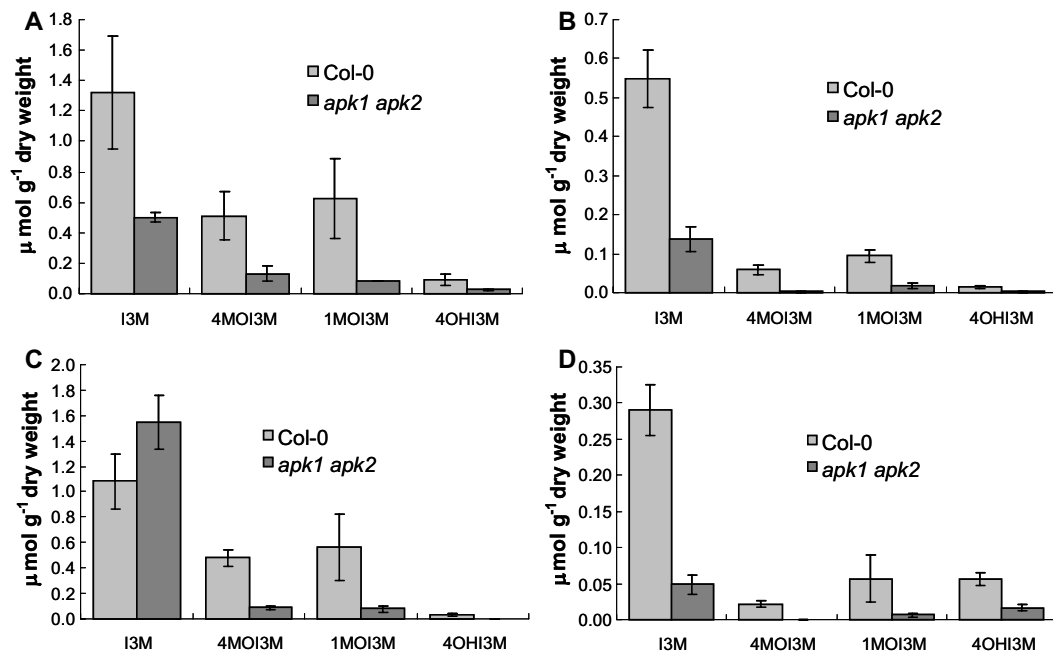
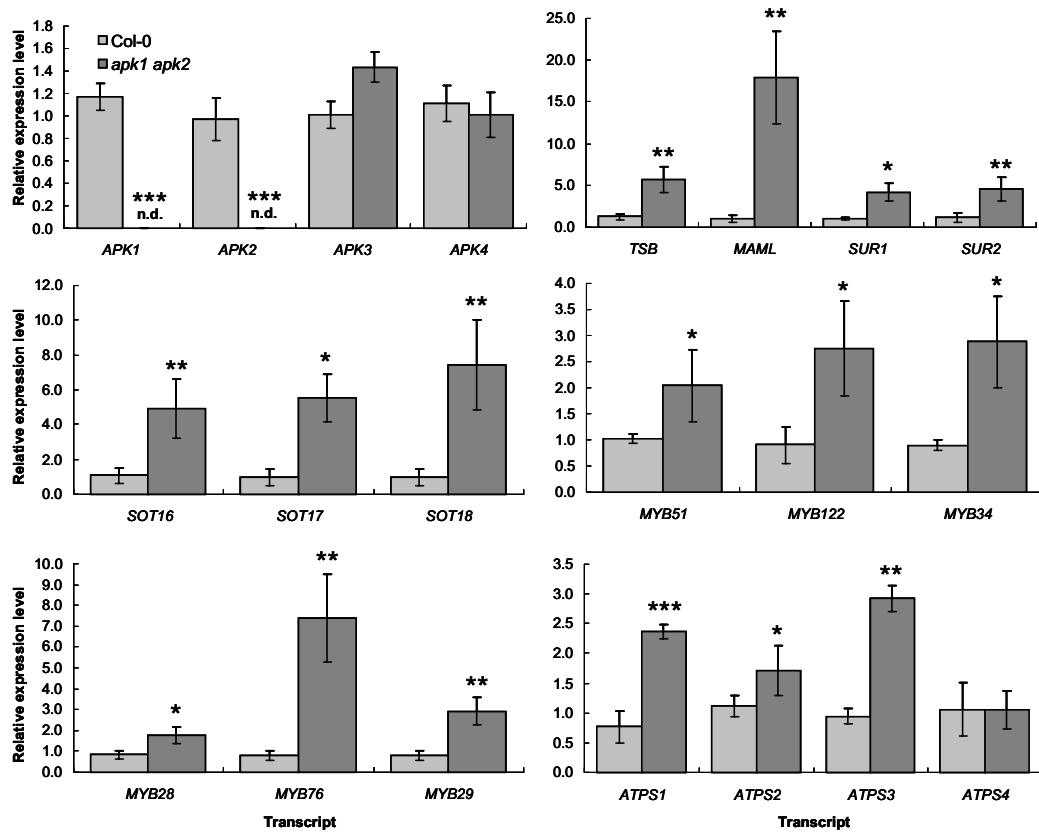


