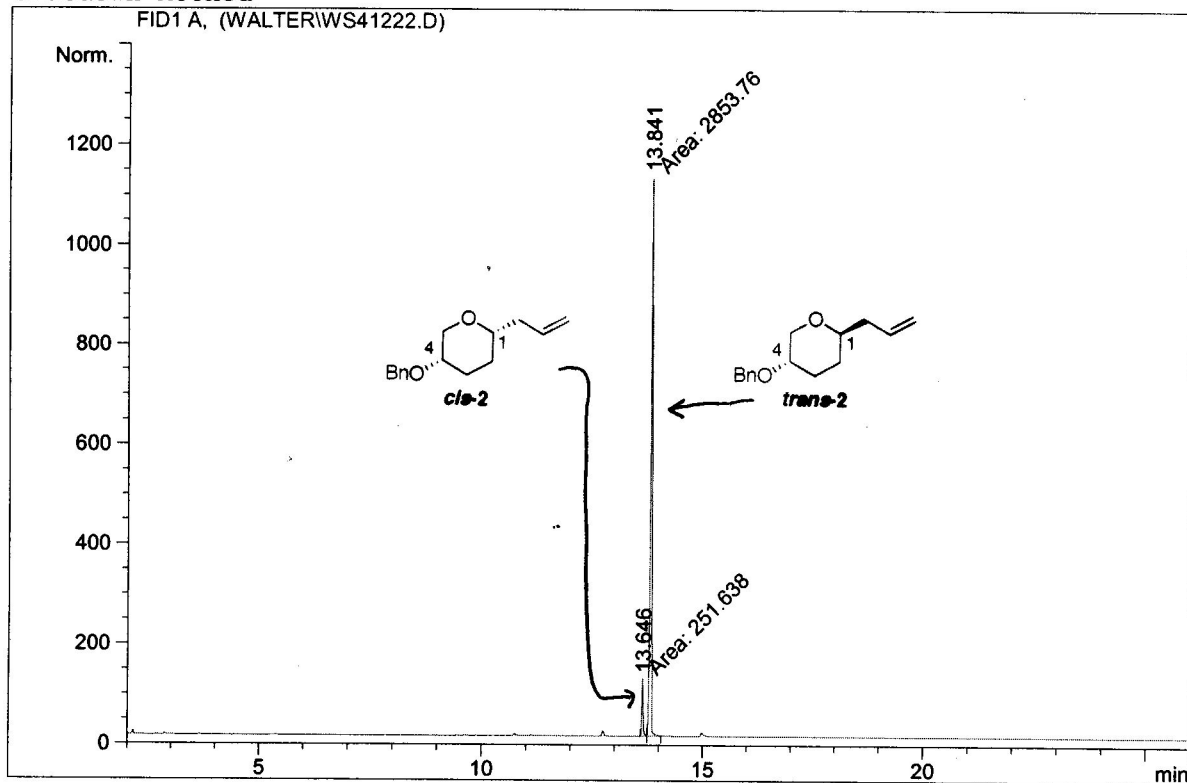
```

=====
Injection Date   : 2/21/2008 10:22:15 PM      Seq. Line   :    1
Sample Name     : WAS-IV-12-22                Location    : Vial 5
Acq. Operator  : jelena                       Inj         :    1
                                           Inj Volume  : 1 µl
                                           Actual Inj Volume : 5 µl

Different Inj Volume from Sequence !
Acq. Method    : C:\HPCHEM\1\METHODS\WALTER.M
Last changed   : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
Last changed   : 6/10/2008 9:29:12 AM by jelena
                (modified after loading)
  
```

Table 2, Entry 1

Shutdown Method



```

=====
Area Percent Report
=====
  
```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
  
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	13.646	MM	0.0365	251.63829	114.98837	8.10325
2	13.841	MM	0.0425	2853.76147	1119.56714	91.89675

```
Totals :                3105.39977 1234.55551
```

Results obtained with enhanced integrator!

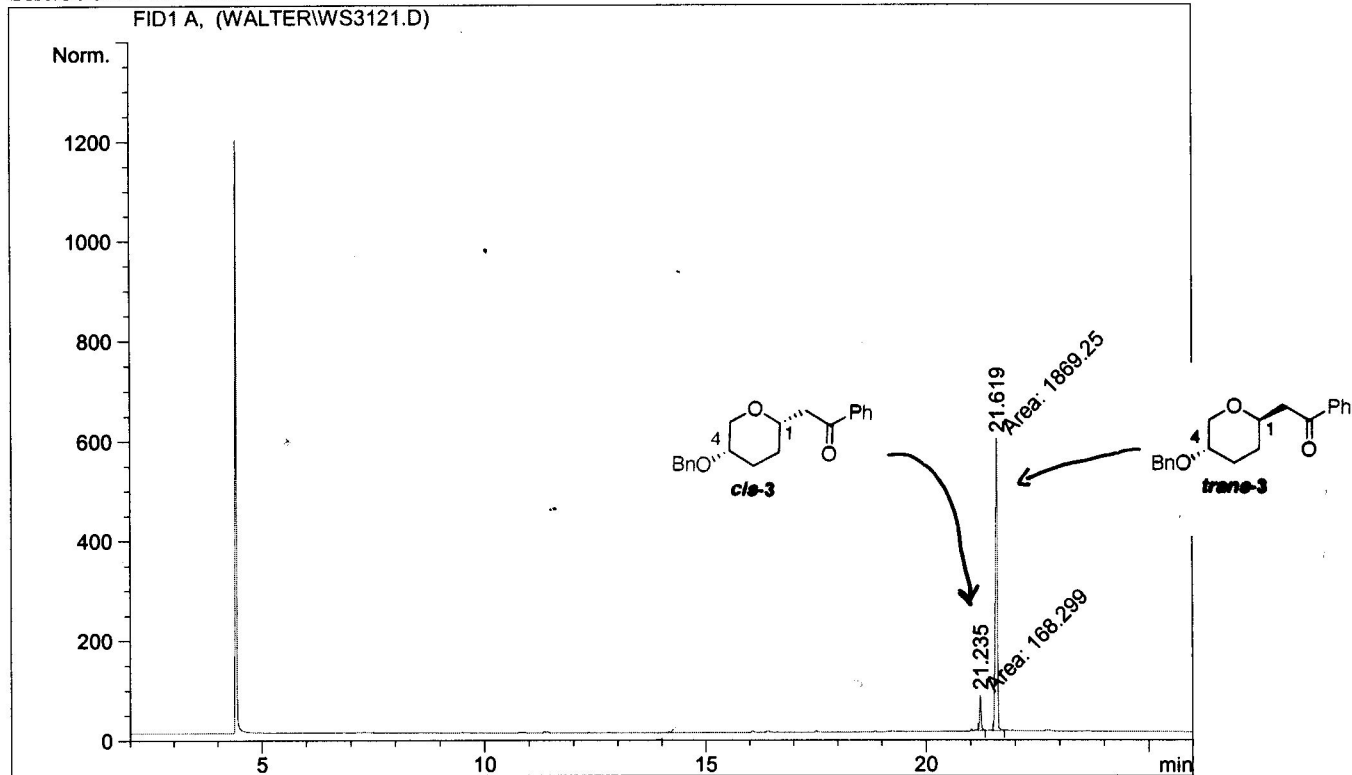
```

=====
*** End of Report ***
  
```

Injection Date : 6/22/2007 3:07:27 PM Seq. Line : 3
 Sample Name : WAS-III-121 Location : Vial 1
 Acq. Operator : jelena Inj : 1
 Inj Volume : 1 µl
 Different Inj Volume from Sequence ! Actual Inj Volume : 5 µl
 Acq. Method : C:\HPCHEM\1\METHODS\WALTER.M
 Last changed : 11/12/2005 3:11:30 PM by Susan
 Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
 Last changed : 6/10/2008 9:31:32 AM by jelena
 (modified after loading)

Table 2, Entry 2

Shutdown Method


 =====
 Area Percent Report
 =====

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	21.235	MM	0.0401	168.29939	70.01180	8.25990
2	21.619	MM	0.0533	1869.24854	584.89423	91.74010

Totals : 2037.54793 654.90602

Results obtained with enhanced integrator!

