

SUPPORTING ONLINE MATERIAL FOR

**Poly(ADP-ribose) Binding to Heterogeneous Nuclear Ribonucleoproteins Modulates
Alternative Splicing Activity**

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This PDF file includes:

Figures 1 to 8.

Figure 1. The expression level of Squid/hrp40 is strongly reduced in the *Squid*/*hrp40* mutant, indicating the specificity of the Squid/hrp40 antibody (8D2). **A.** 10 µg total proteins from the third-instar larvae of the homozygous *Squid*/*hrp40* mutant (*sqd*^{j6E3}) and the wild-type (y,w) fly was immunoblotted with the Squid/hrp40 antibody (8D2). The residual Squid/hrp40 present in the *Squid*/*hrp40* mutant came from the maternal expression of the *Squid*/*hrp40* gene. **B.** The same blot as in (A) was stripped and probed with mouse anti-alpha-tubulin (Sigma) to show the equal loading.

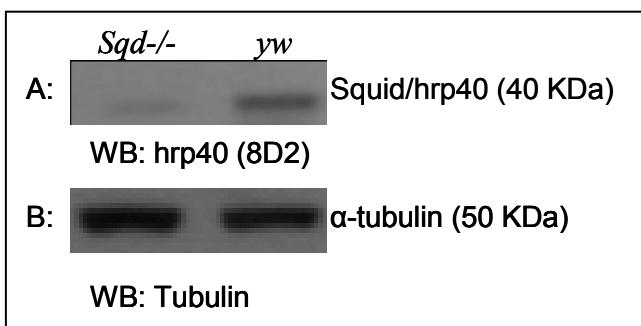
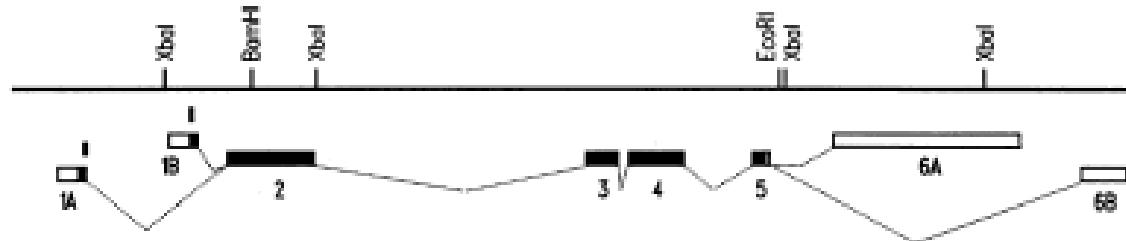
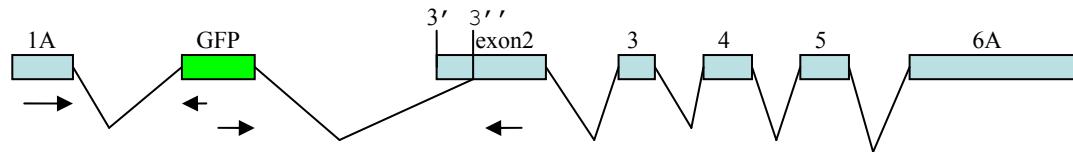


Figure 2. The transcript structure and expression of the *hrp38*:GFP fusion gene.

A. The transcript structure of the *Hrb98DE/hrp38* gene.



B. *Hrb98DE/hrp38*:GFP fusion gene.



GFP was inserted between the exon1A and exon2 of the *Hrb98DE/hrp38* gene. Sequencing the RT-PCR products using indicated primers (arrow) showed that GFP was also spliced in-frame to exon1A and exon2 using the 3'-most (3'') splice site, producing the Hrb98DE-PE:GFP