**Supplemental Figure 2. Accumulation of glucosinolates in 2-week old plants grown in hydroculture.** Sterilised seeds of WT and *apk1 apk2* were placed in 250 ml flasks containing 50 ml of 0.5 x MS. The flasks were placed in a growth chamber under constant light conditions for two weeks. Data are presented as mean  $\pm$  SD from three biological replicates. Asterisks denote values significantly different from WT at  $P < 0.001$ .

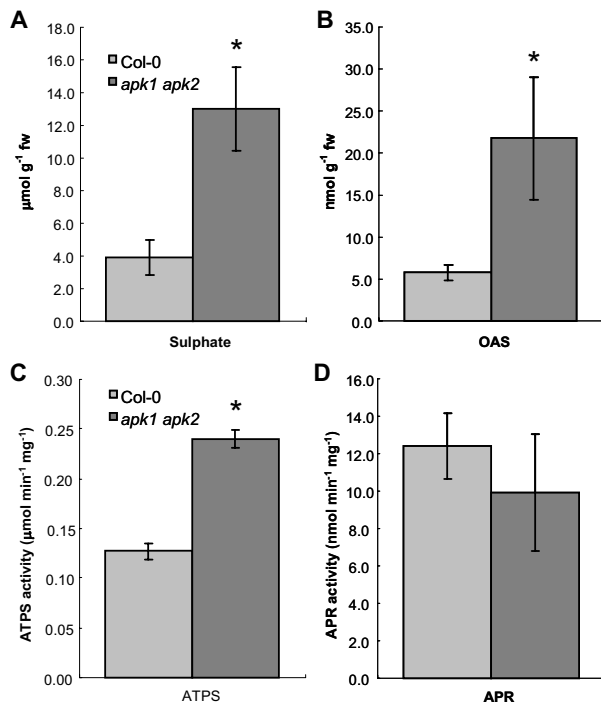


**Supplemental Figure 3. Variation in levels of indolic glucosinolates.**

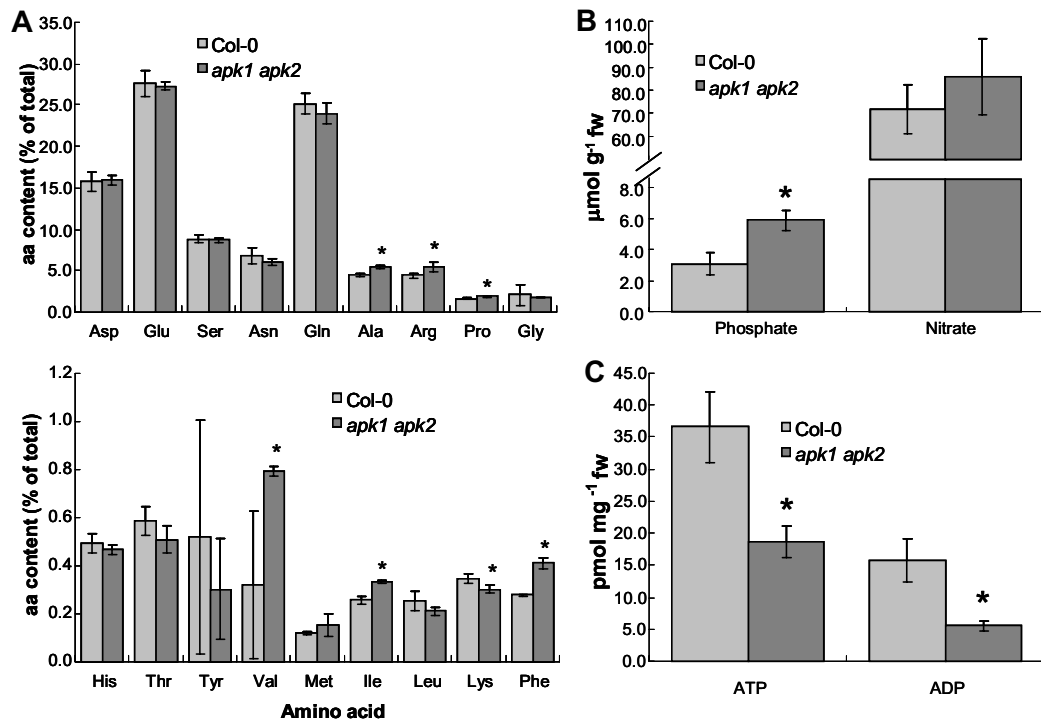
Four independent measurements of indolic GLs in leaves of WT and *apk1 apk2* plants. Plants for each experiment were grown separately. Data are presented as means  $\pm$  SD from five (C) or four (A,B,D) biological replicates. (A) represents the dataset used for Figure 6 in the paper.



