

# **Finding Optimum Solutions for Multiple Target Control in Disease**

## **Related Molecular Network**

Kun Yang<sup>1,2</sup>, Hongjun Bai<sup>1,2</sup>, Qi Ouyang<sup>2</sup>, Luhua Lai<sup>1,2\*</sup>, Chao Tang<sup>2,3</sup>

<sup>1</sup>Beijing National Laboratory for Molecular Sciences, State Key Laboratory for Structural Chemistry of Unstable and Stable Species, College of Chemistry and Molecular Engineering, Peking University, Beijing 100871, China.

<sup>2</sup>Center for Theoretical Biology, Peking University, Beijing 100871, China

<sup>3</sup>Department of Biopharmaceutical Sciences, University of California, San Francisco, CA 94158-2330, USA

\*Corresponding author:

Luhua Lai

College of Chemistry and Molecular Engineering and Center for Theoretical Biology

Peking University

Beijing 100871, China

E-mail: [lh lai@pku.edu.cn](mailto:lh lai@pku.edu.cn)

Tel: 86-10-62757486; Fax: 86-10-62751725

### **Table of Content**

- Supplementary protocol
- Supplementary figures
- Supplementary tables

## Supplementary Protocol

### The construction of arachidonic acid metabolic network

The arachidonic acid metabolic network (AAnetwork) with a multi-cellular ensemble of human polymorphonuclear leukocyte (PMN), endothelial cells (EC) and platelet (PLT) was constructed based on our previous work and published experiments (Claesson & Haeggstrom, 1988; Denzlinger, 1996; Husain & Abdel-Latif, 2001; Inoue *et al*, 2000; Kojima *et al*, 2005; Ohd *et al*, 2000; Tsubouchi *et al*, 2001; Weaver *et al*, 2001; Yang *et al*, 2007). A group of ordinary differential equations (ODE) were written for PMN, EC and PLT, respectively, to simulate the time-course metabolism of arachidonic acid (AA). The productions of metabolites in the network were calculated as the sum of the corresponding output in three types of cells. For example:

$$Output_{PGE2} = C_{PGE2,PMN} \cdot V_{PMN} \cdot n_{PMN} + C_{PGE2,EC} \cdot V_{EC} \cdot n_{EC} + C_{PGE2,PLT} \cdot V_{PLT} \cdot n_{PLT} \quad \text{--- (1)}$$

where  $C_{PGE2,PMN}$  is the concentration of PGE2 in PMN,  $V_{PMN}$  is the volume of PMN and  $n_{PMN}$  is the number of PMN in the model. Parameters and initial concentrations in the model are list in Table SI. Results of MTOI for different parameter sets are summarized in Table SII.

The ODEs for PMN are:

$$\frac{d[AA]}{dt} = \frac{K_{cat,PLA2} \left(1 + \frac{[12-HPETE]}{K_{12-HPETE \rightarrow PLA2}} + \frac{[15-HPETE]}{K_{15-HPETE \rightarrow PLA2}} + \frac{[LTB4]}{K_{LTB4 \rightarrow PLA2}} + \frac{[5-HETE]}{K_{5-HETE \rightarrow PLA2}}\right) [PLA2][PL]}{K_{m,PLA2} \left(1 + \frac{[AA]}{K_i}\right) + [PL]} - \frac{K_{cat,15-LOX} [15-LOX][AA]}{K_{m,15-LOX} \left(1 + \frac{[15-HPETE]}{K_i}\right) + [AA]} - \frac{K_{cat,12-LOX} [12-LOX][AA]}{K_{m,12-LOX} \left(1 + \frac{[12-HPETE]}{K_{i_{12-HPETE \rightarrow 12-LOX}}} + \frac{[15-HETE]}{K_{i_{15-HETE \rightarrow 12-LOX}}}\right) + [AA]} - \frac{K_{cat,5-LOX} [5-LOX][AA]}{K_{m,5-LOX} \left(1 + \frac{[5-HPETE]}{K_i} + \frac{[12-HETE]}{K_{i_{12-HETE \rightarrow 5-LOX}}} + \frac{[15-HETE]}{K_{i_{15-HETE \rightarrow 5-LOX}}} + \frac{[PGE2]}{K_{i_{PGE2 \rightarrow 5-LOX}}} + \frac{[5-HETE]}{K_{i_{5-HETE \rightarrow 5-LOX}}}\right) + [AA]}$$

$$- \frac{K_{cat,COX2}[COX-2][AA]}{K_{m,COX2}(1 + \frac{[PGH2]}{Ki} + \frac{[PGE2]}{Ki_{PGE2 \rightarrow COX2}})} + Kd_{exoAA}[exoAA] - Kd_{AA}[AA]$$

$$\frac{d[15-HPETE]}{dt} = \frac{K_{cat,15-LOX}[15-LOX][AA]}{K_{m,15-LOX}(1 + \frac{[15-HPETE]}{Ki}) + [AA]} - \frac{K_{cat,PHGPx}[PHGPx][15-HPETE]}{K_{m,PHGPx}(1 + \frac{[15-HETE]}{Ki}) + [15-HPETE]} - Kd_{15-HPETE}[15-HPETE]$$

$$\frac{d[15-HETE]}{dt} = \frac{K_{cat,PHGPx}[PHGPx][15-HPETE]}{K_{m,PHGPx}(1 + \frac{[15-HETE]}{Ki}) + [15-HPETE]} - Kd_{15-HETE}[15-HETE]$$

$$\frac{d[12-HPETE]}{dt} = \frac{K_{cat,12-LOX}[12-LOX][AA]}{K_{m,12-LOX}(1 + \frac{[12-HPETE]}{Ki_{12-HPETE \rightarrow 12-LOX}} + \frac{[15-HETE]}{Ki_{15-HETE \rightarrow 12-LOX}})} + [AA] - \frac{K_{cat,PHGPx}[PHGPx][12-HPETE]}{K_{m,PHGPx}(1 + \frac{[12-HETE]}{Ki}) + [12-HPETE]}$$

$$\frac{d[12-HETE]}{dt} = \frac{K_{cat,PHGPx}[PHGPx][12-HPETE]}{K_{m,PHGPx}(1 + \frac{[12-HETE]}{Ki}) + [12-HPETE]}$$

$$\frac{d[PGH2]}{dt} = \frac{K_{cat,COX2}[COX-2][AA]}{K_{m,COX2}(1 + \frac{[PGH2]}{Ki} + \frac{[PGE2]}{Ki_{PGE2 \rightarrow COX2}})} + [AA] - \frac{K_{cat,TXAS}[TXAS][PGH2]}{K_{m,TXAS}(1 + \frac{[TXA2]}{Ki}) + [PGH2]} - \frac{K_{cat,PGES}[PGES][PGH2]}{K_{m,PGES}(1 + \frac{[PGE2]}{Ki} + \frac{[AA]}{Ki_{AA \rightarrow PGES}} + \frac{[15-HETE]}{Ki_{15-HETE \rightarrow PGES}})} + [PGH2]$$

$$\frac{d[PGE2]}{dt} = \frac{K_{cat,PGES}[PGES][PGH2]}{K_{m,PGES}(1 + \frac{[PGE2]}{Ki} + \frac{[AA]}{Ki_{AA \rightarrow PGES}} + \frac{[15-HETE]}{Ki_{15-HETE \rightarrow PGES}})} + [PGH2]$$

$$\frac{d[TXA2]}{dt} = \frac{K_{cat,TXAS}[TXAS][PGH2]}{K_{m,TXAS}(1 + \frac{[TXA2]}{Ki}) + [PGH2]} - Kd_{TXA2}[TXA2]$$

$$\frac{d[TXB2]}{dt} = Kd_{TXA2}[TXA2] - Kd_{TXB2}[TXB2]$$

$$\frac{d[5-HPETE]}{dt} = \frac{K_{cat,5-LOX}[5-LOX][AA]}{K_{m,5-LOX}(1 + \frac{[5-HPETE]}{Ki} + \frac{[12-HETE]}{Ki_{12-HETE \rightarrow 5-LOX}} + \frac{[15-HETE]}{Ki_{15-HETE \rightarrow 5-LOX}} + \frac{[PGE2]}{Ki_{PGE2 \rightarrow 5-LOX}} + \frac{[5-HETE]}{Ki_{5-HETE \rightarrow 5-LOX}})} + [AA]$$

$$-\frac{K_{cat,5-LOX}[5-LOX][5-HPETE]}{K_{m,5-LOX}\left(1+\frac{[LTA4]}{Ki}+\frac{[12-HETE]}{Ki_{12-HETE\rightarrow5-LOX}}+\frac{[15-HETE]}{Ki_{15-HETE\rightarrow5-LOX}}+\frac{[PGE2]}{Ki_{PGE2\rightarrow5-LOX}}+\frac{[5-HETE]}{Ki_{5-HETE\rightarrow5-LOX}}\right)+[5-HPETE]}$$

$$-\frac{K_{cat,PHGPx}[PHGPx][5-HPETE]}{K_{m,PHGPx}\left(1+\frac{[5-HETE]}{Ki}\right)+[5-HPETE]}$$

$$\frac{d[5-HETE]}{dt} = \frac{K_{cat,PHGPx}[PHGPx][5-HPETE]}{K_{m,PHGPx}\left(1+\frac{[5-HETE]}{Ki}\right)+[5-HPETE]} - Kd_{5-HETE}[5-HETE]$$

$$\frac{d[LTA4]}{dt} = \frac{K_{cat,5-LOX}[5-LOX][5-HPETE]}{K_{m,5-LOX}\left(1+\frac{[LTA4]}{Ki}+\frac{[12-HETE]}{Ki_{12-HETE\rightarrow5-LOX}}+\frac{[15-HETE]}{Ki_{15-HETE\rightarrow5-LOX}}+\frac{[PGE2]}{Ki_{PGE2\rightarrow5-LOX}}+\frac{[5-HETE]}{Ki_{5-HETE\rightarrow5-LOX}}\right)+[5-HPETE]}$$

$$-\frac{K_{cat,LTA4H}[LTA4H][LTA4]}{K_{m,LTA4H}\left(1+\frac{[LTA4]}{Ki}\right)+[LTA4]} - Kd_{LTA4}[LTA4]$$

$$\frac{d[LTB4]}{dt} = \frac{K_{cat,LTA4H}[LTA4H][LTA4]}{K_{m,LTA4H}\left(1+\frac{[LTA4]}{Ki}\right)+[LTA4]} - Kd_{LTB2}[LTB2]$$

$$-\frac{K_{cat,CYP4F3}[CYP4F3][LTB4]}{K_{m,CYP4F3}\left(1+\frac{[20-OH-LTB4]}{Ki}+\frac{[12-HETE]}{Ki_{12-HETE\rightarrow CYP4F3}}+\frac{[5-HETE]}{Ki_{5-HETE\rightarrow CYP4F3}}\right)+[LTB4]}$$

$$- Kd_{LTB4}[LTB4]$$

$$\frac{d[20-OH-LTB4]}{dt} = \frac{K_{cat,CYP4F3}[CYP4F3][LTB4]}{K_{m,CYP4F3}\left(1+\frac{[20-OH-LTB4]}{Ki}+\frac{[12-HETE]}{Ki_{12-HETE\rightarrow CYP4F3}}+\frac{[5-HETE]}{Ki_{5-HETE\rightarrow CYP4F3}}\right)+[LTB4]}$$

$$\frac{d[PLA2]}{dt} = 0$$

$$\frac{d[15-LOX]}{dt} = \frac{k_{PGE2\rightarrow15-LOX}[PGE2]^2}{[PGE2]^2 + K_{PGE2\rightarrow15-LOX}} - Kd_{15-LOX}[15-LOX]$$

$$\frac{d[12-LOX]}{dt} = -Ki_{15-HPETE\rightarrow12-LOX}[15-HPETE][12-LOX]$$

$$\frac{d[COX-2]}{dt} = 0$$

$$\frac{d[PGES]}{dt} = 0$$

$$\frac{d[TXAS]}{dt} = -(K_{i_{15-HPETE \rightarrow TXAS}}[15-HPETE] + K_{i_{PGH2 \rightarrow TXAS}}[PGH2])[TXAS]$$

$$\begin{aligned} \frac{d[5-LOX]}{dt} &= (K_{LTB4 \rightarrow 5-LOX}[LTB4] - K_{LTA4 \rightarrow 5-LOX}[LTA4] - K_{i_{5-HPETE \rightarrow 5-LOX}}[5-HPETE] \\ &\quad - K_{i_{15-HPETE \rightarrow 5-LOX}}[15-HPETE])[5-LOX] \end{aligned}$$

$$\frac{d[LTA4H]}{dt} = -\frac{K_{cat,LTA4H}[LTA4H][LTA4]}{129(K_{m,LTA4H} + [LTA4])}$$

$$\frac{d[CYP4F3]}{dt} = 0$$

$$\frac{d[PHGPx]}{dt} = 0$$

The ODEs for EC are:

$$\begin{aligned} \frac{d[AA]}{dt} &= \frac{K_{cat,PLA2}(1 + \frac{[12-HPETE]}{K_{12-HPETE \rightarrow PLA2}} + \frac{[15-HPETE]}{K_{15-HPETE \rightarrow PLA2}} + \frac{[PGF2\alpha]}{K_{PGF2\alpha \rightarrow PLA2}})[PLA2][PL]}{K_{m,PLA2}(1 + \frac{[AA]}{K_i}) + [PL]} \\ &\quad - \frac{K_{cat,15-LOX}[15-LOX][AA]}{K_{m,15-LOX}(1 + \frac{[15-HPETE]}{K_i}) + [AA]} - \frac{K_{cat,12-LOX}[12-LOX][AA]}{K_{m,12-LOX}(1 + \frac{[12-HPETE]}{K_{i_{12-HPETE \rightarrow 12-LOX}}}) + [AA]} \\ &\quad - \frac{K_{cat,COX2}[COX-2][AA]}{K_{m,COX2}(1 + \frac{[PGH2]}{K_i} + \frac{[PGE2]}{K_{i_{PGE2 \rightarrow COX2}}}) + [AA]} \end{aligned}$$

$$\begin{aligned} \frac{d[15-HPETE]}{dt} &= \frac{K_{cat,15-LOX}[15-LOX][AA]}{K_{m,15-LOX}(1 + \frac{[15-HPETE]}{K_i}) + [AA]} - \frac{K_{cat,PHGPx}[PHGPx][15-HPETE]}{K_{m,PHGPx}(1 + \frac{[15-HETE]}{K_i}) + [15-HPETE]} \\ &\quad - Kd_{15-HPETE}[15-HPETE] \end{aligned}$$

