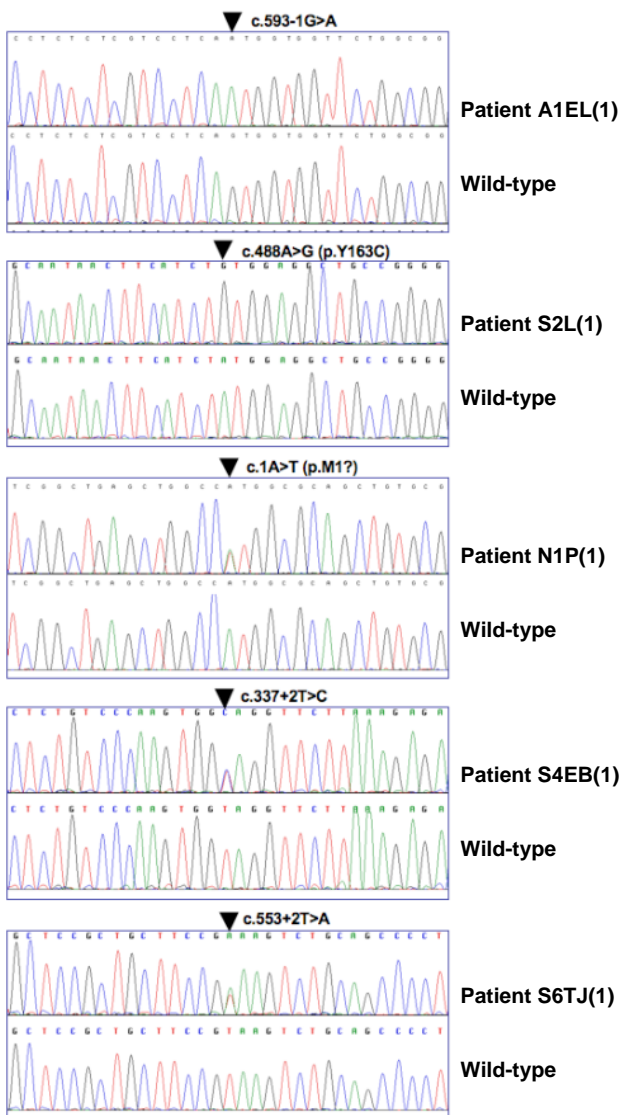


Supplemental Data

Mutations in *SPINT2* Cause a Syndromic Form of Congenital Sodium Diarrhea

Peter Heinz-Erian, Thomas Müller, Birgit Krabichler, Melanie Schranz, Christian Becker, Franz Rüschen-dorf, Peter Nürnberg, Bernard Rossier, Mihailo Vujic, Ian W. Booth, Christer Holmberg, Cisca Wijmenga, Giedre Grigelioniene, C. M. Frank Kneepkens, Stefan Rosipal, Martin Mistrik, Matthias Kappler, Laurent Michaud, Ludwig-Christoph Dóczy, Victoria Mok Siu, Marie Krantz, Heinz Zoller, Gerd Utermann, and Andreas R. Janecke

Figure S1. Detection of *SPINT2* Mutations in Syndromic CSD



Sequence chromatograms revealing 5 distinct *SPINT2* mutations (arrow heads) identified in patients with syndromic CSD.

Figure S2. MLPA Analysis Does Not Reveal *SPINT2* Copy Number Changes in CSD Patients



Multiplex ligation-dependent probe amplification (MLPA) analysis was conducted for each CSD patient without *SPINT2* mutations identified upon genomic sequencing and for patient S2L(1) who was identified with a homozygous p.Y163C mutation, and compared with the results of 8 control samples. Two panels are shown for each patient. Each upper panel displays the raw data of the fragment analysis including 3 probes corresponding to *SPINT2* exons 5, 2, and 7, respectively, and 17 control probes from different chromosomes contained in the MRC-Holland SALSA MLPA P-300-A1 kit. Fragment traces of patient samples are shown in blue and are each compared with the average of 8 control samples (in red). The size of the fragments is given in bases on the X-axis, and the amount of the fragments is displayed on the Y-axis. The results of the MLPA analyses are shown in the lower panels, and the relative copy numbers for each probe are scaled on the Y-axis. Blue bars correspond to *SPINT2* exons 5, 2, and 7, displayed at 100, 105 and 113 bases, respectively, and black bars correspond to control probes. Control probes displayed at 208 and 238 bases correspond to X- and Y-chromosomal loci, respectively.

