

Supporting Information for

Quantum Dot Nanotoxicity Assessment Using the Zebrafish Embryo

Tisha C. King-Heiden^{†,1,2}, Paige N. Wiecinski¹, Andrew N. Mangham³, Kevin M. Metz^{‡,4},
Dorothy Nesbit², Joel A. Pedersen^{1,4}, Robert J. Hamers³, Warren Heideman^{1,2}, and Richard E.
Peterson^{*,1,2}

¹Molecular and Environmental Toxicology Center, ²School of Pharmacy, ³Department of
Chemistry, ⁴Environmental Chemistry and Technology Program, University of Wisconsin,
Madison WI, 53705.

*Corresponding author: School of Pharmacy, 777 Highland Ave, University of Wisconsin,
Madison WI 53705, repeterson@pharmacy.wisc.edu

Author e-mail addresses: king-hei.tish@uwlax.edu, pwiecinski@wisc.edu,
anmangham@wisc.edu, kmmetz@wisc.edu, djnesbit@wisc.edu, japedersen@soils.wisc.edu,
rjhamers@wisc.edu, wheidema@facstaff.wisc.edu

[†] Present address: Biology Department, University of Wisconsin, LaCrosse, WI

[‡] Present address: Chemistry Department, Albion College, Albion, MI

Supporting Information is 6 pages (including the cover sheet) with a description of the sources and purities of chemicals used, details regarding QD synthesis (and references), 1 Table, and 2 supplementary figures.

Experimental Section

Chemicals. Cadmium chloride hemipentahydrate (CdCl_2 , Sigma Aldrich, >98% pure), selenium tetrachloride (SeCl_4 , Fisher, >98% pure), zinc chloride (ZnCl_2 , Sigma Aldrich, >98% pure), poly-L-lysine (Sigma Aldrich, 99% pure), $\text{PEG}_{350}\text{-OCH}_3$ (Diagnostic Chemicals Ltd, >95% pure) and $\text{PEG}_{5000}\text{-COOH}$, -OCH_3 or -NH_3 (RAAP Polymere, >96% pure) were used as obtained. Stock solutions were prepared in distilled deionized water (ddH_2O ; 18 $\text{M}\Omega\text{-cm}$ resistivity).

Quantum Dot Functionalization. Mercaptopropionic acid (MPA), $\text{PEG}_{350}\text{-OCH}_3$, $\text{PEG}_{5000}\text{-COO}^-$, $\text{PEG}_{5000}\text{-OCH}_3$, or $\text{PEG}_{5000}\text{-NH}_3$ were added via a ligand exchange process (1,2). Poly-L-lysine was added to MPA ligands using a charge-stabilized hydrogen bonding polymer wrapping procedure (3). After functionalization and removal of residual solvent by evaporation, QDs were suspended in ddH_2O and filtered through a 0.45- μm syringe filter.

Calculation of Expected QD diameters (d_{calc}). The first exciton peaks of the QDs were centered between at 558 nm for the PLL-wrapped QDs and between 525 and 530 nm for PEGylated QDs, corresponding to 3.2 and 2.6 nm, respectively (4,5). X-ray photoelectron spectroscopy indicated that the ZnS did not exceed 1 nm diameter (6). Core-shell diameter was confirmed by TEM (6). The thickness of the PLL wrapping was calculated from bond lengths assuming a single PLL shell with both the MPA and PLL in fully extended conformation. Application of an empirical relationship between effective diameter and molecular mass for free PEG molecules (7) indicated that the length of attached PEG molecules was 1 nm for PEG_{350} and 4.5 nm for PEG_{5000} . Calculated diameters (d_{calc}) of individual PLL-wrapped and PEGylated QDs (based on estimated contributions of the CdSe core, ZnS shell and PLL coating/extended PEG ligands) are presented in Table 1.

SI Literature Cited

- (1) Kang, E. C.; Ogura, A.; Katoaka, K.; Nagasaki, Y. Preparation of water-soluble pegylated semiconductor nanocrystals. *Chemistry Letters*. **2004**, *33*, 840-841.
- (2) Aldana, J.; Lavelle, N.; Wang, Y.; Peng, X. Size-dependent dissociation pH of thiolate ligands from cadmium chalcogenide nanocrystals. *J Am. Chem Soc.* **2005**, *127*, 2496-2504.
- (3) Liu, Q.; Liu, X.; Zhu, Y.; Tang, D. Synthesis of stable luminescent microspheres by a simple method. *J Colloid Interface Sci.* **2007**, *307*, 563-566.
- (4) Yu, W. W.; Qu, L. H.; Guo, W. Z.; Peng, X. G. Experimental determination of the extinction coefficient of CdTe, CdSe, and CdS nanocrystals. *Chem. of Materials*. **2003**, *15*, 2854-2860.
- (5) Peng, X.; Wickham, J.; Alivisatos, A. Kinetics of II-VI and III-V colloidal semiconductor nanocrystal growth: "Focusing" of size distributions. *J Am. Chem. Soc.* **1998**, *120*, 5343-5344.
- (6) Metz, K.M.; Maghan, A.N.; Bierman, M.J.; Jin, S.; Hamers, R.J.; Pedersen, J.A. Engineered nanomaterial transformation under oxidative environmental conditions: Development of an *in vitro* biomimetic assay. *Environ. Sci. Technol.* (in review).
- (7) Sperling, R.A.; Liedl, T.; Duhr, S.; Kudera, S.; Zanella, M.; Lin, C.A.J.; Chang, W.H.; Braun, D.; Parak, W.J. Size determination of (bio)conjugated water-soluble colloidal nanoparticles: A comparison of different techniques. *J. Phys. Chem. C.* **2007**, *111*, 11552-11559

Table S1. Individual Correlations of Cd Body Burden with Toxicity Score and MT Fold Induction in Zebrafish Exposed to CdCl₂ or Quantum Dots.