**Supplemental Figure 4. Quantitative Real-Time RT-PCR analysis of glucosinolate biosynthetic genes and ATPS gene family in *Col-0* and *apk1 apk2* knockout mutant.** PCR was performed on cDNA synthesised from total RNA extracted from the rosette leaves of 5-week-old plants. Data are presented as means  $\pm$  SD from four biological replicates. Asterisks denote values significantly different from *Col-0* at  $P < 0.05$  (\*),  $P < 0.01$  (\*\*) and  $P < 0.001$  (\*\*\*).

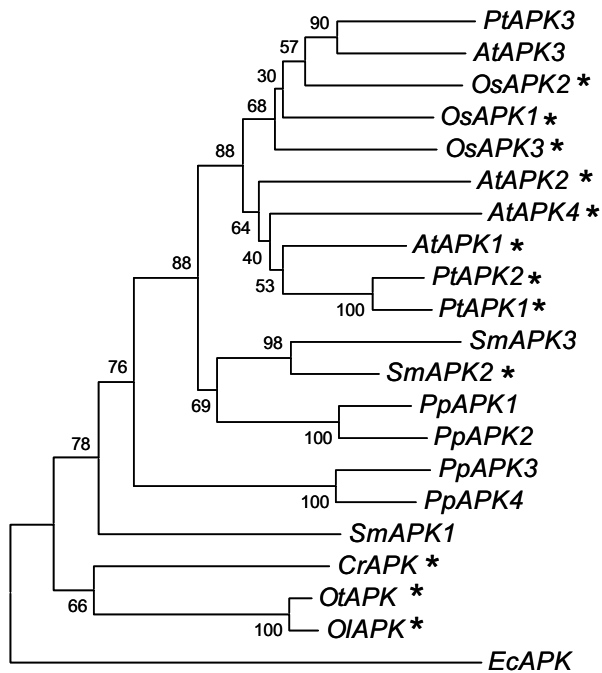


**Supplemental Figure 5. Analysis of sulphate assimilation in *apk1 apk2* plants.** Concentration of (A) sulphate and (B) *O*-acetylserine were determined in rosette leaves of 5-week old plants by HPLC. Enzyme activity of (C) ATPS and (D) APR was measured in the same rosette leaves. Data are presented as means  $\pm$  SD from four biological replicates. Asterisks denote values significantly different from Col-0 at  $P < 0.05$ .



**Supplemental Figure 6. Metabolite analysis of *apk1 apk2* plants.**

Concentration of (A) amino acids, (B) nitrate and phosphate, and (C) ATP and ADP were determined in rosette leaves of 5-week old plants by HPLC. Data are presented as means  $\pm$  SD from three to four biological replicates. Asterisks denote values significantly ( $P < 0.05$ ) different from WT values.



**Supplemental Figure 7. Phylogenetic analysis of APS kinases from plants and green algae.**

Neighbor-joining tree was constructed from alignment of amino acid sequences of APS kinases from *A. thaliana* (At), *Oryza sativa* (Os), *Populus tremuloides* (Pt), *Selaginella moellendorffii* (Sm), *Physcomitrella patens* (Pp), the green algae *Chlamydomonas reinhardtii* (Cr), *Ostreococcus tauri* (Ot) and *Ostreococcus lucimarinus* (Ol), and *Escherichia coli* (Ec) as an outgroup. Numbers represent results of bootstrap analysis with 1000 replicates. Asterisks mark sequences possessing putative plastid targeting peptides.

