

## The HIV sequence sets

The following table shows the number of individual sequences in each gene set from LANL, Virco and both combined. It also shows the number and percentage of LANL sequences that were clade B, as well as the average length of the sequences from each LANL gene set compared to the corresponding gene length from NL4-3. The LANL sequences are available on request.

Gene	LANL (all)	Virco	LANL (+V.)	LANL (B)	LANL % B	LANL len.	NL4-3 len.
LTR	382		382	111	29%	636	634
Gag	1610		1610	748	46%	1497	1503
Pol	1567		1567	438	28%	3007	3012
Vif	2507	105	2612	1253	48%	579	579
Vpr	2319	105	2424	998	41%	291	<b>288 (291) *</b>
Tat (x1)	3262	105	3367	1500	45%	215	215
Rev (x1)	3403	105	3508	1530	44%	76	76
Vpu	2442	105	2547	886	35%	251	<b>243 (246) *</b>
Env	3560		3560	2026	57%	2574	2565
Tat (x2)	5781	83	5864	2961	50%	91	46
Rev (x2)	5469	83	5552	2771	50%	280	275
Nef	4873	83	4956	3106	63%	622	<b>618 (621) *</b>
<b>Totals :</b>	<b>37175</b>	<b>774</b>	<b>37949</b>	<b>18328</b>	<b>45%</b>		

*nb. The Virco data included a small amount of LTR sequence with the Nef data, which as used where possible, but not included in the above sequence tallies.*

*\* the native 3 nt. stop codons (the last 3 nucleotides) were excluded from the individual gene fragment sequences used in this study as it they were not included our reporters, however, the missing 3 nt. sequences were included in overlapping gene fragments.*

## The 19 mer target sets

If the complete genome for the NL4-3 strain was broken into all possible 19 mers there would be **9, 691** sequences (nb. including duplicate LTR sequences). By using individual gene sets we generated **9, 838** 19 mer sequences. This included **936** duplicate and **28** triplicate sequences from partially overlapping genes. There were also **227** sequences missing (or **2.3** % of all possible 19 mers) due to several inter-gene gaps. Notably, this included **173** sequences between the 5' LTR and Gag (617 - 789; NL4-3 co-ordinates) which covered the highly structured packaging signal **psi ( $\Psi$ )**. However, this region was covered by Naito *et. al.* (2007), using full-length genomes to estimate conservations. Excluding overlapping and LTR duplicate sequences, our pool of potential HIV-1 targets contained **8, 846 unique** 19 mer sequences.

Gene	NL4-3 19mers	LANL 19mers	Virco 19mers *	LANL + V 19mers	19:19 Comparisons
LTR	616	235913	85889	321802	198230032
Gag	1485	2381419		2381419	3536407215
Pol	2994	4684143		4684143	14024324142
Vif	561	1407036	131805	1538841	863289801
Vpr	270	633152	131805	764957	206538390
Tat (x1)	197	642158	131805	773963	152470711
Rev (x1)	58	197213	131805	329018	19083044
Vpu	225	567764	131805	699569	157403025
Env	2547	9097846		9097846	23172213762
Tat (x2)	28	422240	85889	508129	14227612
Rev (x2)	257	1431363	85889	1517252	389933764
Nef	600	2943335	85889	3029224	1817534400
<b>Totals:</b>	<b>9838</b>				<b>Total 19mer to 19 mer comparisons: 44551655898</b>
<b>Unique:</b>	<b>8846</b>				<b>Total 19 mers: 24861276</b>

\* The Virco data was generated as two groups of sequence for each sample. The first sequence covered the Vif, Vpr, Tat (exon 1), Rev (exon 1) and Vpu regions (105 sequences and 131805 19 mers), the second sequence covered the Tat (exon 2), Rev (exon 2), and Nef regions including a small amount of overlapping LTR sequence (83 sequences and 85889 19 mers). Each group was only counted once in the tally of total 19 mers analyzed (i.e. the sum of the LANL 19 mers + 131805 + 85889), rather than being counted 5 and 4 times respectively. However, all sequences in each group were compared to the corresponding NL4-3 19 mers and thus were included in the the tally of of total 19 mer : 19 mer comparisons.

## Conservation summary

**8,846** unique 19 mers from NL4-3 were compared against the 19 mers from the LANL and Virco HIV gene fragment sequence sets. The following table categorizes the conservation results, showing the number of targets and the percentage of the total 8,846 NL4-3 19 mer targets that were in each category, for both all clades (LANL and Virco) and clade B only (LANL only).

Conservation	LANL + Virco all clades		LANL subtype B	
	% of targets	# of targets	% of targets	# of targets
100%	0.0	0	0.5	44
> 90 %	0.5	40	1.2	102
> 70 %	2.8	246	13.9	1227
> 50 %	10.4	921	37.1	3286
<b>Total number of 19 mer targets:</b>		<b>8846</b>		

## The selected targets

The following table shows the **96** selected targets, along with sequence at the p -2, -1 and p +1, +2 positions, the loop sequence used, and a termination spacer base (**Tsp.**) if used. For targets that were made into a 20 bp hairpin, the last nucleotide of the loop was selected to be the complement of the last nucleotide of the p+2 position so that if the processed siRNA product(s) included the the last nucleotide of the loop then it too would be matched to the target. For targets made into a 21 bp hairpin, we used our default loop sequence (**ACTCGAGA**). The hairpins in which the last base of the anti-sense stem was 'T' also included a 'termination spacer' (**Tsp.**) so as to prevent premature termination via an early run of 'T's. This nucleotide was always the complement of the first nucleotide of the p-1 position (but never a 'T'), so that if included in the processed siRNA product(s) it was also matched to the target.

#	Target	p-2,1	Core 19 mer (p0)	p+1,2	Loop	Tsp.
1	LTR 63-20	CA	CAAGGCTACTTCCCTGATT	GG	CCTCGAGC	
2	LTR 506-20	AG	GGAACCCACTGCTTAAGCC	TC	GCTCGAGG	
3	LTR 507-20	GG	GAACCCACTGCTTAAGCCT	CA	TCTCGAGT	
4	LTR 509-20	GA	ACCCACTGCTTAAGCCTCA	AA	ACTCGAGA	
5	LTR 510-21	AA	CCCACTGCTTAAGCCTCAA	TA	ACTCGAGA	
6	LTR 516-21	CT	GCTTAAGCCTCAATAAAGC	TT	ACTCGAGA	
7	LTR 522-20	AA	GCCTCAATAAAGCTTGCCT	TG	CCTCGAGC	
8	LTR 527-21	TC	AATAAAGCTTGCCTTGAGT	GC	ACTCGAGA	G
9	LTR 553-20	AA	GTAGTGTGTGCCCGTCTGT	TG	CCTCGAGC	
10	Gag 532-20	AA	GGAGCCACCCACACAAGATT	TA	TCTCGAGT	
11	Gag 532-21	AA	GGAGCCACCCACACAAGATT	TA	ACTCGAGA	
12	Gag 533-20	AG	GAGCCACCCACACAAGATTT	AA	TCTCGAGT	
13	Gag 533-21	AG	GAGCCACCCACACAAGATTT	AA	ACTCGAGA	
14	Gag 534-20	GG	AGCCACCCACACAAGATTTA	AA	TCTCGAGT	C
15	Gag 534-21	GG	AGCCACCCACACAAGATTTA	AA	ACTCGAGA	C
16	Gag 535-20	GA	GCCACCCACACAAGATTTAA	AT	ACTCGAGA	
17	Gag 578-20	GG	GACATCAAGCAGCCATGCA	AA	TCTCGAGT	
18	Gag 698-20	GG	GAAGTGACATAGCAGGAAC	TA	TCTCGAGT	
19	Pol 242-20	AG	ATACAGGAGCAGATGATAC	AG	CCTCGAGC	C
20	Pol 246-21	AC	AGGAGCAGATGATACAGTA	TT	ACTCGAGA	G

#	Target	p-2,1	Core 19 mer (p0)	p+1,2	Loop	Tsp.
21	Pol 247-21	CA	GGAGCAGATGATACAGTAT	TA	ACTCGAGA	
22	Pol 248-20	AG	GAGCAGATGATACAGTATT	AG	CCTCGAGC	
23	Pol 248-21	AG	GAGCAGATGATACAGTATT	AG	ACTCGAGA	
24	Pol 249-20	GG	AGCAGATGATACAGTATTA	GA	TCTCGAGT	C
25	Pol 249-21	GG	AGCAGATGATACAGTATTA	GA	ACTCGAGA	C
26	Pol 250-20	GA	GCAGATGATACAGTATTAG	AA	TCTCGAGT	
27	Pol 250-21	GA	GCAGATGATACAGTATTAG	AA	ACTCGAGA	
28	Pol 251-20	AG	CAGATGATACAGTATTAGA	AG	CCTCGAGC	
29	Pol 404-20	AC	CTACACCTGTCAACATAAT	TG	CCTCGAGC	
30	Pol 529-21	TT	AAACAATGGCCATTGACAG	AA	ACTCGAGA	A
31	Pol 921-20	GG	ATGGAAGGATCACCAGCA	AT	ACTCGAGA	C
32	Pol 925-20	GG	AAAGGATCACCAGCAATAT	TC	GCTCGAGG	C
33	Pol 926-20	GA	AAGGATCACCAGCAATATT	CC	GCTCGAGG	
34	Pol 1161-20	TA	TGAACTCCATCCTGATAAA	TG	CCTCGAGC	
35	Pol 2668-21	CA	GCAGTACAAATGGCAGTAT	TC	ACTCGAGA	
36	Pol 2670-21	GC	AGTACAAATGGCAGTATTC	AT	ACTCGAGA	G
37	Pol 2671-21	CA	GTACAAATGGCAGTATTCA	TC	ACTCGAGA	
38	Pol 2724-20	GG	GTACAGTGCAGGGGAAAGA	AT	ACTCGAGA	
39	Pol 2726-21	GT	ACAGTGCAGGGGAAAGAAT	AG	ACTCGAGA	A
40	Pol 2750-20	AG	ACATAATAGCAACAGACAT	AC	GCTCGAGG	C
41	Pol 2873-21	CT	GGAAAGGTGAAGGGGCAGT	AG	ACTCGAGA	
42	Pol 2874-21	TG	GAAAGGTGAAGGGGCAGTA	GT	ACTCGAGA	
43	Pol 2878-20	AA	GGTGAAGGGGCAGTAGTAA	TA	TCTCGAGT	
44	Pol 2878-21	AA	GGTGAAGGGGCAGTAGTAA	TA	ACTCGAGA	
45	Pol 2879-20	AG	GTGAAGGGGCAGTAGTAAT	AC	GCTCGAGG	
46	Pol 2879-21	AG	GTGAAGGGGCAGTAGTAAT	AC	ACTCGAGA	
47	Pol 2880-20	GG	TGAAGGGGCAGTAGTAATA	CA	TCTCGAGT	
48	Pol 2881-21	GT	GAAGGGGCAGTAGTAATAC	AA	ACTCGAGA	
49	Pol 2882-21	TG	AAGGGGCAGTAGTAATACA	AG	ACTCGAGA	C
50	Pol 2885-20	AG	GGGCAGTAGTAATACAAGA	TA	TCTCGAGT	
51	Vif 9-21	AA	CAGATGGCAGGTGATGATT	GT	ACTCGAGA	
52	Vif 10-21	AC	AGATGGCAGGTGATGATTG	TG	ACTCGAGA	G
53	Vif 11-21	CA	GATGGCAGGTGATGATTGT	GT	ACTCGAGA	

