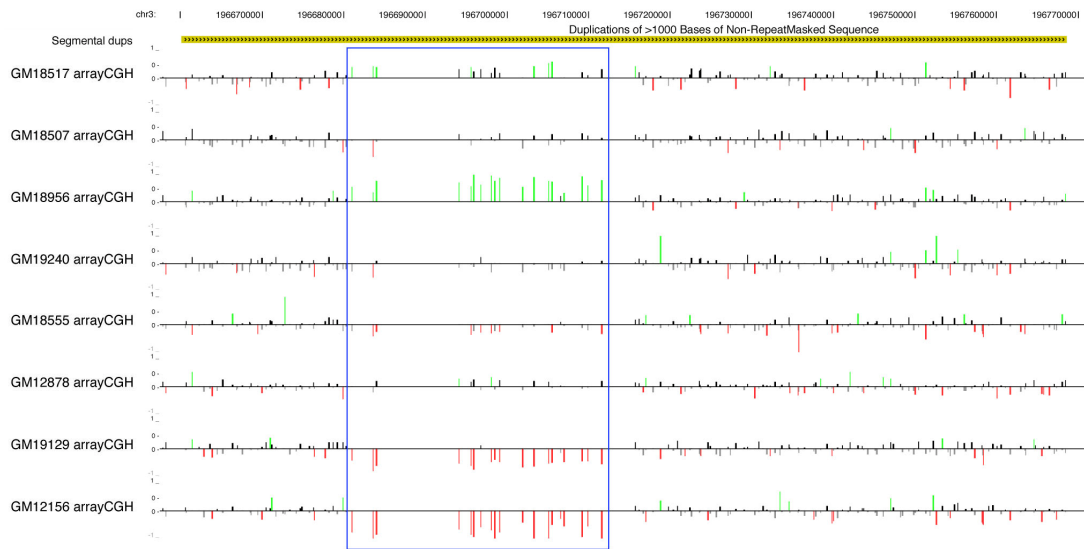
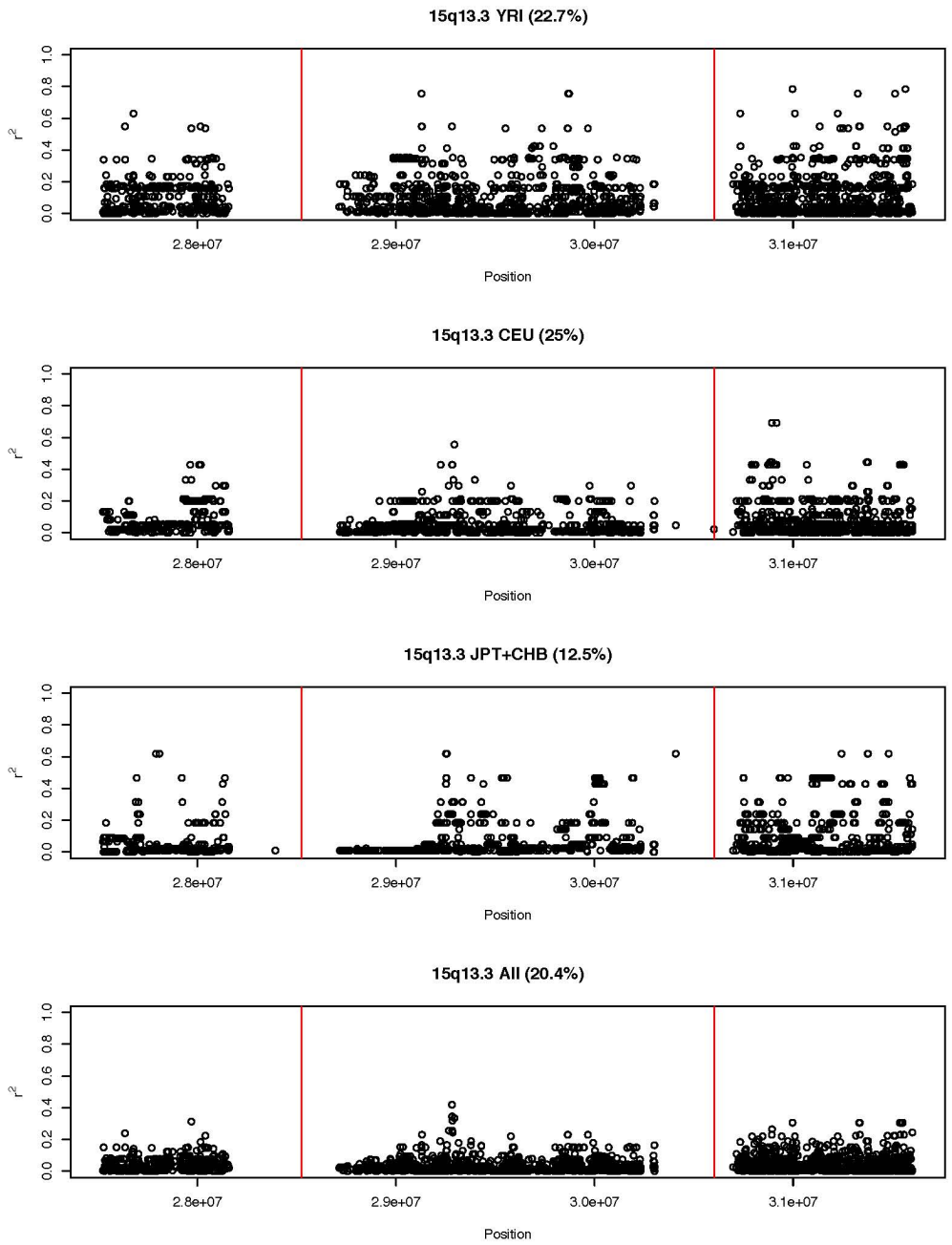
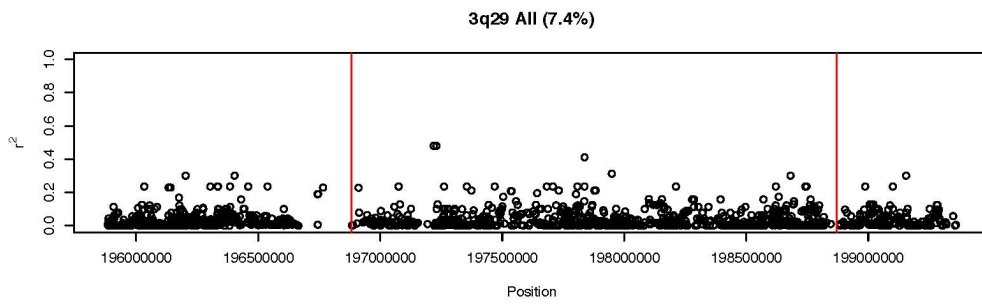
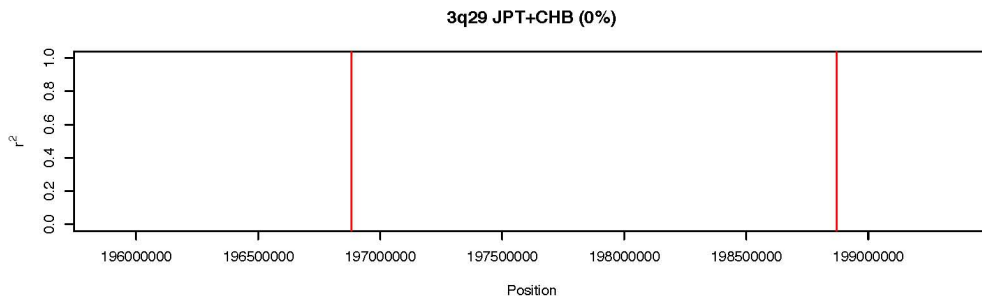
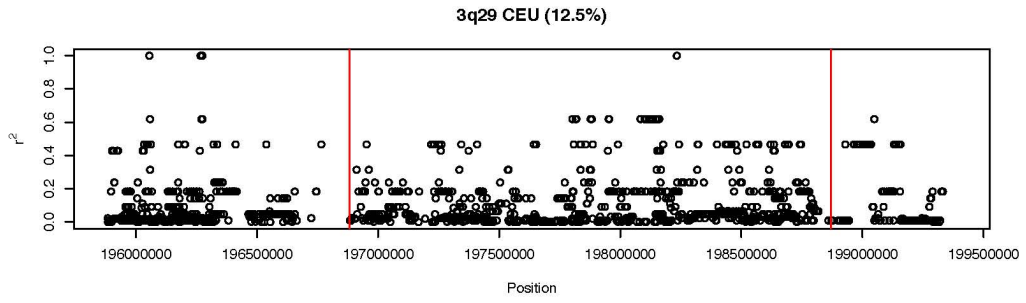
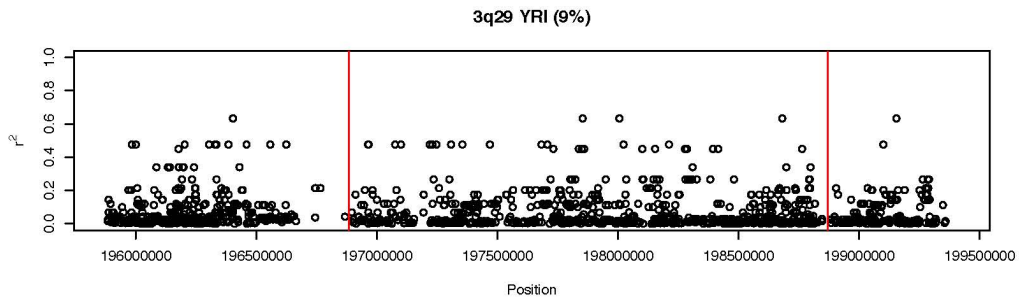


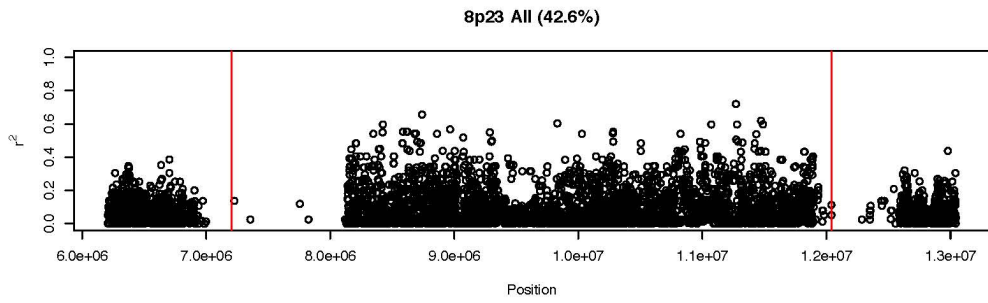
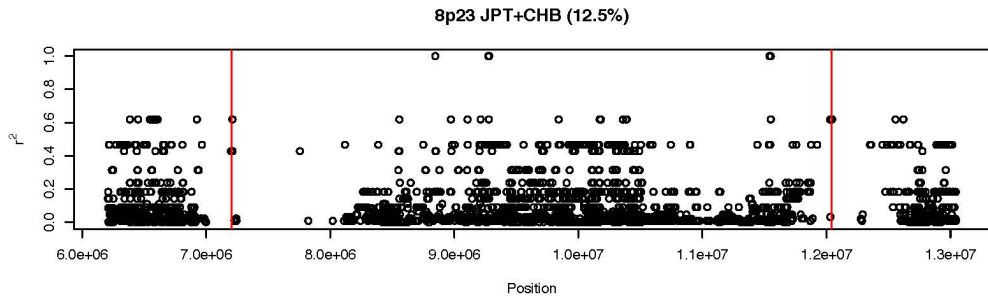
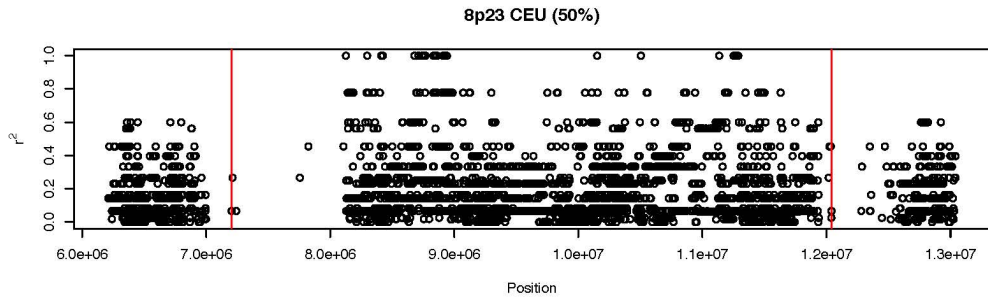
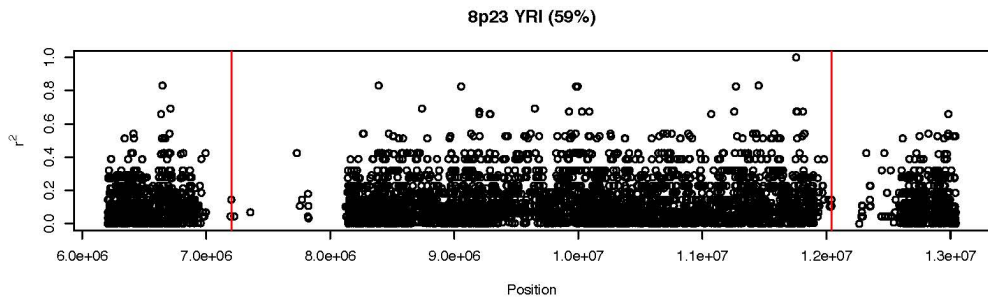
Supplementary Figure 1: Oligonucleotide arrayCGH results for the 17q21.31 inversion distal breakpoint region. Individuals GM12156 (heterozygous H1/H2) and GM12878 (homozygous H1/H1) show a 75-kbp extended duplication (in the blue rectangle) compared to the other individuals. On the right, interphase FISH validation using probe G248P85784G12 (red) mapping in the polymorphic region (chr17:41545746-41586094) and probe G248P8967C6 (green) mapping in the single copy region (chr17:39477456-39515250).