**Supplemental Table 1.** Sequence identity in percent identical residues of Arabidopsis APS kinase isoforms and the APK from *E. coli*. The numbers in top right part of the table represent identity of the mature proteins on amino acid level, while the bottom left part denotes the identity at nucleotide level. The alignments were performed with the program ALIGN of Biology WorkBench 3.2 using the default parameters.

	APK1	APK2	APK3	APK4	EcAPK
APK1		72.5	71.6	68.8	46.6
APK2	60.4		67	65.5	44.9
APK3	51.6	48.3		62	45.2
APK4	58	58.6	48.9		47
EcAPK	44.7	47.2	39.8	44.2	



**Supplemental Table 2.** Fold-changes of transcription levels of genes of primary sulphur metabolism and metabolism of sulphated compounds in *apk1 apk2* mutant. n/e = not expressed, n/r = not represented on Affymetrix chip. Transcripts highlighted in bold are significantly altered in the *apk1 apk2* mutant at  $q < 0.05$  or as determined by qRT-PCR at  $p < 0.05$  (compare Fig. 8 and Supplemental Figure 4).

Gene	AGI code	Description	Fold-change	p value	q value
SULTR1,1	At4g08620	Group 1 sulphate transporters	n/e	-	-
SULTR1,2	At1g78000		1.922	0.374	0.450
SULTR1,3	At1g22150		n/e	-	-
<b>SULTR2,1</b>	<b>At5g10180</b>	Group 2 sulphate transporters	<b>3.317</b>	<b>0.00013</b>	<b>0.0123</b>
SULTR2,2	At1g77990		1.049	0.981	0.680
SULTR3,1	At3g51900	Group 3 sulphate transporters	n/r	-	-
SULTR3,2	At4g02700		n/e	-	-
SULTR3,3	At1g23090		0.612	0.0100	0.113
SULTR3,4	At3g15990		3.258	0.116	0.312
SULTR3,5	At5g19600		0.354	0.374	0.450
SULTR4,1	At5g13550	Group 4 sulphate transporters	1.370	0.0244	0.168
SULTR4,2	At3g12520		1.219	0.273	0.446
SULTR5,1	At1g80310	Group 5 sulphate transporters	1.140	0.134	0.323
SULTR5,2	At2g25680		1.806	0.374	0.450
<b>ATPS1</b>	<b>At3g22890</b>	ATP sulphurylases	<b>1.639</b>	<b>0.00207</b>	<b>0.0552</b>
ATPS2	At1g19920		0.950	0.554	0.526
<b>ATPS3</b>	<b>At4g14680</b>		<b>1.969</b>	<b>0.000687</b>	<b>0.0312</b>
ATPS4	At5g43780		0.673	0.0156	0.137
APR1	At4g04610	APS reductases	1.846	0.00541	0.0825
APR2	At1g62180		0.793	0.0772	0.270
APR3	At4g21990		0.652	0.0240	0.166
SiR	At5g04590	Sulphite reductase	1.379	0.00219	0.0582
SERAT1,1	At5g56760	Serine acetyltransferases	1.165	0.120	0.312
SERAT2,1	At1g55920		1.310	0.00251	0.0601
SERAT2,2	At3g13110		1.134	0.0277	0.178
SERAT3,1	At2g17640		0.894	0.0950	0.296
SERAT3,2	At4g35640		n/r	-	-
<b>OASA1</b>	<b>At4g14880</b>	O-acetylserine (thiol)lyases	<b>1.546</b>	<b>0.000177</b>	<b>0.0151</b>
OASTL	At1g55880		n/e	-	-
OASB	At2g43750		0.835	0.0235	0.164
OASTL	At3g22460		0.646	0.0385	0.205