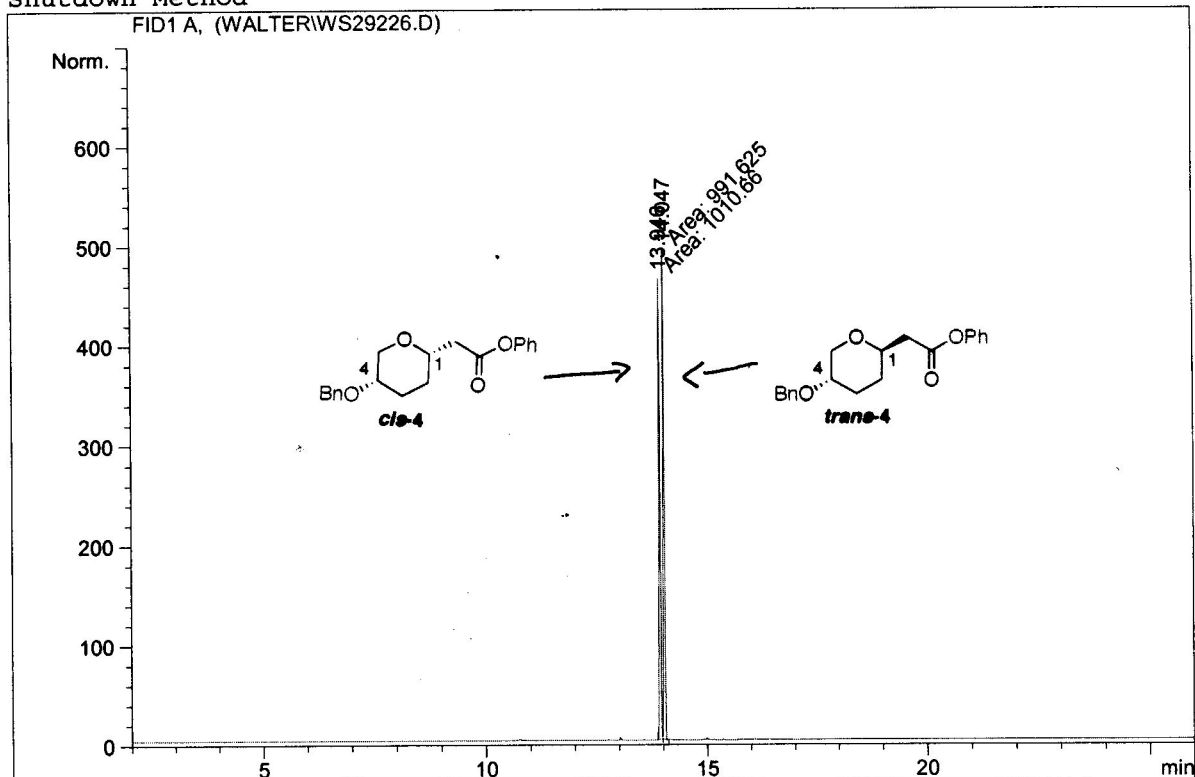
 =====
 *** End of Report ***

```

=====
Injection Date   : 11/4/2005 3:55:37 PM          Seq. Line   :    5
Sample Name     : WAS-II-92-26                  Location    : Vial 2
Acq. Operator   : jelena                        Inj         :    1
                                                    Inj Volume  : 1 µl
                                                    Actual Inj Volume : 5 µl
Different Inj Volume from Sequence !
Acq. Method     : C:\HPCHEM\1\METHODS\WALTER.M
Last changed    : 7/28/2005 3:54:19 PM by jelena
Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
Last changed    : 6/10/2008 9:33:41 AM by jelena
                (modified after loading)
  
```

Table 2, Entry 3

Shutdown Method



```

=====
                          Area Percent Report
=====
  
```

```

Sorted By           :      Signal
Multiplier          :      1.0000
Dilution            :      1.0000
  
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	13.946	MM	0.0364	1010.65692	462.99808	50.47527
2	14.047	MM	0.0337	991.62451	490.82718	49.52473

```
Totals :                      2002.28143  953.82526
```

Results obtained with enhanced integrator!

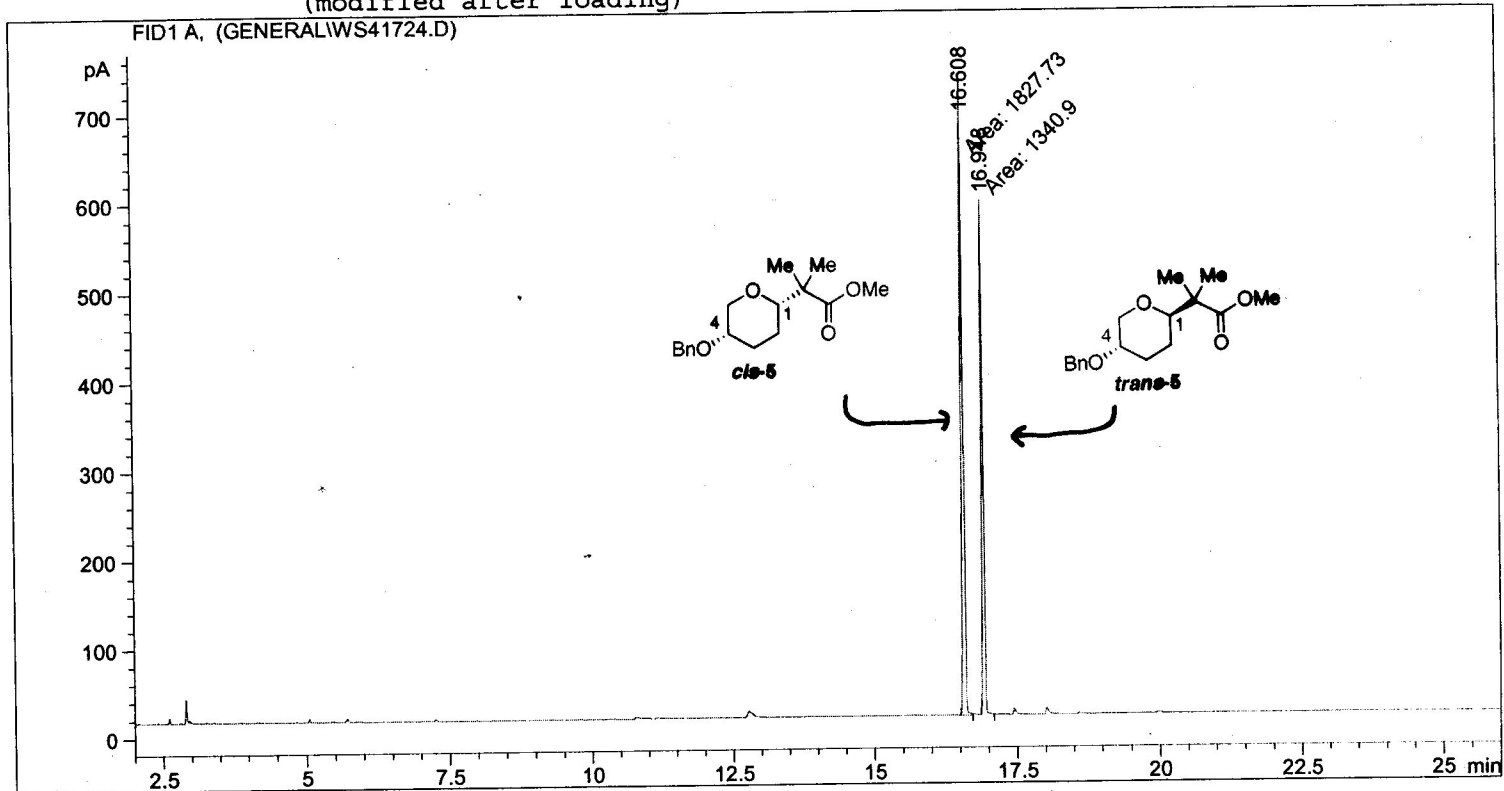
```