$$\frac{d[15-HETE]}{dt} = \frac{K_{cat,PHGPx}[PHGPx][15-HPETE]}{K_{m,PHGPx}(1 + \frac{[15-HETE]}{K_i}) + [15-HPETE]} - Kd_{15-HETE}[15-HETE]$$

$$\frac{d[12-HPETE]}{dt} = \frac{K_{cat,12-LOX}[12-LOX][AA]}{K_{m,12-LOX}(1 + \frac{[12-HPETE]}{K_i}) + [AA]} - \frac{K_{cat,PHGPx}[PHGPx][12-HPETE]}{K_{m,PHGPx}(1 + \frac{[12-HETE]}{K_i}) + [12-HPETE]}$$

$$\frac{d[12 - HETE]}{dt} = \frac{K_{cat,PHGPx}[PHGPx][12 - HPETE]}{K_{m,PHGPx}(1 + \frac{[12 - HETE]}{K_i}) + [12 - HPETE]} - Kd_{12-HETE}[12 - HETE]$$

$$\begin{aligned} \frac{d[PGH2]}{dt} = & \frac{K_{cat,COX2}[COX - 2][AA]}{K_{m,COX2}(1 + \frac{[PGH2]}{K_i} + \frac{[PGE2]}{K_{i_{PGE2 \rightarrow COX2}}}) + [AA]} - \frac{K_{cat,TXAS}[TXAS][PGH2]}{K_{m,TXAS}(1 + \frac{[TXA2]}{K_i}) + [PGH2]} \\ & - \frac{K_{cat,PGIS}[PGIS][PGH2]}{K_{m,PGIS}(1 + \frac{[PGI2]}{K_i} + \frac{[15 - HPETE]}{K_{i_{15-HPETE \rightarrow PGIS}}}) + [PGH2]} - \frac{K_{cat,PGDS}[PGDS][PGH2]}{K_{m,PGDS}(1 + \frac{[PGD2]}{K_i}) + [PGH2]} \\ & - \frac{K_{cat,PGES}[PGES][PGH2]}{K_{m,PGES}(1 + \frac{[PGE2]}{K_i} + \frac{[AA]}{K_{i_{AA \rightarrow PGES}}} + \frac{[15 - HETE]}{K_{i_{15-HETE \rightarrow PGES}}}) + [PGH2]} - Kd_{PGH2}[PGH2] \end{aligned}$$

$$\frac{d[TXA2]}{dt} = \frac{K_{cat,TXAS}[TXAS][PGH2]}{K_{m,TXAS}(1 + \frac{[TXA2]}{K_i}) + [PGH2]} - Kd_{TXA2}[TXA2]$$

$$\frac{d[TXB2]}{dt} = Kd_{TXA2}[TXA2] - Kd_{TXB2}[TXB2]$$

$$\frac{d[PGI2]}{dt} = \frac{K_{cat,PGIS}[PGIS][PGH2]}{K_{m,PGIS}(1 + \frac{[PGI2]}{K_i} + \frac{[15 - HPETE]}{K_{i_{15-HPETE \rightarrow PGIS}}}) + [PGH2]} - Kd_{PGI2}[PGI2]$$

$$\frac{d[6 - keto - PGF1\alpha]}{dt} = Kd_{PGI2}[PGI2] - Kd_{6-keto-PGF1\alpha}[6 - keto - PGF1\alpha]$$

$$\begin{aligned} \frac{d[PGD2]}{dt} = & \frac{K_{cat,PGDS}[PGDS][PGH2]}{K_{m,PGDS}(1 + \frac{[PGD2]}{K_i}) + [PGH2]} - Kd_{PGD2}[PGD2] \\ & - \frac{K_{cat,PGFS}[PGFS][PGD2]}{K_{m,PGFS}(1 + \frac{[11 - epi - PGF2\alpha]}{K_i}) + [PGD2]} \end{aligned}$$

$$\frac{d[PGJ2]}{dt} = Kd_{PGD2}[PGD2] - Kd_{PGJ2}[PGJ2]$$

$$\frac{d[15d - PGJ2]}{dt} = Kd_{PGJ2}[PGJ2]$$

$$\frac{d[PGE2]}{dt} = \frac{K_{cat,PGES}[PGES][PGH2]}{K_{m,PGES}(1 + \frac{[PGE2]}{K_i} + \frac{[AA]}{K_{i_{AA \rightarrow PGES}}} + \frac{[15 - HETE]}{K_{i_{15-HETE \rightarrow PGES}}}) + [PGH2]}$$

$$-\frac{K_{cat,CR}[CR][PGE2]}{K_{m,CR}(1+\frac{[PGF2\alpha]}{Ki})+[PGE2]} - \frac{K_{cat,9-KPR}[9-KPR][PGE2]}{K_{m,9-KPR}(1+\frac{[PGF2\alpha]}{Ki})+[PGE2]}$$

$$\begin{aligned} \frac{d[PGF2\alpha]}{dt} = & \frac{K_{cat,CR}[CR][PGE2]}{K_{m,CR}(1+\frac{[PGF2\alpha]}{Ki})+[PGE2]} + \frac{K_{cat,9-KPR}[9-KPR][PGE2]}{K_{m,9-KPR}(1+\frac{[PGF2\alpha]}{Ki})+[PGE2]} + Kd_{PGH2}[PGH2] \\ & - \frac{K_{cat,15-PGDH}[15-PGDH][PGF2\alpha]}{K_{m,15-PGDH}(1+\frac{[15-keto-PGF2\alpha]}{Ki})+[PGF2\alpha]} \end{aligned}$$

$$\frac{d[15-keto-PGF2\alpha]}{dt} = \frac{K_{cat,15-PGDH}[15-PGDH][PGF2\alpha]}{K_{m,15-PGDH}(1+\frac{[15-keto-PGF2\alpha]}{Ki})+[PGF2\alpha]}$$

$$\frac{d[PLA2]}{dt} = -Ki_{15d-PGJ2 \rightarrow PLA2}[15d-PGJ2][PLA2]$$

$$\frac{d[15-LOX]}{dt} = \frac{k_{PGE2 \rightarrow 15-LOX}[PGE2]^2}{[PGE2]^2 + K_{PGE2 \rightarrow 15-LOX}} - Kd_{15-LOX}[15-LOX]$$

$$\frac{d[12-LOX]}{dt} = 0$$

$$\frac{d[PHGPs]}{dt} = 0$$

$$\frac{d[COX-2]}{dt} = 0$$

$$\frac{d[TXAS]}{dt} = -(Ki_{15-HPETE \rightarrow TXAS}[15-HPETE] + Ki_{PGH2 \rightarrow TXAS}[PGH2])[TXAS]$$

$$\frac{d[PGIS]}{dt} = 0$$

$$\frac{d[PGDS]}{dt} = 0$$

$$\frac{d[PGFS]}{dt} = 0$$

$$\frac{d[PGES]}{dt} = -Ki_{15d-PGJ2 \rightarrow PGES}[15d-PGJ2][PGES]$$

$$\frac{d[CR]}{dt} = 0$$

$$\frac{d[9 - KPR]}{dt} = 0$$

$$\frac{d[15 - PGDH]}{dt} = 0$$

The ODEs for PLT are:

$$\begin{aligned} \frac{d[AA]}{dt} = & \frac{K_{cat,PLA2} \left(1 + \frac{[12 - HPETE]}{K_{12-HPETE \rightarrow PLA2}}\right) [PLA2][PL]}{K_{m,PLA2} \left(1 + \frac{[AA]}{K_i}\right) + [PL]} - \frac{K_{cat,12-LOX} [12 - LOX][AA]}{K_{m,12-LOX} \left(1 + \frac{[12 - HPETE]}{K_i}\right) + [AA]} \\ & - \frac{K_{cat,COX1} [COX - 1][AA]}{K_{m,COX1} \left(1 + \frac{[PGH2]}{K_i}\right) + [AA]} \end{aligned}$$

$$\frac{d[12 - HPETE]}{dt} = \frac{K_{cat,12-LOX} [12 - LOX][AA]}{K_{m,12-LOX} \left(1 + \frac{[12 - HPETE]}{K_i}\right) + [AA]} - \frac{K_{cat,PHGPx} [PHGPx][12 - HPETE]}{K_{m,PHGPx} \left(1 + \frac{[12 - HETE]}{K_i}\right) + [12 - HPETE]}$$

$$\frac{d[12 - HETE]}{dt} = \frac{K_{cat,PHGPx} [PHGPx][12 - HPETE]}{K_{m,PHGPx} \left(1 + \frac{[12 - HETE]}{K_i}\right) + [12 - HPETE]}$$

$$\frac{d[PGH2]}{dt} = \frac{K_{cat,COX1} [COX - 1][AA]}{K_{m,COX1} \left(1 + \frac{[PGH2]}{K_i}\right) + [AA]} - \frac{K_{cat,TXAS} [TXAS][PGH2]}{K_{m,TXAS} \left(1 + \frac{[TXA2]}{K_i}\right) + [PGH2]}$$

$$\frac{d[TXA2]}{dt} = \frac{K_{cat,TXAS} [TXAS][PGH2]}{K_{m,TXAS} \left(1 + \frac{[TXA2]}{K_i}\right) + [PGH2]} - Kd_{TXA2} [TXA2]$$

$$\frac{d[TXB2]}{dt} = Kd_{TXA2} [TXA2] - Kd_{TXB2} [TXB2]$$

$$\frac{d[PLA2]}{dt} = 0$$

$$\frac{d[12 - LOX]}{dt} = 0$$



$$\frac{d[COX - 1]}{dt} = 0$$

$$\frac{d[TXAS]}{dt} = -K_{i_{PGH2 \rightarrow TXAS}}[PGH2][TXAS]$$

$$\frac{d[PHGPx]}{dt} = 0$$

### The construction of simplified AA network model

The simplified AA network model was set up to verify the practicability of MTOI in incomplete network. Only disease-related metabolites and enzymes were included. Like the full model, a group of ODEs were written for PMN, EC and PLT, respectively, to simulate the time-course metabolism of AA. The productions of metabolites in the network were calculated as equation 1. Unknown parameters and initial concentrations were evaluated by parameter fitting (Fig. S1, Table SIII). Results of MTOI are summarized in Table SIV. The ODEs for PMN are:

$$\begin{aligned} \frac{d[AA]}{dt} &= \frac{K_{cat,PLA2}(1 + \frac{[AA]}{K_{AA \rightarrow PLA2}})[PLA2][PL]}{K_{m,PLA2}(1 + \frac{[AA]}{K_i}) + [PL]} - \frac{K_{cat,E1}[E1][AA]}{K_{m,E1}(1 + \frac{[HETEs]}{K_i}) + [AA]} \\ &\quad - \frac{K_{cat,5-LOX}[5-LOX][AA]}{K_{m,5-LOX}(1 + \frac{[LTA4]}{K_i} + \frac{[HETEs]}{K_{i_{HETEs \rightarrow 5-LOX}}}) + [AA]} - \frac{K_{cat,COX2}[COX-2][AA]}{K_{m,COX2}(1 + \frac{[PGH2]}{K_i} + \frac{[HETEs]}{K_{i_{HETEs \rightarrow COX2}}}) + [AA]} \\ \frac{d[HETEs]}{dt} &= \frac{K_{cat,E1}[E1][AA]}{K_{m,E1}(1 + \frac{[HETEs]}{K_i}) + [AA]} - K_{d_{HETEs}}[HETEs] \\ \frac{d[PGH2]}{dt} &= \frac{K_{cat,COX2}[COX-2][AA]}{K_{m,COX2}(1 + \frac{[PGH2]}{K_i} + \frac{[HETEs]}{K_{i_{HETEs \rightarrow COX2}}}) + [AA]} - \frac{K_{cat,PGES}[PGES][PGH2]}{K_{m,PGES}(1 + \frac{[PGE2]}{K_i}) + [PGH2]} \\ &\quad - \frac{K_{cat,TXAS}[TXAS][PGH2]}{K_{m,TXAS}(1 + \frac{[TXS]}{K_i}) + [PGH2]} \end{aligned}$$

$$\frac{d[PGE2]}{dt} = \frac{K_{cat,PGES}[PGES][PGH2]}{K_{m,PGES}(1 + \frac{[PGE2]}{Ki}) + [PGH2]}$$

$$\frac{d[TXS]}{dt} = \frac{K_{cat,TXAS}[TXAS][PGH2]}{K_{m,TXAS}(1 + \frac{[TXS]}{Ki}) + [PGH2]}$$

$$\frac{d[LTA4]}{dt} = \frac{K_{cat,5-LOX}[5-LOX][AA]}{K_{m,5-LOX}(1 + \frac{[LTA4]}{Ki} + \frac{[HETEs]}{Ki_{HETEs \rightarrow 5-LOX}}) + [AA]} - \frac{K_{cat,LTA4H}[LTA4H][LTA4]}{K_{m,LTA4H}(1 + \frac{[LTA4]}{Ki}) + [LTA4]}$$

$$\frac{d[LTA4H]}{dt} = \frac{K_{cat,LTA4H}[LTA4H][LTA4]}{K_{m,LTA4H}(1 + \frac{[LTA4]}{Ki}) + [LTA4]} - Kd_{LTA4H}[LTA4H]$$

$$\frac{d[PLA2]}{dt} = 0$$

$$\frac{d[E1]}{dt} = 0$$

$$\frac{d[COX-2]}{dt} = 0$$

$$\frac{d[PGES]}{dt} = 0$$

$$\frac{d[TXAS]}{dt} = -Ki_{PGH2 \rightarrow TXAS}[PGH2][TXAS]$$

$$\frac{d[5-LOX]}{dt} = -Ki_{LTA4 \rightarrow 5-LOX}[LTA4][5-LOX]$$

$$\frac{d[LTA4H]}{dt} = -Ki_{LTA4 \rightarrow LTA4H}[LTA4][LTA4H]$$

The ODEs for EC are:

$$\frac{d[AA]}{dt} = \frac{K_{cat,PLA2}(1 + \frac{[AA]}{K_{AA \rightarrow PLA2}})[PLA2][PL]}{K_{m,PLA2}(1 + \frac{[AA]}{Ki}) + [PL]} - \frac{K_{cat,E1}[E1][AA]}{K_{m,E1}(1 + \frac{[HETEs]}{Ki}) + [AA]} - \frac{K_{cat,COX2}[COX-2][AA]}{K_{m,COX2}(1 + \frac{[PGH2]}{Ki}) + [AA]}$$