#	Target	p-2,1	Core 19 mer (p0)	p+1,2	Loop	Tsp.
54	Vif 12-20	AG	ATGGCAGGTGATGATTGTG	TG	CCTCGAGC	C
55	Vif 14-21	AT	GGCAGGTGATGATTGTGTG	GC	ACTCGAGA	
56	Vif 15-21	TG	GCAGGTGATGATTGTGTGG	CA	ACTCGAGA	
57	Vif 37-20	AA	GTAGACAGGATGAGGATTA	AC	GCTCGAGG	
58	Tat (x1) 131-20	AG	GCATCTCCTATGGCAGGAA	GA	TCTCGAGT	
59	Tat (x1) 132-21	GG	CATCTCCTATGGCAGGAAG	AA	ACTCGAGA	
60	Tat (x1) 140-21	CT	ATGGCAGGAAGAAGCGGAG	AC	ACTCGAGA	A
61	Tat (x1) 143-21	TG	GCAGGAAGAAGCGGAGACA	GC	ACTCGAGA	
62	Tat (x1) 147-20	AG	GAAGAAGCGGAGACAGCGA	CG	CCTCGAGC	
63	Rev (x1) 9-20	GG	AAGAAGCGGAGACAGCGAC	GA	TCTCGAGT	C
64	Vpu 139-21	TA	GAAAGAGCAGAAGACAGTG	GC	ACTCGAGA	
65	Vpu 140-20	AG	AAAGAGCAGAAGACAGTGG	CA	TCTCGAGT	C
66	Vpu 140-21	AG	AAAGAGCAGAAGACAGTGG	CA	ACTCGAGA	C
67	Vpu 141-20	GA	AAGAGCAGAAGACAGTGGC	AA	TCTCGAGT	
68	Vpu 143-20	AA	GAGCAGAAGACAGTGGCAA	TG	CCTCGAGC	
69	Vpu 144-20	AG	AGCAGAAGACAGTGGCAAT	GA	TCTCGAGT	C
70	Vpu 144-21	AG	AGCAGAAGACAGTGGCAAT	GA	ACTCGAGA	C
71	Vpu 145-20	GA	GCAGAAGACAGTGGCAATG	AG	CCTCGAGC	
72	Vpu 146-20	AG	CAGAAGACAGTGGCAATGA	GA	TCTCGAGT	
73	Vpu 147-21	GC	AGAAGACAGTGGCAATGAG	AG	ACTCGAGA	G
74	Vpu 148-21	CA	GAAGACAGTGGCAATGAGA	GT	ACTCGAGA	
75	Vpu 149-20	AG	AAGACAGTGGCAATGAGAG	TG	CCTCGAGC	C
76	Env 628-20	AG	CCAATTCCCATACATTATT	GT	ACTCGAGA	
77	Env 1423-20	GG	GACAATTGGAGAAGTGAAT	TA	TCTCGAGT	
78	Env 1423-21	GG	GACAATTGGAGAAGTGAAT	TA	ACTCGAGA	
79	Env 1424-21	GG	ACAATTGGAGAAGTGAATT	AT	ACTCGAGA	C
80	Env 1425-20	GA	CAATTGGAGAAGTGAATTA	TA	TCTCGAGT	
81	Env 1425-21	GA	CAATTGGAGAAGTGAATTA	TA	ACTCGAGA	
82	Env 1426-21	AC	AATTGGAGAAGTGAATTAT	AT	ACTCGAGA	G
83	Env 1428-21	AA	TTGGAGAAGTGAATTATAT	AA	ACTCGAGA	
84	Env 1431-21	TG	GAGAAGTGAATTATATAAA	TA	ACTCGAGA	
85	Env 1432-20	GG	AGAAGTGAATTATATAAAT	AT	ACTCGAGA	C
86	Env 1432-21	GG	AGAAGTGAATTATATAAAT	AT	ACTCGAGA	C

#	Target	p-2,1	Core 19 mer (p0)	p+1,2	Loop	Tsp.
87	Env 1433-20	GA	GAAGTGAATTATATAAATA	TA	TCTCGAGT	
88	Env 1433-21	GA	GAAGTGAATTATATAAATA	TA	ACTCGAGA	
89	Env 1434-20	AG	AAGTGAATTATATAAATAT	AA	TCTCGAGT	C
90	Env 1436-20	AA	GTGAATTATATAAATATAA	AG	CCTCGAGC	
91	Env 1437-20	AG	TGAATTATATAAATATAAA	GT	ACTCGAGA	
92	Env 2308-20	TG	AGAGACTTACTCTTGATTG	TA	TCTCGAGT	C
93	Rev (x2) 158-20	CT	TGAGAGACTTACTCTTGAT	TG	CCTCGAGC	
94	Rev (x2) 159-20	TT	GAGAGACTTACTCTTGATT	GT	ACTCGAGA	
95	Nef 350-20	CA	CACAAGGCTACTTCCCTGA	TT	ACTCGAGA	
96	Nef 351-20	AC	ACAAGGCTACTTCCCTGAT	TG	CCTCGAGC	G

*n.b.* The target positions are the co-ordinates within each gene of NL4-3.

## The 22 regions

The following table shows the regions # (**R**), the shRNA # (**hp #**), the number of shRNA within each region (**T**), the starting co-ordinates of the first and last target within each region (and the specific gene product area if applicable), whether or not the region was unique amongst all previously made (**U14**: 14 regions were unique) or previously proposed (**U11**: 11 regions were unique) targets (see **Additional file 1** and Naito *et. at.* (2007)), and the sequence of the region spanning the first nucleotide of the first p0 19 mer target core to the last nucleotide of the last p0 19 mer target core within each region. n.b. the common regions cover the overlapping 19 nt. cores only, i.e. some of the first nucleotides of the p-2 and p-1 positions, and some of the final nucleotides of the p+1 and p+2 positions are not covered (see the individual core data after for these).

R	hp #	T	Target	U14	U11	Sequence region
1	1, 95-96	3	LTR 63 (U3) / Nef 350-52	U	U	CACAAGGCTACTTCCCTGATTG GGAACCCACTGCTTAAGCCTCAATAAAGCTTGCCCTT- GAGTGC
2	2-8.	7	LTR 506-27 (R)			
3	9	1	LTR 553 (U5)	U		GTAGTGTGTGCCCGTCTGTT
4	10-16.	7	Gag 532-35 (P24: CA)	U	U	GGAGCCACCCCACAAGATTTAAA
5	17	1	Gag 578 (P24: CA)	U	U	GACATCAAGCAGCCATGCAA
6	18	1	Gag 698 (P24: CA)	U	U	GAAGTGACATAGCAGGAACT
7	19-28	10	Pol 242-51			ATACAGGAGCAGATGATACAGTATTAGAA
8	29	1	Pol 404	U		CTACACCTGTCAACATAATT
9	30	1	Pol 529 (P51: RT)	U		AAACAATGGCCATTGACAGAA
10	31-33	3	Pol 921-26 (P51: RT)			ATGGAAAGGATCACCAGCAATATTC
11	34	1	Pol 1161 (P51: RT)	U	U	TGAACTCCATCCTGATAAAT
12	35-37	3	Pol 2668-71 (P31: IN)			GCAGTACAAATGGCAGTATTCATC
13	38-39	2	Pol 2724-26 (P31: IN)			GTACAGTGCAGGGGAAAGAATAG
14	40	1	Pol 2750 (P31: IN)	U	U	ACATAATAGCAACAGACATA
15	41-50	10	Pol 2873-85 (P31: IN)			GGAAAGGTGAAGGGGCAGTAGTAATACAAGAT
16	51-56	6	Vif 9-15 (P31: IN, Vif)	U	U	CAGATGGCAGGTGATGATTGTGTGGCA
17	57	1	Vif 37 (P31: IN, Vif-xj)	U	U	GTAGACAGGATGAGGATTAA
18	58-63	6	Tat (x1) 131-43 / Rev (x1) 8-9	U	U	GCATCTCCTATGGCAGGAAGAAGCGGAGACAG- CGACG
19	64-77	12	Vpu 139-49			GAAAGAGCAGAAGACAGTGGCAATGAGAGT
20	76	1	Env 628 (GP120)	U	U	CCAATCCCATACATTATTG



R	hp #	T	Target	U14	U11	Sequence region
21	77-91	15	Env 1423-37 (GP120)			GACAATTGGAGAAGTGAATTATATAAATATAAAG
22	92-94	4	Env 2308 (GP41) / Rev (x2) 158-60	U	U	TGAGAGACTTACTCTTGATTGTGT

*n.b.* CA: Capsid, RT: Reverse Transcriptase, IN: Integrase, x1: exon1, x2: exon2, xj: crosses and ORF junction.

U14: These are the 14 regions that are unique amongst all previously made and published si- and shRNAs.

U11: These are the 11 regions that are unique amongst all previously proposed targets.

## Matched target pairs

The 96 hairpins were comprised of 53 hairpins with 20 bp stems, and 43 hairpins with 21 bp stems, including **14** matched pairs of hairpins with 20 and 21 bp stems designed to the same target site. The pairs are listed in the following with the 3 replicate fluorescence values (as a % of the control) from the fluorescent reporter assays used for statistical analysis shown.

Pair	First shRNA		Replicate % F.			Second shRNA		Replicate % F.		
	#	Target	r1	r2	r3	#	Target	r1	r2	r3
<b>1</b>	10	Gag 532-20	57.9	52.1	59.9	11	Gag 532-21	41.3	36.2	51.4
<b>2</b>	12	Gag 533-20	30.7	23.8	29.4	13	Gag 533-21	31.6	27.9	39.1
<b>3</b>	14	Gag 534-20	46.5	36.8	40.1	15	Gag 534-21	43.7	41.9	49.1
<b>4</b>	22	Pol 248-20	9.0	7.8	9.7	23	Pol 248-21	9.1	7.9	9.1
<b>5</b>	24	Pol 249-20	10.1	7.9	9.5	25	Pol 249-21	9.0	8.0	9.1
<b>6</b>	26	Pol 250-20	11.2	8.6	9.6	27	Pol 250-21	10.3	8.6	10.3
<b>7</b>	43	Pol 2878-20	7.9	8.0	7.2	44	Pol 2878-21	8.6	7.2	8.2
<b>8</b>	45	Pol 2879-20	8.4	8.1	7.9	46	Pol 2879-21	99.6	95.6	93.4
<b>9</b>	65	Vpu 140-20	34.2	27.1	30.4	66	Vpu 140-21	27.2	23.0	23.9
<b>10</b>	69	Vpu 144-20	7.4	5.8	7.0	70	Vpu 144-21	6.5	5.6	5.8
<b>11</b>	77	Env 1423-20	6.1	5.8	6.4	78	Env 1423-21	8.1	7.4	9.5
<b>12</b>	80	Env 1425-20	12.3	10.4	11.4	81	Env 1425-21	7.5	6.8	7.3
<b>13</b>	85	Env 1432-20	12.7	11.8	11.4	86	Env 1432-21	7.2	6.6	7.9
<b>14</b>	87	Env 1433-20	19.3	17.2	18.1	88	Env 1433-21	19.0	18.4	19.3

## Overlapping target pairs

The **19** regions of overlapping hairpins comprised **73** adjacent pairs of hairpins where the starting position of the targets differed by 1 nt. **Forty two** of these had the same stem length (either both 20 or both 21 bp), **31** had different stem lengths. The pairs are listed in the following 2 tables grouped according to pairs with the same, and pairs with different stem lengths. The 3 replicate fluorescence values (as a % of the control) from the fluorescent reporter assays used for statistical analysis are shown.