C. The amino acid sequence of Hrp38-PE:GFP fusion protein:

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MGGHDNWNNNGQNEEQDIHHHHHHGVSKGEELFTGVVPILVELGDVNGHKFVSSGEGEGDATYGKLTLKFI
CTTGKLPVPWPTLVTTLTYGVQCFSRYPDHMKQHDFKSAMPEGYVQERTIFFKDDGNYKTRAEVKFEGDT
LVNRIELKGIDFKEDGNILGHKLEYNYNSHNVYIMADKQKNGIKVNFKIRHNIEDGSVQLAHYQQNTPIGD
GPVLLPDNHYLSTQSALSKDPNEKRDHMVILEFVTAAGITLGMDELYKSTLEDSITEPEHMRKLFIGGLDY
RTTDENLKAHFEKGWGNIVDVVMKDPRTKRSRGFGFITYSHSSMIDEAQKSRPHKIDGRVVEPKRAVPRQD
IDSPNAGATVKKLFVGALKDDHDEQSIRDYFQHFGNIVDINIVIDKETGKKRGFAFVEFDDYDPVDKVVLQ
KQHQLNGKMVDVKALPKQNDQGGGGGGRRGGPGGRAGGNRGNMGGGGNYGNQNGGNWNNGNNWGNRGGN
DNWGNNSFGGGGGGGGYGGGNNSWGNNNPWDNGNGGNFGGGNNWNGNDFGGYQQNYGGPQRGGGN
FNNNRMQPYQGGGGFKAGGGNQGNYGGNNQGFNNNGNNRRY .

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The amino acid sequence of hrp38-PE:GFP fusion protein was derived from the RT-PCR sequences obtained in Figure B and the hrp38-PE sequence in the flybase. The 6X His-GFP insertion is highlighted in green. The pADPr-binding motif is highlighted in yellow. Based on our sequences, it appears that the pPGC construct (ATC-6XHis-GFP-TCGACACTCGAG) (Morin *et al.*, 2001) was inserted into the intron between exon 1A and exon 2 of the *Hrb98DE/hrp38* gene in the *ZCL588* strain.

D. The expression of *hrp38*:GFP fusion protein. Upper panel: 50 µg total proteins from the third-instar larvae of the indicated genotypes was subjected to immunoblotting analysis using anti-GFP antibody, showing that about 55 kDa *hrp38*:GFP fusion protein was detected from the *ZCL588* (*hrp38*:GFP) and *PARP1-DsRed; hrp38:GFP* fly strains, respectively, but not from the wild-type strain (*yw*). Lower panel: The same blot as shown in the upper panel was stripped and probed with the mouse actin antibody (Millipore) as the loading control. It appears that the *ZCL588* (*hrp38*:GFP) strain had an expression level equal to the *hrp38*:GFP fusion protein with the *PARP1-DsRed; hrp38:GFP* strain after normalization with the loading control.

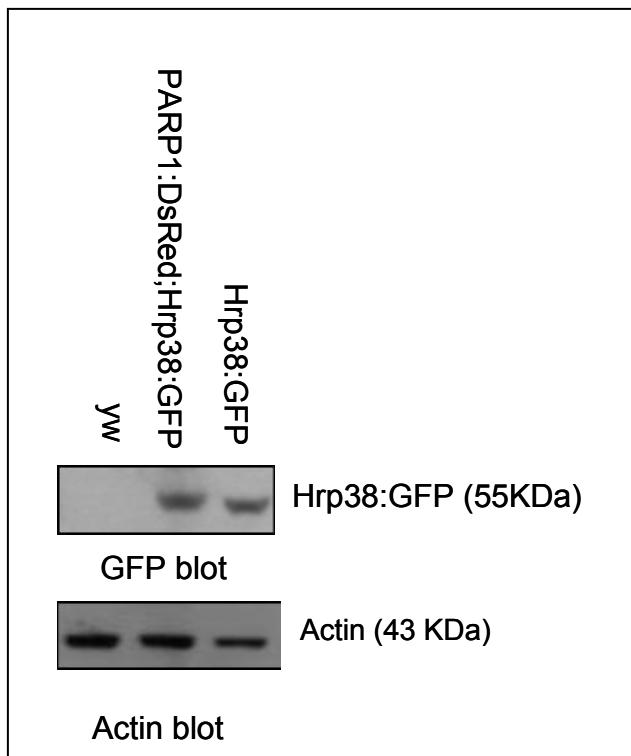
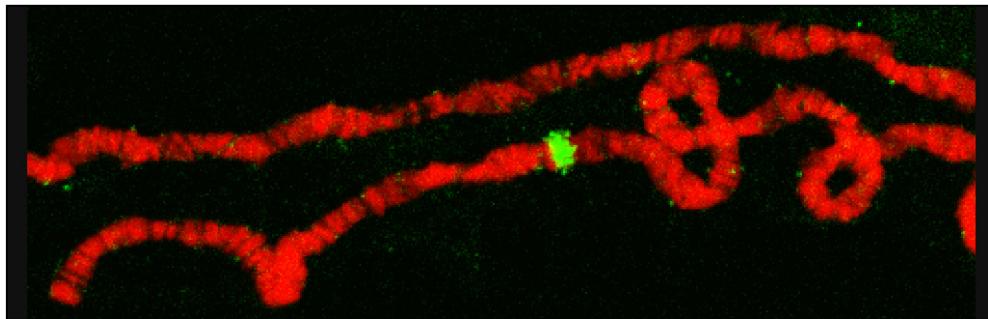