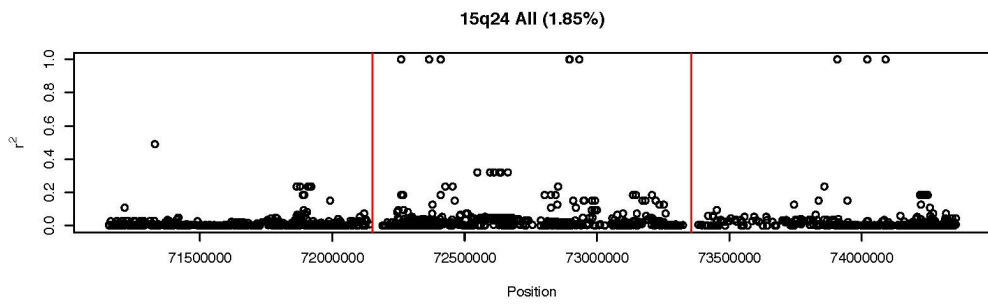
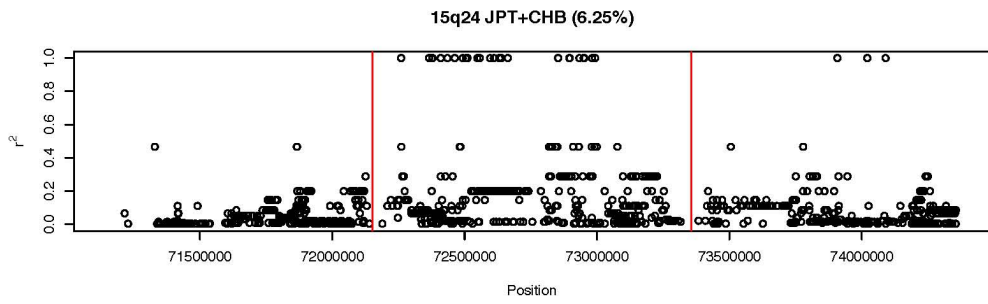
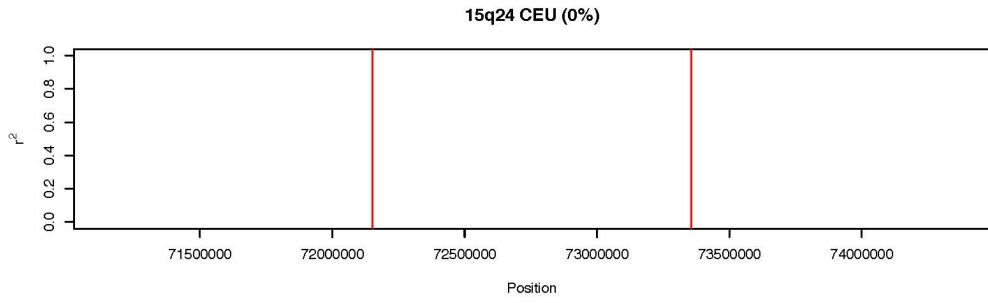
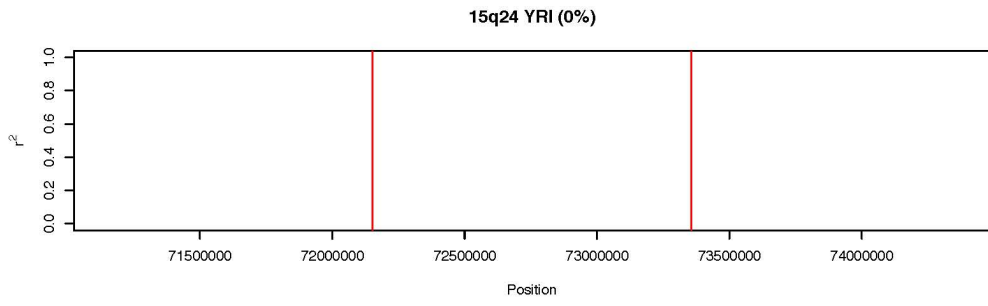


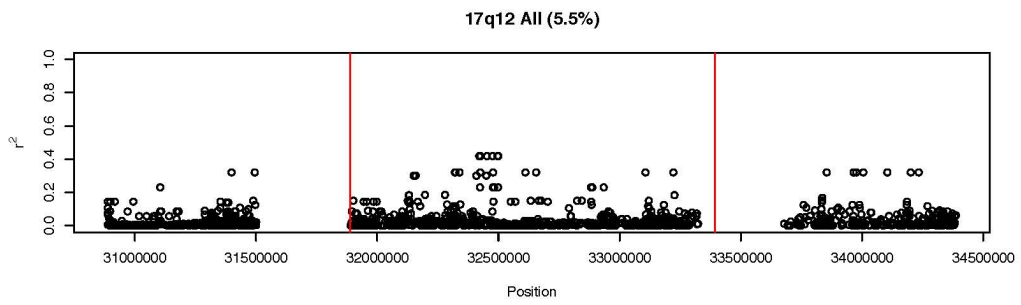
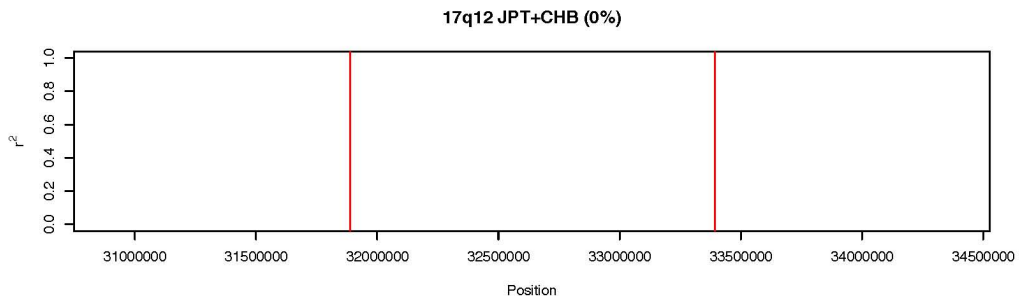
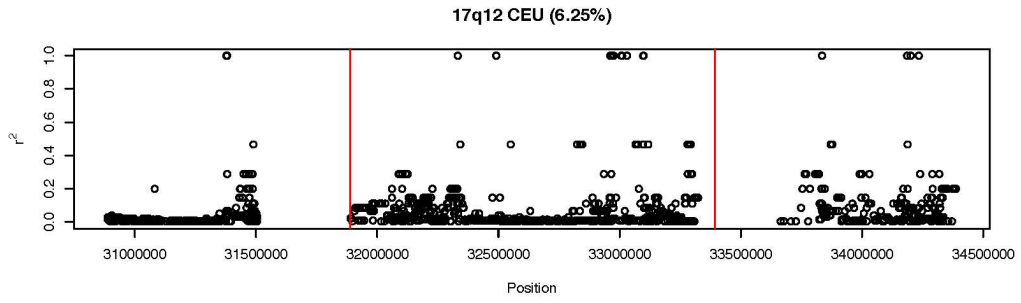
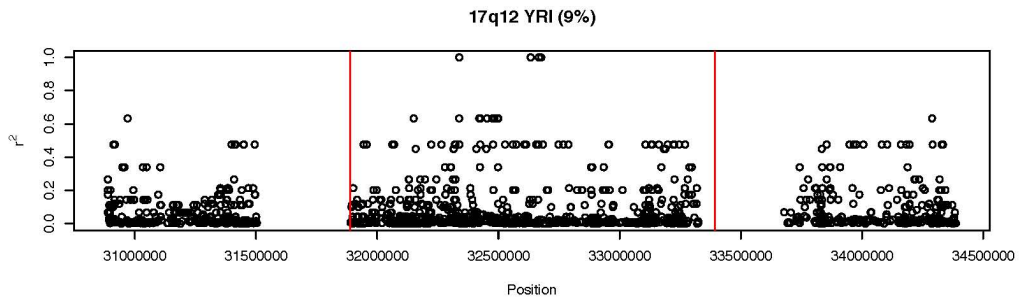
Supplementary Figure 2: Oligonucleotide arrayCGH results show that individuals GM19129 and GM12156, both carriers for the inverted allele, show a 25-kbp copy-number loss within the duplicated sequence at proximal breakpoint of the 3q29 inversion.