$$\frac{d[HETEs]}{dt} = \frac{K_{cat,E1}[E1][AA]}{K_{m,E1}(1 + \frac{[HETEs]}{Ki}) + [AA]} - Kd_{HETEs}[HETEs]$$

$$\frac{d[PGH2]}{dt} = \frac{K_{cat,COX2}[COX-2][AA]}{K_{m,COX2}(1 + \frac{[PGH2]}{Ki}) + [AA]} - \frac{K_{cat,TXAS}[TXAS][PGH2]}{K_{m,TXAS}(1 + \frac{[TXS]}{Ki}) + [PGH2]} - \frac{K_{cat,PGIS}[PGIS][PGH2]}{K_{m,PGIS}(1 + \frac{[PGI2]}{Ki} + \frac{[HETEs]}{Ki_{HETEs \rightarrow PGIS}}) + [PGH2]}$$

$$\begin{aligned}
& - \frac{K_{cat,PGES}[PGES][PGH2]}{K_{m,PGES} \left(1 + \frac{[PGE2]}{K_i} + \frac{[HETEs]}{K_{i_{HETEs \rightarrow PGES}}}\right) + [PGH2]} - Kd_{PGH2}[PGH2] \\
\frac{d[PGE2]}{dt} &= \frac{K_{cat,PGES}[PGES][PGH2]}{K_{m,PGES} \left(1 + \frac{[PGE2]}{K_i} + \frac{[HETEs]}{K_{i_{HETEs \rightarrow PGES}}}\right) + [PGH2]} - Kd_{PGE2}[PGE2] \\
\frac{d[PGI2]}{dt} &= \frac{K_{cat,PGIS}[PGIS][PGH2]}{K_{m,PGIS} \left(1 + \frac{[PGI2]}{K_i} + \frac{[HETEs]}{K_{i_{HETEs \rightarrow PGIS}}}\right) + [PGH2]} \\
\frac{d[TXS]}{dt} &= \frac{K_{cat,TXAS}[TXAS][PGH2]}{K_{m,TXAS} \left(1 + \frac{[TXS]}{K_i}\right) + [PGH2]} \\
\frac{d[PLA2]}{dt} &= 0 \\
\frac{d[E1]}{dt} &= 0 \\
\frac{d[COX-2]}{dt} &= 0 \\
\frac{d[PGES]}{dt} &= 0 \\
\frac{d[PGIS]}{dt} &= 0 \\
\frac{d[TXAS]}{dt} &= -K_{i_{PGH2 \rightarrow TXAS}}[PGH2][TXAS]
\end{aligned}$$

The ODEs for PLT are:

$$\begin{aligned}
\frac{d[AA]}{dt} &= \frac{K_{cat,PLA2} \left(1 + \frac{[AA]}{K_{AA \rightarrow PLA2}}\right) [PLA2][PL]}{K_{m,PLA2} \left(1 + \frac{[AA]}{K_i}\right) + [PL]} - \frac{K_{cat,E1}[E1][AA]}{K_{m,E1} \left(1 + \frac{[HETEs]}{K_i}\right) + [AA]} \\
& - \frac{K_{cat,COX1}[COX-1][AA]}{K_{m,COX1} \left(1 + \frac{[PGH2]}{K_i} + \frac{[HETEs]}{K_{i_{HETEs \rightarrow COX1}}}\right) + [AA]} \\
\frac{d[HETEs]}{dt} &= \frac{K_{cat,E1}[E1][AA]}{K_{m,E1} \left(1 + \frac{[HETEs]}{K_i}\right) + [AA]} - Kd_{HETEs}[HETEs] \\
\frac{d[TXS]}{dt} &= \frac{K_{cat,COX1}[COX-1][AA]}{K_{m,COX1} \left(1 + \frac{[PGH2]}{K_i} + \frac{[HETEs]}{K_{i_{HETEs \rightarrow COX1}}}\right) + [AA]} \\
\frac{d[PLA2]}{dt} &= 0 \\
\frac{d[E1]}{dt} &= 0
\end{aligned}$$

$$\frac{d[COX-1]}{dt} = -K_{i_{TXS \rightarrow COX1}}[TXS][COX-1]$$

### Single parameter sensitivity analysis

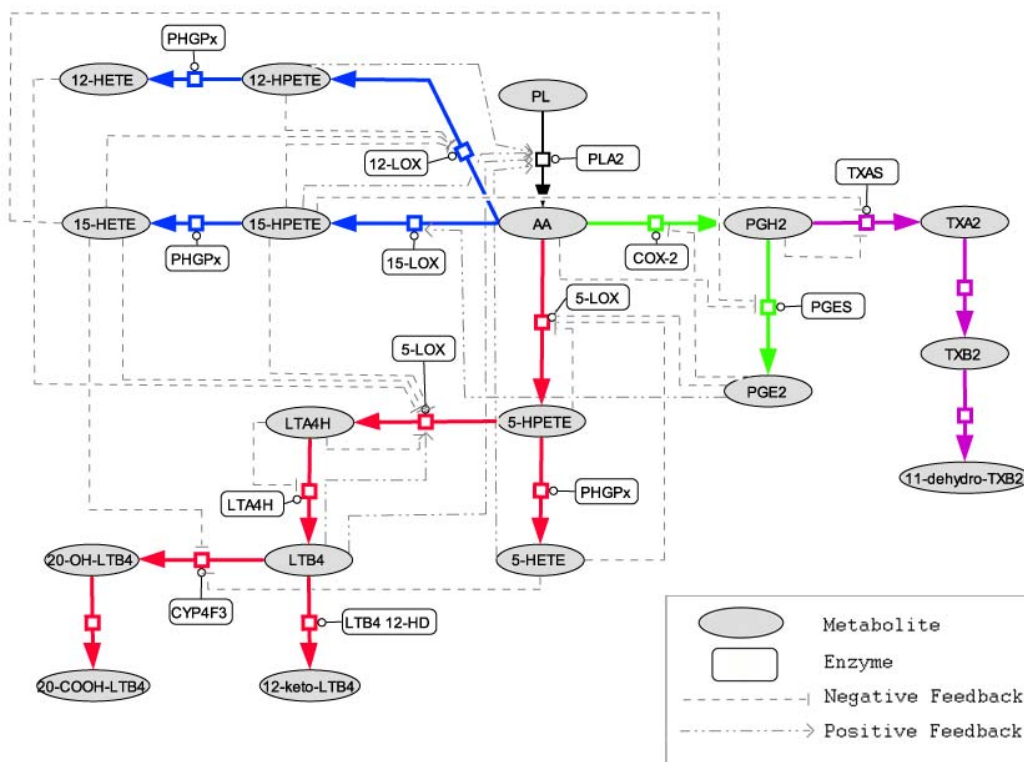
Single parameter sensitivity analysis was performed using the reported method (Pant & Ghosh, 2005). The sensitivity coefficient is calculated as:

$$C = \frac{\Delta F_{obj} / F_{obj}}{\Delta K / K}$$

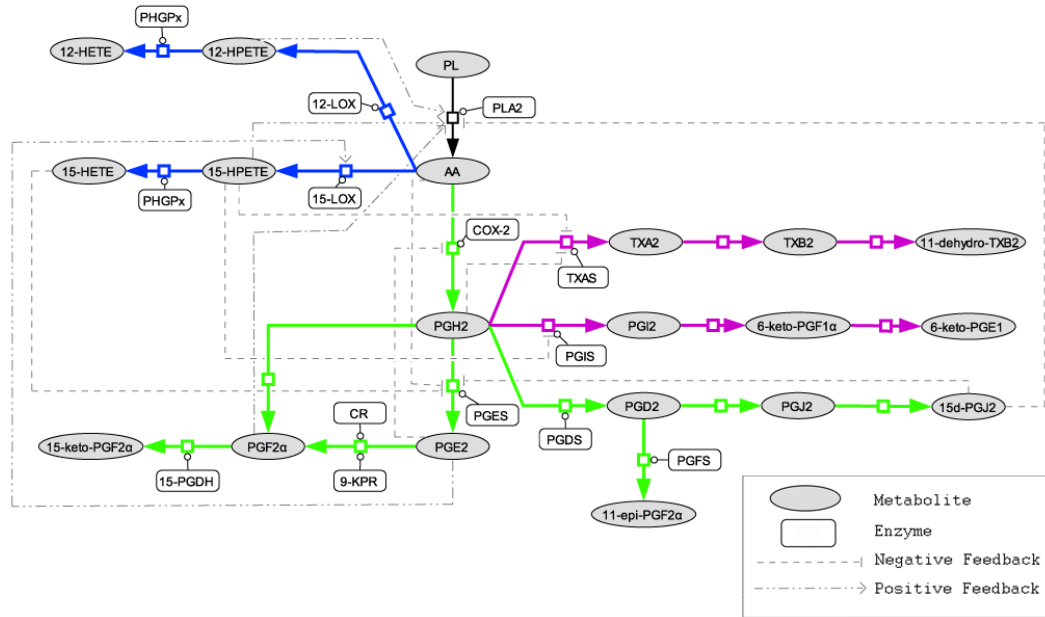
The result of AAnetwork is summarized in Table SV

### Supplementary Figures

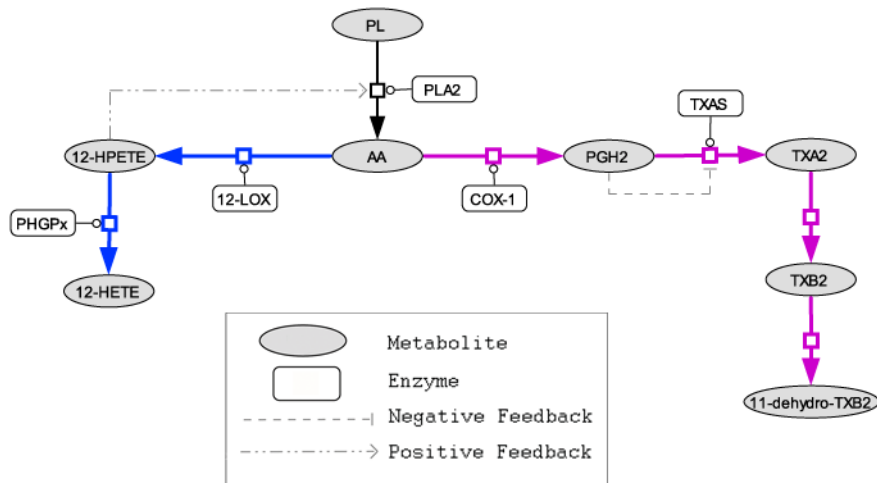
**Figure S1.** The AAnetwork in human PMN (A), EC (B) and PLT (C). Two pathways are responsible to the production of inflammatory mediators: COX-2 pathway (green) and 5-LOX pathway (red). LTB4 and PGE2 are major inflammatory mediators produced in the AAnetwork. The HETEs pathway is in blue, while the pathways of PGI2 and TXA2 production are in purple.



(A)

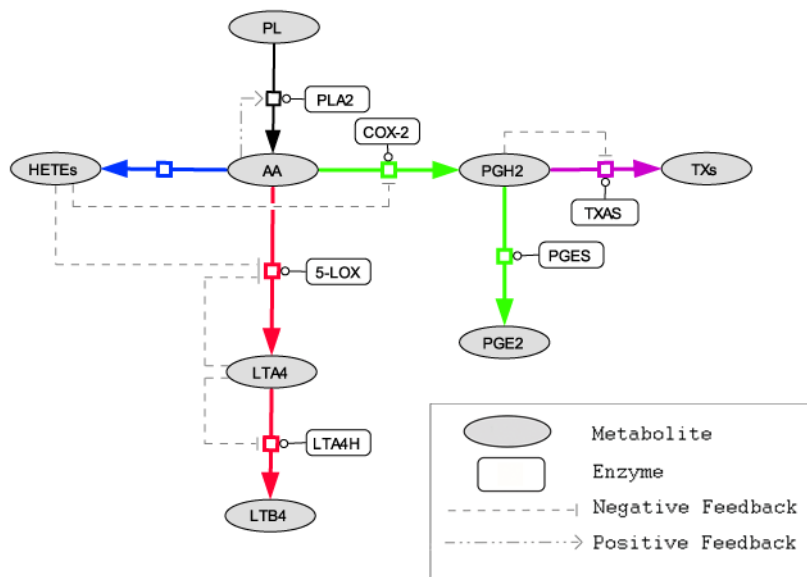


(B)

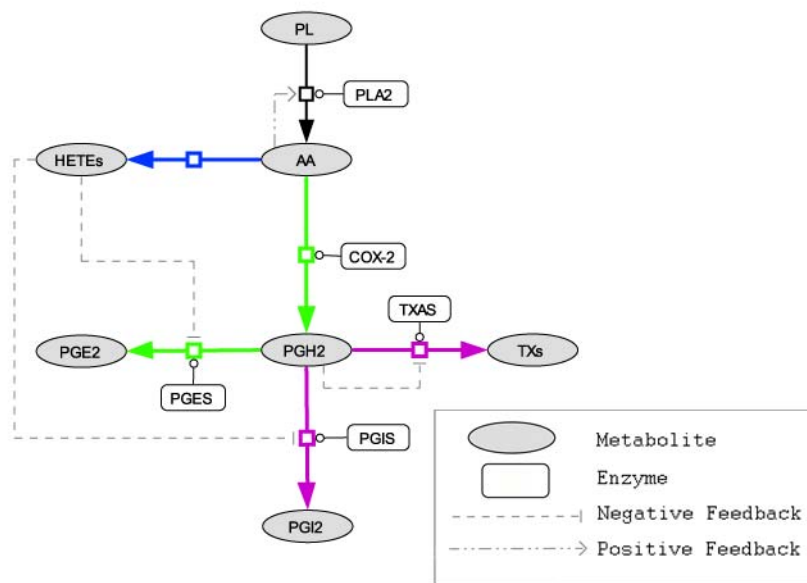


(C)

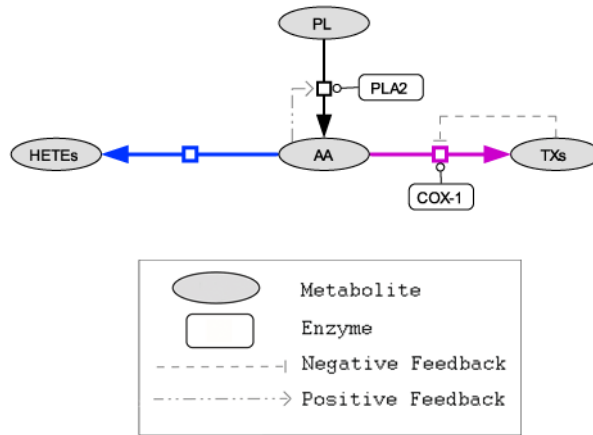
**Figure S2.** The simplified AA network in human PMN (A), EC (B) and PLT (C). The COX-2 pathway is in green, the 5-LOX pathway is in red, the HETEs pathway is in blue, and the pathways of PGI<sub>2</sub> and TXA<sub>2</sub> production are in purple.