### The 42 pairs with matched stem lengths

Pair	First shRNA		Replicate % F.			Second shRNA		Replicate % F.		
	#	Target	r1	r2	r3	#	Target	r1	r2	r3
1	2	Ltr 506-20	66.7	82.4	68.6	3	Ltr 507-20	73.6	85.9	77.5
2	10	Gag 532-20	57.86	52.08	59.92	12	Gag 533-20	30.7	23.8	29.4
3	11	Gag 532-21	41.3	36.2	51.4	13	Gag 533-21	31.6	27.9	39.1
4	12	Gag 533-20	30.69	23.82	29.39	14	Gag 534-20	46.5	36.8	40.1
5	13	Gag 533-21	31.6	27.9	39.1	15	Gag 534-21	43.7	41.9	49.1
6	14	Gag 534-20	46.5	36.8	40.1	16	Gag 535-20	42.7	29.6	34.8
7	20	Pol 0246-21	9.1	7.6	8.5	21	Pol 0247-21	10.0	9.3	10.6
8	21	Pol 0247-21	10.0	9.3	10.6	23	Pol 0248-21	9.1	7.9	9.1
9	22	Pol 0248-20	9.0	7.8	9.7	24	Pol 0249-20	10.1	7.9	9.5
10	23	Pol 0248-21	9.1	7.9	9.1	25	Pol 0249-21	9.0	8.0	9.1
11	24	Pol 0249-20	10.1	7.9	9.5	26	Pol 0250-20	11.2	8.6	9.6
12	25	Pol 0249-21	9.0	8.0	9.1	27	Pol 0250-21	10.3	8.6	10.3
13	26	Pol 0250-20	11.2	8.6	9.6	28	Pol 0251-20	119	120.9	135.2
14	32	Pol 0925-20	14.1	15.0	14.6	33	Pol 0926-20	17.3	23.3	21.6
15	36	Pol 2670-21	9.9	9.3	9.4	37	Pol 2671-21	10.6	9.3	10.0
16	41	Pol 2873-21	38.3	40.4	39.5	42	Pol 2874-21	23.2	19.9	18.2
17	43	Pol 2878-20	7.9	8.0	7.2	45	Pol 2879-20	8.4	8.1	7.9
18	44	Pol 2878-21	8.6	7.2	8.2	46	Pol 2879-21	99.6	95.6	93.4
19	45	Pol 2879-20	8.4	8.1	7.9	47	Pol 2880-20	29.9	30.9	27.7
20	48	Pol 2881-21	12.2	12.4	11.8	49	Pol 2882-21	9.6	8.6	8.3
21	51	Vif 9-21	16.6	16.9	14.5	52	Vif 10-21	13.9	12.8	12.0
22	52	Vif 10-21	13.9	12.8	12.0	53	Vif 11-21	14.2	14.3	13.5

Pair	First shRNA		Replicate % F.			Second shRNA		Replicate % F.		
	#	Target	r1	r2	r3	#	Target	r1	r2	r3
23	55	Vif 14-21	16.8	17.5	15.8	56	Vif 15-21	11.6	11.1	10.5
24	62	Revx1 8-20	18.7	16.4	14.9	63	Revx1 9-20	15.9	14.6	14.4
25	64	Vpu 139-21	10.4	7.5	9.2	66	Vpu 140-21	27.2	23.0	23.9
26	65	Vpu 140-20	34.2	27.1	30.4	67	Vpu 141-20	7.1	5.7	6.8
27	68	Vpu 143-20	7.0	6.1	6.9	69	Vpu 144-20	7.4	5.8	7.0
28	69	Vpu 144-20	7.4	5.8	7.0	71	Vpu 145-20	7.6	5.8	7.1
29	71	Vpu 145-20	7.6	5.8	7.1	72	Vpu 146-20	8.6	6.1	7.3
30	73	Vpu 147-21	6.5	5.7	6.4	74	Vpu 148-21	6.7	5.3	6.0
31	78	Env 1423-21	8.1	7.4	9.5	79	Env 1424-21	6.5	6.1	7.0
32	79	Env 1424-21	6.5	6.1	7.0	81	Env 1425-21	7.5	6.8	7.3
33	81	Env 1425-21	7.5	6.8	7.3	82	Env 1426-21	7.6	6.3	7.0
34	84	Env 1431-21	32.2	22.9	29.2	86	Env 1432-21	7.2	6.6	7.9
35	85	Env 1432-20	12.7	11.8	11.4	87	Env 1433-20	19.3	17.2	18.1
36	86	Env 1432-21	7.2	6.6	7.9	88	Env 1433-21	19.0	18.4	19.3
37	87	Env 1433-20	19.3	17.2	18.1	89	Env 1434-20	20.9	18.6	20.3
38	90	Env 1436-20	44.0	41.3	41.4	91	Env 1437-20	64.9	62.3	61.9
39	93	Revx2 158-20	7.8	7.7	8.4	94	Revx2 159-20	11.3	9.9	12.7
40	94	Revx2 159-20	11.3	9.9	12.7	92	Env 2308-20	20.3	17.1	18.8
41	95	Nef 350-20	29.6	25.5	28.6	96	Nef 351-20	37.7	38.9	38.6
42	96	Nef 351-20	37.7	38.9	38.6	1	Ltr 63-20	57.2	57.5	54.2

### The 31 pairs with mismatched stem lengths

Pair	First shRNA		Replicate % F.			Second shRNA		Replicate % F.		
	#	Target	r1	r2	r3	#	Target	r1	r2	r3
1	4	Ltr 509-20	39.3	35.6	32.7	5	Ltr 510-21	17.3	17.7	21.5
2	10	Gag 532-20	57.86	52.08	59.92	13	Gag 533-21	31.6	27.9	39.1
3	11	Gag 532-21	41.3	36.2	51.4	12	Gag 533-20	30.7	23.8	29.4
4	12	Gag 533-20	30.7	23.8	29.4	15	Gag 534-21	43.7	41.9	49.1
5	13	Gag 533-21	31.6	27.9	39.1	14	Gag 534-20	46.5	36.8	40.1
6	15	Gag 534-21	43.7	41.9	49.1	16	Gag 535-20	42.7	29.6	34.8
7	21	Pol 0247-21	10.0	9.3	10.6	22	Pol 0248-20	9.0	7.8	9.7
8	22	Pol 0248-20	9.0	7.8	9.7	25	Pol 0249-21	9.0	8.0	9.1

Pair	First shRNA		Replicate % F.			Second shRNA		Replicate % F.		
	#	Target	r1	r2	r3	#	Target	r1	r2	r3
9	23	Pol 0248-21	9.1	7.9	9.1	24	Pol 0249-20	10.1	7.9	9.5
10	24	Pol 0249-20	10.1	7.9	9.5	27	Pol 0250-21	10.3	8.6	10.3
11	25	Pol 0249-21	9.0	8.0	9.1	26	Pol 0250-20	11.2	8.6	9.6
12	27	Pol 0250-21	10.3	8.6	10.3	28	Pol 0251-20	119	120.9	135.2
13	43	Pol 2878-20	7.9	8.0	7.2	46	Pol 2879-21	99.6	95.6	93.4
14	44	Pol 2878-21	8.6	7.2	8.2	45	Pol 2879-20	8.4	8.1	7.9
15	46	Pol 2879-21	99.6	95.6	93.4	47	Pol 2880-20	29.9	30.9	27.7
16	47	Pol 2880-20	29.9	30.9	27.7	48	Pol 2881-21	12.2	12.4	11.8
17	53	Vif 11-21	14.2	14.3	13.5	54	Vif 12-20	20.5	25.3	22.8
18	58	Tatx1 131-20	28.8	24.5	25.2	59	Tatx1 132-21	28.2	27.8	24.4
19	64	Vpu 139-21	10.4	7.5	9.2	65	Vpu 140-20	34.2	27.1	30.4
20	66	Vpu 140-21	27.2	23.0	23.9	67	Vpu 141-20	7.1	5.7	6.8
21	68	Vpu 143-20	7.0	6.1	6.9	70	Vpu 144-21	6.5	5.6	5.8
22	70	Vpu 144-21	6.5	5.6	5.8	71	Vpu 145-20	7.6	5.8	7.1
23	72	Vpu 146-20	8.6	6.1	7.3	73	Vpu 147-21	6.5	5.7	6.4
24	74	Vpu 148-21	6.7	5.3	6.0	75	Vpu 149-20	8.9	6.8	7.6
25	77	Env 1423-20	6.1	5.8	6.4	79	Env 1424-21	6.5	6.1	7.0
26	79	Env 1424-21	6.5	6.1	7.0	80	Env 1425-20	12.3	10.4	11.4
27	80	Env 1425-20	12.3	10.4	11.4	82	Env 1426-21	7.6	6.3	7.0
28	84	Env 1431-21	32.2	22.9	29.2	85	Env 1432-20	12.7	11.8	11.4
29	85	Env 1432-20	12.7	11.8	11.4	88	Env 1433-21	19.0	18.4	19.3
30	86	Env 1432-21	7.2	6.6	7.9	87	Env 1433-20	19.3	17.2	18.1
31	88	Env 1433-21	19.0	18.4	19.3	89	Env 1434-20	20.9	18.6	20.3

## Detailed conservation profiles

The following table shows the detailed conservation profiles for the **96** selected targets. Conservations were calculated for LANL and Virco sequences combined, LANL clade B subtypes only, and the Virco sequences only (where applicable). In each group, 6 conservations are given: the conservation for the entire overlapping 23 mer region, and the 5 individual conservations for the 5 overlapping 19 mers that make up the 23 mer profile region. Some of shRNAs were matched to more than 1 gene, however, conservations were only estimated using a single gene data set (corresponding to the shRNA / target name), e.g. hairpin cores 1, 95, and 96 were slightly higher conserved in the Nef data set rather than the LTR data set (LTR conservations shown).