=====
*** End of Report ***
  
```

```

=====
Injection Date   : 3/3/2008 8:33:30 PM      Seq. Line   :    1
Sample Name     : WAS-IV-17-24             Location    : Vial 1
Acq. Operator  : jelena                    Inj         :    1
                                                Inj Volume  : 1 µl
                                                Actual Inj Volume : 5 µl
Different Inj Volume from Sequence !
Acq. Method    : C:\HPCHEM\1\METHODS\WALTER.M
Last changed   : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\OFF.M
Last changed   : 3/4/2008 9:19:13 AM by jelena
                (modified after loading)
    
```

Table 2, Entry 4



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	16.608	MM	0.0424	1827.72974	719.08203	57.68210
2	16.948	MM	0.0385	1340.89600	580.03223	42.31790

Totals : 3168.62573 1299.11426

Results obtained with enhanced integrator!

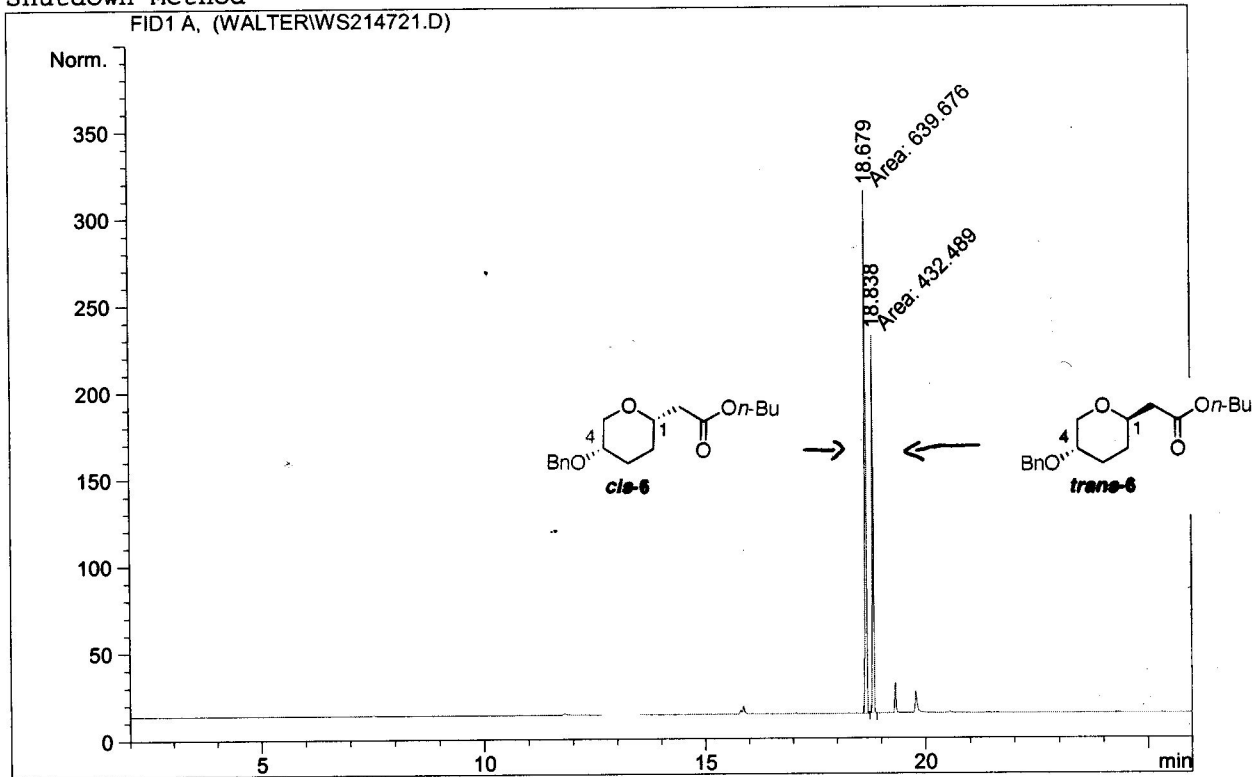
*** End of Report ***

```

=====
Injection Date : 4/12/2006 1:30:20 PM      Seq. Line : 2
Sample Name    : WAS-II-147-21             Location  : Vial 13
Acq. Operator  : Jelena                     Inj      : 1
                                                Inj Volume: 1 µl
                                                Actual Inj Volume: 3 µl
Different Inj Volume from Sequence !
Acq. Method    : C:\HPCHEM\1\METHODS\WALTER.M
Last changed   : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
Last changed   : 6/10/2008 9:36:06 AM by jelena
                (modified after loading)
    