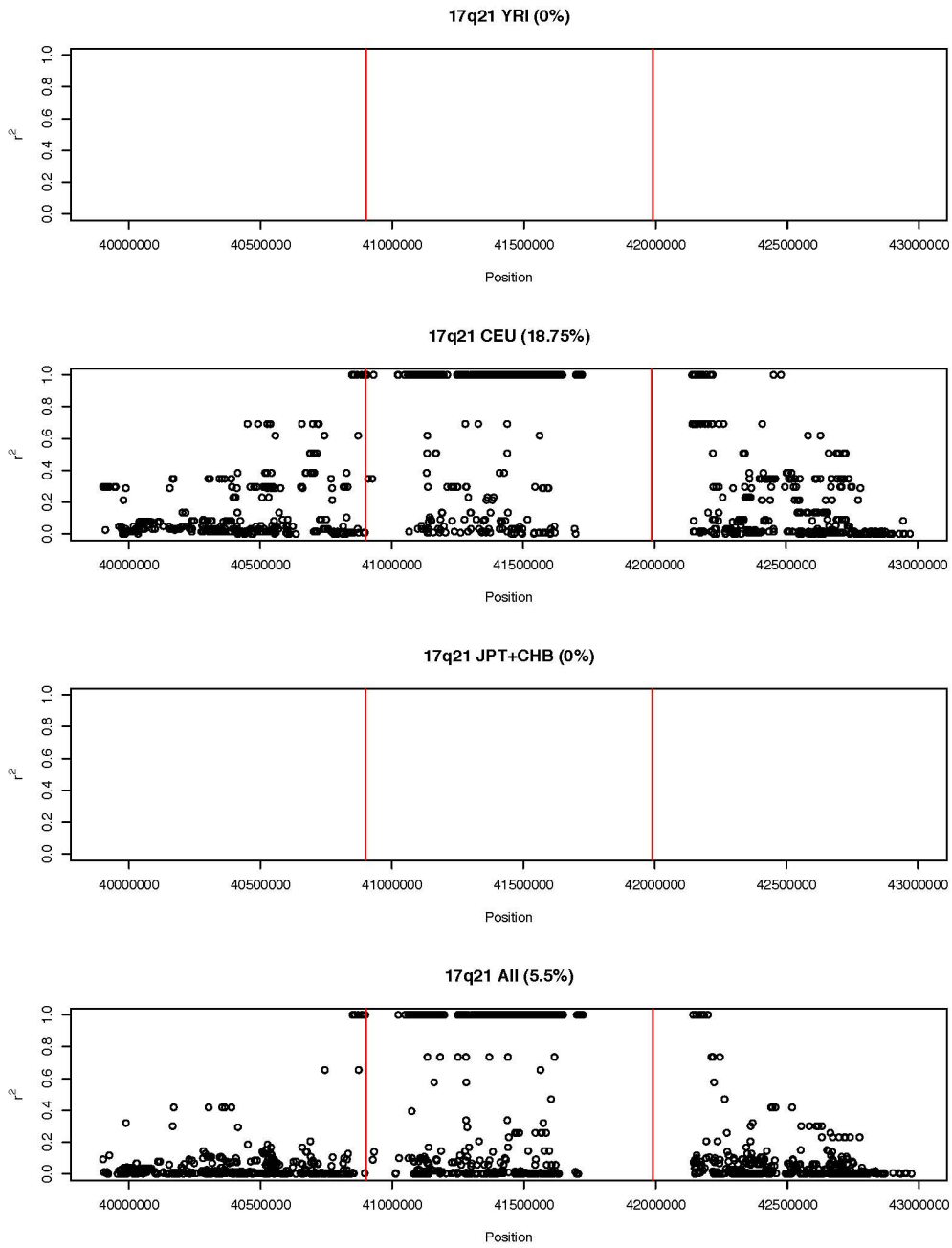




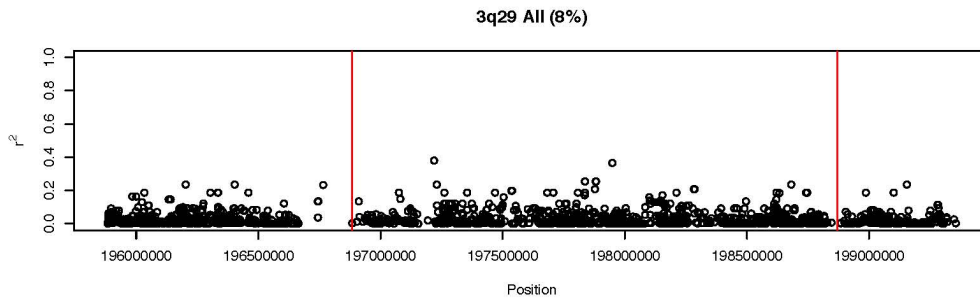
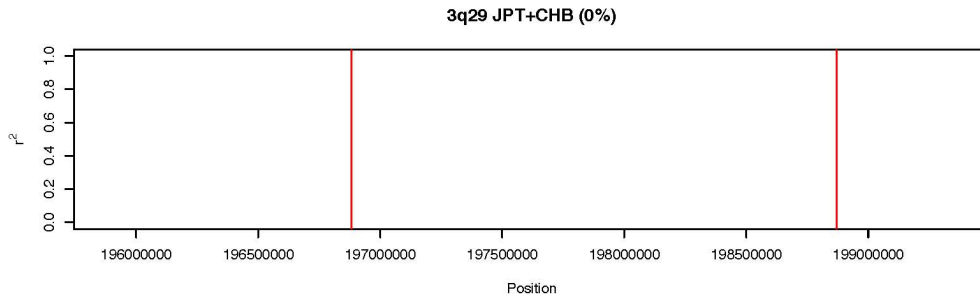
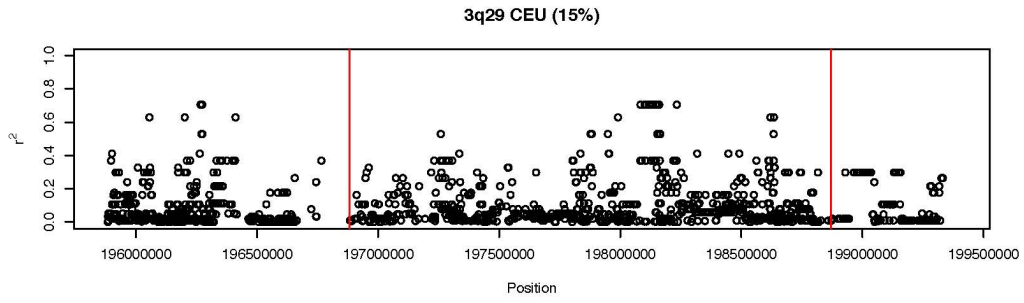
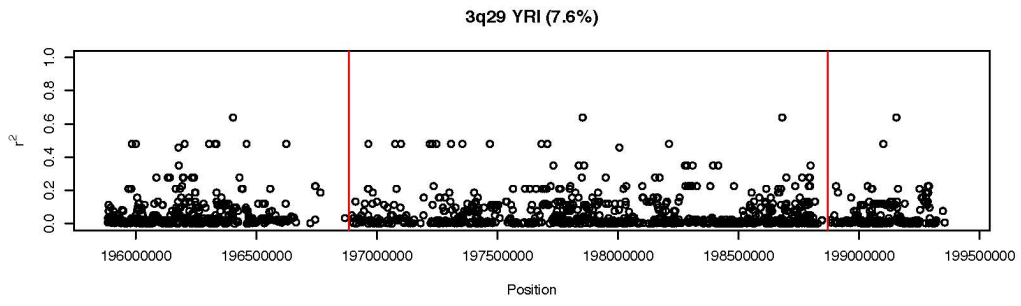


