

Supporting Information

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SI Text

Transfection. A GAL4-thymidine kinase-luciferase reporter (GAL4-TK-Luc) was introduced into HeLa cells along with the indicated amounts of plasmids by using LipofectAmine LTX reagent (Invitrogen). The total mass of DNA transfected was normalized by the addition of the appropriate vector plasmid. The extracts were prepared 40–48 h after transfection. A renilla luciferase expression vector (Dual-Luciferase Reporter Assay System; Promega) was cotransfected to adjust for variation in transfection efficiency, and luciferase assays were performed following the manufacturer's instructions (Promega).

Oligonucleotides for Mouse *Lsd1* shRNA. The top strand was TCGAGAAAAACACAAGGAAAGCTAGAAGATCTCTTGAATCTTCTAGCTTTCCTTGTTGGGG; the bottom strand was GATCCCCCACAAGGAAAGCTAGAAGATTCAAGAGATCTTCTAGCTTTCCTTGTTTTC. They were designed by using the Oligoengine program and cloned into the pSUPER-retro vector (Oligoengine). The retrovirus generation and infection were performed as described (1). Briefly, 5×10^5 exponentially-growing MEL or ES cells were cocultured with virus particles in the presence of 4 μg of Polybrene per mL followed by puromycin selection (1 $\mu\text{g}/\text{mL}$) for 2 weeks.

Plasmid Constructs. LSD1 cDNA was a kind gift from Yang Shi. The C-terminal deleted (amino acids 1–427) LSD1 cDNA were

cloned as described (2). A variety of pGEX-TAL1 constructs, pcDNA-TAL1, pGAL4-TAL1, and GAL4-TK-luciferase reporter plasmids were as described (1). Plasmid expressing GST-TAL1^{142–185} was constructed by subcloning a TAL1 PCR fragment that corresponded to amino acids 142–185 of TAL1 protein into the pGEX-5X-1 vector (GE Healthcare).

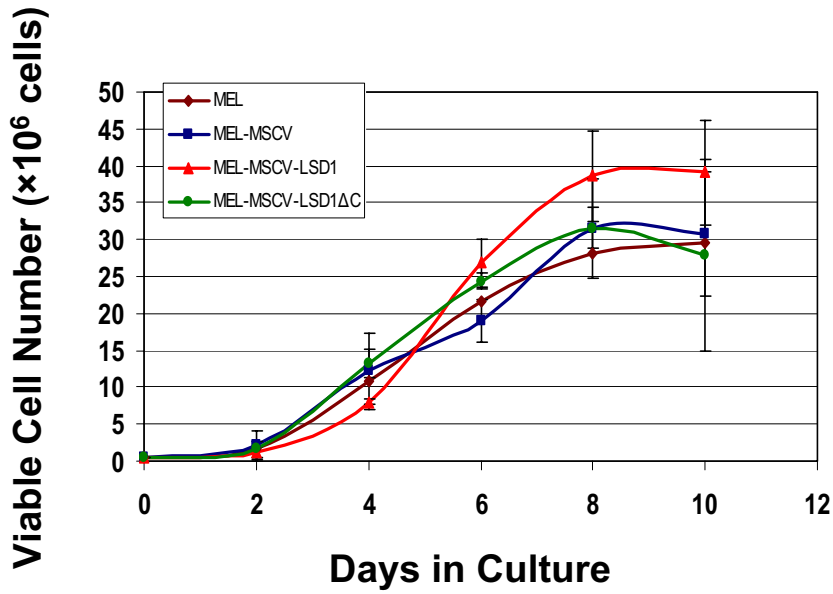
Primers for Quantitative RT-PCR. The primers were: *P4.2* gene, forward TCCCAAACAACCCTCAACCGTC, reverse TGGT-ATGAAACATCTGAACACCCC; *β -globin* gene, forward CACTTTGCCAGCCTCAGTG, reverse GGTTTAGTGGTACTTGTGAGCC; *GATA-1* gene, forward CAGGGATCCCATGGATTTTCTGGTC, reverse TCCACAGTTCACACACTCTCTGGC; *GATA-2* gene, forward TGCAACACACCACCCGATACC, reverse CAATTTGCACAACAGGTGCCC; *β -actin* gene, forward GTGGGCCGCTCTAGGCACCA, reverse: TG-GCCTTAGGGTGCCAGGGGG.

