

1 **Live-cell imaging and measurement of intracellular pH in filamentous fungi using a genetically**
2 **encoded ratiometric probe**
3 **(pH imaging in fungi using a genetically encoded probe)**

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1 **Supplementary material**

2 **Materials and Methods**

3 **RaVC gene construction**

4 Genes encoding for Venus, Citrine and pHluorin were used as templates to construct RaVC. Replacing
5 some fragments of pHluorin with Citrine or Venus enabled substantial codon optimization. The primers
6 were constructed with overhangs to enable consecutive PCR ligation of the obtained PCR fragments
7 derived from Venus, Citrine and pHluorin (Supplementary Table 1). Six fragments: 1-195 bp of
8 Citrine, 196-244 bp of pHluorin, 245-396 bp of Citrine, 397-507 pHluorin, 508-606 bp of Venus, 607-
9 720 pHluorin were PCR amplified. After PCR ligations we obtained a 720 bp product named RaVC
10 (ratiometric pHluorin, Venus, Citrine) that preserved mutations which confer pHluorin with its pH
11 sensitivity and had been partially codon optimized (Supplementary Table 2 and Supplementary Fig. 2).
12 A point mutation at amino acid site 206 was introduced. GFP-A206K was constructed by site-directed
13 mutagenesis, using the QuickChange XL Site-Directed Mutagenesis Kit following the manufacturer's
14 protocol (Stratagene) using the following primers: sense 5' cctgcacacacaatctaagctttcgaaagatc and
15 antisense 5' gggatctttcgaaagcttagattgtgtgtgca.

16 The gene obtained was cloned into the pBlueScript plasmid (Stratagene) using the blunt end *EcoRV*
17 restriction site and transformed into *Escherichia coli* DH5 α for plasmid propagation. Sequencing of
18 RaVC confirmed the expected DNA sequence. The RaVC construct was inserted into the bacterial
19 expression plasmid pRSET (Invitrogen) using *Bam*HI and *Eco*RI restriction sites and was then
20 expressed in *Escherichia coli* BL21(DE3) cells.

1 **Supplementary Table 1. Primers used for PCR amplifications and PCR ligations.** Underlined
 2 bases indicate where primers anneal to template. Red marks label the start and stop codon. The green
 3 fluorescent proteins Citrine, Venus and pHluorin were used as templates.

template	primers: forward and reverse	base position	amino acid position
Citrine	GGATCCGCCACC ATG <u>GTGAGCAAGGGCGAGG</u> ACCATAAGAGAAGGTGGTCACGAGG	1-195	1-65
pHluorin	<u>GTGACCACCTTCTCTTATGGTGTCAATG</u> GAAGTCGTGCCGTTTCATATGATCTG	196-243	66-81
Citrine	CATATGAAACGGC <u>CACGACTTCTTCAAGTC</u> GTTCCATCATCCTTGAAGTCGATGCCC	244-396	82-132
pHluorin	CGACTTCAAGGATGATGGAAACATTATTCTTG <u>CTCGATGTTGTGGTGAACCTGAAAG</u>	397-507	133-169
Venus	CAAGTTCACCACAACATCGAGGACG <u>TTGTGTGTGCAGGTAGTGGTTGTCGG</u>	508-606	170-202
pHluorin	ACCCACTACCTGCACACACAATCTG GAATTC TTA <u>TTTGTATAGTTCATC</u>	607-720	203-240

1 **Supplementary Table 2. Codon usage *Aspergillus niger* and the number of codon triplets in**
 2 **pHluorin and RaVC.**