Figure 3.

A. The accumulation of hrp38:GFP in the 93D puffs after heat-shock treatment.

Drosophila polytene chromosomes of the hrp38:GFP line were immunostained with the anti-GFP antibody: Hrp38:GFP, green; DNA, red.



B. There is no significant difference in the expression levels of hrp38:GFP and Squid between the hrp38:GFP and *Parg*; hrp38:GFP lines after heat-shock treatment. After one hour of heat-shock treatment, 30 µg total proteins from the third-instar larvae of the hrp38: GFP and *Parg*; hrp38:GFP lines was immunoblotted with anti-GFP and anti-Squid antibody (8D2), respectively. The blot was stripped and probed with mouse anti-actin antibody for loading control. HS: heat-shock treatment.

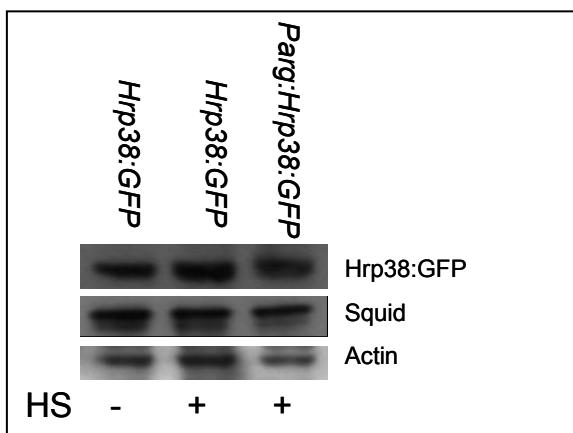
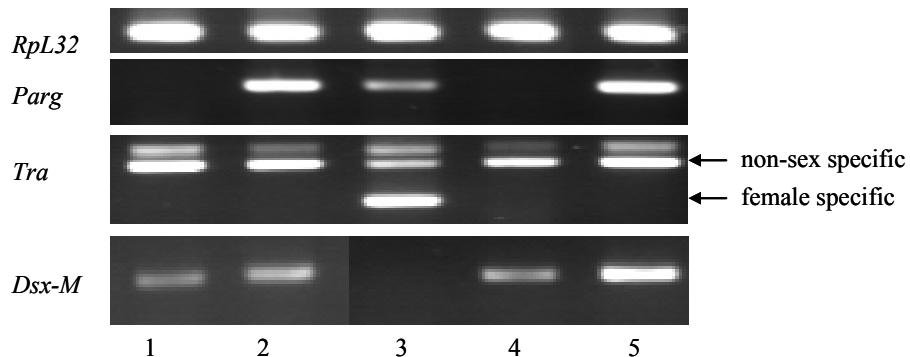


Figure 4.

