Supporting Information

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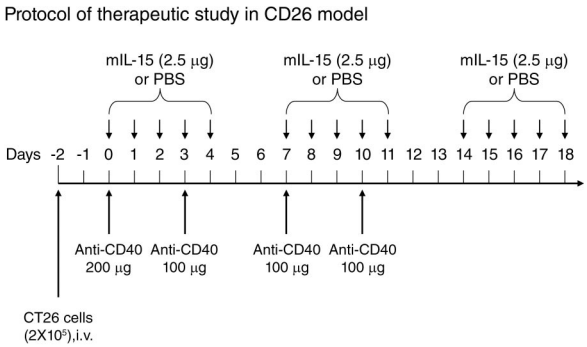

Fig. S1. Treatment schema of the therapeutic study in the CT26 model. Female BALB/c mice were injected with the CT26 cells i.v. and the thearpy started 2 days later. Groups of 10 mice each received mIL-15 i.p., 2.5 μg per mouse, 5 days a week for 3 weeks; the anti-CD40 antibody, 200 μg on day 0, then 100 μg on days 3, 7, and 10; or a combination of mIL-15 with the anti-CD40 antibody at the same doses and dosing schedule as those in mIL-15 and the anti-CD40 antibody groups. An additional group of mice that received PBS solution injections served as a control. The survival of the mice was monitored throughout the experiments.

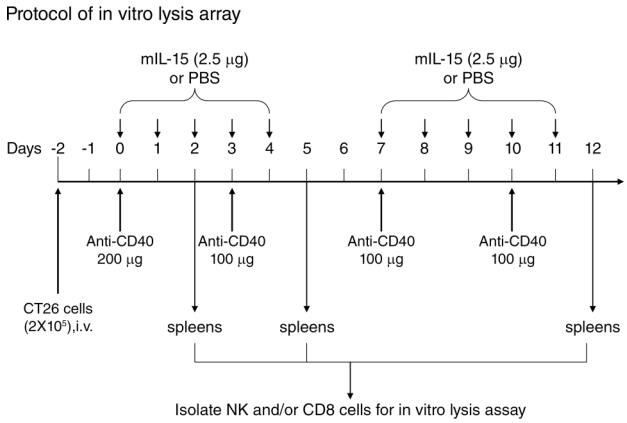

Fig. S2. Treatment schema for the in vitro lysis assay. Female BALB/c mice were injected with the CT26 cells i.v. Two days later, groups of the tumor-bearing mice received mIL-15 or the anti-CD40 antibody alone or their combination. An additional group of the mice was given PBS solution as a control. The mice were killed and the spleens were taken at days 2, 5, or 12 after therapy. NK and $CD8^+$ T cells were isolated from the spleens.

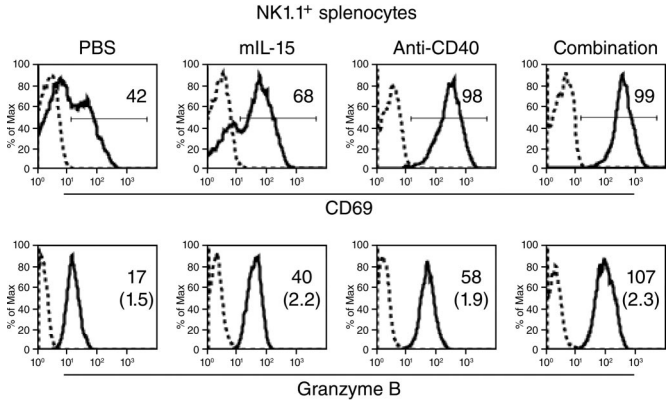

Fig. S3. Activation of NK cells was induced by treatments with mIL-15, the anti-CD40 antibody, and their combination. Surface expression of CD69 (*Upper*) and intracellular expression of granzyme B (*Lower*) on the NK1.1$^+$ population of splenocytes were analyzed by flow cytometry. C57BL/6 mice were treated with mIL-15 (2.5 μg), the anti-CD40 antibody (200 μg), or their combination, or with PBS solution as a control. The splenocytes were separated 24 h later for flow cytometric analysis. Treatment with mIL-15, anti-CD40 antibody, or their combination up-regulated the expression of CD69 (*Upper*) on NK1.1$^+$ splenocytes compared with the PBS solution control group. Treatment with the combination regimen induced the highest level of intracellular granzyme B in the NK1.1$^+$ cells compared with those from either mIL-15-treated or anti-CD40 antibody-treated mice (*Lower*). Dashed lines represent isotype controls. Numbers refer to percentages (*Upper*) and mean fluorescence intensity measurements (*Lower*). Numbers in parentheses (*Lower*) represent isotype controls. The data are representative of 3 separate experiments.