#	Target	LANL + Virco (all)						LANL clade B						Virco (all)					
		23	-2	-1	p0	+1	+2	23	-2	-1	p0	+1	+2	23	-2	-1	p0	+1	+2
1	LTR 63-20	32	34	32	<b>33</b>	33	33	73	78	75	<b>76</b>	75	75	39	44	39	<b>40</b>	40	40
2	LTR 506-20	50	51	64	<b>67</b>	69	69	97	97	97	<b>97</b>	98	100	-	-	-	-	-	-
3	LTR 507-20	64	64	67	<b>69</b>	69	69	97	97	97	<b>98</b>	100	100	-	-	-	-	-	-
4	LTR 509-20	69	69	69	<b>69</b>	70	71	98	98	100	<b>100</b>	100	100	-	-	-	-	-	-
5	LTR 510-21	69	69	69	<b>70</b>	71	71	100	100	100	<b>100</b>	100	100	-	-	-	-	-	-
6	LTR 516-21	71	71	71	<b>71</b>	71	71	100	100	100	<b>100</b>	100	100	-	-	-	-	-	-
7	LTR 522-20	90	93	91	<b>92</b>	92	93	100	100	100	<b>100</b>	100	100	-	-	-	-	-	-
8	LTR 527-21	90	93	93	<b>93</b>	93	91	100	100	100	<b>100</b>	100	100	-	-	-	-	-	-
9	LTR 553-20	48	51	52	<b>51</b>	50	50	90	94	92	<b>92</b>	92	90	-	-	-	-	-	-
10	Gag 532-20	58	64	62	<b>70</b>	70	69	77	79	78	<b>80</b>	80	80	-	-	-	-	-	-
11	Gag 532-21	58	64	62	<b>70</b>	70	69	77	79	78	<b>80</b>	80	80	-	-	-	-	-	-
12	Gag 533-20	58	62	70	<b>70</b>	69	71	77	78	80	<b>80</b>	80	82	-	-	-	-	-	-
13	Gag 533-21	58	62	70	<b>70</b>	69	71	77	78	80	<b>80</b>	80	82	-	-	-	-	-	-
14	Gag 534-20	68	70	70	<b>69</b>	71	71	79	80	80	<b>80</b>	82	82	-	-	-	-	-	-
15	Gag 534-21	68	70	70	<b>69</b>	71	71	79	80	80	<b>80</b>	82	82	-	-	-	-	-	-
16	Gag 535-20	16	70	69	<b>71</b>	71	16	6	80	80	<b>82</b>	82	6	-	-	-	-	-	-
17	Gag 578-20	55	62	63	<b>63</b>	57	58	72	78	79	<b>79</b>	73	73	-	-	-	-	-	-
18	Gag 698-20	67	74	71	<b>70</b>	68	69	74	78	78	<b>76</b>	75	75	-	-	-	-	-	-
19	Pol 242-20	57	58	58	<b>58</b>	57	83	76	77	77	<b>77</b>	77	84	-	-	-	-	-	-
20	Pol 246-21	76	83	83	<b>80</b>	80	80	78	84	84	<b>81</b>	87	87	-	-	-	-	-	-
21	Pol 247-21	75	83	80	<b>80</b>	80	79	77	84	81	<b>87</b>	87	86	-	-	-	-	-	-

#	Target	LANL + Virco (all)					LANL clade B					Virco (all)							
		23	-2	-1	p0	+1	+2	23	-2	-1	p0	+1	+2	23	-2	-1	p0	+1	+2
22	Pol 248-20	75	80	80	80	79	80	77	81	87	87	86	87	-	-	-	-	-	-
23	Pol 248-21	75	80	80	80	79	80	77	81	87	87	86	87	-	-	-	-	-	-
24	Pol 249-20	78	80	80	79	80	80	85	87	87	86	87	87	-	-	-	-	-	-
25	Pol 249-21	78	80	80	79	80	80	85	87	87	86	87	87	-	-	-	-	-	-
26	Pol 250-20	77	80	79	80	80	79	85	87	86	87	87	87	-	-	-	-	-	-
27	Pol 250-21	77	80	79	80	80	79	85	87	86	87	87	87	-	-	-	-	-	-
28	Pol 251-20	75	79	80	80	79	78	85	86	87	87	87	86	-	-	-	-	-	-
29	Pol 404-20	57	58	67	67	67	70	64	64	65	65	65	68	-	-	-	-	-	-
30	Pol 529-21	62	64	64	68	68	66	78	80	80	84	84	81	-	-	-	-	-	-
31	Pol 921-20	58	60	60	58	61	61	75	78	78	76	79	78	-	-	-	-	-	-
32	Pol 925-20	50	61	60	60	60	51	72	78	77	77	77	73	-	-	-	-	-	-
33	Pol 926-20	50	60	60	60	51	53	72	77	77	77	73	74	-	-	-	-	-	-
34	Pol 1161-20	32	36	34	32	32	32	75	77	77	75	75	76	-	-	-	-	-	-
35	Pol 2668-21	72	82	79	78	78	73	73	87	85	84	84	74	-	-	-	-	-	-
36	Pol 2670-21	72	78	78	73	73	74	73	84	84	74	74	74	-	-	-	-	-	-
37	Pol 2671-21	22	78	73	73	74	22	57	84	74	74	74	57	-	-	-	-	-	-
38	Pol 2724-20	69	70	71	71	81	81	62	63	63	67	86	86	-	-	-	-	-	-
39	Pol 2726-21	14	71	81	81	79	18	41	67	86	86	82	50	-	-	-	-	-	-
40	Pol 2750-20	65	66	65	65	65	70	74	75	75	75	74	77	-	-	-	-	-	-
41	Pol 2873-21	77	82	88	88	85	84	80	84	95	95	93	91	-	-	-	-	-	-
42	Pol 2874-21	84	88	88	85	84	84	91	95	95	93	91	91	-	-	-	-	-	-
43	Pol 2878-20	74	84	76	74	74	74	79	91	80	80	79	79	-	-	-	-	-	-
44	Pol 2878-21	74	84	76	74	74	74	79	91	80	80	79	79	-	-	-	-	-	-
45	Pol 2879-20	73	76	74	74	74	74	79	80	80	79	79	80	-	-	-	-	-	-
46	Pol 2879-21	73	76	74	74	74	74	79	80	80	79	79	80	-	-	-	-	-	-
47	Pol 2880-20	74	74	74	74	74	74	79	80	79	79	80	80	-	-	-	-	-	-
48	Pol 2881-21	63	74	74	74	74	64	78	79	79	80	80	79	-	-	-	-	-	-
49	Pol 2882-21	63	74	74	74	64	64	78	79	80	80	79	79	-	-	-	-	-	-
50	Pol 2885-20	39	64	64	64	40	42	67	79	79	79	67	68	-	-	-	-	-	-
51	Vif 9-21	71	71	71	71	71	71	91	91	91	92	92	92	74	75	75	74	74	74
52	Vif 10-21	69	71	71	71	71	70	88	91	92	92	92	89	74	75	74	74	74	75
53	Vif 11-21	69	71	71	71	70	70	88	92	92	92	89	90	74	74	74	74	75	76
54	Vif 12-20	69	71	71	70	70	70	88	92	92	89	90	90	74	74	74	75	76	76
55	Vif 14-21	69	70	70	70	70	70	89	89	90	90	90	90	75	75	76	76	76	76

#	Target	LANL + Virco (all)					LANL clade B					Virco (all)							
		23	-2	-1	p0	+1	+2	23	-2	-1	p0	+1	+2	23	-2	-1	p0	+1	+2
56	Vif 15-21	70	70	70	70	70	70	89	90	90	90	90	90	76	76	76	76	76	76
57	Vif 37-20	2	45	43	71	13	4	3	76	75	87	17	6	2	36	35	67	18	3
58	Tat (x1) 131-20	68	68	68	68	68	70	83	84	84	84	84	85	57	58	58	57	57	57
59	Tat (x1) 132-21	68	68	68	68	70	70	84	84	84	84	85	86	57	58	57	57	57	58
60	Tat (x1) 140-21	68	81	82	73	72	72	73	79	78	77	75	75	62	77	80	77	74	73
61	Tat (x1) 143-21	60	72	68	62	60	60	68	75	74	70	68	68	57	73	66	58	58	57
62	Tat (x1) 147-20	59	60	60	60	59	59	67	68	68	68	67	67	54	58	57	57	56	54
63	Rev (x1) 9-20	57	61	60	60	59	58	66	68	68	68	67	66	52	57	57	56	54	53
64	Vpu 139-21	6	6	44	64	64	56	12	12	70	77	78	72	6	6	49	76	76	66
65	Vpu 140-20	36	44	64	64	56	65	65	70	77	78	72	82	40	49	76	76	66	69
66	Vpu 140-21	36	44	64	64	56	65	65	70	77	78	72	82	40	49	76	76	66	69
67	Vpu 141-20	55	64	64	56	65	66	72	77	78	72	82	83	66	76	76	66	69	69
68	Vpu 143-20	55	56	65	66	65	66	72	72	82	83	83	83	65	66	69	69	68	69
69	Vpu 144-20	65	65	66	65	66	67	82	82	83	83	83	84	68	69	69	68	69	73
70	Vpu 144-21	65	65	66	65	66	67	82	82	83	83	83	84	68	69	69	68	69	73
71	Vpu 145-20	49	66	65	66	67	53	60	83	83	83	84	63	56	69	68	69	73	61
72	Vpu 146-20	48	65	66	67	53	53	59	83	83	84	63	62	56	68	69	73	61	61
73	Vpu 147-21	50	66	67	53	53	53	60	83	84	63	62	62	56	69	73	61	61	61
74	Vpu 148-21	46	67	53	53	53	49	55	84	63	62	62	56	48	73	61	61	61	50
75	Vpu 149-20	48	53	53	53	49	49	56	63	62	62	56	57	50	61	61	61	50	53
76	Env 628-20	45	46	46	61	61	60	63	64	65	85	85	84	-	-	-	-	-	-
77	Env 1423-20	48	53	67	72	74	74	60	66	73	76	77	77	-	-	-	-	-	-
78	Env 1423-21	48	53	67	72	74	74	60	66	73	76	77	77	-	-	-	-	-	-
79	Env 1424-21	63	67	72	74	74	79	70	73	76	77	77	84	-	-	-	-	-	-
80	Env 1425-20	71	72	74	74	79	79	75	76	77	77	84	84	-	-	-	-	-	-
81	Env 1425-21	71	72	74	74	79	79	75	76	77	77	84	84	-	-	-	-	-	-
82	Env 1426-21	67	74	74	79	79	73	75	77	77	84	84	81	-	-	-	-	-	-
83	Env 1428-21	72	79	79	73	75	75	80	84	84	81	82	82	-	-	-	-	-	-
84	Env 1431-21	65	75	75	65	65	65	80	82	82	81	81	81	-	-	-	-	-	-
85	Env 1432-20	62	75	65	65	65	63	78	82	81	81	81	79	-	-	-	-	-	-
86	Env 1432-21	62	75	65	65	65	63	78	82	81	81	81	79	-	-	-	-	-	-
87	Env 1433-20	62	65	65	65	63	64	78	81	81	81	79	80	-	-	-	-	-	-
88	Env 1433-21	62	65	65	65	63	64	78	81	81	81	79	80	-	-	-	-	-	-
89	Env 1434-20	62	65	65	63	64	64	78	81	81	79	80	80	-	-	-	-	-	-