A. RT-PCR confirmed the genotypes for the real-time RT-PCR assay in Figure 8B in the main text and Figure 4B in the Supplemental Materials. Lane 1. *Parg* (-/-), the third-instar larvae male; 2. *yw*, the third-instar larvae male (control); 3. *Parg* (+/-), the third-instar female; 4. *Parg* (-/-), pharate male; 5. *yw* pharate male (control).



B. There is no significant difference in the *RpL32* expression level relative to the wild type among the indicated genotypes and treatment in Figure 7C in the main text. The *RpL32* expression level as an internal control in Figure 8B was measured by real-time RT-PCR. The error bar represents the standard deviation from two independent experiments. Lane 1. *Parg* (-/-), the third-instar larvae male; 2. *yw*, the third-instar larvae male (control); 3. *Parg* (+/-), the third-instar female; 4. *Parg* (-/-), pharate male; 5. *yw* pharate male (control).

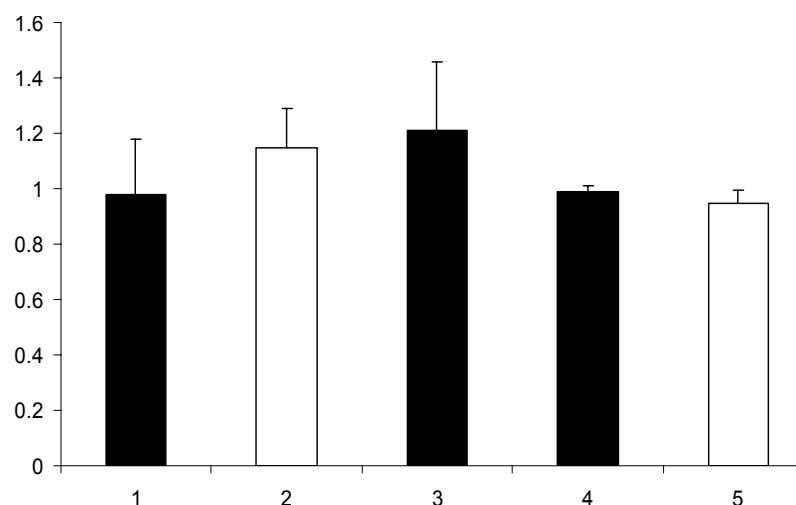
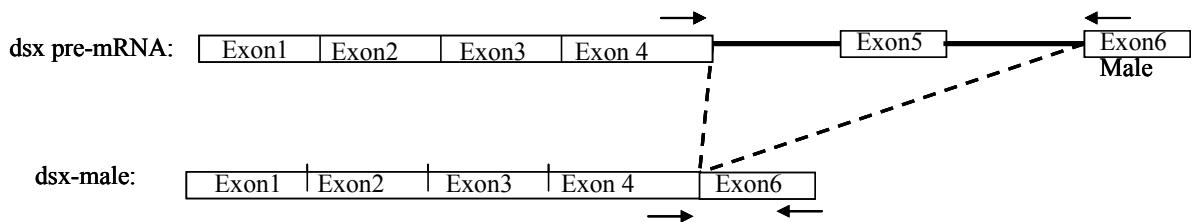


Figure 5. Inhibition of alternative splicing of the *dsx* gene by PARG loss-of-function.

A. The structures of *dsx* pre-mRNA (unspliced) and *dsx*-male (spliced) transcripts in the *doublesex* (*dsx*) locus. The primers (arrows) specific for the *dsx*-male transcript were used for *dsx*-male quantification. **B.** The *dsx*-male expression level relative to *RpL32* in the indicated genotypes measured by real-time RT-PCR. The error bar represents the standard deviation from two independent experiments. Lane 1. *Parg* (-/-), the third-instar larvae male; 2. *yw*, the third-instar larvae male (control); 3. *Parg* (+/-), the third-instar female; 4. *Parg* (-/-), pharate male; 5. *yw* pharate male (control). **: P<0.01.

A.



B.