Table S1. Primer Sequences for Amplification and Sequencing of *SPINT2* in Genomic DNA

| Name | Sequence | fragment length (bp) |
|-------------|-------------------------|----------------------|
| SPINT2 1f | ACCTGATCGCGAGACCCC | 285 |
| SPINT2 1r | GAACGCCATCAAGTAGCCC | |
| SPINT2 2f | GATTGCCCTGCCAAGCTAAC | 242 |
| SPINT2 2rnn | AACAAAAGCTCCA ACTACTGCC | |
| SPINT2 3f | CTGGCAGTCTCTCGAAAGC | 288 |
| SPINT2 3r | CCCATAAGGATGCTGGAGC | |
| SPINT2 4f | GCCCAGCCTCCCTAACAC | 193 |
| SPINT2 4rnn | AGGATGGTCTTGATATCCTGACC | |
| SPINT2 5f | TCAGGCACTTTCTGGCTTGC | 366 |
| SPINT2 5r | GGCTTAGAGGCCTTGCTGC | |
| SPINT2 6f | CCATGGAGGCCCTGGCTG | 265 |
| SPINT2 6rnn | TCACGCAGAAACATGACTTTCTG | |
| SPINT2 7f | ACTCTGGCTGCAACTCCCC | 310 |
| SPINT2 7r | ACTCAAATCCGAGTCAATCCC | |
| SPINT2 7R3 | CACCATCACGAACAGCCCC | 89 |

Table S2. Primer Sequences to Amplify *SPINT2* cDNA in Two Overlapping Fragments

| | |
|------------|-----------------------|
| SPINT2 1f | ACCTGATCGCGAGACCCC |
| SPINT2 c5r | gtgactgcggtggcgggtgc |
| SPINT2 c4f | tcctctgtcccaagtgtcc |
| SPINT2 7r | ACTCAAATCCGAGTCAATCCC |

Table S3. Oligos for *SPINT2* cDNA Cloning

| | |
|---------|---------------------------|
| TopoF | CACCATGGCGCAGCTGTGCGGGC |
| V5R | CAGGACATATGTGTTCTTCACCAGC |
| TopoM1F | CACCTTGGCGCAGCTGTGCGGGC |

Table S4. Synthetic MLPA Probes to Target Six out of the Seven *SPINT2* Exons

| Exon | | | Complete probe length (bases) |
|------|------------|--|-------------------------------|
| # | Name | 5' modification and 5'-3' sequence including primer annealing sequence | |
| 1 | SPINT2 1_5 | FAM-ggggtccctaaggggttgaGGCTGAGGCGGAGCCGGGCGTTTCTCG | 96 |
| | SPINT2 1_3 | Pho-CCCTGCTGGGATCGCTGCTCCTCTCTGtctagattggatcttgctggcac | |
| 2 | SPINT2 2_5 | FAM-ggggtccctaaggggttgaCAATGTCACTGACGGATCCTGCCA | 105 |
| | SPINT2 2_3 | Pho-GCTGTTTGTGTATGGGGGCTGTGACGGAAACAGCAATAAtctagattggatcttgctggcac | |
| 5 | SPINT2 5_5 | FAM-ggggtccctaaggggttgaACGCTGGTACTTTGACGTGGAGAGGAACT | 100 |
| | SPINT2 5_3 | Pho-CCTGCAATAACTTCATCTATGGAGGCTGtctagattggatcttgctggcac | |
| 7 | SPINT2 7_5 | FAM-ggggtccctaaggggttgaAGGAACCAGGAGCGTGCCCTGCGCACCGTCTGGAG | 113 |
| | SPINT2 7_3 | Pho-CTCCGAGATGACAAGGAGCAGCTGGTGAAGAACACtctagattggatcttgctggcac | |
| 4 | SPINT2 4_5 | FAM-ggggtccctaaggggttgaCAGCTCCCAGAAGGCAGGATTCTGAAGACCACTCCAGC | 117 |
| | SPINT2 4_3 | Pho-GATATGTTCAACTATGAAGGTAAAACCTCAAAGAGGCtctagattggatcttgctggcac | |
| 3 | SPINT2 3_5 | FAM-ggggtccctaaggggttgaCTCTTTCCTGTGCTGTTTCTTTGTCCCCTTGCAGAGAATGCCACGGG | 122 |
| | SPINT2 3_3 | Pho-ACGGGTGACCTGGCCACCAGCAGGAATGCAGCtctagattggatcttgctggcac | |

Synthetic probes were designed according to MRC-Holland (Amsterdam, The Netherlands), http://www.mrc-holland.com/pages/support_desing_synthetic_probepag.html. MLPA analysis was performed using standard conditions.