

Supplementary Data

Figure S1 – Tris/Tricine SDS-PAGE analysis of CB elastase digests over 48h. A total amount of 7.5 μ g CB membranes were loaded onto each lane prior to ,6h, 12h, 24h and 48h after elastase was added. The prominent elastase band at approximately 25 kDa height is labelled. The Tris/Tricine gel electrophoresis was performed after the protocol of H. Schägger [Tricine SDS-PAGE (2006). *Nat. Prot.*, 1, 16-22.]

Figure S2 – Reproducibility of three PM elastase digests in 60% methanol. **(A)** spectra of three individual digests corresponding to 25 ng of protein sample each (approx. 1 pmol BR). **(B)** PMF match to BR sequence of the displayed spectra. Minimum S/N ratio set to 20. Peaks matched within 10 ppm. If multiple peptides match to single masses, manual selection based on nLC-MS/MS data was done. **(C)** Chart showing proportions of peptides present in one, two or all of the three replicates.

Figure S3 – nLC-MALDI-MS/MS BR sequence coverage display of tryptic digests. Identified peptide locations indicated by thin bars underneath the protein sequence. **(A)** regular KR specificity defined during MASCOT™ search. **(B)** search without enzyme definition.

Figure S4 – Accuracy dependent PMF matching of 81 MS/MS verified BR AVLIST peptides to BR and two comparable-size false positive protein sequences (Q1LZA1 and P06030). **(A)** specificity considered, only AVLIST cleavages allowed. **(B)** specificity completely ignored, allowing all peptides.

Figure S1

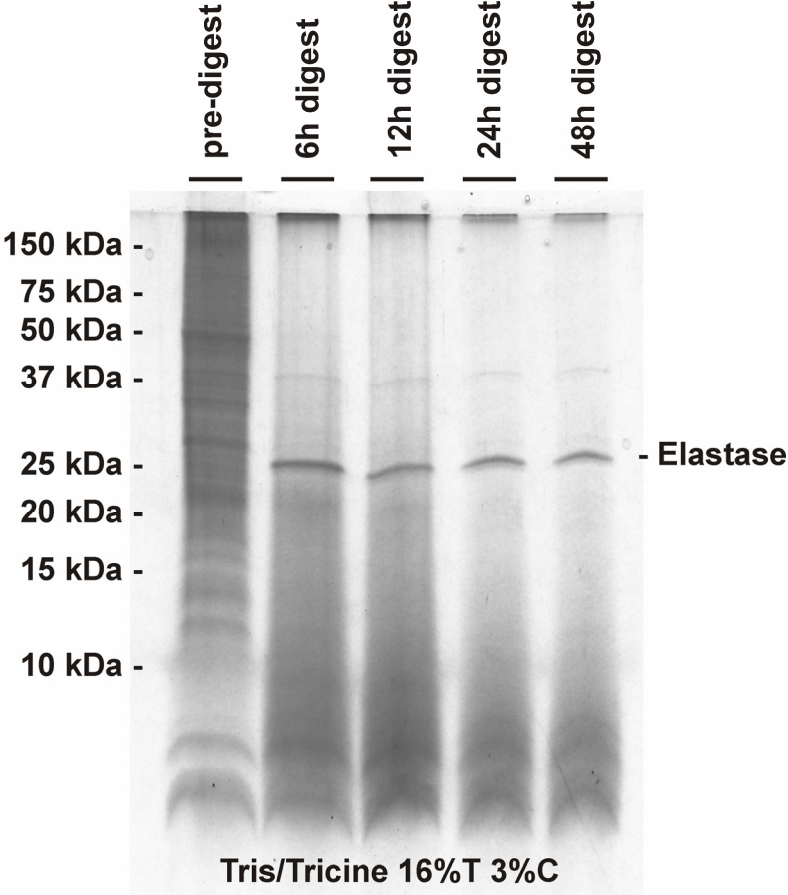
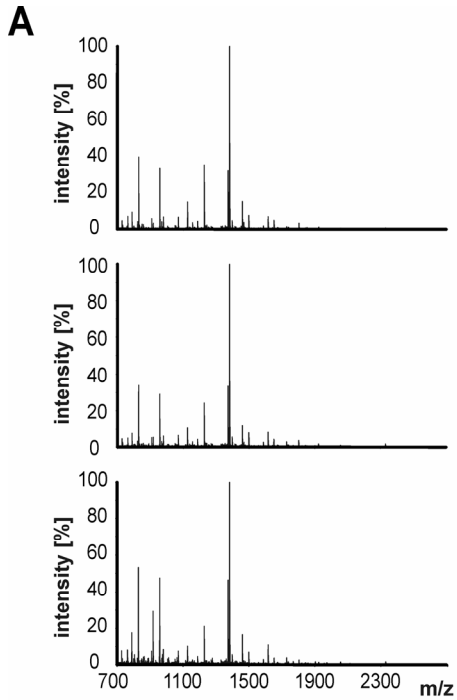