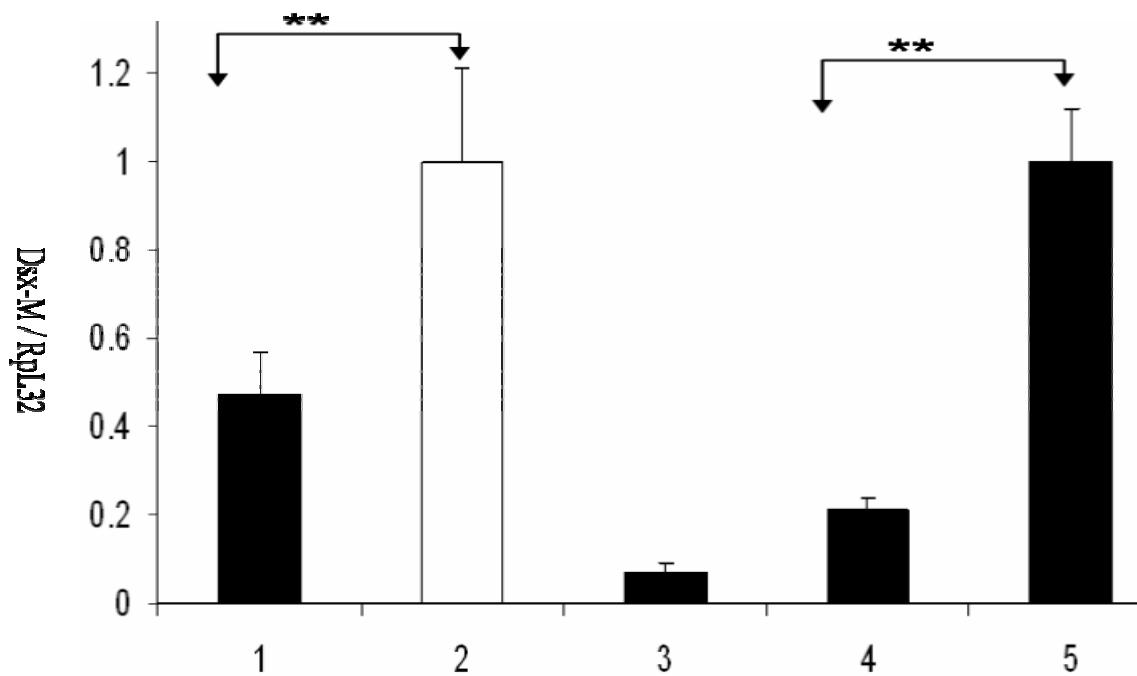


Figure 6. There is no significant difference in the *RpL32* expression level relative to the wild type among the indicated genotypes and treatment in Figure 8D in the main text. The error bar represents the standard deviation from two independent experiments. The expression level of the *RpL32* expression level as an internal control was measured by real-time RT-PCR.

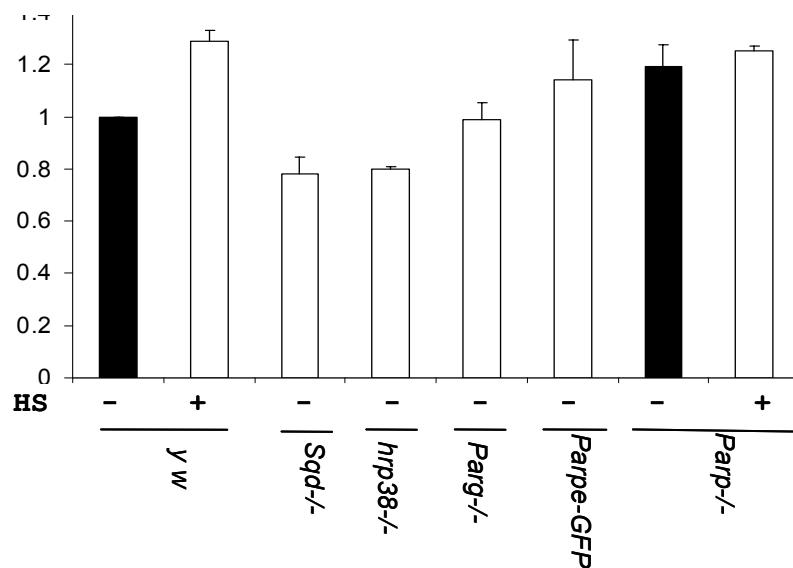


Figure 7. The structure of Hrsr-Omega-RC. Introns in lower case are marked in blue; G triple and quarters in exon 2 are marked in red.