(A)

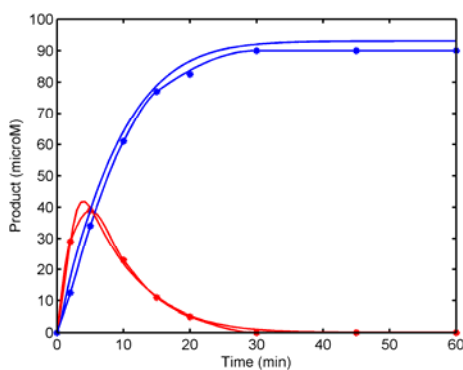


(B)

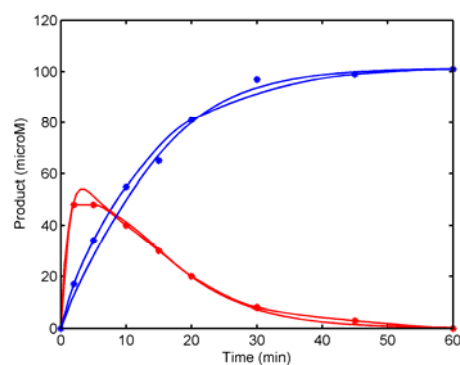


(C)

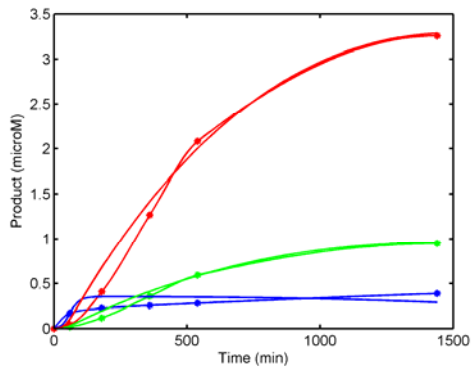
**Figure S3.** Experimental data and Parameter fitting results. Lines with star are experimental curves while the ones without star are model calculations with fitted parameters. **(A)** The production curve of LTB<sub>4</sub> (red) and ω-LTB<sub>4</sub> (blue) in PMN. In the experiment (Shak & Goldstein, 1984), PMN were incubated at 37°C with 10 microM A23187 (a calcium ionophore) added at time zero. **(B)** The same as in **(A)**, but with 10 microM A23187 + 30microM arachidonic acid added at time zero. **(C)** The production curve of PGF<sub>2</sub>α (red), PGE<sub>2</sub> (green) and 6-keto-PGF<sub>1</sub>α (blue) in EC where 10U/mL 1L-1β was added at time zero (Camacho *et al*, 1998). **(D)** The production curve of TXA<sub>2</sub> (red) and TXB<sub>2</sub> (blue) in PLT, where 160 μM exogenous AA was added at time zero (Anderson *et al*, 1978).



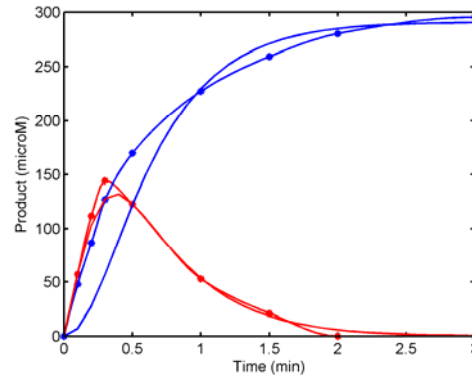
(A)



(B)

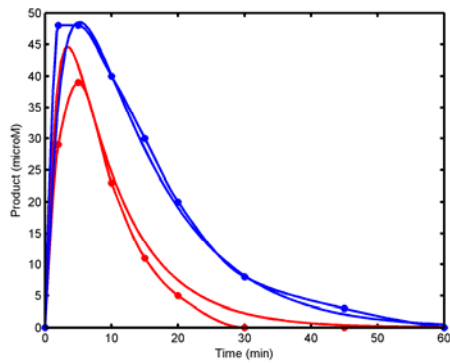


(C)

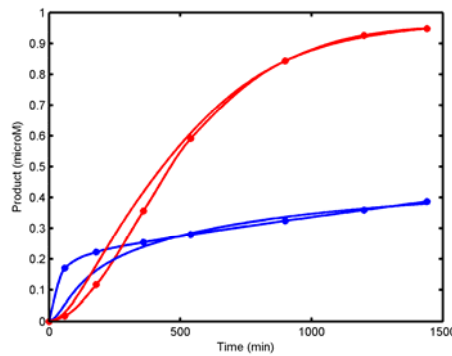


(D)

**Figure S4.** Parameter fitting results of the simplified AAnetwork model. Lines with star are experimental curves while the ones without star are model calculations with fitted parameters. **(A)** The production curve of LTB4 in PMN. In the experiment (Shak *et al*, 1984), PMN were incubated at 37°C with 10 microM A23187 (red) or with 10 microM A23187 + 30microM arachidonic acid (blue) added at time zero. **(B)** The production curve PGE2 (red) and PGI2 (blue) in EC where 10U/mL 1L-1β was added at time zero (Camacho *et al*, 1998). **(C)** The total production curve of TXA2 and TXB2 in PLT, where 160 μM exogenous AA was added at time zero (Anderson *et al*, 1978).

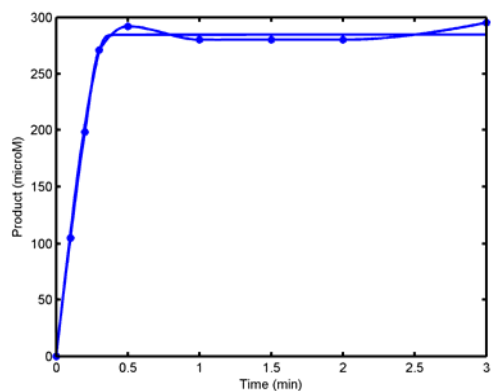


(A)



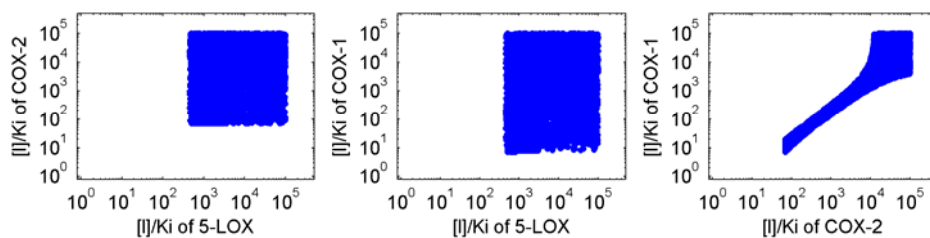
(B)



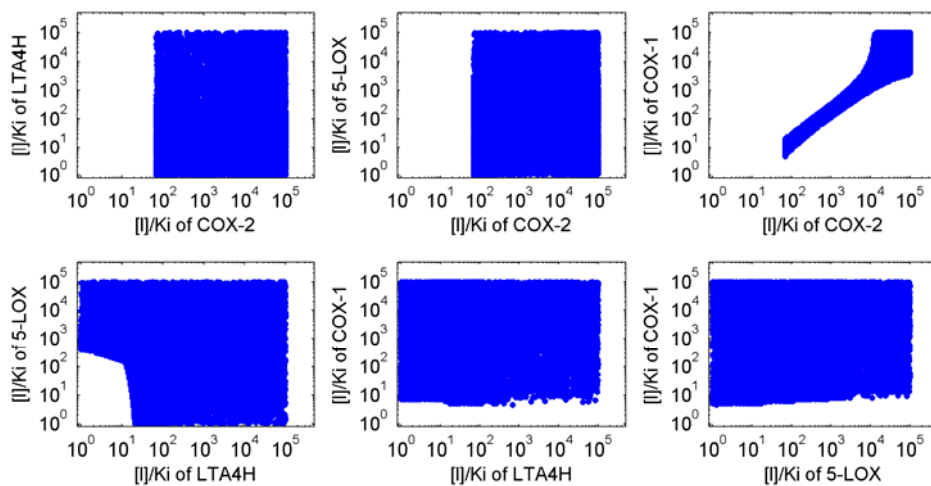


(C)

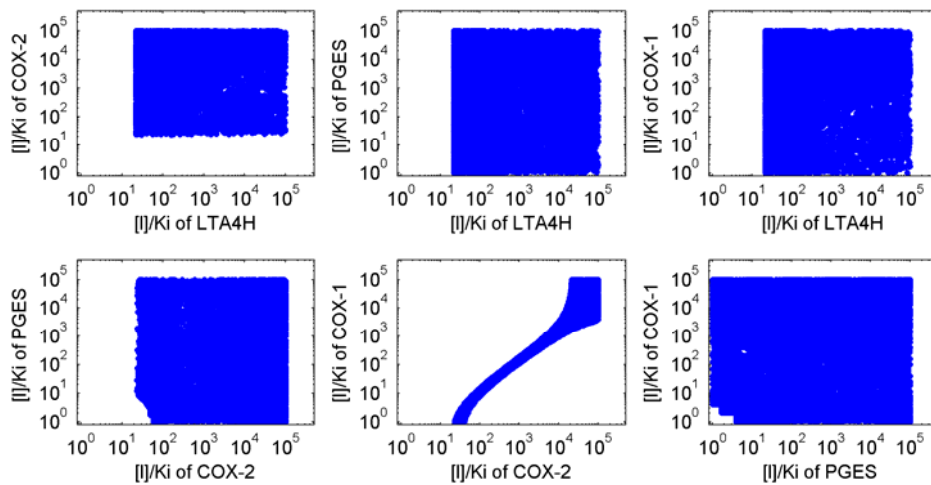
**Figure S5.** The distribution of  $[I]/K_i$  of MTOI solutions: the inhibitor against COX1/2 and 5-LOX (A), the inhibitor against COX1/2, 5-LOX and LTA4H (B), the inhibitor against COX1/2, PGES and LTA4H (C), the inhibitor against COX1/2, PGES and 5-LOX (D), the inhibitor against COX1/2, PGES, 5-LOX and LTA4H (E), the inhibitor against PLA2 and COX1/2 (F), the inhibitor against PLA2, COX1/2 and 5-LOX (G), the inhibitor against PLA2, COX1/2, 5-LOX and LTA4H (H), the inhibitor against PLA2, COX1/2 and PGES (I), the inhibitor against PGES and LTA4H (J), the inhibitor against PLA2, COX1/2 and PGES (I), the inhibitor against PGES and LTA4H (J), the inhibitor against PLA2, COX1/2, PGES and LTA4H (K), the inhibitor against PLA2, COX1/2, PGES and 5-LOX (L), the inhibitor against PLA2, COX-1/2, PGES, 5-LOX and LTA4H (M).



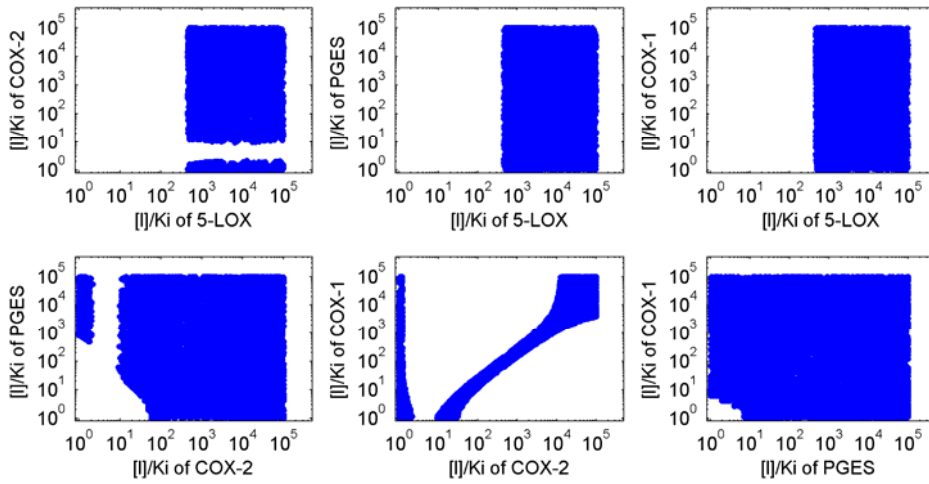
(A)



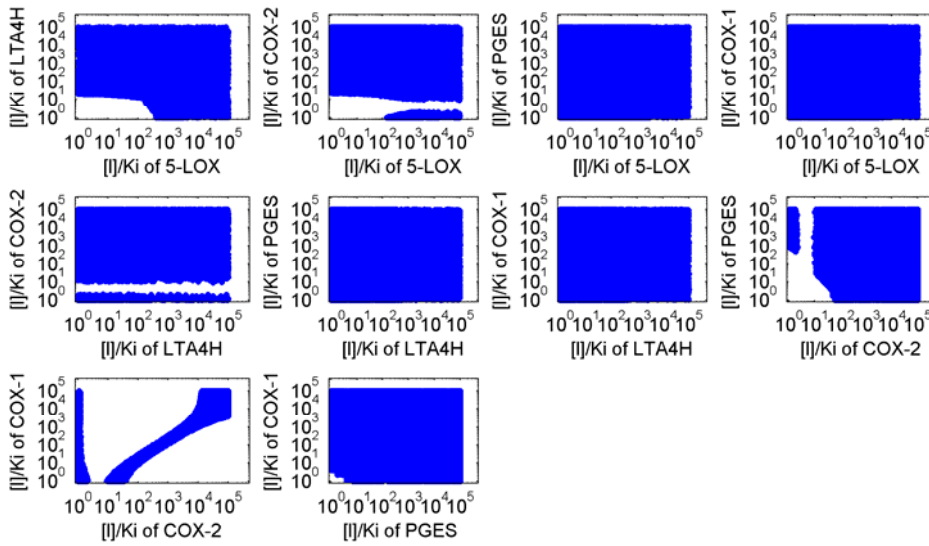
(B)



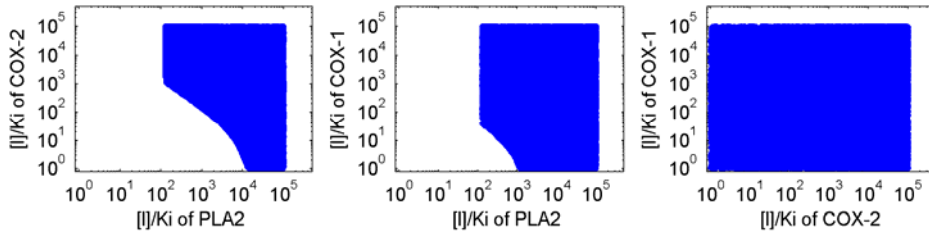
(C)