Figure S2



B

10	20	30	40	50	60
QAQITGRPEW	IWLALGTALM	GLGTLVFLVK	GMGVSDDPAK	KFYAITTLVP	AIAFTMYLSM
70	80	90	100	110	120
LLGYGLMVP	FGGEQNPIYW	ARYADWLETT	PLLLLDLALL	VDADQGTILA	LVGADGIMIG
130	140	150	160	170	180
TGLVGALTRV	YSYRFVWAI	STAMLYILY	VLFEGFTSKA	ESMRPEVAST	FKVLRNVTVV
190	200	210	220	230	240
LWSAYPVVWL	IGSEGAGIVP	LNIETLLEFMV	LDVSAKVGFG	LILLRSRAIF	GEAEAPEPSA
250					
GDGAAATSD					

10	20	30	40	50	60
QAQITGRPEW	IWLALGTALM	GLGTLVFLVK	GMGVSDDPAK	KFYAITTLVP	AIAFTMYLSM
70	80	90	100	110	120
LLGYGLMVP	FGGEQNPIYW	ARYADWLETT	PLLLLDLALL	VDADQGTILA	LVGADGIMIG
130	140	150	160	170	180
TGLVGALTRV	YSYRFVWAI	STAMLYILY	VLFEGFTSKA	ESMRPEVAST	FKVLRNVTVV
190	200	210	220	230	240
LWSAYPVVWL	IGSEGAGIVP	LNIETLLEFMV	LDVSAKVGFG	LILLRSRAIF	GEAEAPEPSA
250					
GDGAAATSD					

10	20	30	40	50	60
QAQITGRPEW	IWLALGTALM	GLGTLVFLVK	GMGVSDDPAK	KFYAITTLVP	AIAFTMYLSM
70	80	90	100	110	120
LLGYGLMVP	FGGEQNPIYW	ARYADWLETT	PLLLLDLALL	VDADQGTILA	LVGADGIMIG
130	140	150	160	170	180
TGLVGALTRV	YSYRFVWAI	STAMLYILY	VLFEGFTSKA	ESMRPEVAST	FKVLRNVTVV
190	200	210	220	230	240
LWSAYPVVWL	IGSEGAGIVP	LNIETLLEFMV	LDVSAKVGFG	LILLRSRAIF	GEAEAPEPSA
250					
GDGAAATSD					

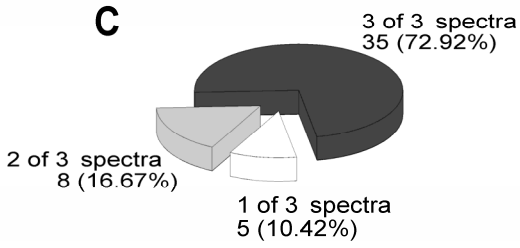


Figure S3

A

nLC-MALDI-MS/MS KR specificity

10	20	30	40	50	60
QAQITGRPEW	IWLALGTALM	GLGTYLFLVK	GMGVSDPDAK	KFYAITTLVP	AIAFTMYLSM
70	80	90	100	110	120
LLGYGLTMVP	FGGEQNPIYW	ARYADWLFTT	PLLLLDLALL	VDADQGTILA	LVGADGIMIG
130	140	150	160	170	180
TGLVGALTKV	YSYRFVWAI	STAAMLYILY	VLFFGFTSKA	ESMRPEVAST	FKVLRNVTVV
190	200	210	220	230	240
LWSAYPVVWL	IGSEGAGIVP	LNIEITLLFMV	LDVSAKVGFG	LILRSRAIF	GEAEPEPSA
250					
GDGAAATSD					

B

nLC-MALDI-MS/MS no specificity

10	20	30	40	50	60
QAQITGRPEW	IWLALGTALM	GLGTYLFLVK	GMGVSDPDAK	KFYAITTLVP	AIAFTMYLSM
70	80	90	100	110	120
LLGYGLTMVP	FGGEQNPIYW	ARYADWLFTT	PLLLLDLALL	VDADQGTILA	LVGADGIMIG
130	140	150	160	170	180
TGLVGALTKV	YSYRFVWAI	STAAMLYILY	VLFFGFTSKA	ESMRPEVAST	FKVLRNVTVV
190	200	210	220	230	240
LWSAYPVVWL	IGSEGAGIVP	LNIEITLLFMV	LDVSAKVGFG	LILRSRAIF	GEAEPEPSA
250					
GDGAAATSD					

Figure S4