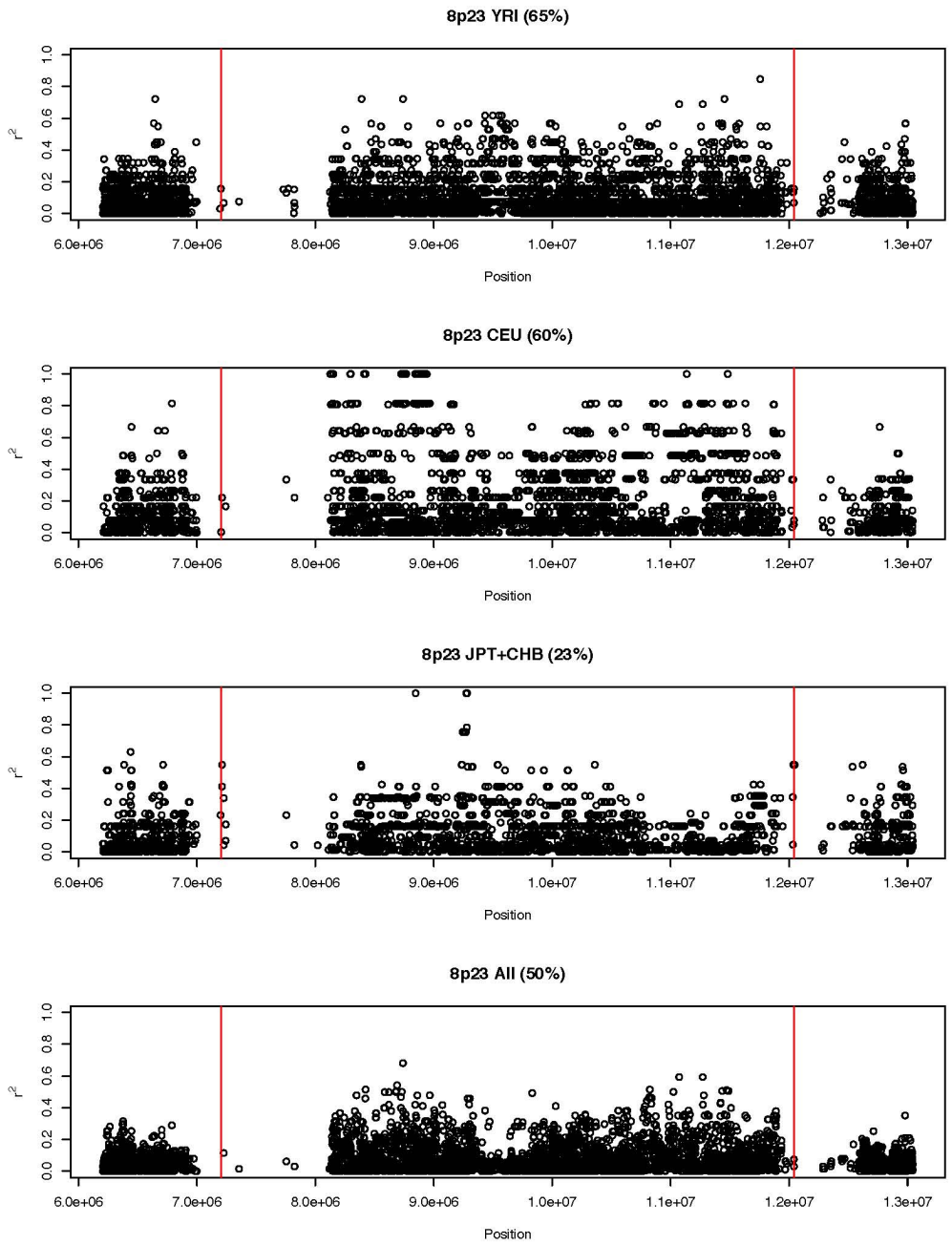


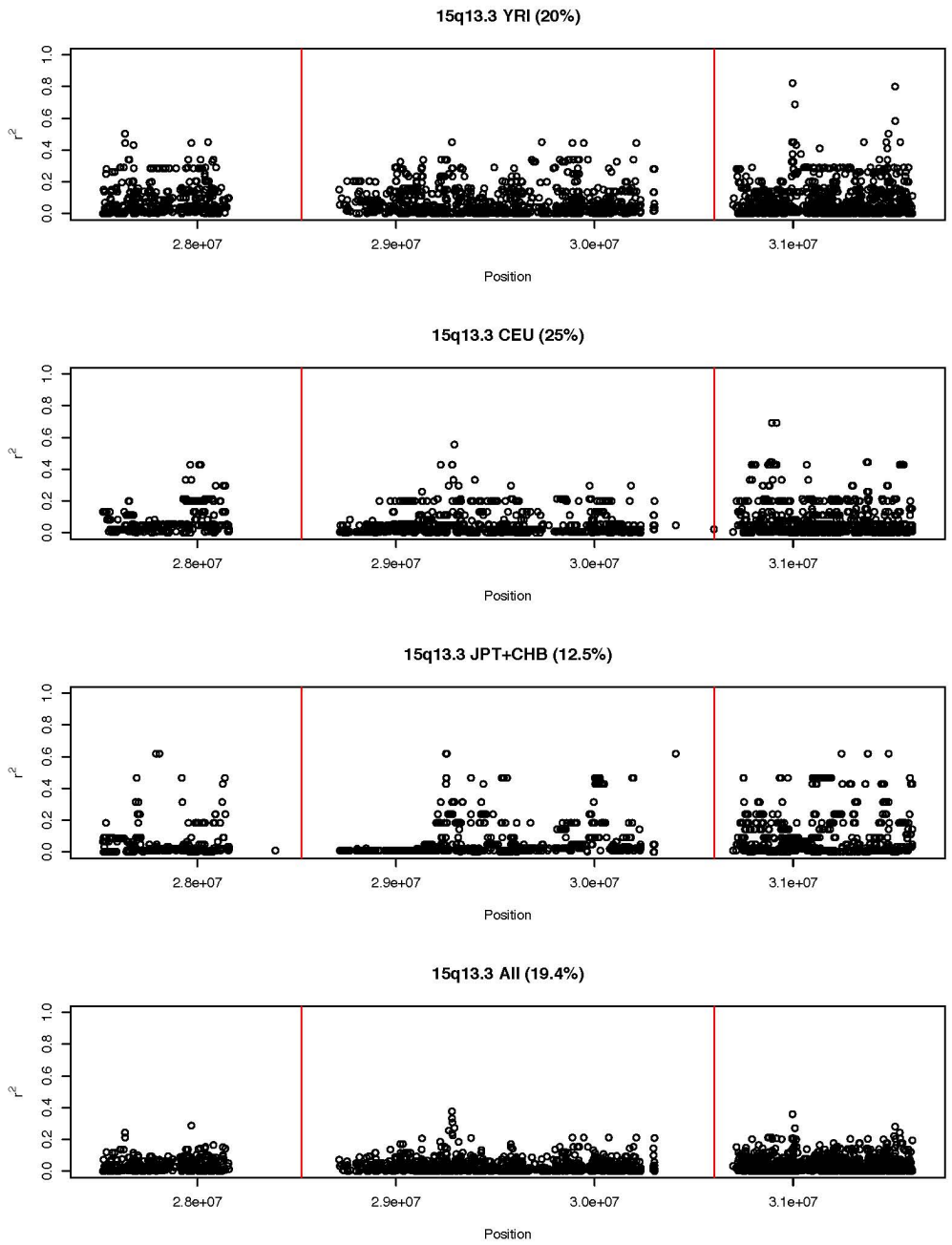


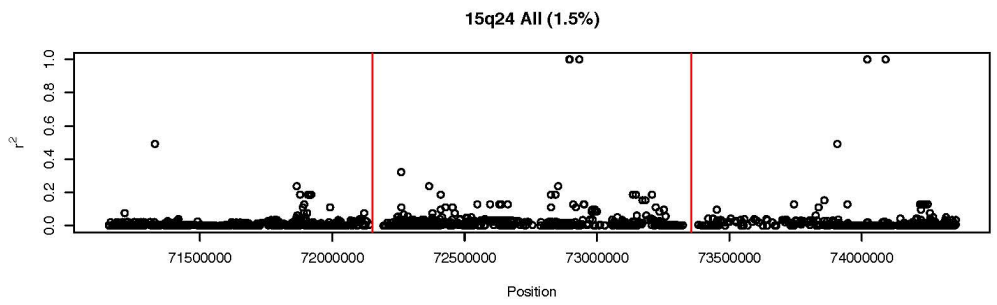
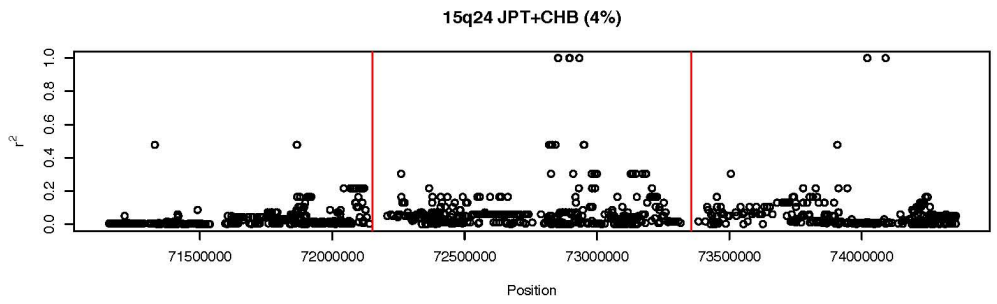
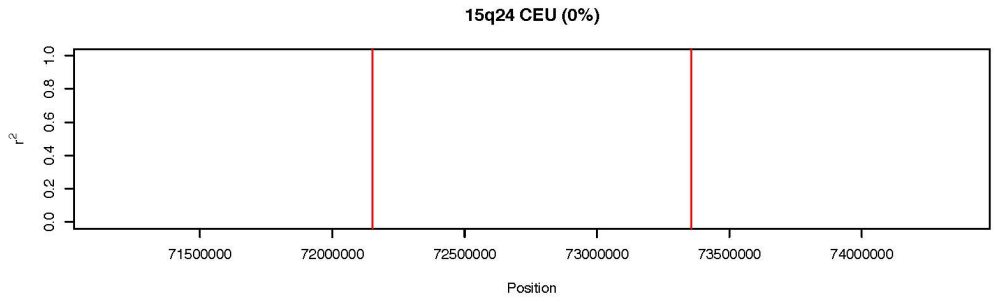
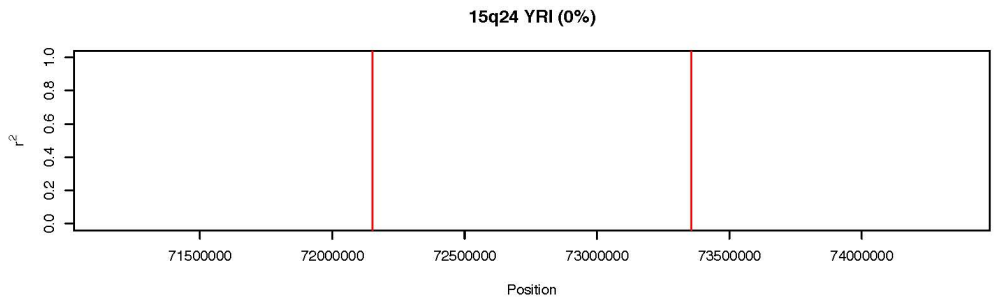


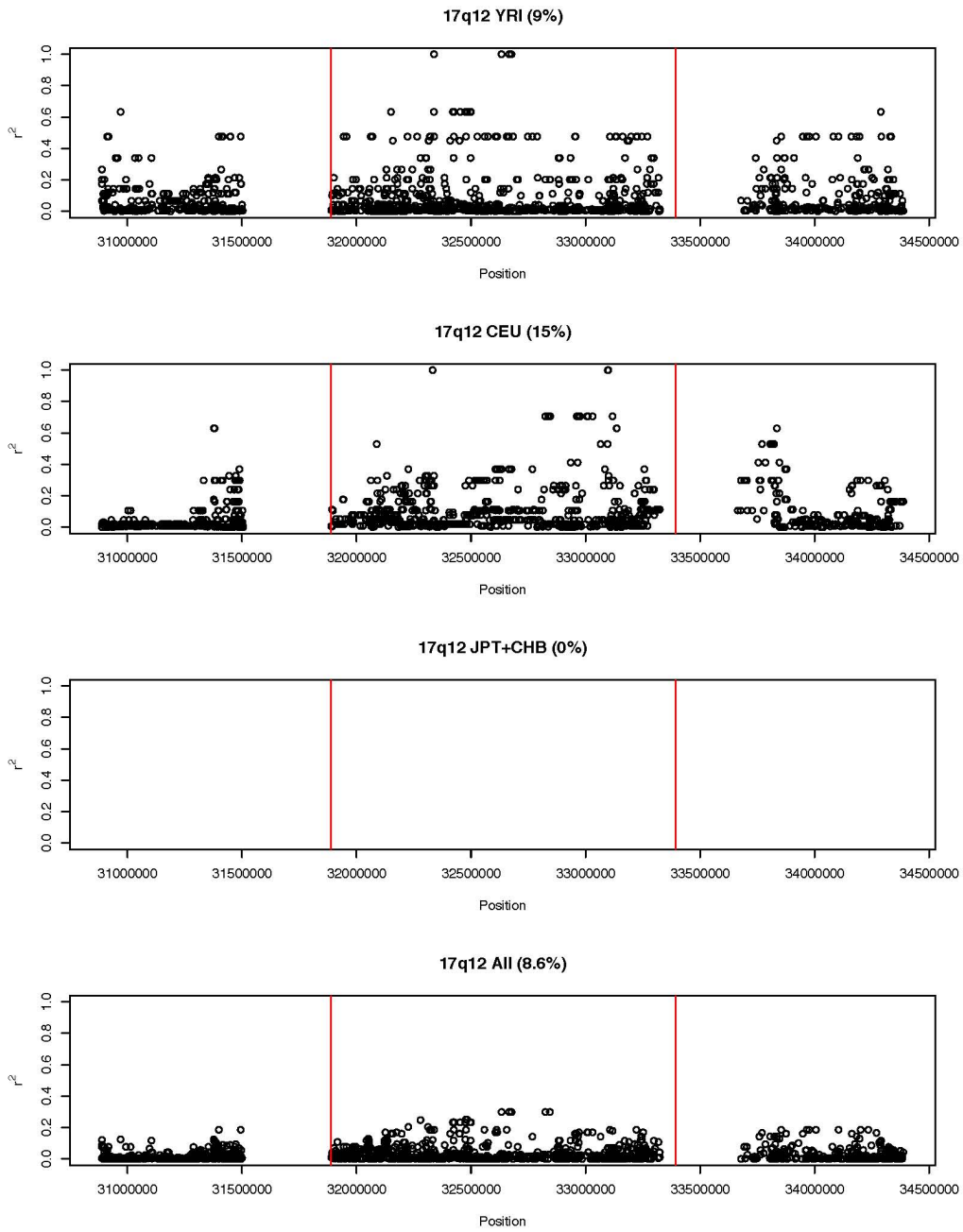
Supplementary Figure 3: Pair-wise r^2 between HapMap SNP and inversion genotypes. The approximate inversion breakpoints are denoted by the red vertical lines. Data are presented from the three HapMap continental groupings individually, as well as for all 27 samples pooled. The frequency of the inversion allele in each sample set is given in parenthesis. For analysis, double heterozygotes were assumed to have minor alleles in a cis configuration (see Materials and Methods).



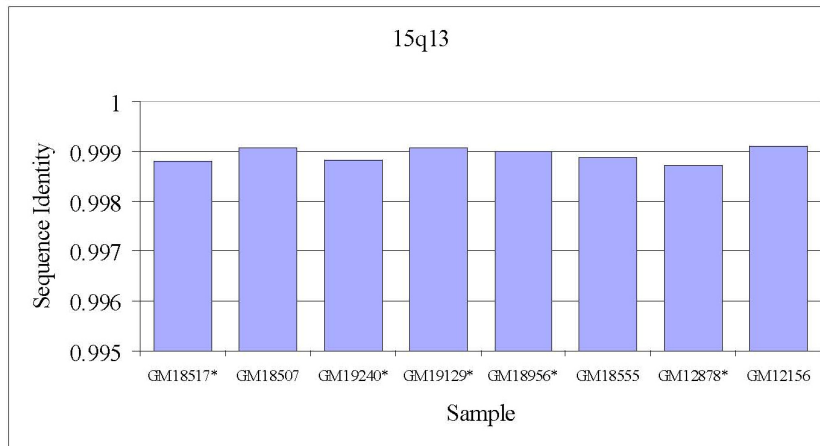
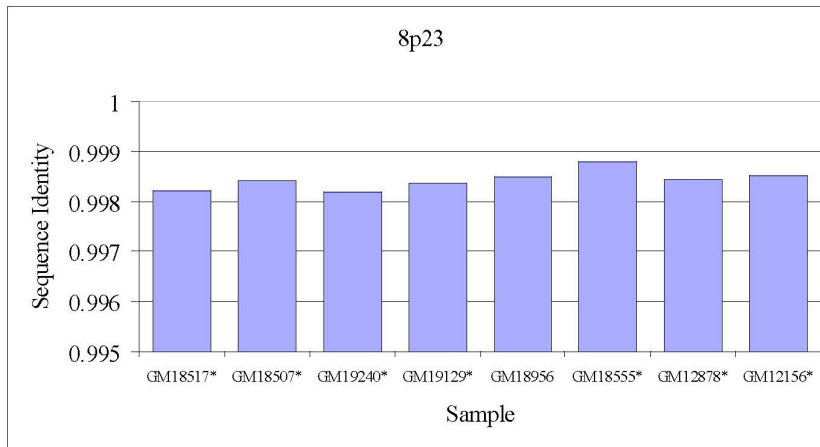
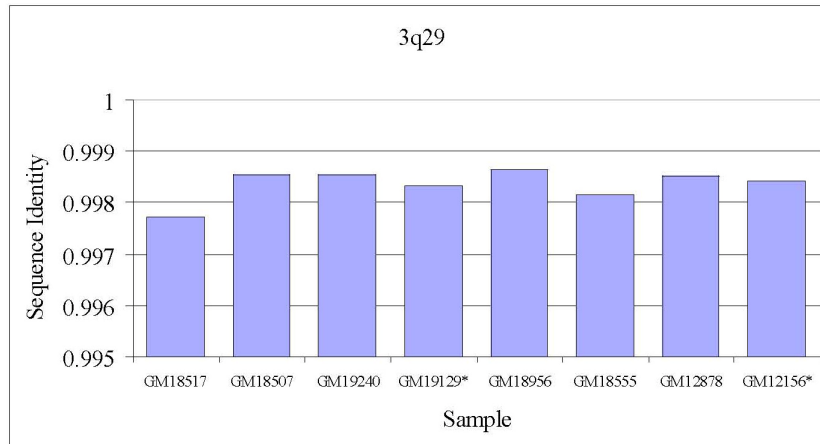