```

Table 2, Entry 5

Shutdown Method



Area Percent Report

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	18.679	MM	0.0355	639.67639	300.33771	59.66213
2	18.838	MM	0.0332	432.48853	217.42862	40.33787

Totals : 1072.16492 517.76633

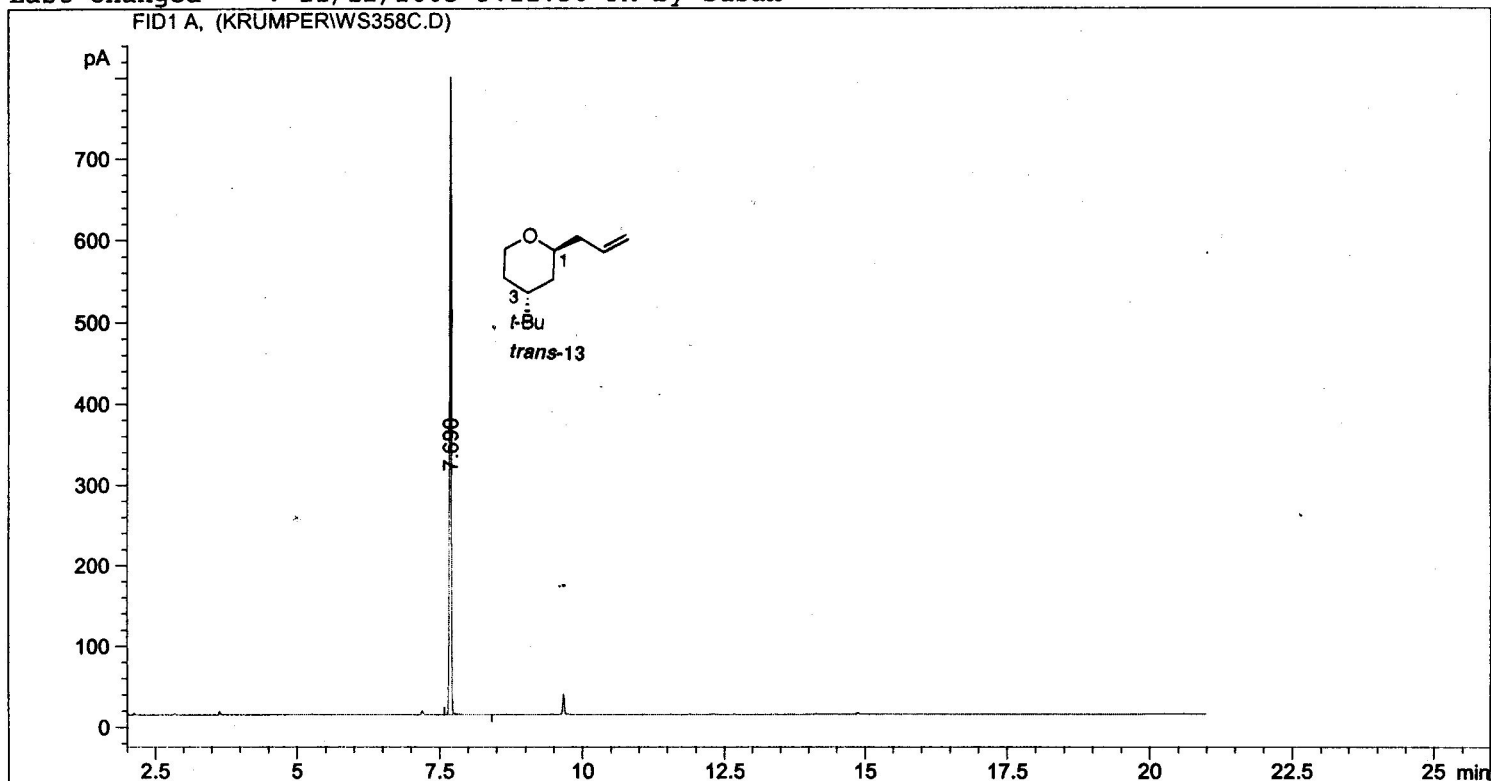
Results obtained with enhanced integrator!

*** End of Report ***

```

=====
Injection Date   : 2/5/2007 11:13:31 AM      Seq. Line   :    1
Sample Name     : WAS-III-58-c              Location    : Vial 1
Acq. Operator  : jennifer                   Inj         :    1
                                           Inj Volume  : 1 µl
                                           Actual Inj Volume : 5 µl
Different Inj Volume from Sequence !
Sequence File   : C:\HPCHEM\1\SEQUENCE\KRUMPER.S
Method          : C:\HPCHEM\1\METHODS\WALTER.M
Last changed    : 11/12/2005 3:11:30 PM by Susan
=====

```

Table 3, Entry 1

```

=====
                          Area Percent Report
=====

```

```

Sorted By           :      Signal
Multiplier          :      1.0000
Dilution            :      1.0000

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	7.690	BP	0.0875	1543.63904	294.10623	1.000e2

```
Totals :                1543.63904  294.10623
```

Results obtained with enhanced integrator!

```

=====
*** End of Report ***
=====

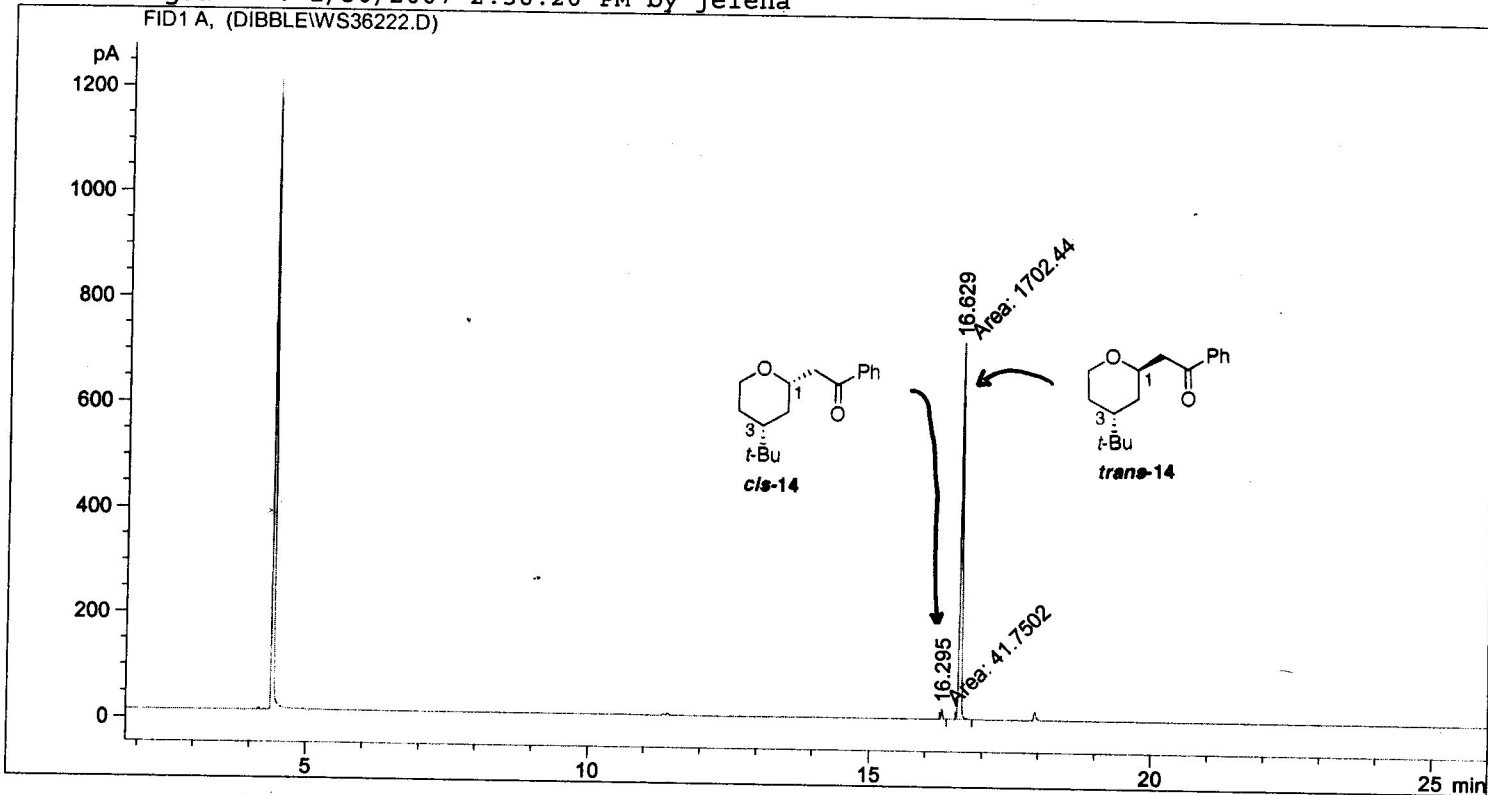
```



```

=====
Injection Date : 2/8/2007 5:43:13 PM      Seq. Line : 5
Sample Name    : WAS-III-62-22            Location  : Vial 11
Acq. Operator  : jennifer                  Inj      : 1
                                           Inj Volume : 1 µl
                                           Actual Inj Volume : 5 µl
Different Inj Volume from Sequence !
Acq. Method    : C:\HPCHEM\1\METHODS\WALTER.M
Last changed   : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\KRUMPER.M
Last changed   : 1/30/2007 2:38:20 PM by jelena
    
```