CS26	At3g03630	Cysteine synthases	1.008	0.981	0.686
CYSD1	At3g04940		0.913	0.438	0.478
CYSC1	At3g61440		1.082	0.0568	0.241
CYSD2	At5g28030		0.990	0.973	0.679
GSH1	At4g23100	Glutathione synthesis	1.398	0.00449	0.0768
GSH2	At5g27380		1.411	0.00233	0.0585
SLIM1/EIL3	At1g73730	Sulphate Limitation transcription factor	0.880	0.320	0.450
SOT1	At5g43690	Sulphotransferases	n/e	-	-
SOT2a	At3g51210		n/e	-	-
SOT2b	At1g62650		n/r	-	-
SOT3	At4g26280		n/e	-	-
SOT4	At2g27570		n/e	-	-
SOT5	At3g45070		n/r	-	-
SOT6	At3g45080		n/r	-	-
SOT7	At1g28170		n/e	-	-
SOT8	At1g13420		n/e	-	-
SOT9	At1g13430		n/r	-	-
SOT10	At2g14920		n/r	-	-
SOT11	At2g03750		0.979	0.725	0.602
SOT12	At2g03760		1.495	0.363	0.450
SOT13	At2g03770		n/e	-	-
<b>SOT14</b>	<b>At5g07000</b>		<b>0.580</b>	<b>0.000415</b>	<b>0.0243</b>
SOT15	At5g07010		0.495	0.0193	0.149
<b>SOT16</b>	<b>At1g74100</b>		<b>2.097</b>	<b>0.00489</b>	<b>0.0805</b>
<b>SOT17</b>	<b>At1g18590</b>		<b>4.186</b>	<b>0.000146</b>	<b>0.0130</b>
<b>SOT18</b>	<b>At1g74090</b>		<b>2.291</b>	<b>0.000278</b>	<b>0.0239</b>
SOT19	At3g50620		0.970	0.753	0.614
SOT20	At2g15730		n/r	-	-
SOT21	At4g34420	n/r	-	-	
AtPSK1	At1g13590	PSK precursors	n/e	-	-
<b>AtPSK2</b>	<b>At2g22860</b>		<b>4.678</b>	<b>0.00164</b>	<b>0.0487</b>
AtPSK3	At3g44735		0.994	0.923	0.670
<b>AtPSK4</b>	<b>At3g49780</b>		<b>2.051</b>	<b>0.0156</b>	<b>0.137</b>
AtPSK5	At5g65870	1.203	0.219	0.408	
AtPSY1	At5g58650	PSY precursors	0.896	0.282	0.450
AtPSY2	At3g47295		0.913	0.617	0.553
AtPSY3	At2g29995		1.671	0.021	0.153

**Supplemental Table 3.** Metabolic pathways over-represented among the altered transcripts - pathways more highly expressed in the *apk1 apk2* mutant at  $p < 0.005$ . The pathways were identified by iterative group analysis using Aracyc metabolic pathways.

<b>Group name</b>	<b>Members</b>	<b>Members changed</b>	<b>p-value</b>
Glucosinolate biosynthesis from tryptophan	3	3	5.08E-05
Glucosinolate biosynthesis from phenylalanine	4	3	0.000198
IAA biosynthesis	8	2	0.000508
Leucine biosynthesis	18	4	0.000984
Glucosinolate biosynthesis from homomethionine	3	2	0.00129
NAD biosynthesis (from aspartate)	6	6	0.00226
Cytokinin-glucoside biosynthesis	77	7	0.00237
Jasmonic acid biosynthesis	14	4	0.00382
Lipoxygenase pathway	11	6	0.00403

**Supplemental Table 4.** Metabolic pathways over-represented among the altered transcripts - pathways more highly expressed in the WT at  $p < 0.005$ . The pathways were identified by iterative group analysis using Aracyc metabolic pathways.

<b>Group name</b>	<b>Members</b>	<b>Members changed</b>	<b>p-value</b>
Cellulose biosynthesis	41	9	0.000926
Cytokinins degradation	6	3	0.000937
Cytokinin-glucoside biosynthesis	77	5	0.00115
Triacylglycerol biosynthesis	5	1	0.00331
Glycine biosynthesis	8	8	0.00341
Glycine degradation	8	8	0.00370