#	Target	LANL + Virco (all)					LANL clade B					Virco (all)							
		23	-2	-1	p0	+1	+2	23	-2	-1	p0	+1	+2	23	-2	-1	p0	+1	+2
90	Env 1436-20	61	63	64	<b>64</b>	66	66	78	79	80	<b>80</b>	83	83	-	-	-	-	-	-
91	Env 1437-20	62	64	64	<b>66</b>	66	67	79	80	80	<b>83</b>	83	83	-	-	-	-	-	-
92	Env 2308-20	27	44	43	<b>43</b>	37	28	48	76	75	<b>74</b>	66	49	-	-	-	-	-	-
93	Rev (x2) 158-20	44	47	49	<b>48</b>	48	46	69	74	77	<b>76</b>	75	72	32	33	38	<b>38</b>	38	37
94	Rev (x2) 159-20	39	49	48	<b>48</b>	46	40	64	77	76	<b>75</b>	72	65	21	38	38	<b>38</b>	37	21
95	Nef 350-20	47	50	51	<b>51</b>	49	49	60	63	64	<b>64</b>	62	62	36	40	44	<b>44</b>	39	40
96	Nef 351-20	48	51	51	<b>49</b>	49	49	61	64	64	<b>62</b>	62	62	39	44	44	<b>39</b>	40	40
<b>Average 96:</b>		59	66	68	68	67	65	74	80	81	81	80	78	52	60	63	63	60	59
<b>Virco equiv. *:</b>		52	59	61	62	59	58	68	77	79	78	74	73	52	60	63	63	60	59

\* *Virco equiv.* = *Virco equivalent sequences*

## Reporter lengths and average activities

Two groups of fluorescent reporters were used in this study. The 'original' reporters made up of GFP fusions to each of the full length HIV genes. The accessory gene reporters were classified as short reporters given their relative shorter lengths compared to the the 3 core genes, which were classified as long reporters. A second group of reporters was for the long core reporters, consisting of shorter fragments spanning overlapping regions of each core gene and containing only that sequence targeted. The following table summarizes each reporter including its length and the average % fluorescence (**A. %F**) of all the shRNAs measured with each reporter. Also shown for the second group of shortened core reporters is the original average % fluorescence of the corresponding shRNA subset (**Cs. A. %F**), i.e. the average % fluorescence (and shRNA #s) for the subset of shRNAs tested with the original long reporters corresponding to each shortened fragment reporter. The last column contains the combined average % fluorescence (**C. A. %F**), that is, the activities measured from the group 1 accessory gene reporters and the group 2 shortened core gene reporters, tallied to a combined average at the bottom.

Rep.	Length	A. %F.	Cs. A. %F.	Shortened Rep.	Length	A. %F.	C. A. %F.
Gag	1503	38.38%	-				-
subset #10 - #18		-	39%	Gag-1 (500 - 1150)	650	37%	37%
Pol	3012	41.64%	-				-
subset #19 - #29		-	41%	Pol-1 (1 - 436)	436	21%	21%
subset #30		-	58%	Pol-2 (400 - 850)	450	51%	51%
subset #31 - #34		-	41%	Pol-3 (800 - 1250)	450	20%	20%
subset #35 - #57		-	42%	Pol-4 (2600 - 3025)	425	27%	27%
Tat (x1-2)	258	18.92%	-				19%
Rev (x1-2)	272	12.73%	-				13%
Vpu	243	10.32%	-				10%
Env	2250	52.20%	-				-
subset #77- #91		-	53%	Env-1 (1300 - 1740)	440	18%	18%
<b>Nef-LTR *</b>	921	44.36%					46%
<b>Combined averages :</b>		<b>37.2%</b>	<b>43.8%</b>			<b>25.0%</b>	<b>24.9%</b>

\* this may be ~ 100 bp shorter than intended due to an internal transcription termination signal within the LTR sequence.

## Ranked selection

We created a scoring method to rank the 96 hairpins for overall performance based on their suppressive activities, non-specific activities and percentage conservations. Each parameter was weighted approximately equally and was awarded a score out of 100 (nb. changing the weighting for any parameter will change the following ranking). Suppressive activities from both assays were tallied as 100 minus the percentage fluorescence value or percentage p24 graphed. Non-specific activity was awarded a top score of 100 for no activity (a normalization factor of 1.0), which was reduced accordingly for increased non-specific activity, e.g. a normalization factor of 1.2 (0.2 fold enhancement) or 0.8 (0.2 fold reduction) was only awarded 80, and so on. The percentage conservations for all strains and the clade B subtypes were also tallied, but only using the p0 position. The top ranking hairpin for each gene is shown in bold. The hairpins sorted in ranked order according to score (the last column, **Rk.**). The original values and the scores for each parameter are shown.

#	Target	%F.		% p24		Non sp.		% cons. ALL (p0)		% cons. B (p0)		C.s.	Rk.
		Val.	Sc.	Val.	Sc.	Val.	Sc.	Val.	Sc.	Val.	Sc.		
5	<b>LTR 510-21</b>	18.7	81.3	1.1	98.9	1.3	70	70	70	100	100	420	1
8	LTR 527-21	23.9	76.1	3.9	96.1	1.5	50	93	93	100	100	415	2
4	LTR 509-20	35.5	64.5	8.5	91.5	1.2	80	69	69	100	100	405	3
83	<b>Env 1428-21</b>	7.3	92.7	6.3	93.8	1.4	60	73	73	81	81	401	4
58	<b>Tat (x1) 131-20</b>	26.3	73.7	16.5	83.5	1.1	90	68	68	84	84	399	5
59	Tat (x1) 132-21	26.8	73.2	26.2	73.8	1.0	100	68	68	84	84	399	6
81	Env 1425-21	7.2	92.8	6.2	93.9	1.4	60	74	74	77	77	398	7
82	Env 1426-21	6.9	93.1	4.3	95.7	1.7	30	79	79	84	84	382	8
6	LTR 516-21	14.1	85.9	0.6	99.4	1.9	10	71	71	100	100	366	9
11	<b>Gag 532-21</b>	40.7	59.3	4.9	95.1	1.4	60	70	70	80	80	364	10
79	Env 1424-21	6.5	93.5	10.7	89.3	1.7	30	74	74	77	77	364	11
7	LTR 522-20	61.0	39.0	47.4	52.6	1.2	80	92	92	100	100	364	12
68	<b>Vpu 143-20</b>	6.7	93.3	10.1	89.9	1.7	30	66	66	83	83	362	13
80	Env 1425-20	11.3	88.7	21.5	78.6	1.6	40	74	74	77	77	358	14
23	<b>Pol 248-21</b>	8.8	91.2	0.3	99.7	3.1	0	80	80	87	87	358	15
22	Pol 248-20	8.9	91.1	0.5	99.5	2.2	0	80	80	87	87	358	16
39	Pol 2726-21	15.4	84.6	4.5	95.5	1.9	10	81	81	86	86	357	17
25	Pol 249-21	8.8	91.2	0.5	99.5	2.7	0	79	79	86	86	356	18

#	Target	%F.		% p24		Non sp.		% cons. ALL (p0)		% cons. B (p0)		C.s.	Rk.
		Val.	Sc.	Val.	Sc.	Val.	Sc.	Val.	Sc.	Val.	Sc.		
27	Pol 250-21	9.8	90.2	1.7	98.3	2.8	0	80	80	87	87	356	19
62	Tat (x1) 147-20	17.0	83.0	5.7	94.3	1.5	50	60	60	68	68	355	20
26	Pol 250-20	9.8	90.2	2.0	98.0	2.5	0	80	80	87	87	355	21
24	Pol 249-20	9.2	90.8	1.0	99.0	3.3	0	79	79	86	86	355	22
37	Pol 2671-21	9.9	90.1	2.7	97.3	1.8	20	73	73	74	74	354	23
21	Pol 247-21	10.0	90.0	2.7	97.3	3.0	0	80	80	87	87	354	24
20	Pol 246-21	8.5	91.5	0.5	99.5	4.2	0	80	80	81	81	352	25
38	Pol 2724-20	13.8	86.2	4.3	95.7	1.7	30	71	71	67	67	350	26
29	Pol 404-20	23.8	76.2	38.7	61.3	0.8	80	67	67	65	65	350	27
64	Vpu 139-21	8.9	91.1	15.0	85.0	1.7	30	64	64	77	77	347	28
44	Pol 2878-21	8.0	92.0	1.5	98.5	2.6	0	74	74	80	80	345	29
43	Pol 2878-20	7.6	92.4	2.5	97.5	2.6	0	74	74	80	80	344	30
45	Pol 2879-20	8.1	91.9	1.2	98.8	2.1	0	74	74	79	79	344	31
57	<b>Vif 37-20</b>	9.5	90.5	4.9	95.1	2.7	0	71	71	87	87	344	32
52	Vif 10-21	11.0	89.0	9.7	90.3	7.2	0	71	71	92	92	342	33
42	Pol 2874-21	20.3	79.7	15.8	84.2	3.1	0	85	85	93	93	342	34
49	Pol 2882-21	8.8	91.2	3.5	96.5	3.4	0	74	74	80	80	342	35
35	Pol 2668-21	17.1	82.9	4.8	95.2	2.6	0	78	78	84	84	340	36
53	Vif 11-21	15.7	84.3	9.4	90.6	3.4	0	71	71	92	92	338	37
48	Pol 2881-21	12.1	87.9	5.7	94.3	3.1	0	74	74	80	80	336	38
36	Pol 2670-21	9.5	90.5	2.0	98.0	2.0	0	73	73	74	74	336	39
71	Vpu 145-20	6.8	93.2	7.7	92.3	2.6	0	66	66	83	83	335	40
13	Gag 533-21	31.1	68.9	5.2	94.8	1.8	20	70	70	80	80	334	41
72	Vpu 146-20	7.3	92.7	11.7	88.3	2.1	0	67	67	84	84	332	42
77	Env 1423-20	6.1	93.9	10.2	89.9	3.0	0	72	72	76	76	332	43
46	Pol 2879-21	95.8	4.2	15.7	84.3	1.1	90	74	74	79	79	332	44
56	Vif 15-21	22.6	77.4	6.4	93.6	2.8	0	70	70	90	90	331	45
55	Vif 14-21	13.9	86.1	15.6	84.4	4.5	0	70	70	90	90	331	46
17	Gag 578-20	35.3	64.7	26.8	73.2	1.5	50	63	63	79	79	330	47
60	Tat (x1) 140-21	14.4	85.6	5.7	94.3	2.6	0	73	73	77	77	330	48
78	Env 1423-21	8.3	91.7	11.5	88.5	2.6	0	72	72	76	76	328	49
70	Vpu 144-21	5.9	94.1	14.0	86.0	2.3	0	65	65	83	83	328	50
47	Pol 2880-20	29.2	70.8	16.0	84.0	1.8	20	74	74	79	79	328	51