Table 3, Entry 2



=====
Area Percent Report
 =====

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	16.295	MM	0.0352	41.75023	19.78161	2.39368
2	16.629	MM	0.0397	1702.43750	715.33228	97.60632

Totals : 1744.18773 735.11388

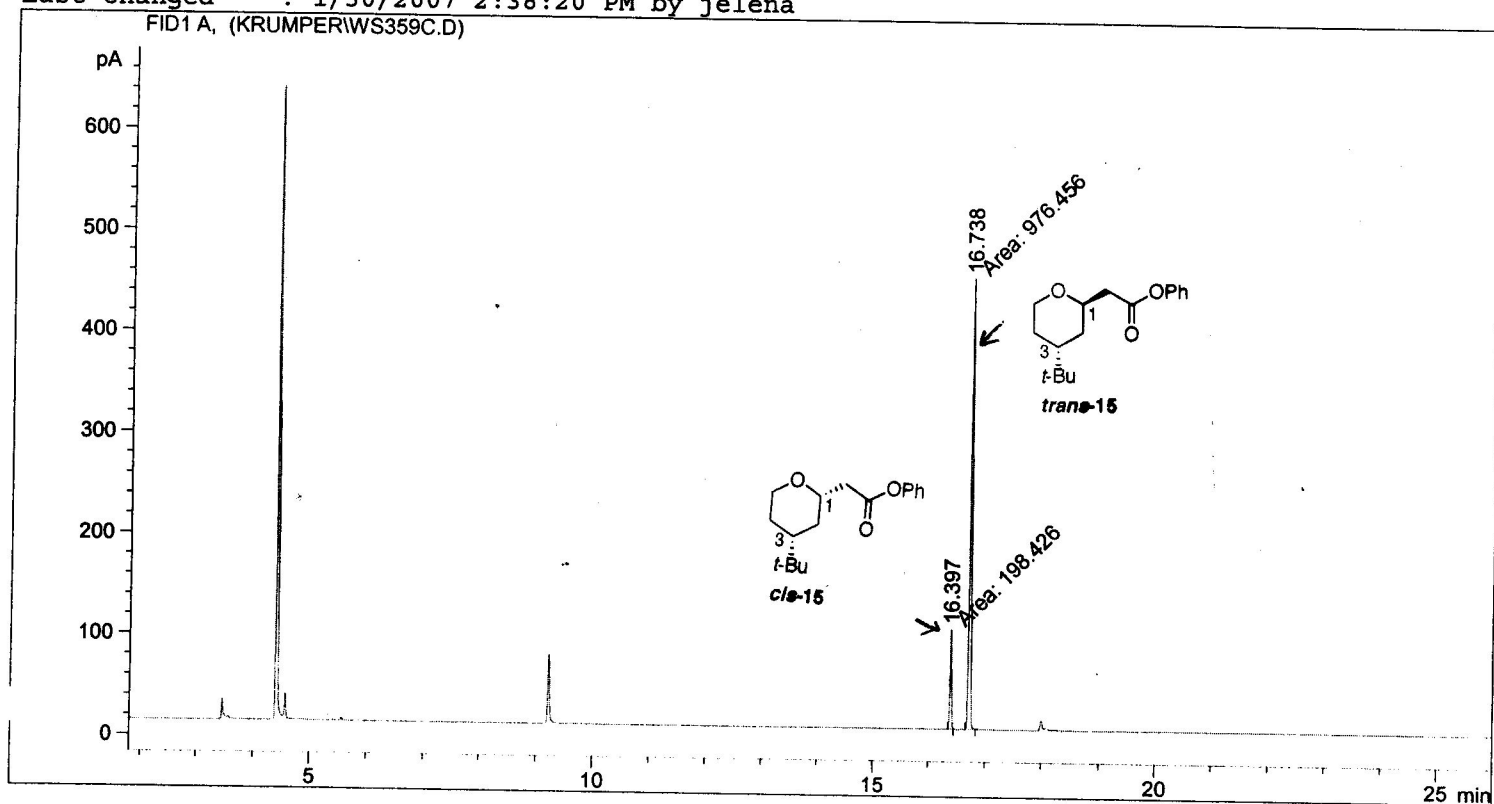
Results obtained with enhanced integrator!

=====
 *** End of Report ***

```

=====
Injection Date   : 2/5/2007 11:49:07 AM      Seq. Line   :    2
Sample Name     : WAS-III-59-c              Location    : Vial 2
Acq. Operator   : jennifer                  Inj         :    1
                                           Inj Volume  : 1 µl
Different Inj Volume from Sequence !      Actual Inj Volume : 5 µl
Acq. Method     : C:\HPCHEM\1\METHODS\WALTER.M
Last changed    : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\KRUMPER.M
Last changed    : 1/30/2007 2:38:20 PM by jelena
    
```

Table 3, Entry 3



Area Percent Report

```

Sorted By      : Signal
Multiplier    : 1.0000
Dilution      : 1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	16.397	MM	0.0338	198.42632	97.87888	16.88904
2	16.738	MM	0.0366	976.45593	444.12595	83.11096

Totals : 1174.88225 542.00483

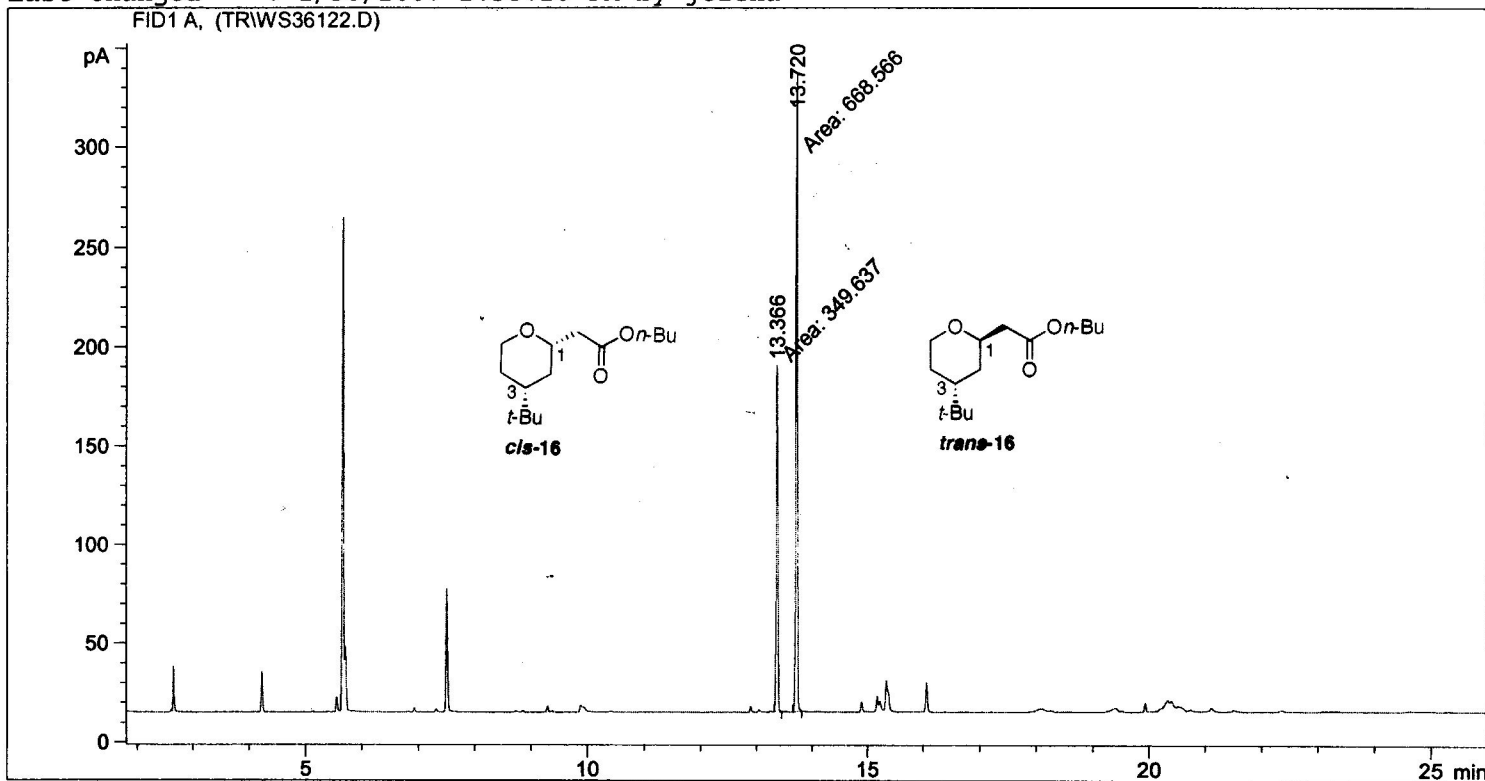
Results obtained with enhanced integrator!

*** End of Report ***