**Supplemental Table 5.** Primer sequences for identification of T-DNA insertions in *apk* mutants

<b>Mutant</b>	<b>Mutant ID</b>	<b>5' Primer</b>	<b>3' Primer</b>
<i>apk1-1</i>	SALK_053427	CCACAACCTGCATATAGACTATGC	GACCCAAATCACACATCC
<i>apk1-2</i>	SALK_034586	CCACAACCTGCATATAGACTATGC	GACCCAAATCACACATCC
<i>apk2-1</i>	SALK_093072	TAACGTCTCTGCTCAAGC	ATGTTTTTCGGTGAGGTGC
<i>apk2-2</i>	SALK_077590	TAACGTCTCTGCTCAAGC	ATGTTTTTCGGTGAGGTGC
<i>apk3-1</i>	SALK_115182	GCGCATATGTCGACAGTGGGAAATTC	CTACTACTACATATGAGACACC
<i>apk3-2</i>	SALK_115507	CCGTGATTATCTCTGTCAGC	TTCCACTCTATCCTCTGC
<i>apk4-1</i>	SALK_035815	ACTTCCGTCCGATCATGG	GCTCGATCATCTGCTTCG
<i>apk4-2</i>	SALK_068072	ACTTCCGTCCGATCATGG	GCTCGATCATCTGCTTCG
LBb1	all	GCGTGGACCGCTTGCTGCAACTC	

**Supplemental Table 6.** Primer sequences for RT-PCR analysis of APK transcripts in T-DNA lines

<b>Gene</b>	<b>5' Primer</b>	<b>3' Primer</b>
<i>APK1</i>	GCTTTCGATGGCTTCTTC	GACCCAAATCACACATCC
<i>APK2</i>	TAACGTCTCTGCTCAAGC	ATGTTTTTCGGTGAGGTGC
<i>APK3</i>	AACTGAAAGGCAGAAGTTG	TTCCACTCTATCCTCTGC
<i>APK4</i>	ACTTCCGTCCGATCATGG	GCTCGATCATCTGCTTCG

**Supplemental Table 7.** Primer sequences for expression analysis by qPCR

<b>gene</b>	<b>5' primer</b>	<b>3' primer</b>
actin2	ATGGAAGCTGCTGGAATCCAC	TTGCTCATACGGTCAGCGATG
PSK2	ATGGTAGACGAAAAGTGGTGC	TCAGTATGAGCGACCAAAGTCCT
PSK3	CCTGGCAGTTCTTCTCCTCATTTT	TCTCTTGATCTTCTTTCTCTTCG
PSK4	AGAAGAAAGCTGCAACGGAATT	AATCGGTGTGAAGAACAAGGCT
APK1	CCTTACGAGCCACCATTGAACTG	GCCATTTTCGATAGGAGAAGTTCCT
APK2	CAAAATCAAAGGCTTCACTGGAATC	TGTTTCAGCACTACCTCGCAATT
APK3	GTTGAAAGAGAAAGAGGGAGAGTGTC	AGAGATCACTTCTCAGCCATAGC
APK4	TCTTCTTCTCTGTGTGAAATGGCAG	TTCTTCAGGTATCCATTTTGGTCC
TSB	CGTTGACACTCTTCTTCTCTAACC	AGCCTCAGGCACCTCTGCTACTT
MAM-L	TTCGTTACTTCTCACATCGTCGAG	GGAGCAACATGAGACGAACAGAGT
CYP83B1	GGCAACAAACCATGTCGTATCAAG	CGTTGACACTCTTCTTCTCTAACC
SUR2	CAAGACGAGCCGTTGCTGAT	GGACGTGGGAGCAAGATGTTT
SOT16	CGAAGTCGTCGAACTCACAGAGTT	AAAGACCTTCGAGGAGACATTCTTG
SOT17	GGAATCCAAAACCATAAACGACG	CGGATCTTTTGGTCTCCAGCC
SOT18	CCCTACCGAGTCACGACGAGA	GGTAGCCACCAGTAACCACCATACT
MYB51	CTACAAGTGTTTCCGTTGACTCTGAA	ACGAAATTATCGCAGTACATTAGAGGA
MYB122	GTCCGTTGAGTCTTGTTTGGAGAATA	TGTCATCCCTTCACAGGAGTCAT
MYB34	CACGACTGTCGATAATTTTGGGTTT	CATATTGTCATCTTCGTTCCAGGAA
MYB28	TCCCTGACAAATACTCTTGCTGAAT	CATTGTGGTTATCTCCTCCGAATT
MYB76	ACGTTTTGGACGATCGAGCTCTAC	TGATTGAGAGAACGAGTCTGGGAGT
MYB29	GAAATATGATGCTTCCTTGAGCTCC	ACGGTGTAGAGCTGATCAAGGTTT
ATPS1	CCCGCCAGACGGTTTTATGT	ACTTCTGGTAGTCTACCATTACCCGC
ATPS2	CAGATCCTCTTACGTTTCTCACATCA	GGTTGGTGAAAGAGGATGGTTTTG
ATPS3	GAGAGAACCCACCAGATGGATTTA	TCCGGTTAGTGTCAAAGTGTCTGAG
ATPS4	TTCTTCAGCAGCAGCCATCG	ACGAGAAGCATGATGGTTATGGA