The Primers for Quantitative ChIP Assay. The primers were: P4.2 gene locus, –6 Kb, forward CTAAGCGGCATCATGAGAGA, reverse CTCCCTCCCTGCTGCTATCT; Promoter, forward ACAGATGGTTTCTGACATGC, reverse ACTCTGAACCTCCTGCCTCA; 3' UTR, forward AGAACCTGACCGGCTACAGA, reverse: AGACGGTTGAGGGTTGTTTG.

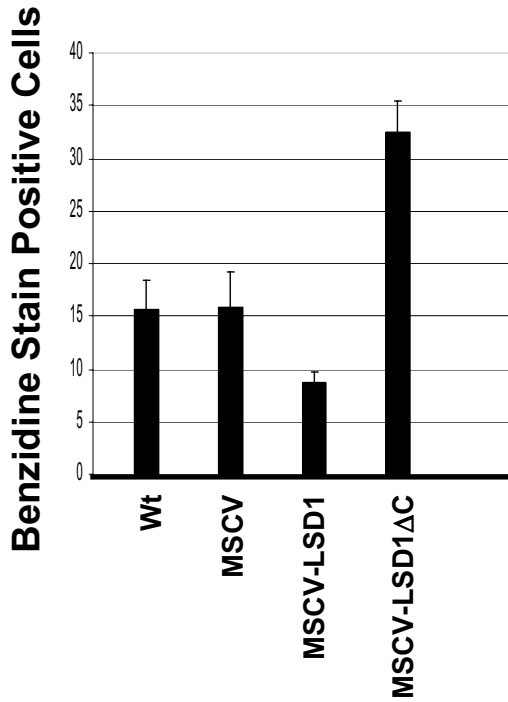
1. Huang S, Brandt SJ (2000) mSin3A regulates murine erythroleukemia cell differentiation through association with the TAL1 (or SCL) transcription factor. *Mol Cell Biol* 20:2248–2259.

2. Shi Y, et al. (2004) Histone demethylation mediated by the nuclear amine oxidase homolog LSD1. *Cell* 119:941–953.

A



B



C

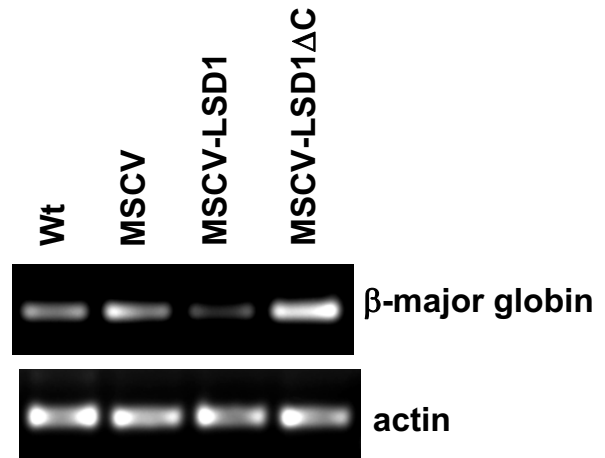


Fig. 54. The role of LSD1 in DMSO-induced MEL cell differentiation. (A) MEL cells were stably infected with retrovirus encoding the vector control, LSD1, and LAS1 Δ C mutant. A total of 5×10^6 cells were inoculated into medium. The number of viable cells was counted in 24-h intervals for 10 days. The data were collected from 3 independent experiments. (B) After 3-day DMSO induction, the cells were collected for analysis of hemoglobin expression by using benzidine staining. (C) Total RNAs from transduced cells incubated with DMSO for 3 days were prepared and analyzed for RT-PCR.