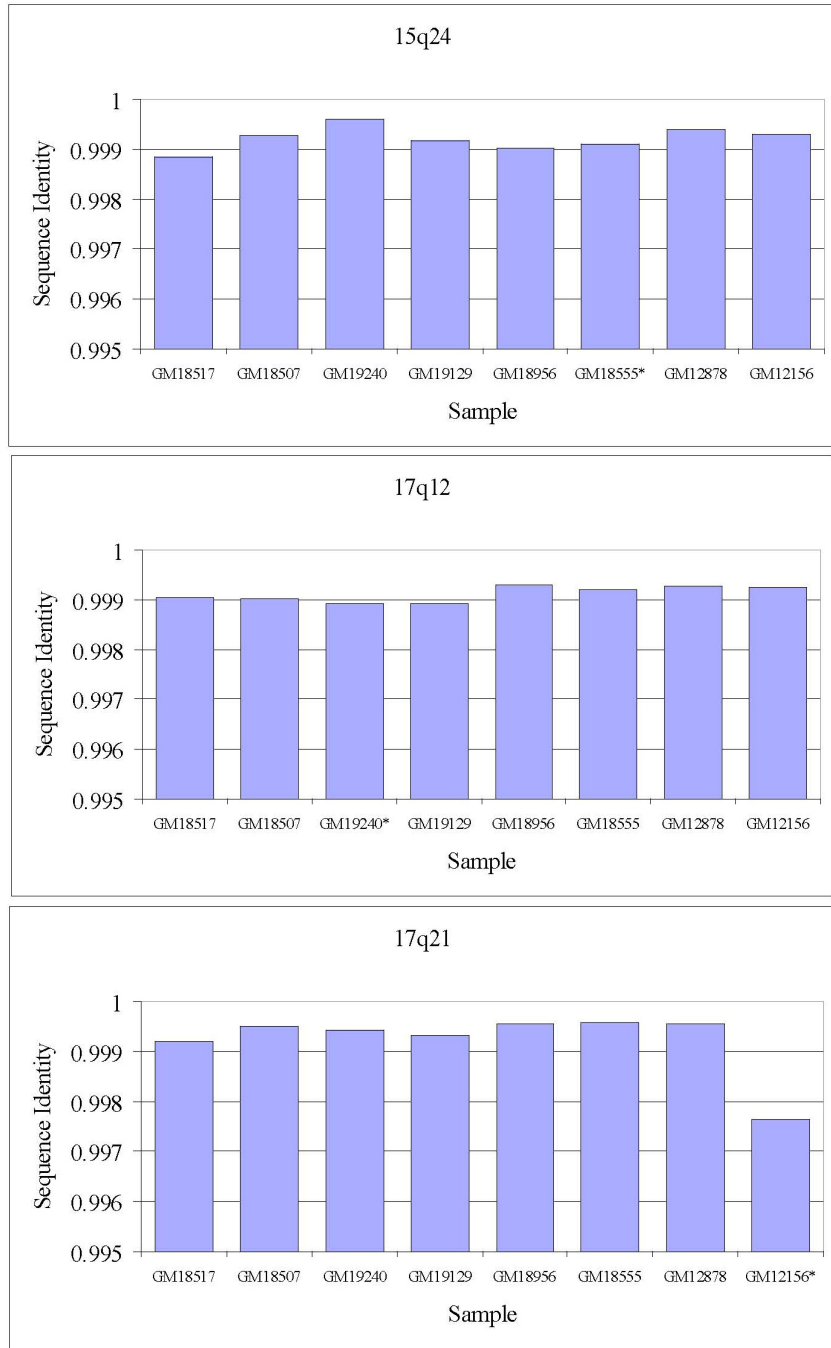




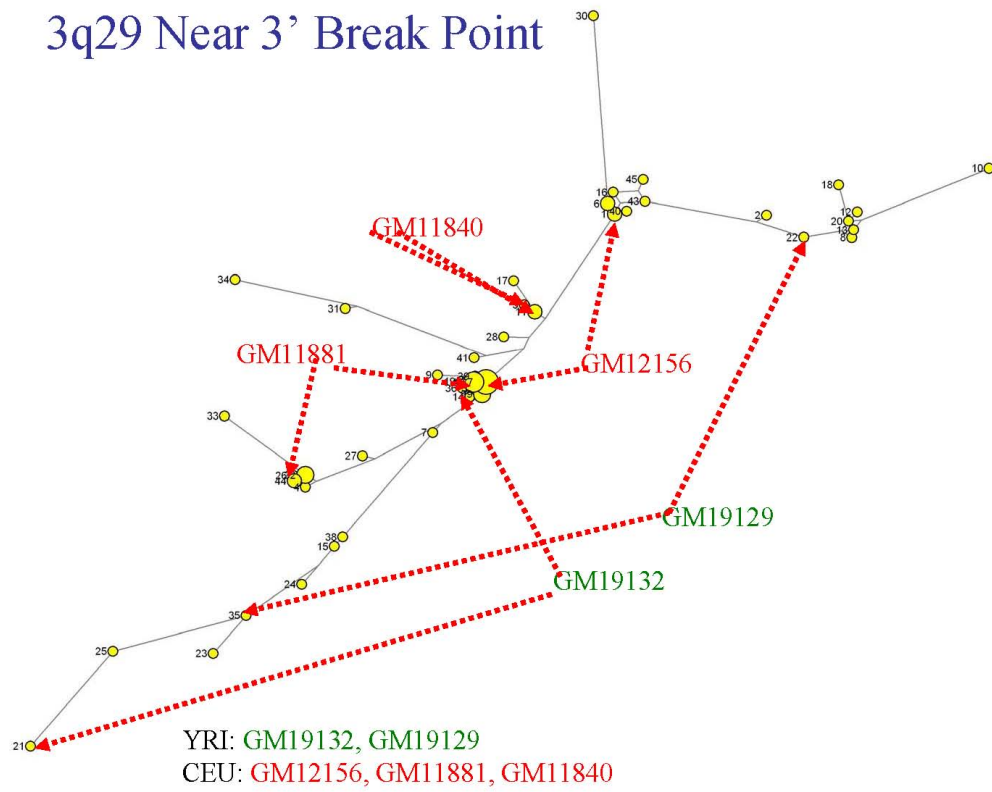
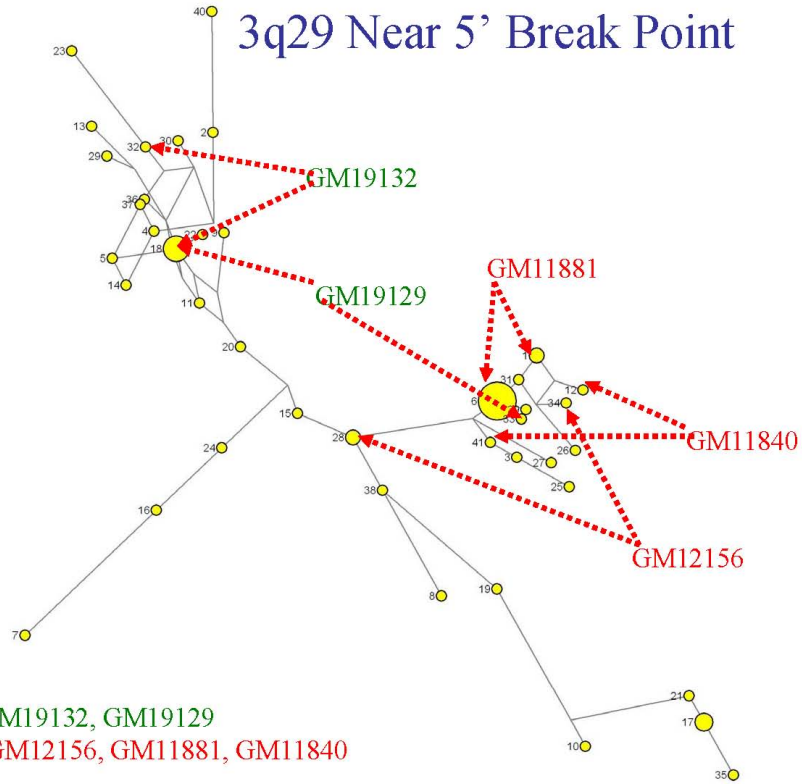


Supplementary Figure 4: R^2 values as in Figure S1, but including genotype results from the samples predicted to carry inversions (see Table 3).

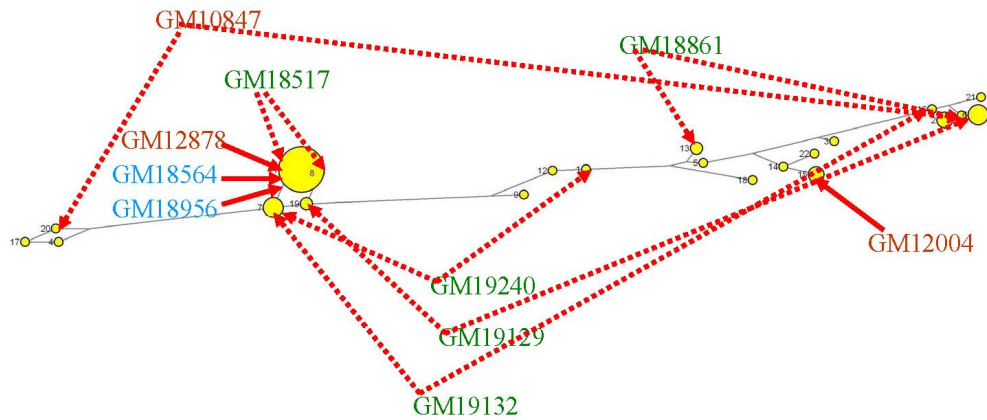




Supplementary Figure 5: Sequence divergence of inversion intervals. The sequence identity of eight HapMap individuals relative to the reference genome assembly (build 35) is shown for the unique portion of each inversion interval. Sequence identity was calculated based on all uniquely mapping fosmid end sequences from each sample. Only mapped positions having a sequence quality of at least Q20 were considered. Individual samples carrying an inversion are indicated by an asterisk.