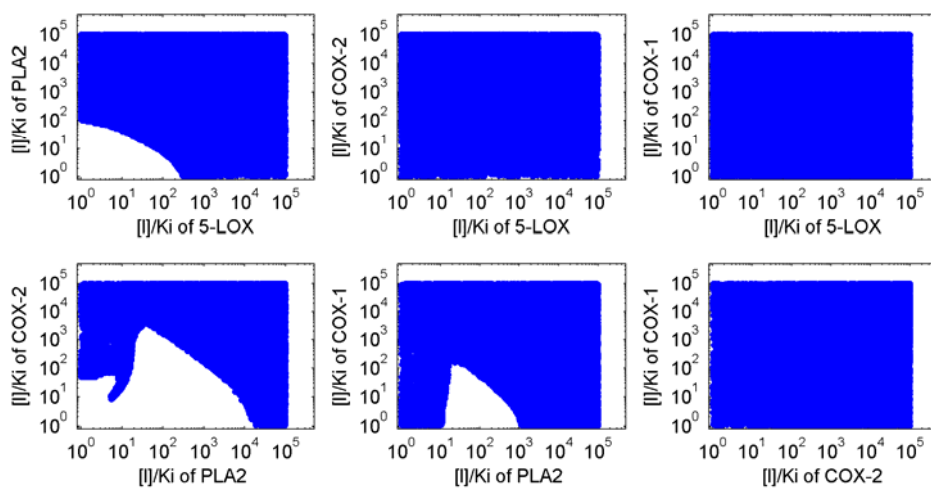
(D)



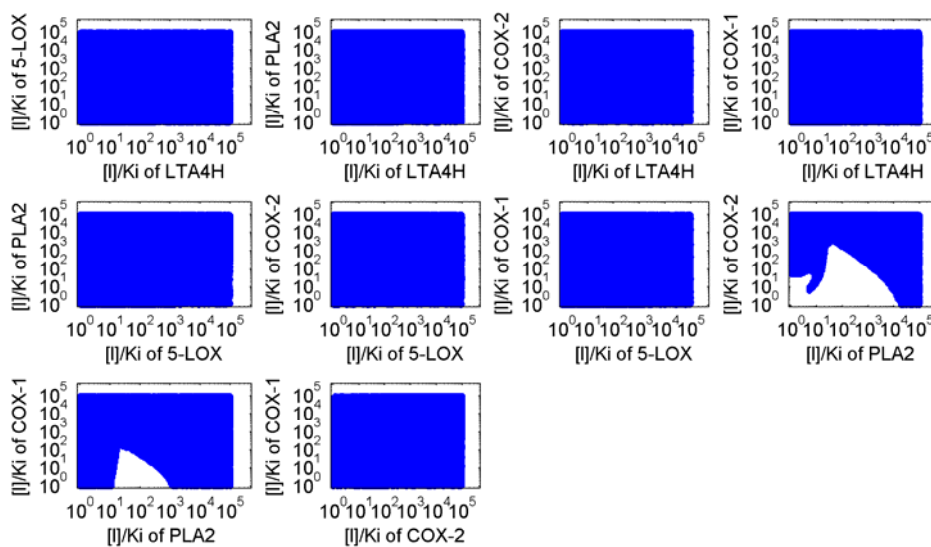
(E)



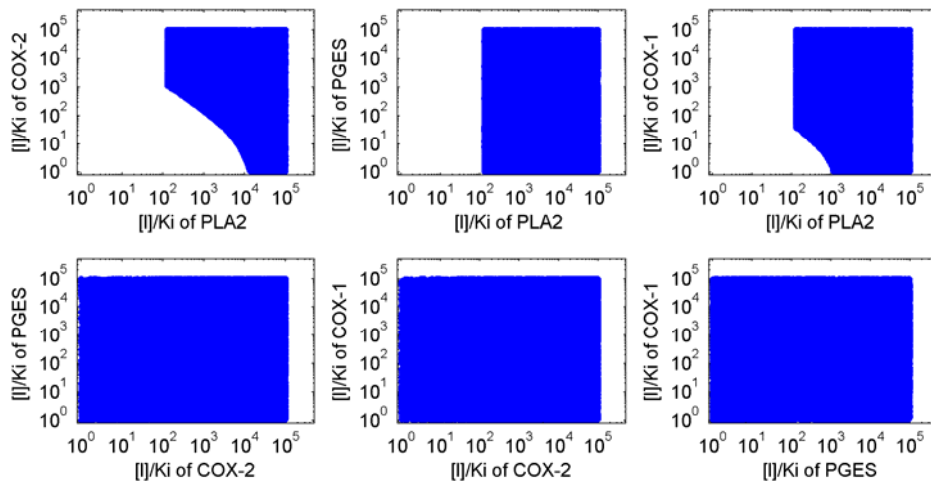
(F)



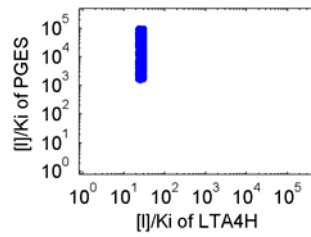
(G)



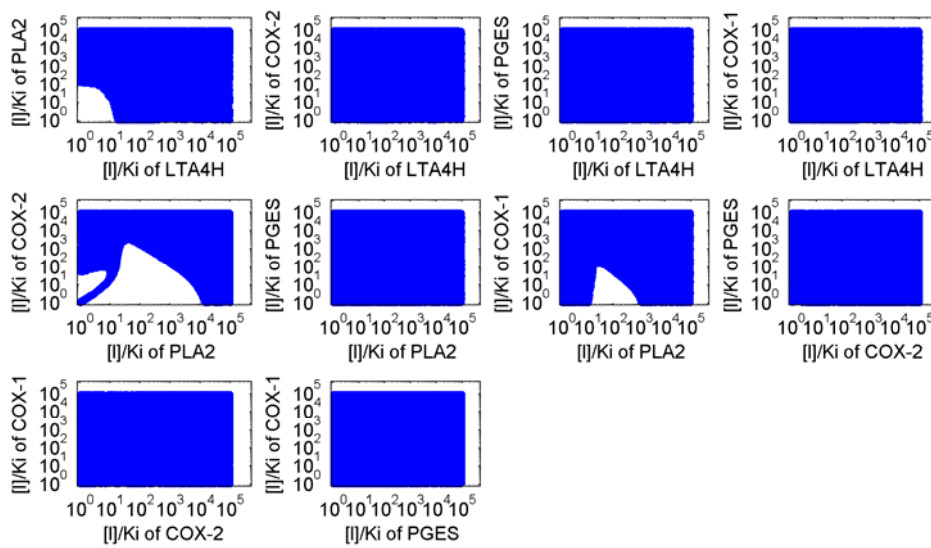
(H)



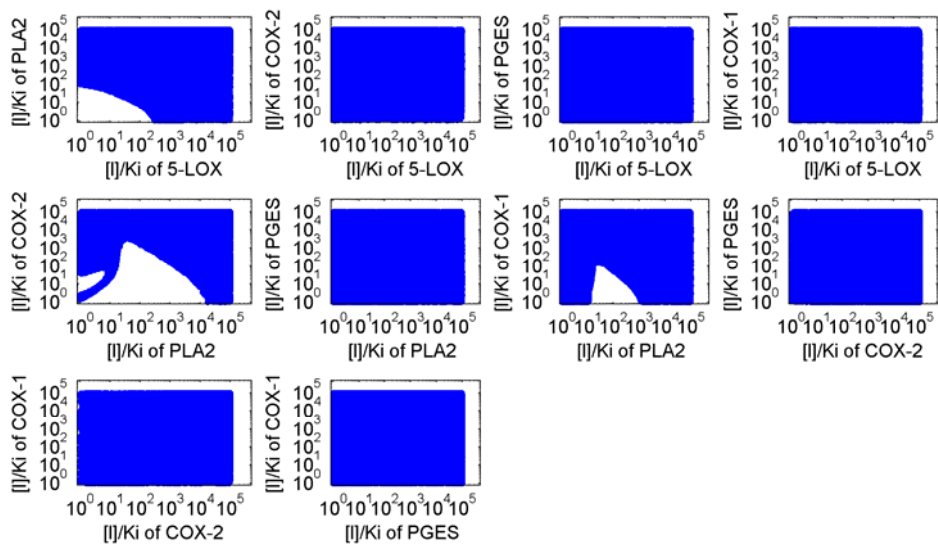
(I)



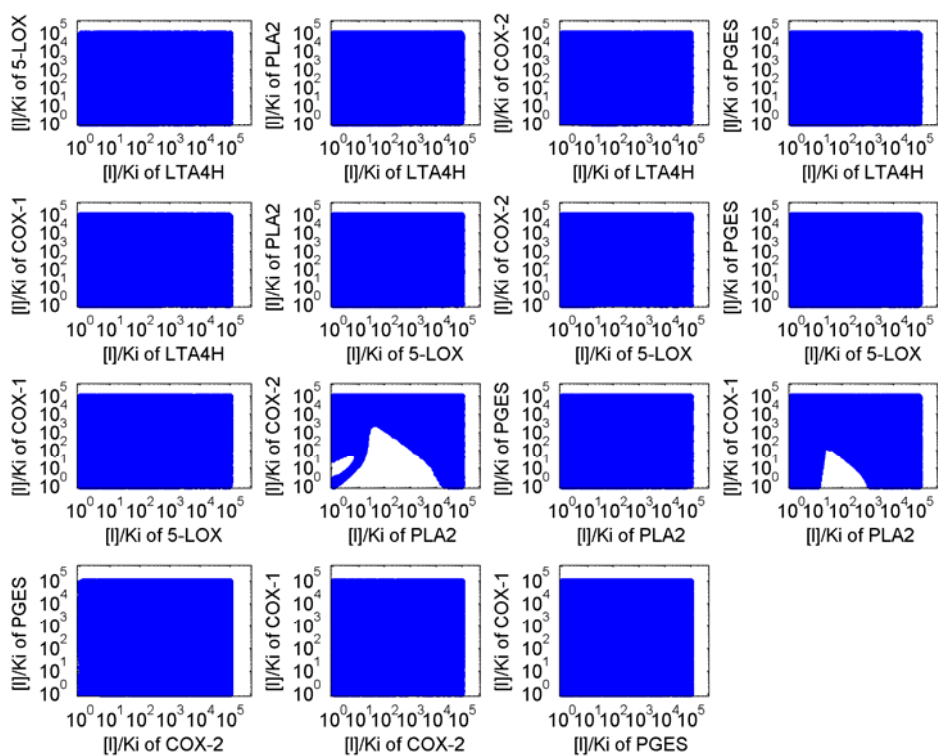
(J)



(K)



(L)



(M)

## Reference

Anderson MW, Crutchley DJ, Tainer BE, Eling TE (1978) Kinetic studies on the conversion of prostaglandin endoperoxide PGH<sub>2</sub> by thromboxane synthase. *Prostaglandins* **16**: 563-570.

Camacho M, Lopez-Belmonte J, Vila L (1998) Rate of vasoconstrictor prostanoids released by endothelial cells depends on cyclooxygenase-2 expression and prostaglandin I synthase activity. *Circ Res* **83**: 353-365.

Claesson HE, Haeggstrom J (1988) Human endothelial cells stimulate leukotriene synthesis and convert granulocyte released leukotriene A4 into leukotrienes B4, C4, D4 and E4. *Eur J Biochem* **173**: 93-100.

Denzlinger C (1996) Biology and pathophysiology of leukotrienes. *Crit Rev Oncol Hematol* **23**: 167-223.

Husain S, Abdel-Latif AA (2001) Effects of prostaglandin F(2alpha) and carbachol on MAP kinases, cytosolic phospholipase A(2) and arachidonic acid release in cat iris sphincter smooth muscle cells. *Exp Eye Res* **72**: 581-590.

Inoue H, Tanabe T, Umesono K (2000) Feedback control of cyclooxygenase-2 expression through PPARgamma. *J Biol Chem* **275**: 28028-28032.

Kojima F, Kato S, Kawai S (2005) Prostaglandin E synthase in the pathophysiology of arthritis. *Fundam Clin Pharmacol* **19**: 255-261.

Ohd JF, Wikstrom K, Sjolander A (2000) Leukotrienes induce cell-survival signaling in intestinal epithelial cells. *Gastroenterology* **119**: 1007-1018.

Pant D, Ghosh A (2005) Automated oncogene detection in complex protein networks with applications to the MAPK signal transduction pathway. *Biophys Chem* **113**: 275-288.

Shak S, Goldstein IM (1984) Omega-oxidation is the major pathway for the catabolism of leukotriene B4 in human polymorphonuclear leukocytes. *J Biol Chem* **259**: 10181-10187.

Tsubouchi Y, Kawahito Y, Kohno M, Inoue K, Hla T, Sano H (2001) Feedback control of the arachidonate cascade in rheumatoid synoviocytes by 15-deoxy-Delta(12,14)-prostaglandin J2. *Biochem Biophys Res Commun* **283**: 750-755.

Weaver JA, Maddox JF, Cao YZ, Mullarky IK, Sordillo LM (2001) Increased 15-HPETE production decreases prostacyclin synthase activity during oxidant stress in aortic endothelial cells. *Free Radic Biol Med* **30**: 299-308.

Yang K, Ma W, Liang H, Ouyang Q, Tang C, Lai L (2007) Dynamic Simulations on the Arachidonic Acid Metabolic Network. *PLoS Comput Biol* **3**: e55.

Table SI-I. The feedback parameters used in the ODEs. Italics means the corresponding parameter has no direct value from experiments and is derived from parameter fitting. The parameter set used in the main manuscript is referred as set 1.