	Toxicity Score			MT Fold Induction		
	<i>R</i> ²	slope	<i>p</i>	<i>R</i> ²	slope	<i>p</i>
CdCl ₂	0.90	0.5	0.04	0.99	0.189	<0.001
CdSe _{core} /ZnS _{shell} -PLL	0.68	1.9	0.50	0.02	0.024	0.12
CdSe _{core} /ZnS _{shell} -PEG ₃₅₀ -OCH ₃	0.10	0.01	0.61	0.27	0.159	0.48
CdSe _{core} /ZnS _{shell} -PEG ₅₀₀₀ -NH ₂	0.98	0.2	0.009	0.99	0.167	0.003
CdSe _{core} /ZnS _{shell} -PEG ₅₀₀₀ -COO ⁻	0.97	0.3	0.01	0.002	0.002	0.96
CdSe _{core} /ZnS _{shell} -PEG ₅₀₀₀ -OCH ₃	0.95	0.2	0.01	0.96	0.340	0.02

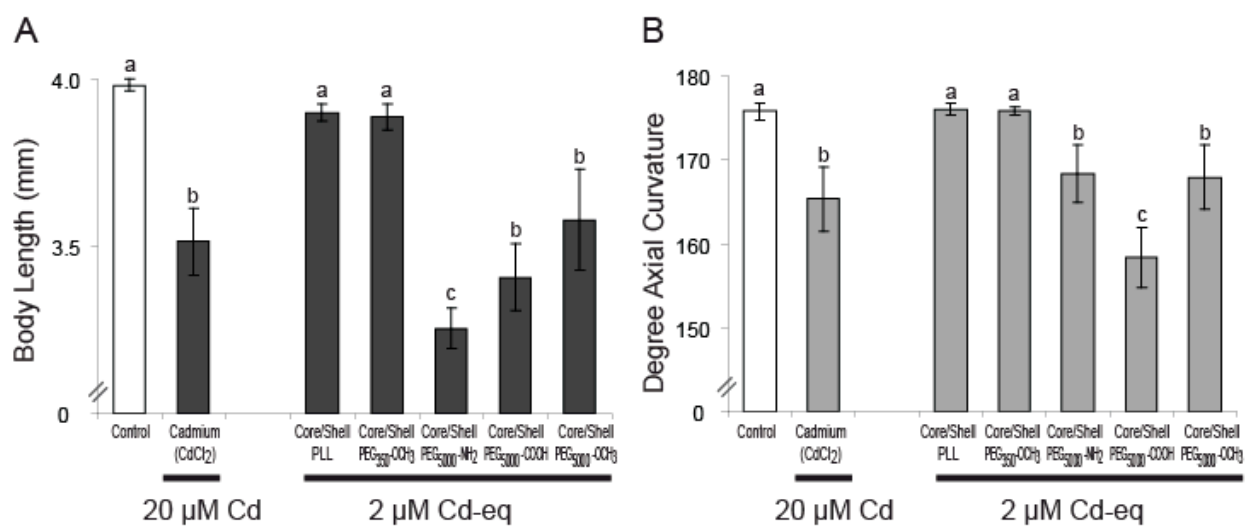


Figure S1. Exposure to an equally effective, mortality-inducing concentration (3-6% mortality) of QDs or CdCl₂ for 120 h (A) decreased zebrafish larvae growth, and (B) increased the curvature of bent spines. Values represent mean ± SE (*n* = 12); letters denote significant differences across treatments (*p* ≤ 0.05). Cores were composed of CdSe; shells were ZnS.

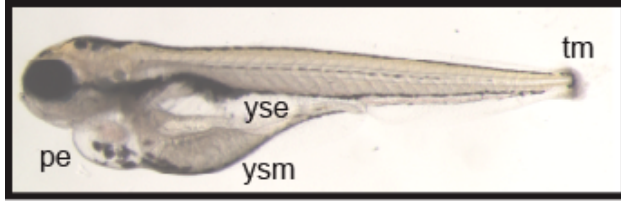


Figure S2. Representative photomicrograph demonstrating toxicity induced in zebrafish larvae by 120-h exposure to $\geq 200 \mu\text{M}$ selenium (as SeCl_4). Abbreviations: pe = pericardial edema; yse = yolk sac edema; ysm = yolk sac malformation (reduced absorption of yolk and opaque tissue indicating apparent necrosis); tm = tail fin malformation.