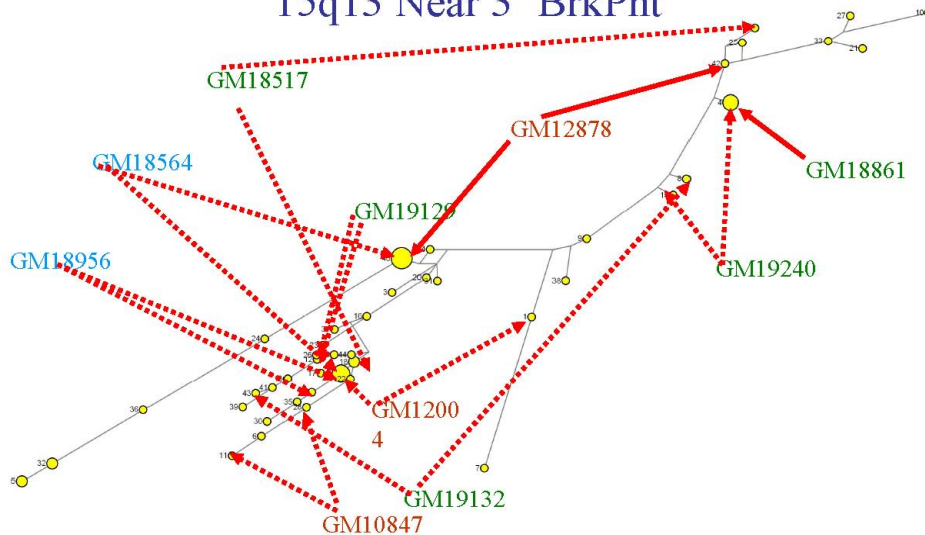


15q13 Near 5' Break Point



YRI: GM18517, GM18861, GM19129,
GM19132, GM19240
CEU: GM10847, GM12004, GM12878
JPT+CHB: GM18564, GM18956

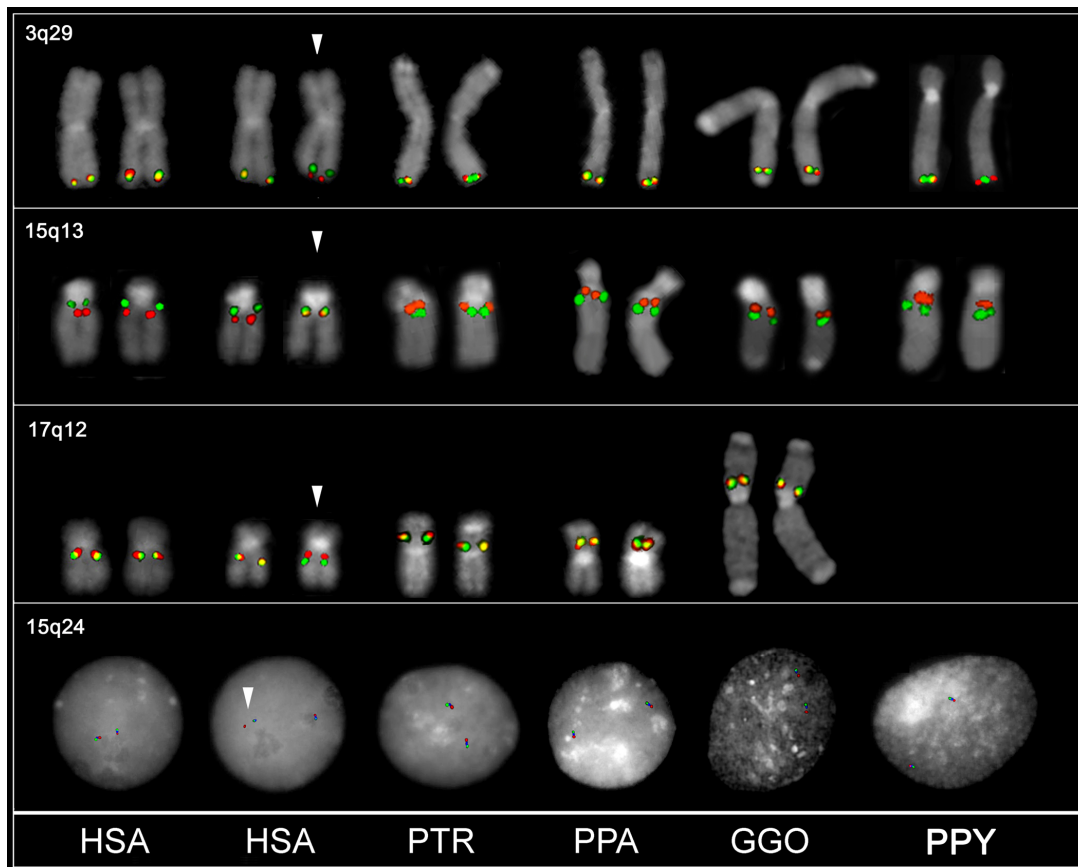
15q13 Near 3' BrkPnt



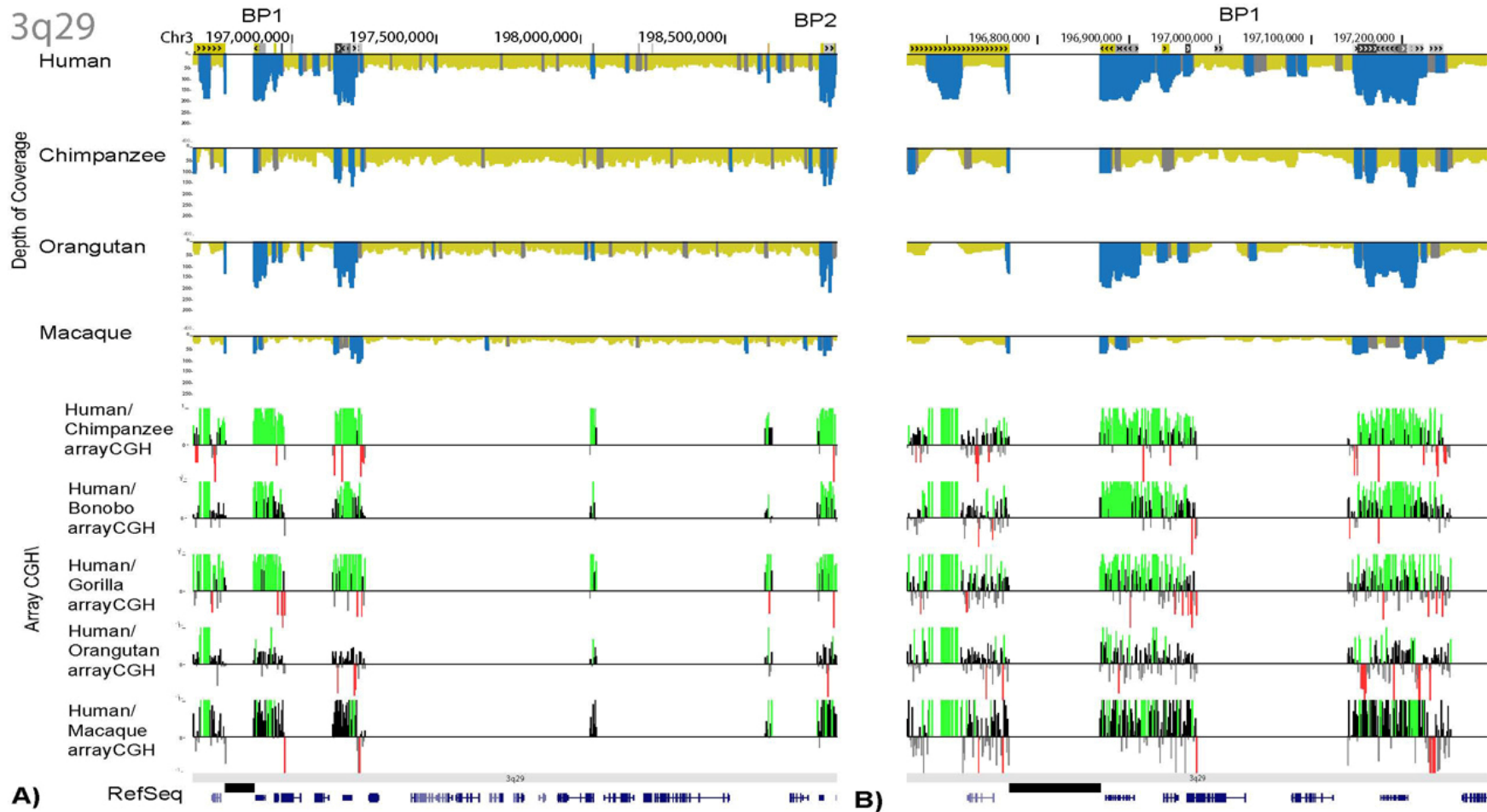
YRI: GM18517, GM18861, GM19129,
GM19132, GM19240
CEU: GM10847, GM12004, GM12878
JPT+CHB: GM18564, GM18956

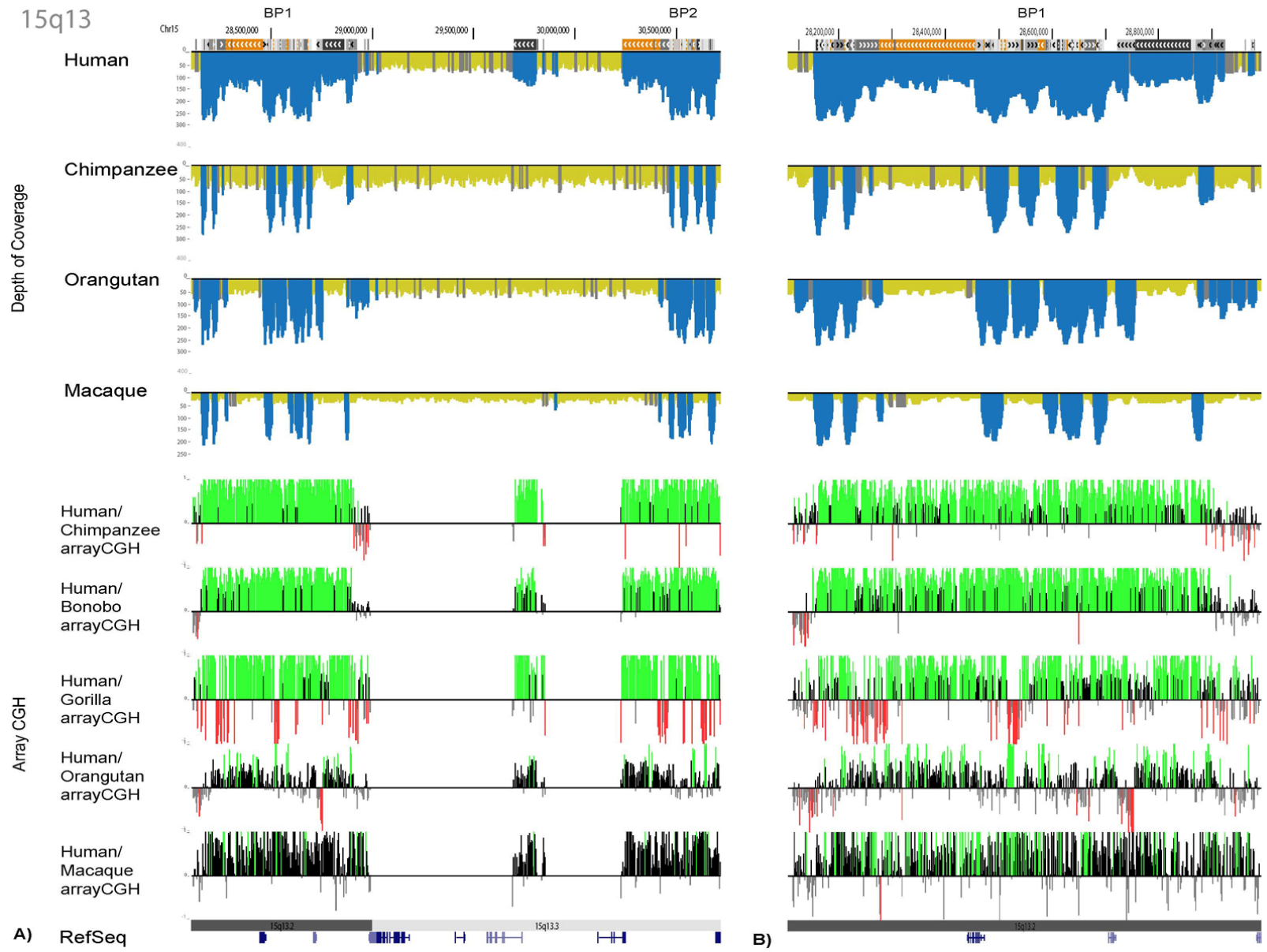
Supplementary Figure 6: Median-joining haplotype networks constructed from HapMap phased haplotypes. Networks were constructed separately for each breakpoint for the 3q29, 17q12 and 15q13.3 inversions (Table S6). Haplotype groups are denoted by yellow circles. The size of the circles is proportional to the number of chromosomes carrying the represented haplotype. The