**Supplemental Table 8.** Primer sequences for the pET14b constructs for heterologous expression of recombinant APKs

Gene	Cloning site	5'- Primer <sup>a</sup>	3'- Primer
<i>APK1</i>	NdeI / XhoI	GCGCATATGGTTTCTATGGATGGATCTC	CGCTCGAGTTATGCTTGAAGATAACC
<i>APK2</i>	NdeI / BamHI	GCGCATATGGCCGTTAACGTCTCTGCTC	GCGGATCCTTAGCCCTCAAGATAACC
<i>APK3</i>	NdeI / BamHI	GCGCATATGTCGACAGTGGGAAATTC	GCGGATCCTTACTCGTTTTGAAGGAAACC
<i>APK4</i>	XhoI / BamHI	GCGCTCGAGTCCGGTGAAACGAAGCAAATC	GCGGATCCTTACATACAATTACGTGATTTTG

<sup>a</sup> Underlined sequences correspond to the indicated restriction sites used for cloning into the pET14b vector

**Supplemental Table 9.** Primer sequences for the chimeric gene constructs of CaMV35S promoter-APK cDNA-GFP

Construct	5' Primer <sup>a</sup>	3' Primer
<i>APK1</i> (N-term)	ACGCGT <u>CGAC</u> ATGATCGCCGCCGGAGC	CATGCCATGGCCTGTCTATCAACTTCTCAAC
<i>APK1</i> (full)	ACGCGT <u>CGAC</u> ATGATCGCCGCCGGAGC	CATGCCATGGCTGCTTGAAGATAACCCTTG
<i>APK2</i> (N-term)	ACGCGT <u>CGAC</u> ATGGAAGGATTAGCTATCAG	CATGCCATGGCTTGTCTGTCGCATCTGC
<i>APK2</i> (full)	ACGCGT <u>CGAC</u> ATGGAAGGATTAGCTATCAG	CATGCCATGGCGCCCTCAAGATAACCTTTG
<i>APK3</i> (full)	CCGCTCGAGATGTCGACAGTGGGAAA	CATGCCATGGCCTCGTTTTGAAGGAAAC
<i>APK4</i> (N-term)	ACGCGT <u>CGAC</u> ATGGATGTTGCCGCGATG	CATGCCATGGCTTGCCTGTCGGATTTAG
<i>APK4</i> (full)	ACGCGT <u>CGAC</u> ATGGATGTTGCCGCGATG	CATGCCATGGCCATACAATTACGTGATTTTG

<sup>a</sup> Underlined sequences correspond to the *Sa*I and *Nco*I restriction sites used for fusion construction.

**Supplemental Table 10.** Primer sequences for creation of transgenic plants expressing APK promoter-coding sequence-GFP

<b>Promoter/ gene</b>	<b>5' Primer<sup>a</sup></b>	<b>3' Primer</b>
APK1	<u>ACTAGTTTC</u> AGTAGCAATCTAAGTATGG	GAATTCTGCTTGAAGATAACCCTTGTTATC
APK2	<u>CTCGAGTTT</u> GGAGGAGGAGACTAACAAC	<u>TCTAGAGCCCT</u> CAAGATAACCTTTGTTTTG
APK3	<u>ACTAGTAAAT</u> CTGGTTTACTTCATGC	<u>GGATCCCTC</u> GTTTTGAAGGAAACCTTTG
APK4	<u>GTCGACCCTT</u> ACAGGTTTTGACAACAGTC	<u>GTCGACCATA</u> CAATTACGTGATTTTGTGG
GFP	<u>GAATTCATGGT</u> GAGCAAGGGCGAGGAGCTG	<u>GAGCTCTTACTT</u> GTACAGCTCGTC

<sup>a</sup> The underlined sequences correspond to the restriction sites used for fusion construction.