```

=====
Injection Date   : 2/6/2007 5:33:45 PM           Seq. Line   :    2
Sample Name     : WAS-III-61-22                 Location    : Vial 8
Acq. Operator   : jennifer                       Inj         :    1
                                                    Inj Volume  : 1 µl
                                                    Actual Inj Volume : 5 µl
Different Inj Volume from Sequence !
Acq. Method     : C:\HPCHEM\1\METHODS\WALTER.M
Last changed    : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\KRUMPER.M
Last changed    : 1/30/2007 2:38:20 PM by jelena
=====

```

Table 3, Entry 4

```

=====
Area Percent Report
=====

```

```

Sorted By       :      Signal
Multiplier      :      1.0000
Dilution        :      1.0000

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	13.366	MM	0.0333	349.63651	174.95299	34.33861
2	13.720	MM	0.0348	668.56580	319.88483	65.66139

```
Totals :                1018.20230  494.83781
```

Results obtained with enhanced integrator!

```

=====
*** End of Report ***
=====

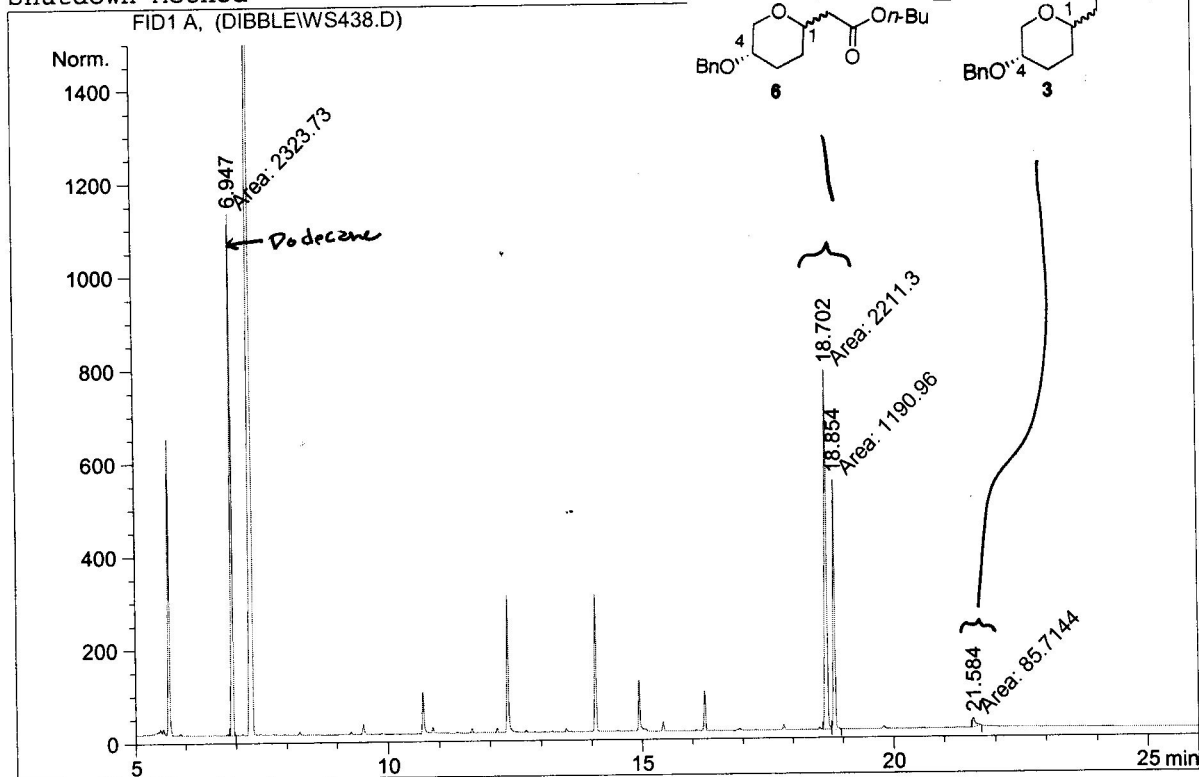
```

```

=====
Injection Date : 5/20/2008 10:44:47 AM      Seq. Line : 1
Sample Name    : WAS-IV-38                  Location  : Vial 2
Acq. Operator  : jelena                     Inj      : 1
                                                Inj Volume: 1 µl
                                                Actual Inj Volume: 5 µl
Different Inj Volume from Sequence !
Acq. Method    : C:\HPCHEM\1\METHODS\WALTER.M
Last changed   : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
Last changed   : 5/20/2008 11:49:48 AM by jelena
                (modified after loading)
    
```

Table 4, Entry 1

Shutdown Method



Area Percent Report

```

=====
Sorted By      : Signal
Multiplier    : 1.0000
Dilution      : 1.0000
    
```

Handwritten notes:

$\frac{2323}{1.0} : \frac{3401}{1.1} : \frac{86}{1.25}$

$2323 : 3092 : 69$

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	6.947	MM	0.0346	2323.73486	1119.60278	39.98368
2	18.702	MM	0.0479	2211.30249	769.95667	38.04910
3	18.854	MM	0.0370	1190.95605	536.26324	20.49236
4	21.584	MM	0.0728	85.71442	19.61160	1.47486

prod 6 : prod 3 = 98 : 2

GC yield prod 6 = 67%
prod 3 = 1%

cis-6 : trans-6
65 : 35

Totals : 5811.70783 2445.43429

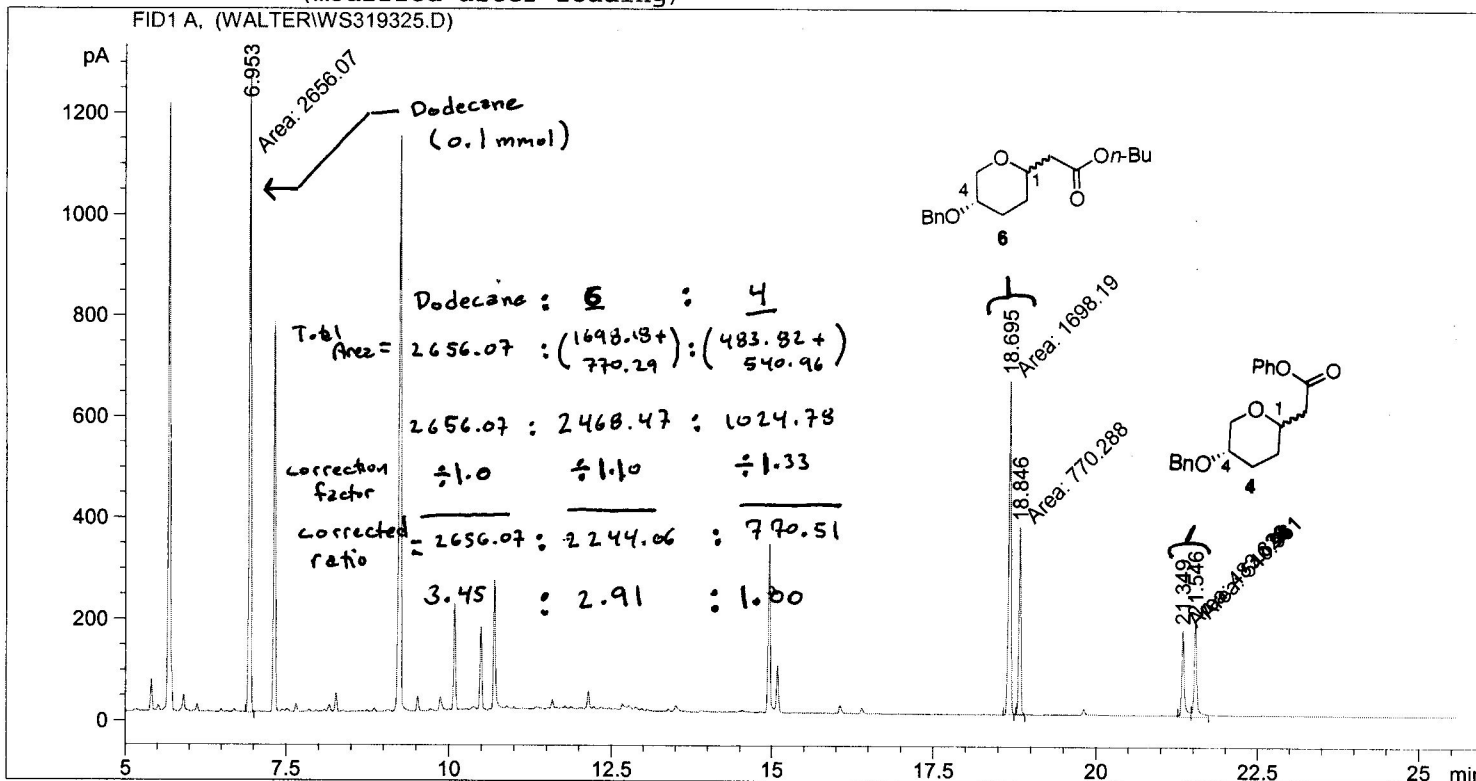
Results obtained with enhanced integrator!

*** End of Report ***

```

Injection Date : 1/4/2008 6:25:26 PM           Seq. Line :    3
Sample Name    : WAS-III-193-24                 Location  : Vial 7
Acq. Operator  : jelena                          Inj      :    1
                                                Inj Volume : 1 µl
                                                Actual Inj Volume : 5 µl
Different Inj Volume from Sequence !
Acq. Method   : C:\HPCHEM\1\METHODS\WALTER.M
Last changed  : 11/12/2005 3:11:30 PM by Susan
Analysis Method : C:\HPCHEM\1\METHODS\BEAVER1.M
Last changed  : 5/14/2008 12:38:03 PM by jelena
                (modified after loading)
    
```

Table 4, Entry 2



Area Percent Report

```

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
    
```

Dodecane : 5 : 4

corrected ratio 3.45 : 2.91 : 1.00

Corrected ratio 5 : 4
74 : 26

Theor yield was 0.20 mmol
Based on Dodecane (0.10 mmol added)

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	6.953	MM	0.0352	2656.06934	1258.35022	43.19284
2	18.695	MM	0.0430	1698.18530	658.24811	27.61579 ← cis-5
3	18.846	MM	0.0347	770.28845	369.60751	12.52639 ← trans-5
4	21.349	MM	0.0476	483.82300	169.45120	7.86790 ← cis-4
5	21.546	MM	0.0483	540.96106	186.53477	8.79708 ← trans-4

GC Yield 5 = 42%
GC Yield 4 = 14%

Totals : 6149.32715 2642.19182

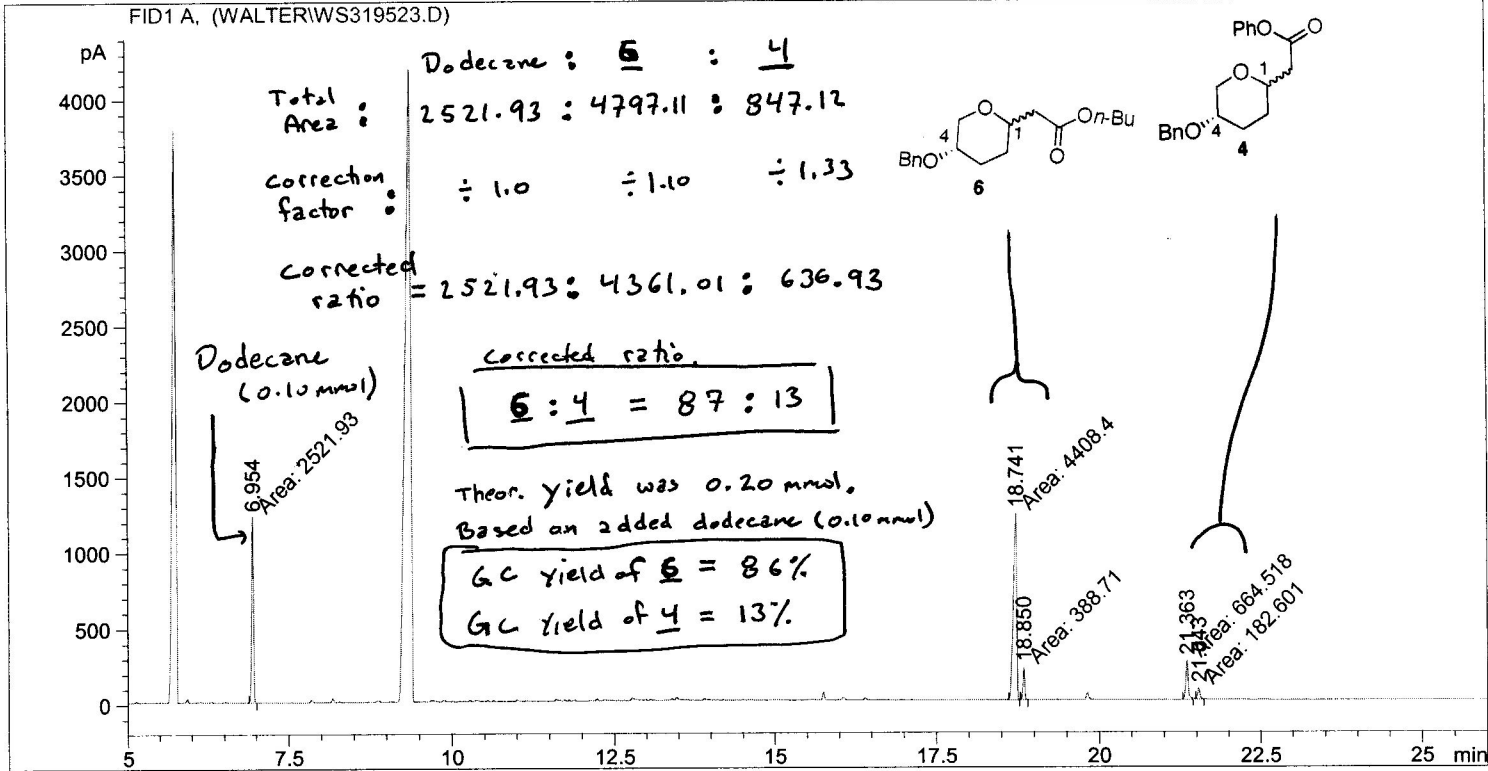
cis-5 : trans-5 = 69 : 31
cis-4 : trans-4 = 47 : 53