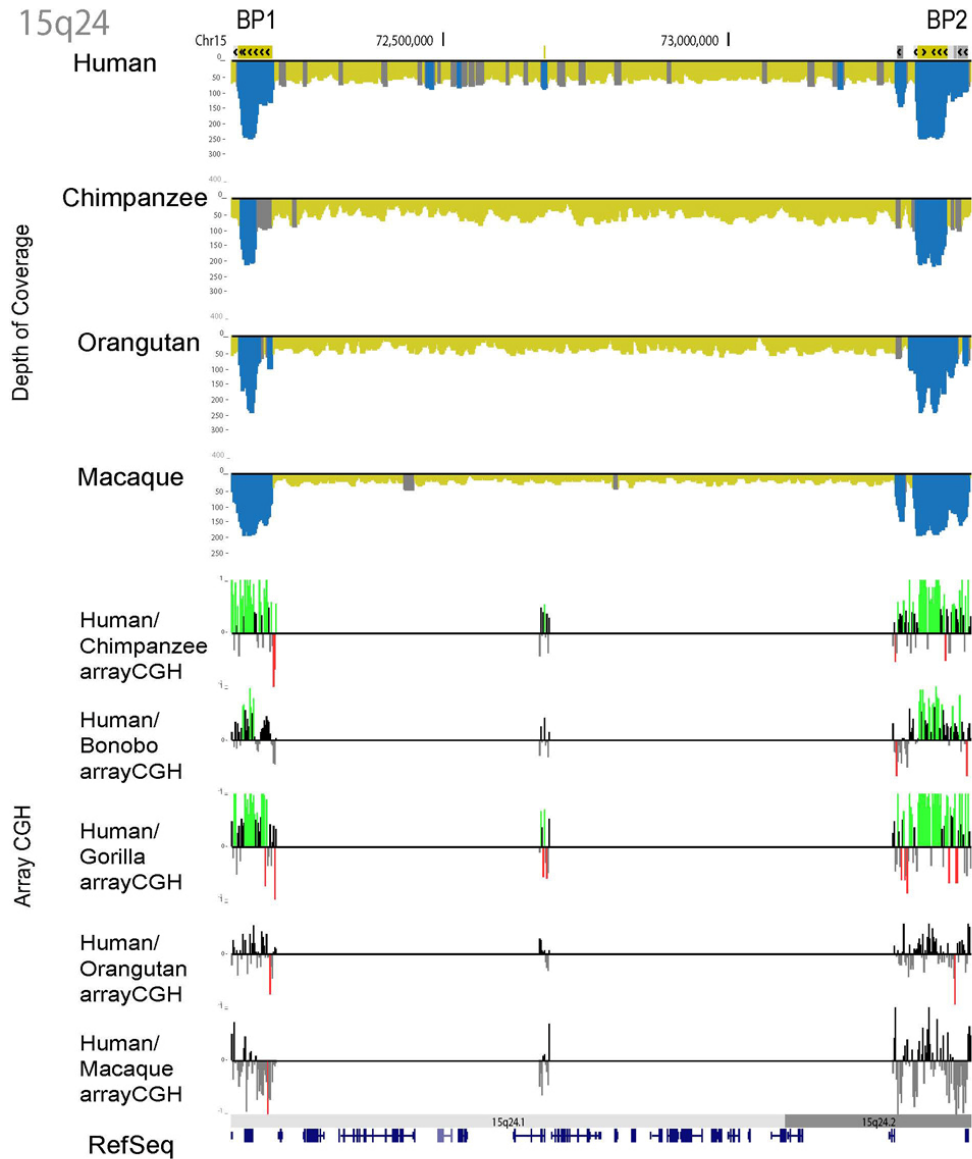
distance between haplotype groups is proportional to the number of SNP mutational events separating each haplotype. The haplotypes carried by individuals known to also carry an inversion allele are indicated by dashed and solid lines. For heterozygotes (dashed lines), only one of the two indicated haplotypes corresponds to the inversion. For homozygotes (solid lines), all indicated haplotypes must be present in the inverted orientation. Configurations where it is not possible to define closely related haplotype groups, which account for all inversion chromosomes, suggest recurrent mutation or substantial exchange between inverted and non-inverted haplotypes.



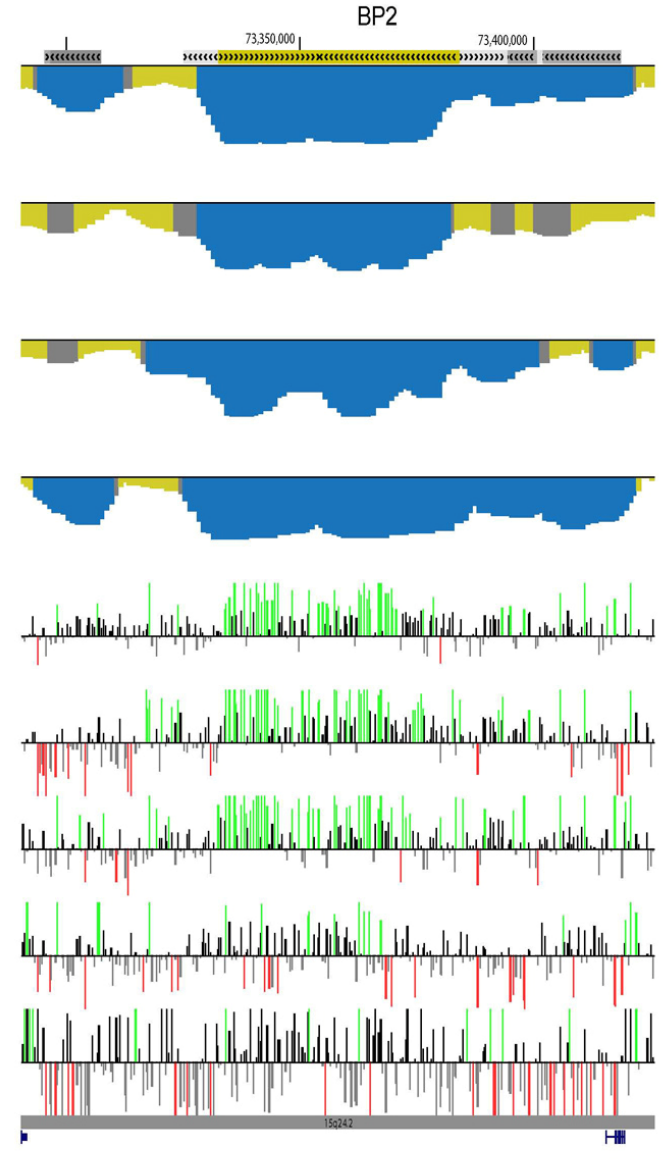
Supplementary Figure 7: Evolutionary origin of 3q29, 15q13.3, 17q12 and 15q24 inversions. None of these individuals were found to be inverted, suggesting that the direct (H1) configuration represents the ancestral state. Arrows indicate inverted chromosomes. Metaphase FISH-based assay could not be used to resolve the 17q12 inversion in PPY as the fosmid probe WIBR2-1900P13 maps within a larger evolutionary paracentric inversion in orangutan (2).





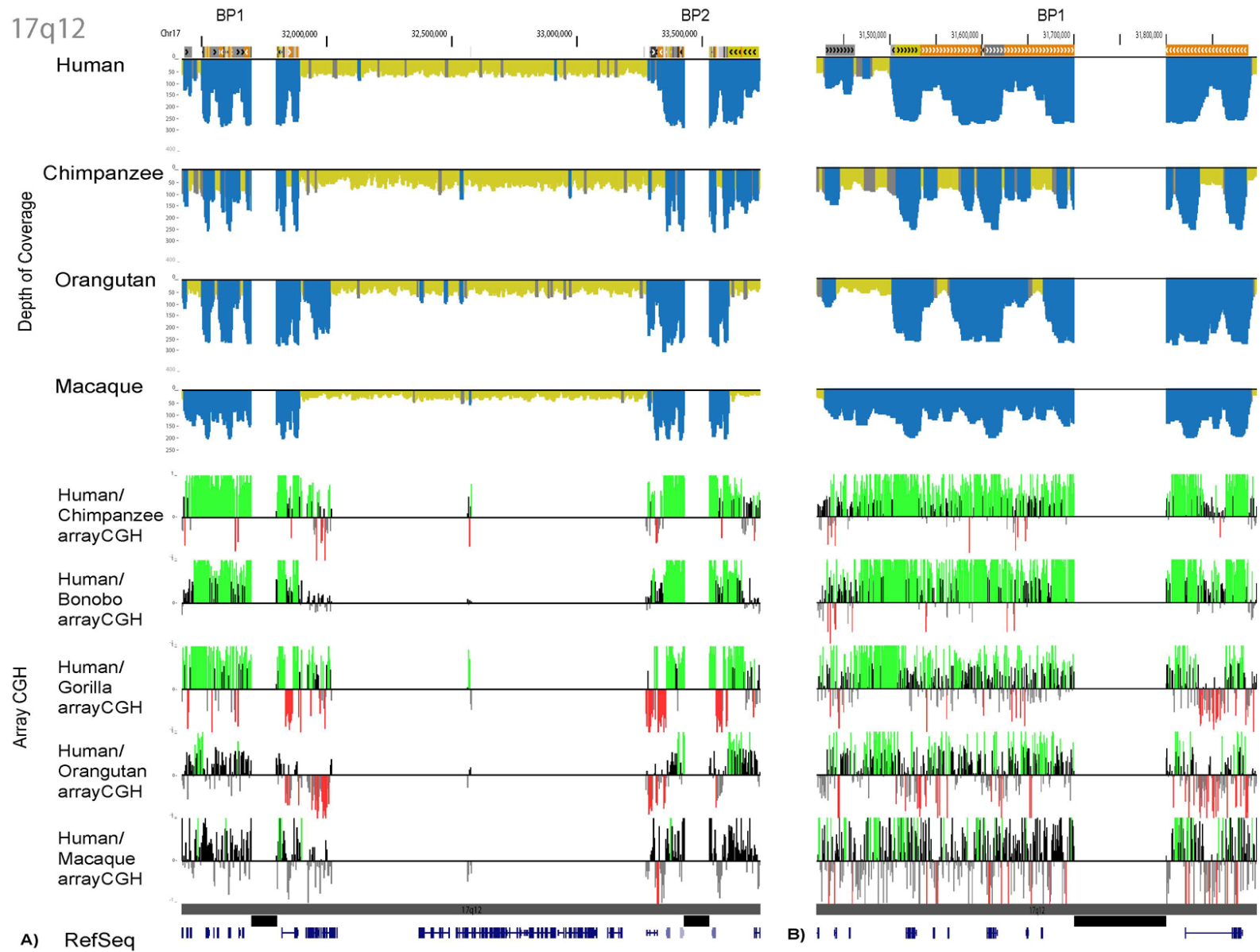


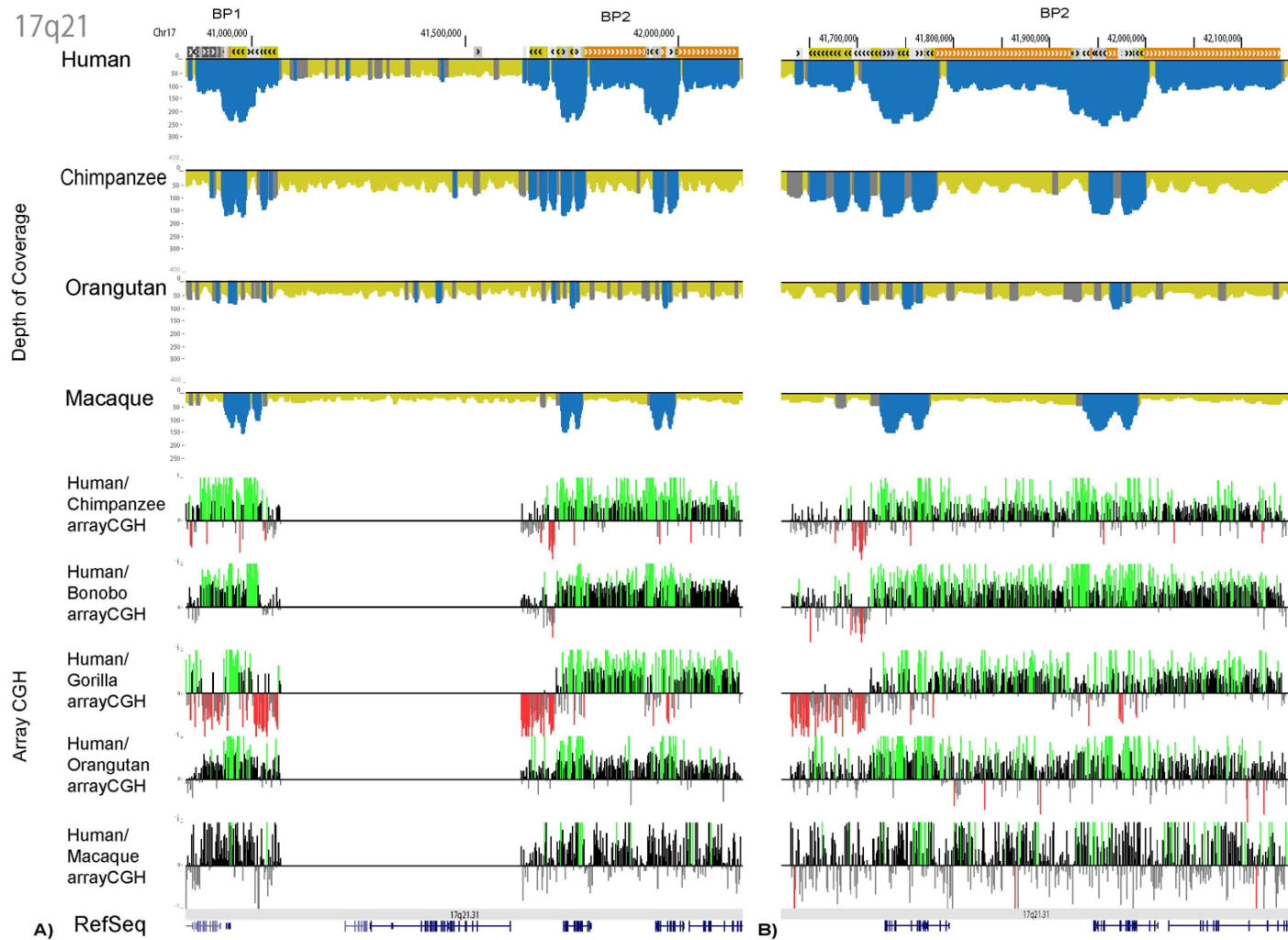
A)