#	Target	%F.		% p24		Non sp.		% cons. ALL (p0)		% cons. B (p0)		C.s.	Rk.
		Val.	Sc.	Val.	Sc.	Val.	Sc.	Val.	Sc.	Val.	Sc.		
51	Vif 9-21	16.6	83.4	18.9	81.1	6.7	0	71	71	92	92	328	52
86	Env 1432-21	7.3	92.7	32.0	68.0	1.8	20	65	65	81	81	327	53
19	Pol 242-20	8.0	92.0	0.5	99.5	2.0	0	58	58	77	77	327	54
14	Gag 534-20	40.1	59.9	14.7	85.3	1.7	30	69	69	80	80	324	55
30	Pol 529-21	51.2	48.8	29.2	70.8	1.5	50	68	68	84	84	322	56
69	Vpu 144-20	6.8	93.2	21.4	78.6	2.5	0	65	65	83	83	320	57
32	Pol 925-20	14.5	85.5	2.7	97.3	3.6	0	60	60	77	77	320	58
12	Gag 533-20	27.1	72.9	4.1	95.9	2.1	0	70	70	80	80	319	59
10	Gag 532-20	55.4	44.6	46.0	54.1	1.3	70	70	70	80	80	319	60
9	LTR 553-20	61.5	38.5	33.0	67.0	1.3	70	51	51	92	92	319	61
96	<b>Nef 351-20</b>	38.5	61.5	36.3	63.7	1.2	80	49	49	62	62	316	62
61	Tat (x1) 143-21	14.0	86.0	2.3	97.7	2.1	0	62	62	70	70	316	63
54	Vif 12-20	12.8	87.2	30.8	69.2	5.1	0	70	70	89	89	315	64
89	Env 1434-20	19.9	80.1	78.2	21.9	1.3	70	63	63	79	79	314	65
67	Vpu 141-20	6.5	93.5	8.5	91.5	2.4	0	56	56	72	72	313	66
18	Gag 698-20	20.9	79.1	12.2	87.9	5.0	0	70	70	76	76	313	67
15	Gag 534-21	44.0	56.0	33.7	66.4	1.6	40	69	69	80	80	311	68
33	Pol 926-20	20.3	79.7	8.5	91.5	2.2	0	60	60	77	77	308	69
73	Vpu 147-21	6.2	93.8	5.8	94.2	2.7	0	53	53	63	63	304	70
1	LTR 63-20	56.4	43.6	48.8	51.2	1.0	100	33	33	76	76	304	71
90	Env 1436-20	42.3	57.7	88.1	12.0	1.1	90	64	64	80	80	304	72
63	<b>Rev (x1) 9-20</b>	15.0	85.0	11.6	88.4	2.9	0	60	60	68	68	301	73
74	Vpu 148-21	6.0	94.0	8.2	91.8	2.7	0	53	53	62	62	301	74
66	Vpu 140-21	24.6	75.4	67.4	32.6	1.5	50	64	64	78	78	300	75
16	Gag 535-20	33.9	66.1	51.4	48.6	1.7	30	71	71	82	82	298	76
41	Pol 2873-21	39.5	60.5	45.8	54.2	3.4	0	88	88	95	95	298	77
75	Vpu 149-20	7.8	92.2	10.3	89.7	2.2	0	53	53	62	62	297	78
87	Env 1433-20	18.2	81.8	89.9	10.1	1.5	50	65	65	81	81	288	79
31	Pol 921-20	27.2	72.8	19.0	81.0	4.0	0	58	58	76	76	288	80
88	Env 1433-21	18.9	81.1	72.6	0.0	1.4	60	65	65	81	81	287	81
3	LTR 507-20	78.7	21.3	102	-1.8	1.0	100	69	69	98	98	287	82
91	Env 1437-20	63.0	37.0	99.9	0.0	1.0	100	66	66	83	83	286	83
34	Pol 1161-20	19.6	80.4	6.5	93.5	2.3	0	32	32	75	75	281	84

#	Target	%F.		% p24		Non sp.		% cons. ALL (p0)		% cons. B (p0)		C.s.	Rk.
		Val.	Sc.	Val.	Sc.	Val.	Sc.	Val.	Sc.	Val.	Sc.		
76	Env 628-20	47.7	52.3	68.3	31.8	1.5	50	61	61	85	85	280	85
95	Nef 350-20	27.6	72.4	28.9	0.0	0.9	90	51	51	64	64	277	86
92	Env 2308-20	18.8	81.2	35.6	64.4	2.2	0	43	43	74	74	263	87
2	LTR 506-20	72.0	28.0	85.1	14.9	1.6	40	67	67	97	97	247	88
85	Env 1432-20	12.0	88.0	40.5	0.0	2.0	0	65	65	81	81	234	89
28	Pol 251-20	125	-25	88.7	11.3	0.8	80	80	80	87	87	234	90
40	Pol 2750-20	122	-22	0.8	99.2	1.9	10	65	65	75	75	227	91
93	Rev (x2) 158-20	8.0	0.0	2.5	97.5	2.1	0	48	48	76	76	222	92
84	Env 1431-21	28.4	71.6	118	-18	1.8	20	65	65	81	81	220	93
65	Vpu 140-20	30.3	69.7	156	-56	1.4	60	64	64	78	78	215	94
94	Rev (x2) 159-20	11.4	0.0	10.1	89.9	3.4	0	48	48	75	75	213	95
50	Pol 2885-20	103	-2.6	212	-112	0.8	80	64	64	79	79	108	96

## Additional methods

### Oligonucleotide templates for Phi-29 extension

The coding region for each hairpin was contained within a single oligonucleotide, organized in the following general format: 5'-GCGCGGATCC | core(20 - 21) | NCTCGAGN | core(20 - 21) | [G/A/C] | TTTTTTGGGAAGCTT-3'. Each oligonucleotide encoded a digestion 'seat' (for the RE to better bind the oligonucleotide), a 5' RE recognition site (*Bam* HI), the shRNA sense stem region, loop sequence, anti-sense stem, a termination spacer (if required), a pol III termination signal consisting of a run of at least 4 'T's, a 3' RE recognition site (*Hind* III) and a 3' digestion seat. The termination spacer was included only if the last base of the anti-sense stem was 'T', so as to prevent premature termination via an early run of 'T's. A short primer was designed to bind at the 3' end of the template oligo and introduced the 3' seat (5'-CGCGAAGCTTCCAAAAA-3').

Oligos were ordered at the minimal synthesis and purification scales (0.05  $\mu$ M and desalt, Sigma-Genosys) and were re-suspended in water (1 - 10  $\mu$ g /  $\mu$ l). Twenty picomoles of each oligo was used in the extension reaction (1x reaction buffer, 2x BSA, 20  $\mu$ M dNTPs (1  $\mu$ l of a 10 mM stock), 10 units of Phi29 (New England Biolabs) and water to 20  $\mu$ l), which was incubated at 30 °C for ~10 min., then 65 °C for 10 min. (to disable the polymerase). The extension product was digested (*Bam* HI plus *Hind* III), purified using the Nucleotide Removal kit (Qiagen), ligated to the expression plasmid and used to transform electrocompetent GT116 *E.coli*. Positive clones were confirmed by automated sequencing using our loop digestion method (see below). The following table lists the **93** oligonucleotide template sequences ordered as Phi-29 extension templates.

Hairpin #	Target (NL4-3)	Oligonucleotide template (5' to 3')
1	LTR 63-20	GCGCGGATCCCAAGGCTACTTCCCTGATTGCCTCGAGCCAATCAGG GAAGTAGCCTTGTTTTTTGGGAAGCTT
2	LTR 506-20	GCGCGGATCCGGAACCCACTGCTTAAGCCTGCTCGAGGAGGCTTAA GCAGTGGGTTCTTTTTTGGGAAGCTT
3	LTR 507-20	GCGCGGATCCGAACCCACTGCTTAAGCCTCTCTCGAGTGAGGCTTAA GCAGTGGGTTCTTTTTTGGGAAGCTT
4	LTR 509-20	GCGCGGATCCACCCACTGCTTAAGCCTCAAACCTCGAGATTGAGGCTT AAGCAGTGGGTTTTTTTTGGGAAGCTT
5	LTR 510-21	GCGCGGATCCCCACTGCTTAAGCCTCAATAACTCGAGATATTGAGG CTTAAGCAGTGGGTTTTTTTTGGGAAGCTT
6	LTR 516-21	Overlapping oligonucleotide pair used instead, see below
7	LTR 522-20	Overlapping oligonucleotide pair used instead, see below

Hairpin #	Target (NL4-3)	Oligonucleotide template (5' to 3')
8	LTR 527-21	Overlapping oligonucleotide pair used instead, see below
9	LTR 553-20	GCGCGGATCCGTAGTGTGCCCCGTCTGTTCCCTCGAGCAACAGACG GGCACACACTACTTTTTTGAAGCTT
10	Gag 532-20	GCGCGGATCCGGAGCCACCCCAAGATTTCTCGAGTAAATCTTGT GGGGTGGCTCCTTTTTTGAAGCTT
11	Gag 532-21	GCGCGGATCCGGAGCCACCCCAAGATTTAACTCGAGATAAATCTT GTGGGGTGGCTCCTTTTTTGAAGCTT
12	Gag 533-20	GCGCGGATCCGAGCCACCCCAAGATTTATCTCGAGTAAATCTTGT GGGGTGGCTCCTTTTTTGAAGCTT
13	Gag 533-21	GCGCGGATCCGAGCCACCCCAAGATTTAACTCGAGATAAATCTT GTGGGGTGGCTCCTTTTTTGAAGCTT
14	Gag 534-20	GCGCGGATCCAGCCACCCCAAGATTTAATCTCGAGTTAAATCTTG TGGGGTGGCTCCTTTTTTGAAGCTT
15	Gag 534-21	GCGCGGATCCAGCCACCCCAAGATTTAAACTCGAGATTTAAATCT TGTGGGGTGGCTCCTTTTTTGAAGCTT
16	Gag 535-20	GCGCGGATCCGCCACCCCAAGATTTAAACTCGAGATTTAAATCTT GTGGGGTGGCTCCTTTTTTGAAGCTT
17	Gag 578-20	GCGCGGATCCGACATCAAGCAGCCATGCAATCTCGAGTTTGCATGGC TGCTTGATGTCTTTTTTGAAGCTT
18	Gag 698-20	GCGCGGATCCGAAGTGACATAGCAGGAATTCTCGAGTAGTTCCTGC TATGTCACCTCTTTTTTGAAGCTT
19	Pol 242-20	GCGCGGATCCATACAGGAGCAGATGATACACCTCGAGCTGTATCATCT GCTCCTGTATCTTTTTTGAAGCTT
20	Pol 246-21	GCGCGGATCCAGGAGCAGATGATACAGTATTACTCGAGAAATACTGTA TCATCTGCTCCTTTTTTGAAGCTT
21	Pol 247-21	GCGCGGATCCGGAGCAGATGATACAGTATTAACTCGAGATAATACTGTA TCATCTGCTCCTTTTTTGAAGCTT
22	Pol 248-20	GCGCGGATCCGAGCAGATGATACAGTATTACCTCGAGCTAATACTGTAT CATCTGCTCTTTTTTGAAGCTT
23	Pol 248-21	GCGCGGATCCGAGCAGATGATACAGTATTAGACTCGAGACTAATACTG TATCATCTGCTCTTTTTTGAAGCTT
24	Pol 249-20	GCGCGGATCCAGCAGATGATACAGTATTAGTCTCGAGTCTAATACTGTA TCATCTGCTCTTTTTTGAAGCTT
25	Pol 249-21	GCGCGGATCCAGCAGATGATACAGTATTAGAACTCGAGATCTAATACT GTATCATCTGCTCTTTTTTGAAGCTT
26	Pol 250-20	GCGCGGATCCGCAGATGATACAGTATTAGATCTCGAGTTCTAATACTGT ATCATCTGCTCTTTTTTGAAGCTT
27	Pol 250-21	GCGCGGATCCGCAGATGATACAGTATTAGAACTCGAGATTCTAATACT GTATCATCTGCTCTTTTTTGAAGCTT
28	Pol 251-20	GCGCGGATCCCAGATGATACAGTATTAGAACCTCGAGCTTCTAATACT GTATCATCTGTTTTTGAAGCTT