Results obtained with enhanced integrator!

*** End of Report ***

Injection Date : 1/9/2008 6:14:17 PM Seq. Line : 1
 Sample Name : WAS-III-195-23 Location : Vial 7
 Acq. Operator : jelena Inj : 1
 Inj Volume : 1 µl
 Different Inj Volume from Sequence ! Actual Inj Volume : 5 µl
 Acq. Method : C:\HPCHEM\1\METHODS\WALTER.M
 Last changed : 11/12/2005 3:11:30 PM by Susan
 Analysis Method : C:\HPCHEM\1\METHODS\BEAVER1.M
 Last changed : 5/14/2008 12:39:43 PM by jelena
 (modified after loading)

Table 4, Entry 3



Area Percent Report

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	6.954	MM	0.0344	2521.93359	1221.22583	30.88273
2	18.741	MM	0.0599	4408.39844	1226.80090	53.98373 ← cis-5
3	18.850	MM	0.0307	388.71021	211.15019	4.76001 ← trans-5
4	21.363	MM	0.0431	664.51801	256.71835	8.13746 ← cis-4
5	21.543	MM	0.0413	182.60135	73.71474	2.23607 ← trans-4

Totals : 8166.16159 2989.61002

Results obtained with enhanced integrator!

Handwritten Ratios:
 cis-5 : trans-5 = 92 : 8
 cis-4 : trans-4 = 78 : 22

*** End of Report ***

jrk-2-21.a crude GC after workup with dodecane IS

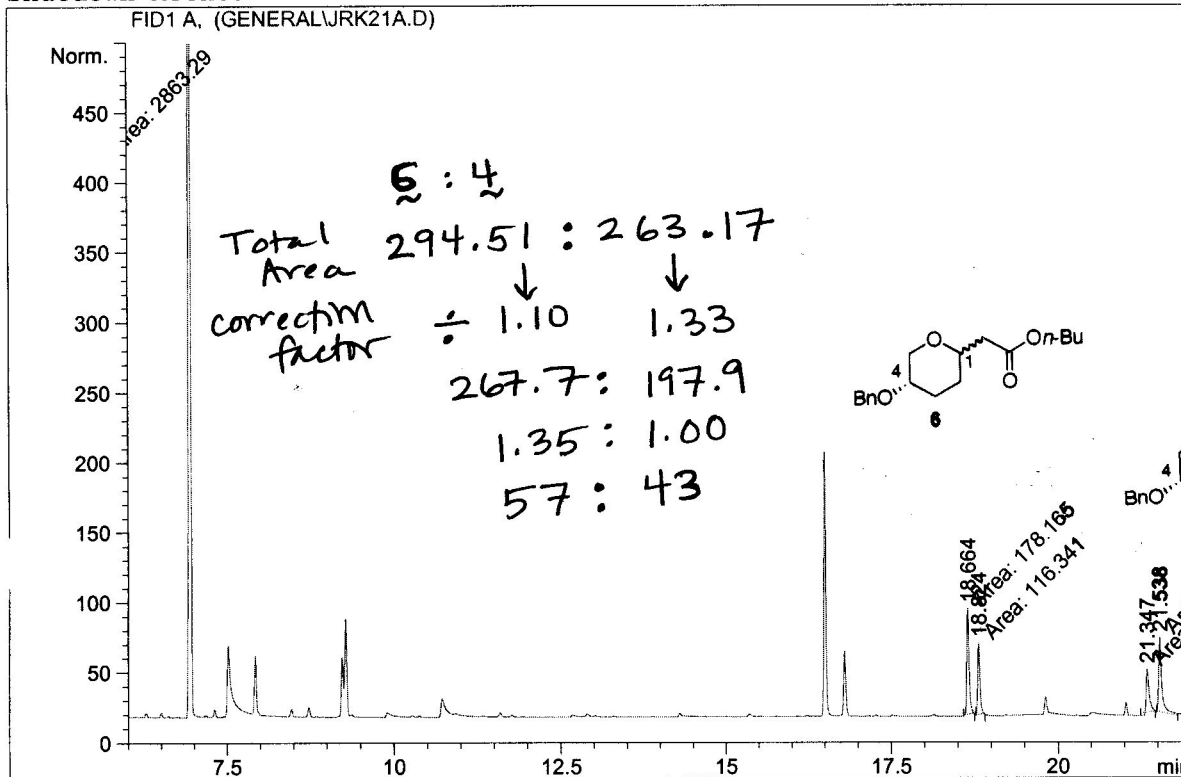
```

=====
Injection Date : 2/1/2008 5:55:12 PM      Seq. Line : 1
Sample Name    : jrk-2-21a                Location  : Vial 21
Acq. Operator  : jelena                    Inj      : 1
                                           Inj Volume: 1 µl
                                           Actual Inj Volume: 5 µl

Different Inj Volume from Sequence !
Acq. Method   : C:\HPCHEM\1\METHODS\KRUMPER.M
Last changed  : 9/5/2007 11:13:31 AM by jelena
Analysis Method : C:\HPCHEM\1\METHODS\SHUTDOWN.M
Last changed  : 5/20/2008 12:24:08 PM by jelena
                (modified after loading)
    
```

eq 1

Shutdown Method



Area Percent Report

```

Sorted By      : Signal
Multiplier    : 1.0000
Dilution      : 1.0000
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	5.698	MM	0.0354	2863.28760	1349.37781	83.69802
2	18.664	MM	0.0396	178.16505	75.03540	5.20802
3	18.824	MM	0.0382	116.34081	50.79527	3.40081
4	21.347	MM	0.0500	91.66496	30.53539	2.67950
5	21.538	MM	0.0541	171.51556	52.82352	5.01365

Handwritten notes: cis : trans 60 : 40 (for peaks 2 and 3), cis : trans 35 : 65 (for peaks 4 and 5).

Totals : 3420.97398 1558.56738

Results obtained with enhanced integrator!

*** End of Report ***