PMN:						
Feedbacks	Parameters	Set 1	Set 2	Set 3	Set 4	Set 5
AA→PGES (Quraishi O., et al., 2002)	Ki <sub>AA→PGES</sub>	0.3μM	0.3μM	0.3μM	0.3μM	0.3μM
15-HETE→PGES (Quraishi O., et al., 2002)	Ki <sub>15-HETE→PGES</sub>	<i>0.53μM</i>	<i>30μM</i>	<i>1.4μM</i>	<i>0.29μM</i>	<i>1.4μM</i>
PGE2→COX2 (Nathan C., 2002)	Ki <sub>PGE2→COX2</sub>	<i>2.8μM</i>	<i>30μM</i>	<i>0.13μM</i>	<i>5.2μM</i>	<i>0.13μM</i>
15-HPETE→TXAS (Jones D. A.&Fitzpatrick F. A., 1991)	Ki <sub>15-HPETE→TXAS</sub>	<i>0.52(μM*min)<sup>-1</sup></i>	<i>0.6(μM*min)<sup>-1</sup></i>	<i>0.73(μM*min)<sup>-1</sup></i>	<i>0.031(μM*min)<sup>-1</sup></i>	<i>0.73(μM*min)<sup>-1</sup></i>
PGH2→TXAS (Jones D. A.&Fitzpatrick F. A., 1990)	Ki <sub>PGH2→TXAS</sub>	<i>0.029(μM*min)<sup>-1</sup></i>	<i>0.1(μM*min)<sup>-1</sup></i>	<i>0.018(μM*min)<sup>-1</sup></i>	<i>3.4(μM*min)<sup>-1</sup></i>	<i>0.018(μM*min)<sup>-1</sup></i>
15-HPETE→5-LOX (Cashman J. R., et al., 1988)	Ki <sub>15-HPETE→5-LOX</sub>	0.01(μM*min) <sup>-1</sup>	0.01(μM*min) <sup>-1</sup>	0.01(μM*min) <sup>-1</sup>	0.01(μM*min) <sup>-1</sup>	0.01(μM*min) <sup>-1</sup>
12-HETE→5-LOX (Vanderhoek J. Y., et al., 1985)	Ki <sub>12-HETE→5-LOX</sub>	30 μM	30 μM	30 μM	30 μM	30 μM
15-HETE→5-LOX (Vanderhoek J. Y., et al., 1985)	Ki <sub>15-HETE→5-LOX</sub>	4 μM	4 μM	4 μM	4 μM	4 μM
LTA4→5-LOX (Lepley R. A.&Fitzpatrick F. A., 1994)	Ki <sub>LTA4→5-LOX</sub>	0.175(μM*min) <sup>-1</sup>	0.175(μM*min) <sup>-1</sup>	0.175(μM*min) <sup>-1</sup>	0.175(μM*min) <sup>-1</sup>	0.175(μM*min) <sup>-1</sup>
5-HPETE→5-LOX (Aharony D., et al., 1987)	Ki <sub>5-HPETE→5-LOX</sub>	<i>0.26(μM*min)<sup>-1</sup></i>	<i>0.01(μM*min)<sup>-1</sup></i>	<i>5.8(μM*min)<sup>-1</sup></i>	<i>0.019(μM*min)<sup>-1</sup></i>	<i>5.8(μM*min)<sup>-1</sup></i>
PGE2→5-LOX (Levy B. D., et al., 2001)	Ki <sub>PGE2→5-LOX</sub>	<i>72μM</i>	<i>15μM</i>	<i>88μM</i>	<i>25μM</i>	<i>88μM</i>
5-HETE→5-LOX (Aharony D., et al., 1987)	Ki <sub>5-HETE→5-LOX</sub>	6.3μM	6.3μM	6.3μM	6.3μM	6.3μM
LTA4→LTA4H (Orning L., et al., 1992)	Ki <sub>LTA4→LTA4H</sub>	129 (turnover/inactivation)	129 (turnover/inactivation)	129 (turnover/inactivation)	129 (turnover/inactivation)	129 (turnover/inactivation)
12-HETE→CYP4F3 (Soberman R. J., et al., 1987)	Ki <sub>12-HETE→CYP4F3</sub>	<i>0.29μM</i>	<i>0.2μM</i>	<i>0.46μM</i>	<i>0.12μM</i>	<i>0.46μM</i>
5-HETE→CYP4F3 (Soberman R. J., et al., 1987)	Ki <sub>5-HETE→CYP4F3</sub>	<i>0.8μM</i>	<i>0.86μM</i>	<i>2μM</i>	<i>2μM</i>	<i>2μM</i>
15-HETE→12-LOX (Kishimoto K., et al., 1996)	Ki <sub>15-HETE→12-LOX</sub>	<i>1.5μM</i>	<i>10μM</i>	<i>0.8μM</i>	<i>58μM</i>	<i>0.8μM</i>
15-HPETE→12-LOX (Kishimoto K., et al., 1996)	Ki <sub>15-HPETE→12-LOX</sub>	<i>0.23(μM*min)<sup>-1</sup></i>	<i>10(μM*min)<sup>-1</sup></i>	<i>2.2(μM*min)<sup>-1</sup></i>	<i>3.4(μM*min)<sup>-1</sup></i>	<i>2.2(μM*min)<sup>-1</sup></i>
12-HPETE→12-LOX (Kishimoto K., et al., 1996)	Ki <sub>12-HPETE→12-LOX</sub>	<i>1.6μM</i>	<i>10μM</i>	<i>16μM</i>	<i>1.1μM</i>	<i>16μM</i>
12-HPETE→PLA2 (Balboa M. A., et al., 2003)	Ki <sub>12-HPETE→PLA2</sub>	<i>260μM</i>	<i>500μM</i>	<i>150μM</i>	<i>530μM</i>	<i>150μM</i>
15-HPETE→PLA2 (Balboa M. A., et al., 2003)	Ki <sub>15-HPETE→PLA2</sub>	<i>430μM</i>	<i>200μM</i>	<i>920μM</i>	<i>210μM</i>	<i>920μM</i>
LTB4→PLA2 (Wijkander J., et al., 1995)	K <sub>LTB4→PLA2</sub>	<i>180μM</i>	<i>500μM</i>	<i>320μM</i>	<i>480μM</i>	<i>320μM</i>
5-HETE→PLA2 (Wijkander J., et al., 1995)	K <sub>5-HETE→PLA2</sub>	<i>240μM</i>	<i>500μM</i>	<i>970μM</i>	<i>990μM</i>	<i>970μM</i>
LTB4→5-LOX (Serio K. J., et al., 1997)	K <sub>LTB4→5-LOX</sub>	<i>0.022(μM*min)<sup>-1</sup></i>	<i>0.053(μM*min)<sup>-1</sup></i>	<i>0.0153(μM*min)<sup>-1</sup></i>	<i>0.013(μM*min)<sup>-1</sup></i>	<i>0.0153(μM*min)<sup>-1</sup></i>
PGE2→15-LOX (Levy B. D., et al., 2001)	K <sub>PGE2→15-LOX</sub>	0.15	0.15	0.15	0.15	0.15
Product inhibition	Ki <sub>PGE2→15-LOX</sub>	0.000023μM	0.000023μM	0.000023μM	0.000023μM	0.000023μM
EC:						
PGE2→COX2 (Akarasereonont P., et al., 2001)	Ki <sub>PGE2→COX2</sub>	30μM	30μM	30μM	30μM	30μM
15-HPETE→TXAS (Jones D. A.&Fitzpatrick F. A., 1991)	Ki <sub>15-HPETE→TXAS</sub>	<i>0.9(μM*min)<sup>-1</sup></i>	<i>0.15(μM*min)<sup>-1</sup></i>	<i>0.9(μM*min)<sup>-1</sup></i>	<i>1.2(μM*min)<sup>-1</sup></i>	<i>0.9(μM*min)<sup>-1</sup></i>
PGH2→TXAS (Jones D. A.&Fitzpatrick F. A., 1990)	Ki <sub>PGH2→TXAS</sub>	<i>0.18(μM*min)<sup>-1</sup></i>	<i>0.35(μM*min)<sup>-1</sup></i>	<i>0.18(μM*min)<sup>-1</sup></i>	<i>0.033(μM*min)<sup>-1</sup></i>	<i>0.18(μM*min)<sup>-1</sup></i>
15-HPETE→PGIS (Weaver J. A., et al., 2001)	Ki <sub>15-HPETE→PGIS</sub>	<i>0.18μM</i>	<i>0.01μM</i>	<i>0.18μM</i>	<i>0.46μM</i>	<i>0.18μM</i>
AA→PGES (Quraishi O., et al., 2002)	Ki <sub>AA→PGES</sub>	0.3μM	0.3μM	0.3μM	0.3μM	0.3μM
15-HETE→PGES (Quraishi O., et al., 2002)	Ki <sub>15-HETE→PGES</sub>	<i>0.91μM</i>	<i>500μM</i>	<i>0.91μM</i>	<i>5.9μM</i>	<i>0.91μM</i>
15d-PGJ2→PGES (Kojima F., et al., 2005)	Ki <sub>15d-PGJ2→PGES</sub>	<i>0.018(μM*min)<sup>-1</sup></i>	<i>0.01(μM*min)<sup>-1</sup></i>	<i>0.018(μM*min)<sup>-1</sup></i>	<i>0.01(μM*min)<sup>-1</sup></i>	<i>0.018(μM*min)<sup>-1</sup></i>
15d-PGJ2→PLA2 (Tsubouchi Y., et al., 2001)	Ki <sub>15d-PGJ2→PLA2</sub>	<i>0.018(μM*min)<sup>-1</sup></i>	<i>0.015(μM*min)<sup>-1</sup></i>	<i>0.018(μM*min)<sup>-1</sup></i>	<i>0.28(μM*min)<sup>-1</sup></i>	<i>0.018(μM*min)<sup>-1</sup></i>
12-HPETE→PLA2 (Balboa M. A., et al., 2003)	Ki <sub>12-HPETE→PLA2</sub>	<i>140μM</i>	<i>500μM</i>	<i>140μM</i>	<i>730μM</i>	<i>140μM</i>
15-HPETE→PLA2 (Balboa M. A., et al., 2003)	Ki <sub>15-HPETE→PLA2</sub>	<i>120μM</i>	<i>200μM</i>	<i>120μM</i>	<i>510μM</i>	<i>120μM</i>
PGF2→PLA2 (Husain S.&Abdel-Latif A. A., 2001)	K <sub>PGF2→PLA2</sub>	<i>210μM</i>	<i>500μM</i>	<i>210μM</i>	<i>200μM</i>	<i>210μM</i>
PGE2→15-LOX (Levy B. D., et al., 2001)	K <sub>PGE2→15-LOX</sub>	0.15	0.15	0.15	0.15	0.15
Product inhibition	Ki <sub>PGE2→15-LOX</sub>	0.000023μM	0.000023μM	0.000023μM	0.000023μM	0.000023μM
PLT:						
PGH2→TXAS (Jones D. A.&Fitzpatrick F. A., 1990)	Ki <sub>PGH2→TXAS</sub>	<i>0.025 (μM*min)<sup>-1</sup></i>	<i>0.08 (μM*min)<sup>-1</sup></i>	<i>0.025 (μM*min)<sup>-1</sup></i>	<i>0.035 (μM*min)<sup>-1</sup></i>	<i>0.031 (μM*min)<sup>-1</sup></i>
12-HPETE→PLA2 (Balboa M. A., et al., 2003)	Ki <sub>12-HPETE→PLA2</sub>	<i>310μM</i>	<i>500μM</i>	<i>230μM</i>	<i>610μM</i>	<i>150μM</i>
Product inhibition	Ki	<i>500μM</i>	<i>500μM</i>	<i>500μM</i>	<i>500μM</i>	<i>500μM</i>

Table SI-II. The Km and Kcat of enzymes used in the ODEs. Italics means the corresponding parameter has no direct value from experiments and is derived from parameter fitting.

PMN:																
Enzyme Name	EC number	Set 1			Set 2			Set 3			Set 4			Set 5		
		K <sub>cat</sub> (1/min)	K <sub>m</sub> (μM)	K <sub>cat</sub> (1/min)	K <sub>m</sub> (μM)	K <sub>cat</sub> (1/min)	K <sub>m</sub> (μM)	K <sub>cat</sub> (1/min)	K <sub>m</sub> (μM)	K <sub>cat</sub> (1/min)	K <sub>m</sub> (μM)	K <sub>cat</sub> (1/min)	K <sub>m</sub> (μM)			
15-LOX (Schomburg I., et al., 2004)	1.13.11.33	<i>5000</i>	<i>13</i>	<i>1000</i>	<i>70</i>	<i>4200</i>	<i>11</i>	<i>460</i>	<i>0.69</i>	<i>4200</i>	<i>11</i>					
5-LOX (Schomburg I., et al., 2004)	1.13.11.34	<i>6000</i>	<i>1.4</i>	<i>5000</i>	<i>5</i>	<i>3500</i>	<i>3.3</i>	<i>520</i>	<i>2.3</i>	<i>3500</i>	<i>3.3</i>					
LTA4H (Schomburg I., et al., 2004)	3.3.2.6	<i>150</i>	<i>20</i>	<i>125</i>	<i>20</i>	<i>7600</i>	<i>20</i>	<i>210</i>	<i>20</i>	<i>7600</i>	<i>20</i>					
CYP4F3A (Christmas P., et al., 1999)	1.14.13.30	150	3.9	150	3.9	150	3.9	150	3.9	150	3.9					
PHGPx (Schomburg I., et al., 2004)	1.11.1.12	<i>2000</i>	<i>58</i>	<i>500</i>	<i>70</i>	<i>600</i>	<i>0.14</i>	<i>290</i>	<i>0.12</i>	<i>600</i>	<i>0.14</i>					
COX-2 (Schomburg I., et al., 2004)	1.14.99.1	1000	33	1000	50	1000	0.52	1000	4	1000	0.52					
PGES (Schomburg I., et al., 2004)	5.3.99.3	3000	160	3000	160	3000	160	3000	160	3000	160					
TXAS (Schomburg I., et al., 2004)	5.3.99.5	1599	4	1599	4	1599	4	1599	4	1599	4					
PLA2 (Schomburg I., et al., 2004)	3.1.1.4	3600	2600	3600	2600	3600	2600	3600	2600	3600	2600					
12-LOX (Schomburg I., et al., 2004)	1.13.11.31	<i>9500</i>	<i>160</i>	<i>1000</i>	<i>50</i>	<i>1800</i>	<i>36</i>	<i>580</i>	<i>14</i>	<i>1800</i>	<i>36</i>					
EC:																
15-LOX (Schomburg I., et al., 2004)	1.13.11.33	<i>360</i>	<i>91</i>	<i>1000</i>	<i>70</i>	<i>360</i>	<i>91</i>	<i>160</i>	<i>120</i>	<i>360</i>	<i>91</i>					
PHGPx (Schomburg I., et al., 2004)	1.11.1.12	<i>160</i>	<i>250</i>	<i>500</i>	<i>50</i>	<i>160</i>	<i>250</i>	<i>6900</i>	<i>1</i>	<i>160</i>	<i>250</i>					
COX-2 (Schomburg I., et al., 2004)	1.14.99.1	<i>250</i>	<i>5</i>	<i>9000</i>	<i>5</i>	<i>250</i>	<i>5</i>	<i>100</i>	<i>5</i>	<i>250</i>	<i>5</i>					
PGES (Schomburg I., et al., 2004)	5.3.99.3	3000	160	3000	160	3000	160	3000	160	3000	160					
TXAS (Schomburg I., et al., 2004)	5.3.99.5	1599	4	1599	4	1599	4	1599	4	1599	4					
PLA2 (Schomburg I., et al., 2004)	3.1.1.4	3600	2600	3600	2600	3600	2600	3600	2600	3600	2600					