Hairpin #	Target (NL4-3)	Oligonucleotide template (5' to 3')
29	Pol 404-20	GCGCGGATCCCTACACCTGTCAACATAATTCCTCGAGCAATTATGTTG ACAGGTGTAGTTTTTTGGAAGCTT
30	Pol 529-21	GCGCGGATCCAACAATGGCCATTGACAGAAACTCGAGATTCTGTCA ATGGCCATTGTTATTTTTGGAAGCTT
31	Pol 921-20	GCGCGGATCCATGGAAGGATCACCAGCAAACCTCGAGATTGCTGGTG ATCCTTTCCATCTTTTTTGAAGCTT
32	Pol 925-20	GCGCGGATCCAAGGATCACCAGCAATATTGCTCGAGGAATATTGCTG GTGATCCTTTCTTTTTGGAAGCTT
33	Pol 926-20	GCGCGGATCCAAGGATCACCAGCAATATTGCTCGAGGAATATTGCT GGTGATCCTTTTTTTGGAAGCTT
34	Pol 1161-20	GCGCGGATCCTGAACTCCATCCTGATAAATCCTCGAGCATTATCAGG ATGGAGTTCATTTTTGGAAGCTT
35	Pol 2668-21	GCGCGGATCCGAGTACAAATGGCAGTATTCACTCGAGAGAATACTG CCATTTGACTGCTTTTTTGAAGCTT
36	Pol 2670-21	GCGCGGATCCAGTACAAATGGCAGTATTCATACTCGAGAATGAATACT GCCATTTGACTGTTTTTGAAGCTT
37	Pol 2671-21	GCGCGGATCCGTACAAATGGCAGTATTCATACTCGAGAGATGAATAC TGCCATTTGACTTTTTTGAAGCTT
38	Pol 2724-20	GCGCGGATCCGTACAGTGCAGGGGAAAGAAACTCGAGATTCTTTCCC CTGCACTGTACTTTTTTGAAGCTT
39	Pol 2726-21	GCGCGGATCCACAGTGCAGGGGAAAGAATAGACTCGAGACTATTCTT TCCCCTGCACTGTATTTTTGGAAGCTT
40	Pol 2750-20	GCGCGGATCCACATAATAGCAACAGACATAGCTCGAGGTATGTCTGTT GCTATTATGTCTTTTTTGAAGCTT
41	Pol 2873-21	GCGCGGATCCGAAAGGTGAAGGGGCAGTAGACTCGAGACTACTGC CCCTTCACCTTTCCTTTTTTGAAGCTT
42	Pol 2874-21	GCGCGGATCCGAAAGGTGAAGGGGCAGTAGTACTCGAGAACTACTG CCCCTTCACCTTCTTTTTTGAAGCTT
43	Pol 2878-20	GCGCGGATCCGGTGAAGGGGCAGTAGTAATTCTCGAGTATTACTACTG CCCCTTCACCTTTTTTGAAGCTT
44	Pol 2878-21	GCGCGGATCCGGTGAAGGGGCAGTAGTAATAACTCGAGATATTACTAC TGCCCCTTCACCTTTTTTGAAGCTT
45	Pol 2879-20	GCGCGGATCCGTGAAGGGGCAGTAGTAATAGCTCGAGGTATTACTACT GCCCTTCACTTTTTTGGAAGCTT
46	Pol 2879-21	GCGCGGATCCGTGAAGGGGCAGTAGTAATACTCGAGAGTATTACTA CTGCCCTTCACTTTTTTGGAAGCTT
47	Pol 2880-20	GCGCGGATCCTGAAGGGGCAGTAGTAATACTCTCGAGTGTATTACTAC TGCCCCTTCATTTTTTGAAGCTT
48	Pol 2881-21	GCGCGGATCCGAAGGGGCAGTAGTAATACAAACTCGAGATTGTATTAC TACTGCCCTTCTTTTTTGAAGCTT
49	Pol 2882-21	GCGCGGATCCAAGGGGCAGTAGTAATACAAGACTCGAGACTTGATTATTA CTACTGCCCTTCTTTTTTGAAGCTT

Hairpin #	Target (NL4-3)	Oligonucleotide template (5' to 3')
50	Pol 2885-20	GCGCGGATCCGGGCAGTAGTAATACAAGATTCTCGAGTATCTTGATTA CTACTGCCCTTTTTTGAAGCTT
51	Vif 9-21	GCGCGGATCCCAGATGGCAGGTGATGATTGTACTIONGAGAACAATCAT CACCTGCCATCTGTTTTTGAAGCTT
52	Vif 10-21	GCGCGGATCCAGATGGCAGGTGATGATTGTGACTCGAGACACAATCA TCACCTGCCATCTGTTTTTGAAGCTT
53	Vif 11-21	GCGCGGATCCGATGGCAGGTGATGATTGTGACTCGAGAACACAATC ATCACCTGCCATCTTTTTTGAAGCTT
54	Vif 12-20	GCGCGGATCCATGGCAGGTGATGATTGTGCTCGAGCACACAATCA TCACCTGCCATCTTTTTTGAAGCTT
55	Vif 14-21	GCGCGGATCCGGCAGGTGATGATTGTGGCACTCGAGAGCCACAC AATCATCACCTGCCCTTTTTTGAAGCTT
56	Vif 15-21	GCGCGGATCCGAGGTGATGATTGTGGCAACTCGAGATGCCACA CAATCATCACCTGCTTTTTTGAAGCTT
57	Vif 37-20	GCGCGGATCCGTAGACAGGATGAGGATTAAGCTCGAGGTTAATCCTC ATCCTGTCTACTTTTTTGAAGCTT
58	Tat (exon 1) 131-20	GCGCGGATCCGCATCTCCTATGGCAGGAAGTCTCGAGTCTTCCTGCC ATAGGAGATGCTTTTTTGAAGCTT
59	Tat (exon 1) 132-21	GCGCGGATCCCATCTCCTATGGCAGGAAGAACTCGAGATTCTTCCT GCCATAGGAGATGTTTTTGAAGCTT
60	Tat (exon 1) 140-21	GCGCGGATCCATGGCAGGAAGAAGCGGAGACACTCGAGAGTCTCC GCTTCTCCTGCCATATTTTTTGAAGCTT
61	Tat (exon 1) 143-21	GCGCGGATCCGAGGAAGAAGCGGAGACAGCACTCGAGAGCTGTC TCCGCTTCTCCTGCTTTTTTGAAGCTT
62	Tat (exon 1) 147-20	GCGCGGATCCGAAGAAGCGGAGACAGCGACCCTCGAGCGTCGCTG TCTCCGCTTCTTTTTTGAAGCTT
63	Rev (exon 1) 9-20	GCGCGGATCCAAGAAGCGGAGACAGCGACTCTCGAGTCGTCGCT GTCTCCGCTTCTTTTTTGAAGCTT
64	Vpu 139-21	GCGCGGATCCGAAAGAGCAGAAGACAGTGGCACTCGAGAGCCACT GTCTTCTGCTCTTCTTTTTTGAAGCTT
65	Vpu 140-20	GCGCGGATCCAAGAGCAGAAGACAGTGGCTCTCGAGTGCCACTGT CTTCTGCTCTTCTTTTTTGAAGCTT
66	Vpu 140-21	GCGCGGATCCAAGAGCAGAAGACAGTGGCAACTCGAGATGCCACT GTCTTCTGCTCTTCTTTTTTGAAGCTT
67	Vpu 141-20	GCGCGGATCCAAGAGCAGAAGACAGTGGCATCTCGAGTTGCCACTG TCTTCTGCTCTTTTTTGAAGCTT
68	Vpu 143-20	GCGCGGATCCGAGCAGAAGACAGTGGCAATCCTCGAGCATTGCCAC TGTCTTCTGCTCTTTTTTGAAGCTT
69	Vpu 144-20	GCGCGGATCCAGCAGAAGACAGTGGCAATGTCTCGAGTCATTGCCA CTGTCTTCTGCTCTTTTTTGAAGCTT
70	Vpu 144-21	GCGCGGATCCAGCAGAAGACAGTGGCAATGAACTCGAGATCATTGC CACTGTCTTCTGCTCTTTTTTGAAGCTT

Hairpin #	Target (NL4-3)	Oligonucleotide template (5' to 3')
71	Vpu 145-20	GCGCGGATCCGCAGAAGACAGTGGCAATGACCTCGAGCTCATTGCC ACTGTCTTCTGCTTTTTTGAAGCTT
72	Vpu 146-20	GCGCGGATCCCAGAAGACAGTGGCAATGAGTCTCGAGTCTCATTGC CACTGTCTTCTGTTTTTGAAGCTT
73	Vpu 147-21	GCGCGGATCCAGAAGACAGTGGCAATGAGAGACTCGAGACTCTCATT GCCACTGTCTTCTGTTTTTGAAGCTT
74	Vpu 148-21	GCGCGGATCCGAAGACAGTGGCAATGAGAGTACTCGAGAACTCTCAT TGCCACTGTCTTCTTTTTTGAAGCTT
75	Vpu 149-20	GCGCGGATCCAAGACAGTGGCAATGAGAGTCTCGAGCACTCTCATT GCCACTGTCTTCTTTTTTGAAGCTT
76	Env 628-20	GCGCGGATCCCCAATTCCCATACATTATTGACTCGAGACAATAATGTAT GGGAATTGGTTTTTGAAGCTT
77	Env 1423-20	GCGCGGATCCGACAATTGGAGAAGTGAATTTCTCGAGTAATTCAC TTC TCCAATTGTCTTTTTTGAAGCTT
78	Env 1423-21	GCGCGGATCCGACAATTGGAGAAGTGAATTA ACTCGAGATAATTCACT TCTCCAATTGTCTTTTTTGAAGCTT
79	Env 1424-21	GCGCGGATCCACAATTGGAGAAGTGAATTATACTCGAGAATAATTCACT TCTCCAATTGTCTTTTTTGAAGCTT
80	Env 1425-20	GCGCGGATCCCCAATTGGAGAAGTGAATTATTCTCGAGTATAATTCACT CTCCAATTGTTTTTGAAGCTT
81	Env 1425-21	GCGCGGATCCCCAATTGGAGAAGTGAATTATAACTCGAGATATAATTCAC TTCTCCAATTGTTTTTGAAGCTT
82	Env 1426-21	GCGCGGATCCAATTGGAGAAGTGAATTATACTCGAGAATAATAATTCAC CTTCTCCAATTGTTTTTGAAGCTT
83	Env 1428-21	GCGCGGATCCTTGGAGAAGTGAATTATATAAACTCGAGATTATATAATTC ACTTCTCCAATTTTTTGAAGCTT
84	Env 1431-21	GCGCGGATCCGAGAAGTGAATTATATAAAATAACTCGAGATATTTATATAAT TCACTTCTTTTTTGAAGCTT
85	Env 1432-20	GCGCGGATCCAGAAGTGAATTATATAAAATAACTCGAGATATTTATATAATT CACTTCTTTTTTGAAGCTT
86	Env 1432-21	GCGCGGATCCAGAAGTGAATTATATAAATAACTCGAGAATAATTTATATAA TTCACTTCTTTTTTGAAGCTT
87	Env 1433-20	GCGCGGATCCGAAGTGAATTATATAAATATTCTCGAGTATATTTATATAAAT CACTTCTTTTTTGAAGCTT
88	Env 1433-21	GCGCGGATCCGAAGTGAATTATATAAATAACTCGAGATATATTTATATA ATTCACTTCTTTTTTGAAGCTT
89	Env 1434-20	GCGCGGATCCAAGTGAATTATATAAATATATCTCGAGTTATATTTATATAAT TCACTTCTTTTTTGAAGCTT
90	Env 1436-20	GCGCGGATCCGTGAATTATATAAATAAACCTCGAGCTTTATATTTATAT AATTCACTTTTTTGAAGCTT
91	Env 1437-20	GCGCGGATCCTGAATTATATAAATAAAGACTCGAGACTTTATATTTATA TAATTCATTTTTTGAAGCTT