Amino acids	Codon triplet	Codon usage frequency <i>A. niger</i> [%]	pHluorin-R [amino acids number]	RaVC [amino acids number]	Amino acids	Codon triplet	Codon usage frequency <i>A. niger</i> [%]	pHluorin-R [amino acids number]	RaVC [amino acids number]
Ala	GCA	12	3	1	Thr	ACA	11	6	3
	GCC	43	2	5		ACC	50	3	13
	GCG	16				ACG	17		
	GCT	29	4	3		ACT	21	7	
Arg	CGA	8			Val	GTA	5	2	2
	CGC	36		1		GTC	45	6	3
	CGG	15	1	3		GTG	28	1	11
	CGT	27	1			GTT	22	8	2
	AGA	7	4	2	His	CAC	66	7	10
	AGG	7				CAT	34	5	2
Asn	AAC	76	7	10	Gln	CAA	24	7	4
	AAT	24	4	1		CAG	76	1	4
Lys	AAA	16	14	6	Gly	GGA	18	8	2
	AAG	84	5	13		GGC	40	4	17
Asp	GAC	58	6	13		GGG	9	3	2
	GAT	42	13	6		GGT	34	8	2
Glu	GAA	29	12	4	Leu	CTA	5	3	1
	GAG	71	4	12		CTC	29		2
Cys	TGC	67	2	2		CTG	33	1	11
	TGT	33				CTT	15	10	3
Trp	TGG	100	1	1		TTA	2	3	
Ser	TCA	6	1	1		TTG	16	2	2
	TCC	30		2	Phe	TTC	75	4	10
	TCG	16	1	1		TTT	25	10	4
	TCT	17	3	2	Pro	CCA	12	6	1
	AGC	21		2		CCC	42	2	9
	AGT	10	3			CCG	21		
Ile	ATA	4	1		CCT	25	2		
	ATC	66	3	9	Met	ATG	100	6	6
	ATT	30	7	2		STOP	TAA	39	1
Try	TAC	73	7	9	TAG	31			
	TAT	27	4	2	TGA	30			

1		1 2
2	gfp-1	M-GKGEELFTGVVPIILVELDGDVNGHKFSVSSEGEEDATYGKLTLLKFICTTGKLPVPWPT
3	pHluorin	M-SKGEELFTGVVPIILVELDGDVNGHKFSVSSEGEEDATYGKLTLLKFICTTGKLPVPWPT
4	Venus	MVSKGEELFTGVVPIILVELDGDVNGHKFSVSSEGEEDATYGKLTLLKFICTTGKLPVPWPT
5	Citrine	MVSKGEELFTGVVPIILVELDGDVNGHKFSVSSEGEEDATYGKLTLLKFICTTGKLPVPWPT
6	RaVC	MVSKGEELFTGVVPIILVELDGDVNGHKFSVSSEGEEDATYGKLTLLKFICTTGKLPVPWPT
7		60
8	gfp-1	LVTTF ^S YGVQCF ^S SRYPDHMKR ^H HDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTL
9	pHluorin	LVTTF ^S YGVQCF ^S SRYPDHMKR ^H HDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTL
10	Venus	LVTTF ^S YGLMCFARYPDHMKQ ^H HDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTL
11	Citrine	LVTTF ^S YGLMCFARYPDHMKQ ^H HDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTL
12	RaVC	LVTTF ^S YGVQCF ^S SRYPDHMKR ^H HDFFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDTL
13		120
14	gfp-1	VNRIELKGIDFKEDGNILGHKLEYNYN ^S SHNVYIMADKQKNGTKVNFKIRHNIEDGSVQLA
15	pHluorin	VNRIELKGIDFK ^D DDGNILGHKLEYN ^N NEHLVYIMADKQKNGTKAIFQVHHNIEDGGVQLA
16	Venus	VNRIELKGIDFKEDGNILGHKLEYNYN ^S SHNVYIMADKQKNGIKVNFKIRHNIEDGGVQLA
17	Citrine	VNRIELKGIDFKEDGNILGHKLEYNYN ^S SHNVYIMADKQKNGIKVNFKIRHNIEDGSVQLA
18	RaVC	VNRIELKGIDFK ^D DDGNILGHKLEYN ^N NEHLVYIMADKQKNGTKAIFQVHHNIEDGGVQLA
19		180
20	gfp-1	DHYQQNTPIGDGPVLLPDNHYLSTQSALS ^S KDPNEKRDHMLLEFVTAAGITHGMDELYK
21	pHluorin	DHYQQNTPIGDGPVLLPDNHYLHTQSALS ^S KDPNEKRDHMLLEFVTAAGITHGMDELYK
22	Venus	DHYQQNTPIGDGPVLLPDNHYLSYQSALS ^S KDPNEKRDHMLLEFVTAAGITLGMDELYK
23	Citrine	DHYQQNTPIGDGPVLLPDNHYLSYQSALS ^S KDPNEKRDHMLLEFVTAAGITLGMDELYK
24	RaVC	DHYQQNTPIGDGPVLLPDNHYLHTQSALS ^S KDPNEKRDHMLLEFVTAAGITHGMDELYK

25 **Supplementary FIG. 1. Alignment of *gfp-1*, *pHluorin*, *Venus*, *Citrine* and *RaVC*.**

26 Amino acids marked in gray were used to construct *RaVC*; yellow indicates mutations characteristic
 27 for *pHluorin*; red indicates point mutation known to prevent di- and oligo-merization of GFP.