PGIS (Schomburg I., et al., 2004)	5.3.99.4	147	9	147	9	147	9	147	9	147	9
PGDS (Schomburg I., et al., 2004)	5.3.99.2	6900	4	200	4	6900	4	240	4	6900	4
PGFS (Schomburg I., et al., 2004)	1.1.1.188	6000	3.4	6000	3.4	6000	3.4	1100	3.4	6000	3.4
CR (Schomburg I., et al., 2004)	1.1.1.184	110	100	100	100	110	100	130	100	110	100
9-KPR (Schomburg I., et al., 2004)	1.1.1.189	120	90	100	90	120	90	130	90	120	90
15-PGDH (Schomburg I., et al., 2004)	1.1.1.196	1700	400	1000	400	1700	400	950	400	1700	400
12-LOX (Schomburg I., et al., 2004)	1.13.11.31	840	8	840	8	840	8	840	8	840	8
PLT:											
PHGPx (Schomburg I., et al., 2004)	1.11.1.12	550	0.83	1000	5	120	4.4	5200	0.16	6900	1.4
COX-1 (Schomburg I., et al., 2004)	1.14.99.1	6000	4.5	1000	4.5	9500	4.5	1100	4.5	9500	4.5
TXAS (Schomburg I., et al., 2004)	5.3.99.5	1599	4	1599	4	1599	4	1599	4	1599	4
PLA2 (Schomburg I., et al., 2004)	3.1.1.4	3600	2600	3600	2600	3600	2600	3600	2600	3600	2600
12-LOX (Schomburg I., et al., 2004)	1.13.11.31	840	8	840	8	840	8	840	8	840	8

Table SI-III. The decay rate of molecules used in ODEs. All the decay rate has no direct value from experiments and is derived from parameter fitting

PMN:					
Decay rate	Set 1 (1/min)	Set 2 (1/min)	Set 3 (1/min)	Set 4 (1/min)	Set 5 (1/min)
Kd <sub>15-HPETE</sub>	0.36	0.05	0.1	8.7	0.1
Kd <sub>15-HETE</sub>	0.1	0.1	6.00E-05	0.1	6.00E-05
Kd <sub>TXA</sub>	1.00E-04	0.8	0.55	2	0.55
Kd <sub>TXB</sub>	1.30E-03	0.5	2.80E-04	2	2.80E-04
Kd <sub>5-HETE</sub>	4.20E-04	0.001	3.50E-05	1.80E-04	3.50E-05
Kd <sub>LTA4</sub>	1.70E-03	0.07	0.02	8.30E-04	0.02
Kd <sub>LTB4</sub>	6.30E-04	0.01	5.50E-04	6.00E-05	5.50E-04
Kd <sub>15-LOX</sub>	8.30E-04	0	5.50E-04	1.1	5.50E-04
Kd <sub>exoAA</sub>	0.017	0.5	1.30E-03	2.5	1.30E-03
Kd <sub>AA</sub>	0.14	0.1	0.16	2.5	0.16
EC:					
Kd <sub>15-HPETE</sub>	1.10E-03	0.5	1.10E-03	0.01	1.10E-03
Kd <sub>15-HETE</sub>	0.55	0.1	0.55	0.03	0.55
Kd <sub>12-HETE</sub>	2.10E-04	0.1	2.10E-04	2.90E-03	2.10E-04
Kd <sub>PGH2</sub>	1.2	0.25	1.2	6.40E-03	1.2
Kd <sub>TXA</sub>	3	3.2	3	5	3
Kd <sub>TXB</sub>	3	3.2	3	5	3
Kd <sub>PGI2</sub>	3.8	0.003	3.8	1.90E-03	3.8
Kd <sub>PGF2</sub>	0.048	0.04	0.048	0.011	0.048
Kd <sub>PGD2</sub>	9.50E-04	0.025	9.50E-04	7.90E-05	9.50E-04
Kd <sub>PGJ2</sub>	2.40E-04	0.04	2.40E-04	0.18	2.40E-04
Kd <sub>15-LOX</sub>	1.7	0.01	1.7	5.80E-03	1.7
PLT:					
Kd <sub>TXA</sub>	2.5	2.3	2.5	2.5	2.5
Kd <sub>TXB</sub>	6.90E-05	1.00E-04	0.01	4.40E-03	0.02

Table SI-IV. Initial concentrations used in ODEs. All the initial concentration has no direct value from experiments and is derived from parameter fitting

PMN:					
Initial concentrations	Set 1 (μM)	Set 2 (μM)	Set 3 (μM)	Set 4 (μM)	Set 5 (μM)
[AA] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[15-HPETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[15-HETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[12-HPETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[12-HETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PGH2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PGE2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[TXA2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[TXB2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[5-HPETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[5-HETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[LTA4] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[LTB4] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[20-OH-LTB4] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[15-LOX] <sub>0</sub>	6.30E-03	1.5	4.00E-03	0.11	4.00E-03
[12-LOX] <sub>0</sub>	2.5	0.5	6.9	0.69	6.9
[TXAS] <sub>0</sub>	0.083	0.1	0.13	2.10E-03	0.13
[5-LOX] <sub>0</sub>	1.1	5	0.69	0.58	0.69
[LTA4H] <sub>0</sub>	0.83	0.76	1.1	0.76	1.1
[lin_PMN] <sub>0</sub>	4.4	12	2.2	17	2.2
[PLA2] <sub>0</sub>	4.8	1.5	9.1	2.5	9.1
[COX-2] <sub>0</sub>	0.63	0.8	0.012	1	0.012
[PGES] <sub>0</sub>	5.20E-03	0.5	1.9	4.00E-03	1.9
[CYP4F3] <sub>0</sub>	0.13	0.07	0.14	0.12	0.14
[PHGPx] <sub>0</sub>	0.069	0.8	5.20E-03	3.30E-03	5.20E-03
[exoAA] <sub>0</sub>	0	0	0	0	0
	30	30	30	30	30

EC:					
[AA] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[15-HPETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[15-HETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[12-HPETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[12-HETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PGH2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[TXA2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[TXB2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PGI2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[6-Keto-PGF1 $\alpha$ ] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PGD2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PGJ2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[15d-PGJ2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PGE2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PGF2 $\alpha$ ] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[15-Keto-PGF2 $\alpha$ ] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PLA2] <sub>0</sub>	0.08	0.5	0.08	0.16	0.08
[15-LOX] <sub>0</sub>	5.70E-03	1.5	5.70E-03	8	5.70E-03
[TXAS] <sub>0</sub>	7.2	0.7	7.2	0.01	7.2
[PGES] <sub>0</sub>	0.025	0.0034	0.025	3.10E-04	0.025
[lin_EC] <sub>0</sub>	5	5	5	16	5
[12-LOX] <sub>0</sub>	3.20E-04	1.5	3.20E-04	2.3	3.20E-04
[COX-2] <sub>0</sub>	0.11	3	0.11	1.4	0.11
[PGIS] <sub>0</sub>	0.35	0.5	0.35	6.00E-04	0.35
[PGDS] <sub>0</sub>	0.064	2.5	0.064	6.40E-03	0.064
[PGFS] <sub>0</sub>	1.80E-03	5	1.80E-03	0.025	1.80E-03
[CR] <sub>0</sub>	3.00E-04	6.00E-04	3.00E-04	2.00E-04	3.00E-04
[9-KPR] <sub>0</sub>	5.00E-04	6.50E-04	5.00E-04	4.00E-04	5.00E-04
[15-PGDH] <sub>0</sub>	3.50E-04	8.00E-04	3.50E-04	2.50E-04	3.50E-04
[PHGPx] <sub>0</sub>	0.57	0.05	0.57	3.20E-03	0.57
PLT:					
[AA] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[12-HPETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[12-HETE] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[PGH2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[TXA2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[TXB2] <sub>0</sub>	0.001	0.001	0.001	0.001	0.001
[TXAS] <sub>0</sub>	0.51	0.635	0.58	0.54	0.62
[lin_PLT] <sub>0</sub>	1	12	20	0.5	16.5
[PLA2] <sub>0</sub>	0.19	0.1	3.10E-03	0.15	7.50E-03
[12-LOX] <sub>0</sub>	0.82	0.5	0.039	0.44	0.084
[PHGPx] <sub>0</sub>	2.20E-03	0.5	9.20E-03	0.17	6.10E-03
[COX-1] <sub>0</sub>	0.19	0.85	0.15	0.94	0.15

#### Reference

- Aharony D., Redkar-Brown D. G., Hubbs S. J., Stein R. L. (1987) Kinetic studies on the inactivation of 5-lipoxygenase by 5(S)-hydroperoxyeicosatetraenoic acid. *Prostaglandins* 33: 85-100
- Akarasereonont P., Chotewuttakorn S., Techatrasak K., Thaworn A. (2001) The effects of COX-metabolites on cyclooxygenase-2 induction in LPS-treated endothelial cells. *J Med Assoc Thai* 84 Suppl 3: S696-709
- Balboa M. A., Perez R., Balsinde J. (2003) Amplification mechanisms of inflammation: paracrine stimulation of arachidonic acid mobilization by secreted phospholipase A2 is regulated by cytosolic phospholipase A2-derived hydroperoxyeicosatetraenoic acid. *J. Immunol.* 171: 989-994
- Cashman J. R., Lambert C., Sigal E. (1988) Inhibition of human leukocyte 5-lipoxygenase by 15-HPETE and related eicosanoids. *Biochem. Biophys. Res. Commun.* 155: 38-44
- Christmas P., Ursino S. R., Fox J. W., Soberman R. J. (1999) Expression of the CYP4F3 gene. tissue-specific splicing and alternative promoters generate high and low K(m) forms of leukotriene B(4) omega-hydroxylase. *J. Biol. Chem.* 274: 21191-21199
- Husain S., Abdel-Latif A. A. (2001) Effects of prostaglandin F(2alpha) and carbachol on MAP kinases, cytosolic phospholipase A(2) and arachidonic acid release in cat iris sphincter smooth muscle cells. *Exp Eye Res* 72: 581-590
- Jones D. A., Fitzpatrick F. A. (1990) "Suicide" inactivation of thromboxane A2 synthase. Characteristics of mechanism-based inactivation with isolated enzyme and intact platelets. *J. Biol. Chem.* 265: 20166-20171
- Jones D. A., Fitzpatrick F. A. (1991) Thromboxane A2 synthase. Modification during "suicide" inactivation. *J. Biol. Chem.* 266: 23510-23514
- Kishimoto K., Nakamura M., Suzuki H., Yoshimoto T., Yamamoto S., Takao T., Shimonishi Y., Tanabe T. (1996) Suicide inactivation of porcine leukocyte 12-lipoxygenase associated with its incorporation of 15-hydroperoxy-5,8,11,13-eicosatetraenoic acid derivative. *Biochim. Biophys. Acta.* 1300: 56-62
- Kojima F., Kato S., Kawai S. (2005) Prostaglandin E synthase in the pathophysiology of arthritis. *Fundam Clin Pharmacol* 19: 255-261
- Lepley R. A., Fitzpatrick F. A. (1994) Irreversible inactivation of 5-lipoxygenase by leukotriene A4. Characterization of product inactivation with purified enzyme and intact leukocytes. *J. Biol. Chem.* 269: 2627-2631
- Levy B. D., Clish C. B., Schmidt B., Gronert K., Serhan C. N. (2001) Lipid mediator class switching during acute inflammation: signals in resolution. *Nat. Immunol.* 2: 612-619
- Nathan C. (2002) Points of control in inflammation. *Nature* 420: 846-852
- Orning L., Gierse J., Duffin K., Bild G., Krivi G., Fitzpatrick F. A. (1992) Mechanism-based inactivation of leukotriene A4 hydrolase/aminopeptidase by leukotriene A4. Mass spectrometric and kinetic characterization. *J. Biol. Chem.* 267: 22733-22739
- Quraishi O., Mancini J. A., Riendeau D. (2002) Inhibition of inducible prostaglandin E(2) synthase by 15-deoxy-Delta(12,14)-prostaglandin J(2) and polyunsaturated fatty acids. *Biochem. Pharmacol.* 63: 1183-1189
- Schomburg I., Chang A., Ebeling C., Gremse M., Heldt C., Huhn G., Schomburg D. (2004) BRENDA, the enzyme database: updates and major new developments. *Nucleic Acids Res.* 32: D431-433
- Serio K. J., Baker J. R., Ring W. L., Riddick C. A., Bigby T. D. (1997) Leukotriene B4 costimulates 5-lipoxygenase activity in neutrophils via increased 5-lipoxygenase translocation. *Am. J. Physiol.* 272: C1329-1334
- Soberman R. J., Okita R. T., Fitzsimmons B., Rokach J., Spur B., Austen K. F. (1987) Stereochemical requirements for substrate specificity of LTB4 20-hydroxylase. *J. Biol. Chem.* 262: 12421-12427
- Tsubouchi Y., Kawahito Y., Kohno M., Inoue K., Hla T., Sano H. (2001) Feedback control of the arachidonate cascade in rheumatoid synoviocytes by 15-deoxy-Delta(12,14)-prostaglandin J2. *Biochem Biophys Res Commun* 283: 750-755
- Vanderhoek J. Y., Karmin M. T., Ekborg S. L. (1985) Endogenous hydroxyeicosatetraenoic acids stimulate the human polymorphonuclear leukocyte 15-lipoxygenase pathway. *J. Biol. Chem.* 260: 15482-15487
- Weaver J. A., Maddox J. F., Cao Y. Z., Mullarky I. K., Sordillo L. M. (2001) Increased 15-HPETE production decreases prostacyclin synthase activity during oxidant stress in aortic endothelial cells. *Free Radic Biol Med* 30: 299-308
- Wijkander J., O'Flaherty J. T., Nixon A. B., Wykle R. L. (1995) 5-Lipoxygenase products modulate the activity of the 85-kDa phospholipase A2 in human neutrophils. *J. Biol. Chem.* 270: 26543-26549