Hairpin #	Target (NL4-3)	Oligonucleotide template (5' to 3')
92	Env 2308-20	GCGCGGATCCAGAGACTTACTCTTGATTGTTCTCGAGTACAATCAAGA GTAAGTCTCTCTTTTTTGGGAAGCTT
93	Rev (exon 2) 158-20	GCGCGGATCCTGAGAGACTTACTCTTGATTCTCGAGCAATCAAGAG TAAGTCTCTCATTTTTTGGGAAGCTT
94	Rev (exon 2) 159-20	GCGCGGATCCGAGAGACTTACTCTTGATTGACTCGAGACAATCAAGA GTAAGTCTCTCTTTTTTGGGAAGCTT
95	Nef 350-20	GCGCGGATCCCACAAGGCTACTTCCCTGATACTCGAGAATCAGGGAA GTAGCCTTGTGTTTTTGGGAAGCTT
96	Nef 351-20	GCGCGGATCCACAAGGCTACTTCCCTGATTCTCGAGCAATCAGGGA AGTAGCCTTGTGTTTTTGGGAAGCTT

*n.b.* The target positions are the co-ordinates within each gene of NL4-3.

### Oligonucleotide pairs for making synthetic inserts with offset ends

Three hairpin templates were created from standard complementary oligonucleotide pairs with offset ends as each hairpin core sequence contained an internal *Hind* III site making it incompatible with the Phi-29 extension method. These templates were made as synthetic duplexes with overhanging ends identical to those created by restriction enzyme (RE) digestion (*Bam* HI at the 5' and *Hind* III at the 3'). The coding region for each hairpin was contained within a single oligonucleotide (upper oligo: 5'-GATCC | core(20 - 21) | NCTCGAGN | core(20 - 21) | [G/A/C] | TTTTTTGGGA-3') and its complementary equivalent (lower oligo: 5'-AGCTTCCAAAAA | [G/A/C] | core(20 - 21) | NCTCGAGN | core(20 - 21) | G-3'). Both oligos were synthesized and re-suspended as previously described. 1  $\mu$ l from each was added to 98  $\mu$ l of annealing solution (10 mM Tris pH 8.0, 50 mM NaCl, 1 mM EDTA), heated to 100 °C for 5 minutes, slowly equilibrated to room temperature and diluted up to 10,000 fold for ligation. The inserts and plasmids were ligated, and used to transform electrocompetent GT116 *E.coli* (Invivogen). The following table lists the 3 hairpin templates ordered as complementary oligonucleotide pairs.

Hairpin #	Target	Oligonucleotide template (5' to 3')
6 (lower)	LTR 516-21	AGCTTTCCAAAAAGCTTAAGCCTCAATAAAGCTTTCTCGAGTAAGCTT TATTGAGGCTTAAGCG
6 (upper)	"	GATCCGCTTAAGCCTCAATAAAGCTTACTCGAGAAAGCTTTATTGAGG CTTAAGCTTTTTTGGGA
7 (lower)	LTR 522-20	AGCTTTCCAAAAAGCCTCAATAAAGCTTGCCTTGCTCGAGGAAGGC AAGCTTTATTGAGGCG
7 (upper)	"	GATCCGCCTCAATAAAGCTTGCCTTCTCGAGCAAGGCAAGCTTTATT GAGGCTTTTTTGGGA
8 (lower)	LTR 527-21	AGCTTTCCAAAAACAATAAAGCTTGCCTTGAGTGCTCTCGAGTGAC TCAAGGCAAGCTTTATTG

Hairpin #	Target	Oligonucleotide template (5' to 3')
8 (upper)	“	GATCCAATAAAGCTTGCCTTGAGTGCACTCGAGAGCACTCAAGGCAA GCTTTATTGTTTTTTGGA

### Oligonucleotide templates for the off-target control shRNA

The off-target hairpin plasmid was taken from previous work, but was originally constructed using standard complementary oligonucleotide pairs as described above. The following table lists the corresponding complementary oligonucleotide pairs, with the stem region indicated in light grey.

Hairpin #	Target	Oligonucleotide template (5' to 3')
off target (upper)	none	GATCCAAGACAGTCCAACACACGCCACCTGTCTCTCTCGAGTGAGA CAGGTGGCGTGTGTTGGACTGTCTTTTTTTTGGGA
off target (lower)	“	AGCTTTCCAAAAAAGACAGTCCAACACACGCCACCTGTCTCACTC GAGAGAGACAGGTGGCGTGTGTTGGACTGTCTTG

### shRNA plasmid sequence confirmation by loop digestion

Each construct was digested and sequenced in two reactions, one containing the forward primer, the other containing the reverse. The primers bound to the expression plasmid backbone approximately 100 bases away from the region encoding the base of the hairpin stem. Each reaction contained: 1x RE buffer (NEB-2, New England Biolabs), 1x BSA, ~ 500 ng of template plasmid, 10 pmol of sequencing primer, ~ 10 units XhoI and water to a total volume of 16  $\mu$ l, and was incubated at 37 °C for 30 - 60 min. prior to shipping, without purification, to an automated sequencing facility (Australian Genome Research Facility, AGRF).

## The gene sequences

The HIV-1 hairpins used in this project were designed to target the common laboratory strain, NL4-3 [Genbank:**AF324493**]. The gene sequences from this strain were as follows:

### LTR

tggaagggctaattcactcccaagaagacaagatatccttgatctgtggatctaccacacacaaggctactccctgattgg  
cagaactacacaccagggccaggggtcagatatccactgaccttggatggtgctacaagctagtaccagttgagccagat  
aaggtagaagaggccaataaaggagagaacaccagctgttacacctgtgagcctgcatggaatggatgacctgagag  
agaagtgttagagtggaggttgacagccgctagctttcatcacgtggcccgagagctgcatccggagtactcaagaac  
tgctgacatcgagctgtacaagggactttccgctggggactttccagggaggcgtggcctgggaggactggggagtgg  
cgagccctcagatgctgcatataagcagctgctttgctgtactgggtctctctggttagaccagatctgagcctgggagctc  
tctggctaactaggaacccactgctaagcctcaataaagcttgcttgagtgttcaagtagtgtgtgccgctgtgtgtga  
ctctggtaactagagatccctcagacccttttagtcagtggtgaaaatctctagca

### Gag

atgggtgagagcgtcggattaagcgggggagaattagataaatgggaaaaattcggtaaggccagggggaaagaa  
acaatataaactaaaacatatagatgggcaagcaggagctagaacgattcgagtaatcctggccttttagagacatcag  
aaggctgtagacaataactgggacagctacaaccatccctcagacaggatcagaagaacttagatcattatataatacaata  
gcagctctctattgtgtgcatcaaaggatagatgtaaagacaccaaggaagccttagataagatagaggaagagcaaac  
aaaagtaagaaaaaggcacagcaagcagcagctgacacaggaaacaacagccaggtcagccaaaattaccctatagtg  
cagaacctccaggggcaaatggtacatcaggccatcacctagaactttaaagcagtggttaaagtagtagaagagaag  
gctttcagcccagaagtaataacctgtttcagcattatcagaaggagccaccccacaagatttaaataccatgctaaacac  
agtggggggacatcaagcagccatgcaaatgttaaagagaccatcaatgaggaagctgcagaatgggtagattgcatc  
cagtgcatgcagggcctattgcaccaggccagatgagagaaccaaggggaagtacatagcaggaactactagaccctt  
caggaacaaataggatggatgacacataatccacctatcccagtaggagaaatctataaaagatggataatcctgggattaa  
ataaaatagtaagaatgtatagccctaccagcattctggacataagacaaggaccaaaggaaccccttagagactatgtaga  
ccgattctataaaactctaagagccgagcaagcttacaagaggtaaaaaattggatgacagaaacctgttggtccaaaat  
gcaacccagattgtaagactatftaaagcattgggaccaggagcgcactagaagaaatgatgacagcatgtcagggga  
gtggggggaccggccataaagcaagagtttggctgaagcaatgagccaagtaacaaatccagctaccataatgataca  
gaaaggcaatfttaggaaccaaagaaagactgttaagtttcaattgtggcaagaaggccacatagccaaaattgcagg  
gcccctaggaaaaaggctgttggaatgtgaaaggaaggacaccaaataagattgtactgagagacaggcctaattttt  
tagggaagatctggcctcccacaaggaagccagggaaattttctcagagcagaccagagccaacagccccaccaga  
agagagcttcaggttggggaagagacaacaactccctctcagaagcaggagccgatagacaaggaactgtatcctttagc  
ttccctcagatcactctttggcagcagcccctcgtcacaataa

### Pol

tttttagggaagatctggcctcccacaaggaaggccagggaaattttctcagagcagaccagagccaacagccccacc  
agaagagagcttcaggttggggaagagacaacaactccctctcagaagcaggagccgatagacaaggaactgtatccttt

agctccctcagatcactcttggcagcgacccctcgtcacaataagataggggggcaattaaaggaagctctattagatac  
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 actaacagaagtagtaccactaacagaagaagcagagctagaactggcagaaaacaggagattctaaagaaccggtg  
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 agaggcagtagcaaaaaatagccacagaaagcatagtaatatgggaaagactcctaaattaaattaccatacaaaagga  
 aacatgggaagcatggtggacagagattggcaagccactggattcctgagtgagggttgtcaatccccctcccttagtga  
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## Vif

Atggaaaacagatggcaggtgatgattgtgtggcaagtagacaggttaggattaacacatggaaaagattagtaaacac  
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 g

**Vpr**

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 tactcgacagagggagagcaagaaatggagccagtagatcctag

**Tat (x1)**

atggagccagtagatcctagactagagccctggaagcatccaggaagtcagcctaaaactgcttgaccaattgctattgtaa  
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**Rev (x1)**

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**Vpu**

atgcaacctataatagtagcaatagtagcattagtagtagcaataataatagcaatagttgtgtggtccatagtaatcatagaata  
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 tgaaggagaagatcagcacttgtggagatgggggtggaaatggggcaccatgctccttgggatattgatgatctgtag

**Env**

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**Tat (x2)**

accacctcccaatcccgaggggacccgacaggcccgaaggaatag

**Rev (x2)**

accacctcccaatcccgaggggacccgacaggcccgaaggaatagaagaagaaggtggagagagagacagagaca  
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**Nef**

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