LEGENDS TO SUPPLEMENTARY FIGURES

Fig. S1. Amplification of cDNAs by the primers designed on the basis of the EST information. The cDNA fragments for *CURS* (A) and DCS (B) were amplified.

Fig. S2. Multiple alignment of the amino acid sequences of plant type III PKSs. The red box indicates the three amino acids forming the Cys-His-Asn catalytic triad that is essential for starter substrate loading and malonyl-CoA condensation. DCS, diketide-CoA synthase; CURS, curcumin synthase; MsCHS, a chalcone synthase from *Medicago sativa* (DNA

accession no. AAA02824); RhBAS, a benzalacetone synthase from *Rheum palmatum* (AAK82824); WtPKS1, a polyketide synthase from *Wachendorfia thyrsiflora* (AAY727928).

Fig. S3. Expression of *DCS* (A) and *CURS* (B) in turmeric. The relative expression was quantified by qPCR and normalized to 18S rRNA.

<u>Fig. S4.</u> SDS-PAGE of purified DCS and CURS by Ni-NTA column. Lane 1, marker; lane 2, purified DCS and CURS.

<u>Fig. S5.</u> Analysis of the compound derived by hydrolysis of feruloyl-diketide-CoA and *trans*-5-(4-hydroxy-3-methoxyphenyl)-3-oxopent-4-enoic acid methyl ester. The alkaline hydrolysis of feruloyl-diketide-CoA [**3b**] (*A*) and *trans*-5-(4-hydroxy-3-methoxyphenyl)-3-oxopent-4-enoic acid methyl ester (*B*) resulted in the formation of the same compound, *trans*-5-(4-hydroxy-3-methoxyphenyl)-3-oxopent-4-enoic acid [**3c**]. *Trans*-5-(4-Hydroxy-3-methoxyphenyl)-3-oxopent-4-enoic acid [**3c**]. *Trans*-5-(4-Hydroxy-3-methoxyphenyl)-3-oxopent-4-enoic acid [**3c**] is readily decarboxylated to yield dehydrozingerone [**3d**]. A methyl ester synthesized from *trans*-5-(4-hydroxy-3-methoxyphenyl)-3-oxopent-4-enoic acid, as a reference, was hydrolyzed by 1 M KOH just before LC-ESIMS analysis. The UV (*C*), MS (*E*), and MS/MS (*G*) spectra of the compound derived by hydrolysis of feruloyl-diketide-CoA are shown, together with the UV (*D*), MS (*F*), and MS/MS (*H*) spectra, of the hydrolysis product of the control methyl ester.

Fig. S6. HPLC analysis of the products after co-incubation of DCS and CURS in the presence of feruloyl-CoA [3a], *p*-coumaroyl-CoA [2a], and malonyl-CoA (*A*) and a similar HPLC analysis of an ethyl acetate extract of the rhizome of turmeric (*B*). Curcumin [3e], demethoxycurcumin [4e], and bisdemethoxycurcumin [2e] were detected in both analyses, although the abundance ratios of curcumin [3e] to demethoxycurcumin [4e] and bisdemethoxycurcumin [2e] were different.

Gene	Primer sequence (5' to 3')
DCS (forward)	CAACAGCACGCCCCAGTCGA
DCS (reverse)	GTGCTGTTCATCCTGGACGAG
CURS (forward)	TCAGCTCATCCATCACGAAGTACAC
CURS (reverse)	CATCATTGACGCCATCGAAGC
18S rRNA (forward)	CCTTCCTCTAAATGATAAGGTTCAATGG
18S rRNA (reverse)	GATTGAATGGTCCGGTGAAGTGTT

Supplementary Table 1. Primers used for quantitative real time PCR



Fig. S1. Katsuyama et al.