Table SII-I. Drug target search result by MTOI. The desired state was defined as a state where the cumulative output of LTB4 and PGE2 should be smaller than 10% of that in the disease state, respectively

Set 2: Rank	Enzyme	abs(MD/SD)	MD	Set 3: Enzyme	abs(MD/SD)	MD	Set 4: Enzyme	abs(MD/SD)	MD	Set 5: Enzyme	abs(MD/SD)	MD
1	PLA2	1.1092	-1.04	COX-2	1.2919	-1.08	PGES	0.7292	-0.76	COX-2	1.1868	-1.08
2	LTA4H	0.9709	-0.92	PLA2	0.9308	-1.16	LTA4H	0.7206	-0.76	PLA2	0.7961	-1
3	COX-2	0.3992	-0.44	5-LOX	0.577	-0.56	PLA2	0.6673	-0.76	PGES	0.6302	-0.68
4	5-LOX	0.3636	-0.44	PGES	0.541	-0.6	5-LOX	0.5284	-0.56	5-LOX	0.5163	-0.52
5	15-LOX	0.2394	0.28	LTA4H	0.428	-0.48	COX-2	0.4202	-0.44	PHGPx	0.4223	0.48
6	TXAS	0.2082	0.24	PHGPx	0.3431	0.4	PHGPx	0.3091	0.36	LTA4H	0.3968	-0.44
7	CYP4F3	0.1715	0.2	PGDS	0.1404	0.16	TXAS	0.1029	0.12	PGDS	0.1032	0.12
8	PGES	0.171	-0.2	TXAS	0.1058	0.12	15-LOX	0.1024	0.12	15-LOX	0.1024	0.12
9	PGDS	0.1052	0.12	15-LOX	0.1025	-0.12	CYP4F3	0.0692	0.08	CYP4F3	0.1014	0.12
10	PHGPx	0.0678	0.08	9-KPR	0.0696	0.08	PGIS	0.0349	0.04	TXAS	0.0696	0.08
11	9-KPR	0.0345	0.04	PGIS	0.0348	0.04	PGDS	0.0347	0.04	PGFS	0.0346	-0.04
12	PGIS	0.0343	-0.04	CR	0.0346	-0.04	9-KPR	0.0347	-0.04	CR	0.0346	0.04
13	12-LOX	0.0342	0.04	12-LOX	0.0345	0.04	CR	0.0346	0.04	12-LOX	0.0343	0.04
14	PGFS	0	0	COX-1	0	0	12-LOX	0.0346	-0.04	COX-1	0	0
15	COX-1	0	0	CYP4F3	0	0	COX-1	0	0	15-PGDH	0	0
16	12-PGDH	0	0	15-PGDH	0	0	15-PGDH	0	0	9-KPR	0	0
17	CR	0	0	PGFS	0	0	PGFS	0	0	PGIS	0	0

Table SII-II. Drug target search result by MTOI. The desired state was defined as a state where the cumulative output of LTB4 and PGE2 should be smaller than 20% of that in the disease state, respectively

Set 1: Rank	Enzyme	abs(MD/SD)	MD	Set 2: Enzyme	abs(MD/SD)	MD	Set 3: Enzyme	abs(MD/SD)	MD	Set 4: Enzyme	abs(MD/SD)	MD	Set 5: Enzyme	abs(MD/SD)	MD
1	PLA2	0.7123	-0.8	PLA2	0.8923	-0.96	COX-2	0.9421	-0.88	PGES	0.6489	-0.68	COX-2	1.0209	-0.92
2	PGES	0.6948	-0.72	LTA4H	0.7178	-0.68	PLA2	0.8185	-1	LTA4H	0.5744	-0.6	PLA2	0.6993	-0.88
3	LTA4H	0.6244	-0.64	COX-2	0.3739	-0.4	PGES	0.5058	-0.56	PLA2	0.5576	-0.64	PGES	0.4687	-0.52
4	COX-2	0.5711	-0.6	PGES	0.2408	-0.28	PHGPx	0.3432	0.4	5-LOX	0.41	-0.44	LTA4H	0.3164	-0.36
5	5-LOX	0.4513	-0.52	5-LOX	0.2376	-0.28	5-LOX	0.3323	-0.32	COX-2	0.3226	-0.36	5-LOX	0.2864	-0.28
6	CYP4F3	0.3482	0.4	12-LOX	0.2035	0.24	LTA4H	0.212	-0.24	PHGPx	0.2059	0.24	PHGPx	0.2748	0.32
7	PHGPx	0.3191	0.36	CYP4F3	0.1723	0.2	TXAS	0.1762	0.2	15-LOX	0.1379	0.16	TXAS	0.1398	0.16
8	TXAS	0.1431	0.16	PGDS	0.1048	0.12	PGDS	0.105	0.12	CYP4F3	0.0694	0.08	CYP4F3	0.1033	0.12
9	PGDS	0.1035	0.12	PHGPx	0.0683	0.08	15-LOX	0.07	-0.08	TXAS	0.0348	0.04	9-KPR	0.0699	0.08
10	9-KPR	0.0348	-0.04	COX-1	0	0	CYP4F3	0.0697	0.08	PGDS	0.0347	0.04	15-LOX	0.0353	-0.04
11	PGIS	0.034	0.04	15-PGDH	0	0	12-LOX	0.068	0.08	12-LOX	0.0341	0.04	PGDS	0.0348	0.04
12	15-LOX	0	0	9-KPR	0	0	9-KPR	0.0348	0.04	COX-1	0	0	12-LOX	0.0341	-0.04
13	COX-1	0	0	CR	0	0	PGIS	0	0	15-PGDH	0	0	PGFS	0	0
14	15-PGDH	0	0	PGFS	0	0	COX-1	0	0	9-KPR	0	0	COX-1	0	0
15	CR	0	0	PGIS	0	0	15-PGDH	0	0	CR	0	0	15-PGDH	0	0
16	PGFS	0	0	TXAS	0	0	CR	0	0	PGFS	0	0	CR	0	0
17	12-LOX	0	0	12-LOX	0	0	PGFS	0	0	PGIS	0	0	PGIS	0	0

Table SIII-I. The feedback parameters used in the ODEs. Italics means the corresponding parameter has no direct value from experiments and is derived from parameter fitting.

PMN:		
Feedbacks	Parameters	value
HETEs→COX2	$K_{i_{\text{HETEs} \rightarrow \text{COX2}}}$	<i>0.61</i> $\mu\text{M}$
HETEs→5-LOX	$K_{i_{\text{HETEs} \rightarrow 5\text{-LOX}}}$	<i>0.78</i> $\mu\text{M}$
LTA4→5-LOX	$K_{i_{\text{LTA4} \rightarrow 5\text{-LOX}}}$	<i>0.012</i> ( $\mu\text{M} \cdot \text{min}$ ) <sup>-1</sup>
LTA4→LTA4H	$K_{i_{\text{LTA4} \rightarrow \text{LTA4H}}}$	<i>0.016</i> ( $\mu\text{M} \cdot \text{min}$ ) <sup>-1</sup>
PGH2→TXAS	$K_{i_{\text{PGH2} \rightarrow \text{TXAS}}}$	<i>8.8</i> ( $\mu\text{M} \cdot \text{min}$ ) <sup>-1</sup>
AA→PLA2	$K_{i_{\text{AA} \rightarrow \text{PLA2}}}$	<i>480</i> $\mu\text{M}$
Product inhibition	$K_i$	<i>500</i> $\mu\text{M}$
EC:		
HETEs→PGES	$K_{i_{\text{HETEs} \rightarrow \text{PGES}}}$	<i>500</i> $\mu\text{M}$
HETEs→PGIS	$K_{i_{\text{HETEs} \rightarrow \text{PGIS}}}$	<i>0.12</i> $\mu\text{M}$
PGH2→TXAS	$K_{i_{\text{PGH2} \rightarrow \text{TXAS}}}$	<i>1.2</i> ( $\mu\text{M} \cdot \text{min}$ ) <sup>-1</sup>
AA→PLA2	$K_{i_{\text{AA} \rightarrow \text{PLA2}}}$	<i>900</i> $\mu\text{M}$
Product inhibition	$K_i$	<i>500</i> $\mu\text{M}$
PLT:		
PGH2→COX1	$K_{i_{\text{PGH2} \rightarrow \text{COX1}}}$	<i>0.013</i> ( $\mu\text{M} \cdot \text{min}$ ) <sup>-1</sup>
AA→PLA2	$K_{\text{AA} \rightarrow \text{PLA2}}$	<i>220</i> $\mu\text{M}$
Product inhibition	$K_i$	<i>500</i> $\mu\text{M}$

Table SIII-II. The  $K_m$  and  $K_{cat}$  of enzymes used in the ODEs. Italics means the corresponding parameter has no direct value from experiments and is derived from parameter fitting.

PMN:			
Enzyme Name	EC Number	$K_{cat}$ (1/min)	$K_m$ ( $\mu\text{M}$ )
E1	---	<i>290</i>	<i>28</i>
5-LOX	1.13.11.34	<i>2900</i>	<i>0.11</i>
LTA4H	3.3.2.6	<i>4200</i>	<i>20</i>

COX-2	1.14.99.1	1000	76
PGES	5.3.99.3	3000	160
TXAS	5.3.99.5	1599	4
PLA2	3.1.1.4	3600	2600
<hr/>			
EC:			
E1	---	600	58
COX-2	1.14.99.1	220	5
PGES	5.3.99.3	3000	160
TXAS	5.3.99.5	1599	4
PLA2	3.1.1.4	3600	2600
PGIS	5.3.99.4	147	9
<hr/>			
PLT:			
COX-1	1.14.99.1	1300	4.5
PLA2	3.1.1.4	3600	2600
E1	---	910	69

Table SIII-III. The decay rate of molecules used in ODEs. All the decay rate has no direct value from experiments and is derived from parameter fitting

<hr/>	
PMN:	
Decay rate	value (1/min)
$Kd_{HETES}$	0.01
$Kd_{LTB4}$	0.12
<hr/>	
EC:	
$Kd_{HETES}$	1.00E-04
$Kd_{PGH2}$	1.00E-04
$Kd_{PGE2}$	0.0066
<hr/>	
PLT:	
$Kd_{HETES}$	2.00E-05

Table SIII-IV. Initial concentrations used in ODEs. All the initial concentration has no direct value from experiments and is derived from parameter fitting

PMN:	
Initial concentrations	value ( $\mu\text{M}$ )
[HETEs] <sub>0</sub>	0.001
[PGH2] <sub>0</sub>	0.001
[PGE2] <sub>0</sub>	0.001
[TXs] <sub>0</sub>	0.001
[LTA4] <sub>0</sub>	0.001
[LTB4] <sub>0</sub>	0.001
[5-LOX] <sub>0</sub>	0.083
[LTA4H] <sub>0</sub>	0.014
[TXAS] <sub>0</sub>	2.1
[lin_PMN] <sub>0</sub>	5.5
[PLA2] <sub>0</sub>	10
[COX-2] <sub>0</sub>	0.0044
[PGES] <sub>0</sub>	0.025
[E1] <sub>0</sub>	5.2
[AA] <sub>0</sub>	0.001
	30
EC:	
[AA] <sub>0</sub>	0.001
[HETEs] <sub>0</sub>	0.001
[PGH2] <sub>0</sub>	0.001
[TXs] <sub>0</sub>	0.001
[PGI2] <sub>0</sub>	0.001
[PGE2] <sub>0</sub>	0.001
[PLA2] <sub>0</sub>	0.022
[E1] <sub>0</sub>	1.30E-04

[TXAS] <sub>0</sub>	0.0023
[PGES] <sub>0</sub>	0.067
[lin_EC] <sub>0</sub>	2.2
[COX-2] <sub>0</sub>	3.20E-06
[PGIS] <sub>0</sub>	0.195
<hr/>	
PLT:	
<hr/>	
[AA] <sub>0</sub>	0.001
[HETEs] <sub>0</sub>	0.001
[TXs] <sub>0</sub>	0.001
[lin_PLT] <sub>0</sub>	340
[PLA2] <sub>0</sub>	0.0011
[E1] <sub>0</sub>	0.72
[COX-1] <sub>0</sub>	0.88
<hr/>	

Table SIV Multi-target intervention solutions found by MTOI for the simplified AAnetwork model. “---” denotes no regulation and “√” denotes that the corresponding enzyme is inhibited.

	PLA2	COX-2	PGES	5-LOX	LTA4H	COX-1
No. 1	√	---	---	---	√	---
No. 2	√	√	√	√	---	√
No. 3	---	√	√	---	√	√
No. 4	√	√	√	---	√	√
No. 5	---	√	---	√	√	√
No. 6	√	---	---	√	√	---
No. 7	√	√	---	---	√	√
No. 8	---	√	√	√	√	√
No. 9	√	---	√	√	√	---
No. 10	√	---	√	---	√	---
No. 11	√	√	---	√	---	√
No. 12	---	√	---	---	√	√
No. 13	√	√	√	√	√	√
No. 14	√	√	---	√	√	√



Table SV. The result of single parameter sensitivity analysis. Top 30 parameters are listed in the table. Kcat is turnover number, Km is Michaelis-Menten constant, con is concentration. “K(PGE2→15-LOX)” denotes the feedback constant of PGE2 to 15-LOX.

Rank	Parameter	Sensitivity
1	Km(PLA2)	0.7393
2	Kcat(PLA2)	0.7369
3	con(PLA2)	0.7014
4	Km(PGES)	0.5656
5	Kcat(PGES)	0.552
6	con(PGES)	0.4593
7	Kcat(15-LOX)	0.3875
8	K(PGE2→15-LOX)	0.3869
9	Km(15-LOX)	0.3831
10	K(LTA4→LTA4H)	0.359
11	con(LTA4H)	0.2838
12	con(COX-2)	0.268
13	K(15-HETE→PGES)	0.2672
14	Kcat(COX)	0.261
15	Km(COX)	0.2337
16	con(PHGPx)	0.232
17	Km(PHGPx)	0.2264
18	Kcat(PHGPx)	0.2257
19	K(LTB4→PLA2)	0.1674
20	K(15HPETE→TXAS)	0.112
21	con(TXAS)	0.0843
22	K(15HPETE→PLA2)	0.0819
23	Kcat(TXAS)	0.0723
24	Km(TXAS)	0.0715
25	Kcat(PGDS)	0.0699
26	con(PGDS)	0.0693
27	Km(PGDS)	0.0693
28	Kcat(5-LOX)	0.069
29	con(5-LOX)	0.069
30	Kcat(LTA4H)	0.0657