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1 ACTCTCAAAT GAAAAGTGT CAAGTGCATT CAAAGTGAAG CTGAAAAAAT AACCAAGTTAA
61 AAATAGTACA AAGAAATTTT CTTTCTTGCA ATTGCAAGC AGTAGCTACA ACCAAAAATG
121 GAAAAGTGT AAAATCGTGT CCCAGCAGAC GAGCAGCAGC AGTACGAGTA TTGCAAAATG
181 CAGGGGCAAG GGCCCACGTA GTATTTTCC ACGTCGGGCA TTTAATGCTC TCGAGTTGGA
241 AACAAATGAAA CCATACGCAA ACCCCCTGGA AAAGATGTGA TTAGTCATCG ATTTGCTGG
301 TCAGCGTCGG GGAATTCGC AATGCAGCAG GCAGTTTCT CAAATTTGGC TAGAAAGTGA
361 CCCACTAGGC AGTCTGAGGC AGTTATCCAG GATGTAAGGA TGTGTGCAGT ACTCTCTGTG
421 TCGAATGGCG ATGGCCCTGA CCTGCGGATT TTGAAAGATT GAAATAGGAA GCCAGgtata
481 catacacagc aacgaatgca gcgtgttcta tattcttaat tcgtaagcca aacctatgtt
541 taagacgttag tttatataac cgtacatatg tatgtagcga aacacgtttt caaacgtcgc
601 atcgaagcca tttcttctgc agccattgca atttcatcg tttgcgtaat cgcaaattgt
661 aaatgtgtcc ctactacttc acatttgtgt gtatccaagt cgaaagtctt cgaactattc
721 tcgtgagcaa ccccccggatc gaaaggccac caactggctc gttctctctc tccagaaaca
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841 tatacactca gagacacccc aatccccac catacatatg gcgagatgtg tgaatgtatc
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1201 GTTGATATCG ATATCGATCC GTGAAAAGTC GATAACCCTGC GCAAGCATGG GGCGGCATAT
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1681 CACACCAACC ATGTGTCACT TATGTTCTTA AATACCAGAA ACTGTTAAA TTTTGTTCT
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1801 CAAAAATAGA AAAAATGAAT AACAAAAGAG CTCTATAACA GAAAAGCCAC AGAAAAAGTG
1861 AATAAAAATT CAAAAATTCA AAAAATAAAT TTCGAAACAT TT

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Figure 8. The structure of the *Ddc*(*dopa decarboxylase*) gene. Introns in lower case are marked in blue; G triple and quarters in intron 1 and 2 are marked in red.

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1 TCAGTTAAGA GGAGAACGCC AAGCGCACAG CAATCAGCAC CGAAATATCA GCATCGAAAT
61 ATCAGCAAAT AAATATTAGC TGTTCTAAC CAGGAGGGCA AACTGAACCTT GGAGCAAAGA
121 TTTAGTCGG AACGGAAGTA AAGCTCGGCA ACAAGTGCAA ACAATTAAAAA GCAGTTAACT
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841 GTTCATATCA TTGCAAAAGT CAAACGAAAA GTAAATCTCT GAAATGAGCC ACATACCCAT
901 TAGTAACACA ATTCCAACAA AACAAACTGA TGGTAAATGGT AAAGCTAACAA TTTGCCCGGA
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2341 CAACTCGTAT CCAGCGATCG TTGCGGACAT GCTGAGTGGA GCGATTGCT GCATCGGATT
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