DCS CURS	1	MEANGYRITHSADGPATILAIGTANPTNVVDQNAYPDFYFRVTNSEY MANLHALRREQRAQGPATIMAIGTATPPNLYEQSTFPDFYFRVTNSDD
RhBAS	1	MVSVSEIRKAORAEGPATILAIGTANPANCVEOSTMPDFYFKITNSEH MATEEMKKIATVMAIGTANPPNCVYOADFPDFYFRVTNSDH
WtPKS1	1	MAPTTTMGSALYPLGEMRRSORADGLAAVLAIGTANPPNCVTOEEIPDFYFRVTNSDH MASTEGIQAYRNNMAEGPATIMAIGTANPPNVVDASTFPDYYWRVTNSEH
DCS	48	LQ - ELKAKFRRICEKAAIRKRHLYLTEEILRENPSLLAPMAPSFDARQAIVVEAVPKLAK
CURS	49	KQ - ELKKKFRRMCEKTMVKKRYLHLTEEILKERPKLCSYKEASFDDRQDIVVEEIPRLAK
MSCHS	49	KT - ELKEKFORMCDKSMIKBRYMYLTEEILKENPNYCEYMAPSLDARODMVVEVPRLGK
RhBAS	42	LI – NLKQKFKRLCENSRIEKRYLHVTEEILKENPNIAAYEATSLNVRHKMQVKGVAELGK
OsCUS	59	LT – ALKDKFKRICQEMGVQRRYLHHTEEMLSAHPEFVDRDAPSLDARLDIAADAVPELAA
WtPKS1	51	LSPEYRVKLKRICERSSIRKRHLVLTEQLLKENPTLTTYVDASYDERQSIVLDAVPKLAC
DCS	107	EAAEKAIKEWGRPKSDITHLVFCSASGIDMPGSDLQLLKLLGLPPSVNRVMLYNVGCHAG
CURS	108	EAAEKAIKEWGRPKSEITHLVFCSISGIDMPGADYRLATLLGLPLTVNRLMIYSQACHMG
Machs	108	EAAWKAIKEWGORKSKITHLUVCTTSGVDMPGADYOLTKLLGLPRYVVPYMMYOOGGEAG
RhBAS OsCUS WtPKS1	101 118 111	EAALKAIKEWGQPKSKITHLIVCTIDGVDMFGADIQHIKLLGLNFIVKKIMHIQQGCFAG EAALKAIKEWGQPKSKITHLIVCCLAGVDMPGADYQLTKLLDLDPSVKRFMFYHLGCYAG EAAKKAIAEWGRPAADITHLVVTTNSGAHVPGVDFRLVPLLGLRPSVRRTMLHLNGCFAG EAAAKAIKEWGRPKTDITHMVVCTGAGVDVPGVDYKMMNLLGLPPTVNRVMLYNVGCHAS
DCS	167	GTALRVAKDLAENNRGARVLAVCSEVTVLSYRGPHPAHIESLFVQALFGDGAAALVVGSD
CURS	168	AAMLRIAKDLAENNRGARVLVVACEITVLSFRGPNEGDFEALAGQAGFGDGAGAVVVGAD
MsCHS	168	GTVLRLAKDLAENNKGARVLVVCSEVTAVTFRGPSDTHLDSLVGQALFGDGAAALIVGSD
RhBAS	161	GTVLRLAKDIAENNKGARVLIVCSEMTTTCFRGPSETHLDSMIGQAILGDGAAAVIVGAD
OsCUS	178	CAALRLAKDLAENSRGARVLVVAAELTLMYFTGPDEGCFRTLLVQGLFGDGAAAVIVGAD
WtPKS1	171	GTVLRIAKDLAENNKGARVLVVSSEVSVMFFRGPAEGDVEILLGQALFGDGSAAIIVGAD
DCS	227	PVDGVERPIFEIASASQVMLPESAEAVGGHLREIGLTFHLKSQLPSIIASNIEQSLTTAC
CURS	228	PLEGIEKPIYEIAAAMQETVAESQGAVGGHLRAFGWTFYFLNQLPAIIADNLGRSLERAL
MsCHS	228	PVPEIEKPIFEMVWTAQTIAPDSEGAIDGHLREAGLTFHLLKDVPGIVSKNITKALVEAF
RhBAS	221	PDLTVERPIFELVSTAQTIVPESHGAIEGHLLESGLSFHLYKTVPTLISNNIKTCLSDAF
OsCUS	238	- ADDVERPLFEIVSAAQTIIPESDHALNMRFTERRLDGVLGRQVPGLIGDNVERCLLDMF
WtPKS1	231	PIEGVEKPIFQIFSASQMTLPEGEHLVAGHLRELGLTFHLKPQLPNTVSSNIHKPLKKAF
DCS	287	SPLGLSD WNOLFWAVHPGGRAILDQVEARLGLEKDRLAATRHVLSEYGNMQSATVL
CURS	288	APLGVRE WNDVFWVAHPGNWAIIDAIEAKLQLSPDKLSTARHVFTEYGNMQSATVY
MsCHS	288	EPLGISD YNSIFWIAHPGGPAILDQVEQKLALKPEKMNATREVLSEYGNMSSACVL
RhBAS	281	TPLNISD WNSLFWIAHPGGPAILDQVTAKVGLEKEKLKVTRQVLKDYGNMSSATVF
OsCUS	297	GPLLGGDGGGGWNDLFWAVHPGSSTIMDQVDAALGLEPGKLAASRRVLSDYGNMSGATVI
WtPKS1	291	EPLNITD WNSIFWIVHPGGRAILDQVQEKIGLEENKLDVSRYVLAENGNMMSASVF
DCS	343	FILDEMRNRSAAEGHATTGEGLDWGVLLGFGPGLSIETVVLHSCRLN-
CURS	344	FVMDELRKRSAVEGRSTTGDGLQWGVLLGFGPGLSIETVVLRSMPL
MsCHS	344	FILDEMRKKSTQNGLKTTGEGLEWGVLFGFGPGLTIETVVLRSVAI
RhBAS	337	FIMDEMRKKSLENGQATTGEGLEWGVLFGFGPGITVETVVLRSVPVIS
OsCUS	357	FALDELRRQRKEAAAAGEWPELGVMMAFGPGMTVDAMLLHATSHVN
WtPKS1	347	FIMDEMRKRSAAQGCSTTGEGHEWGVLFGFGPGLSIETVVLHSVPLSI

Fig. S2. Katsuyama et al.



Fig. S3. Katsuyama et al.



Fig. S4 Katsuyama et al.



Fig. S5. Katsuyama et al.



Fig. S6. Katsuyama et al.