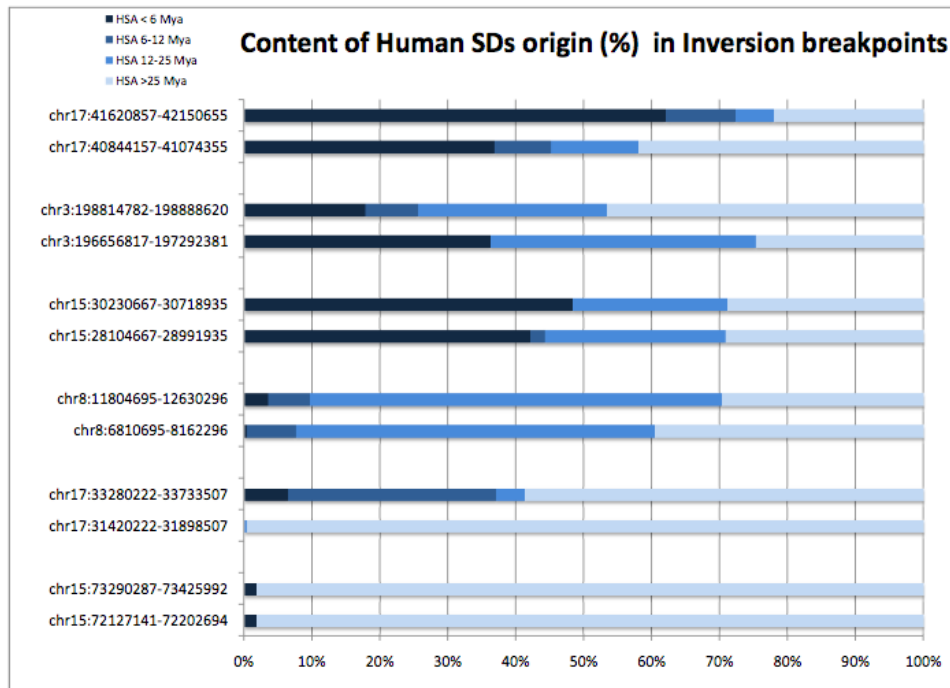
B)

17q12

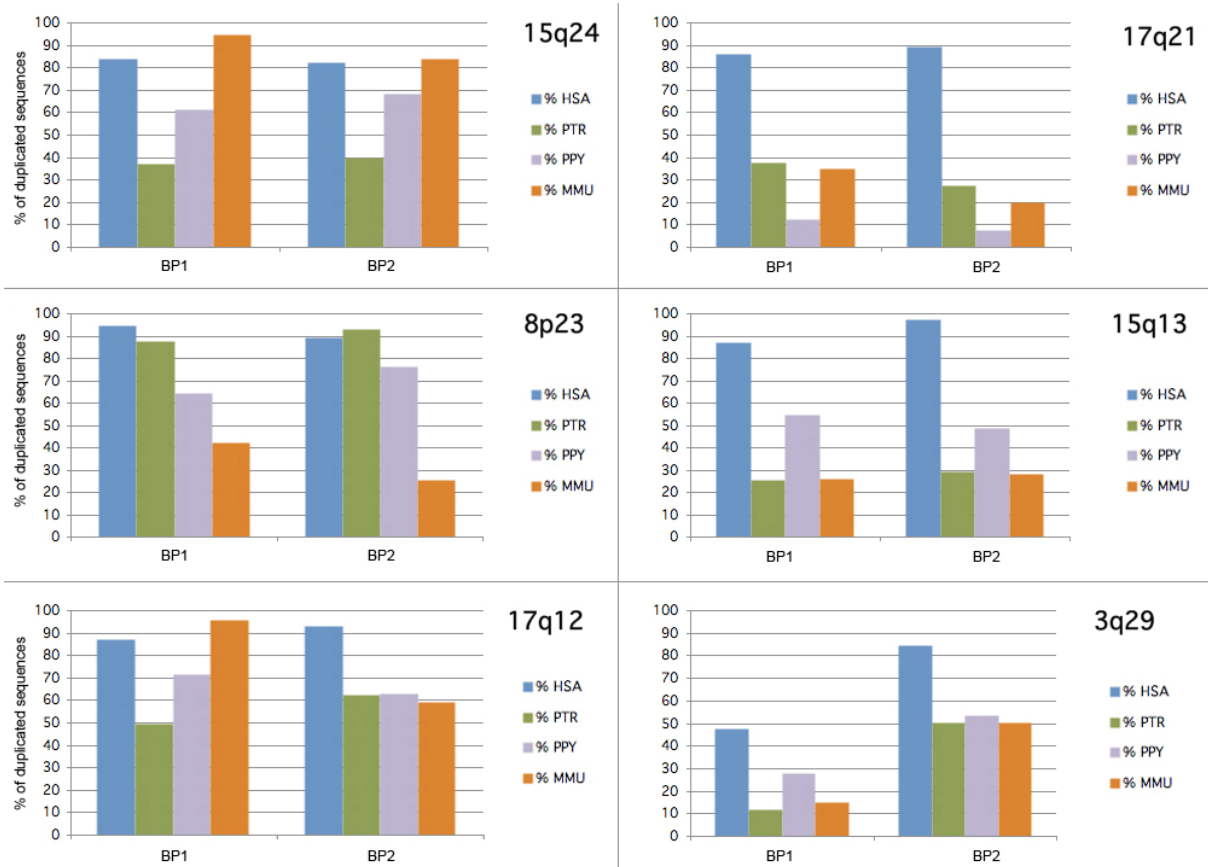




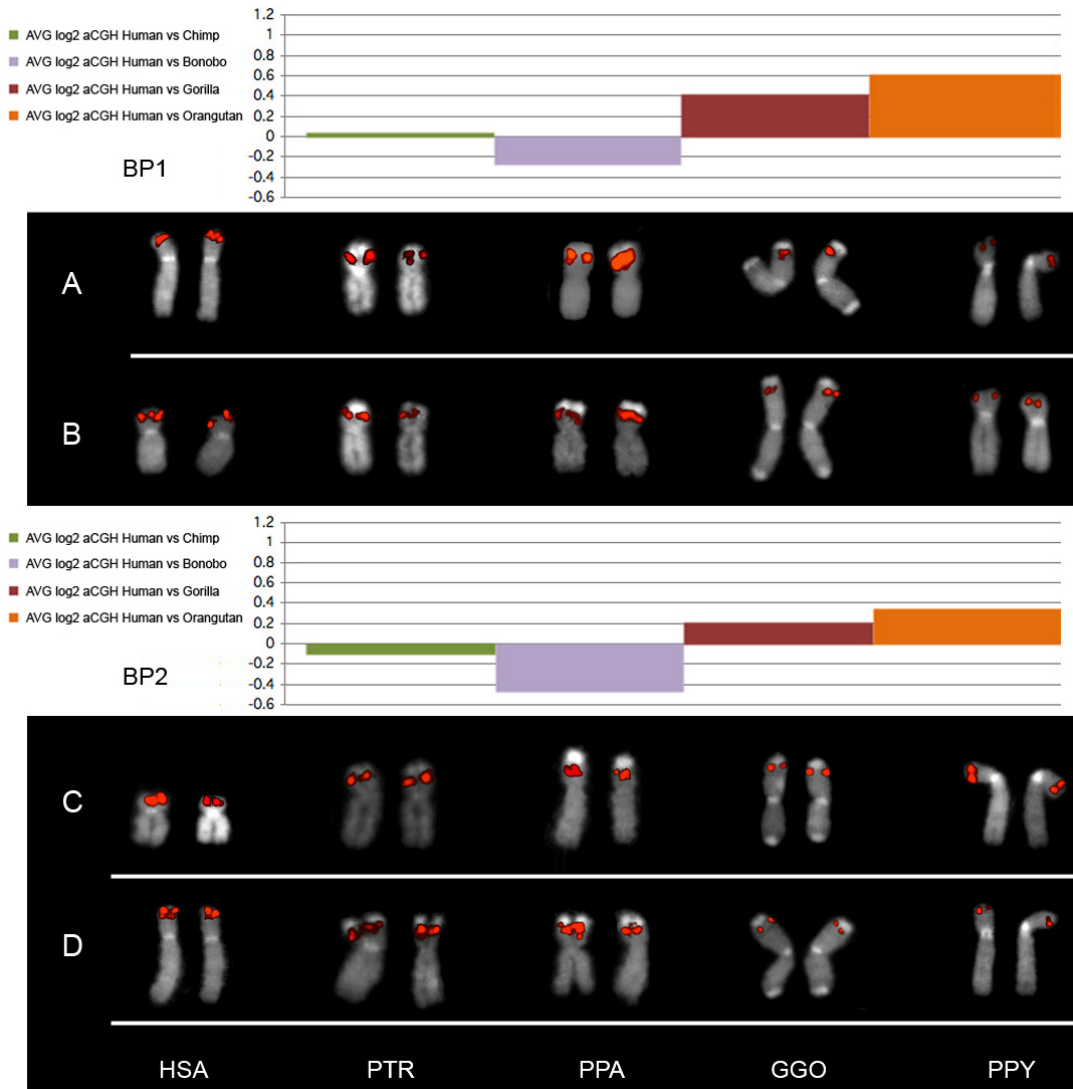
Supplementary Figure 8: (A) Comparative segmental duplication analysis of the 3q29, 15q13.3, 17q12, 15q24 and 17q21.31 inversion regions. Regions of excess WGS depth of coverage (duplicated regions) are shown in blue. ArrayCGH experiments using human as a reference are shown for chimpanzee, bonobo, gorilla, orangutan and macaque. (B) Enlargement of the distal or proximal breakpoints is shown.



Supplementary Figure 9: Estimated age of human segmental duplications at the breakpoints of the inversions is shown as a percentage of the total. Human segmental duplications were categorized into different evolutionary timepoints based on a parsimonious four-way comparison of macaque, orangutan, chimpanzee and human segmental duplications. A human-specific segmental duplication was set to younger than 6 million years (Myr); 6–12 Myr for duplications only shared by human and chimpanzee; 12–25 Myr for those shared by human, chimpanzee and orangutan; and finally, a duplication shared by all four species (human, chimpanzee, orangutan and macaque) was set to be older than 25 Myr.



Supplementary Figure 10: Primate duplication content. The percentage of sequence detected as duplicated at each inversion breakpoint (BP1 and BP2) is shown for each breakpoint region.



Supplementary Figure 11: A bonobo expansion of 8p23 segmental duplications. The average log₂ ratio by arrayCGH for segmental duplications at the 8p23 inversion breakpoint is compared against FISH using clones ABC13-48886900D12 (A), ABC13-48857200H3 (B), WIBR2-1720G15 (C) and WIBR2-1318J03 (D). Note, both arrayCGH and FISH predict excess copy number of segmental duplications in bonobo when compared to human.

Supplementary References

1. Bailey, J.A., Yavor, A.M., Massa, H.F., Trask, B.J. and Eichler, E.E. (2001) Segmental duplications: organization and impact within the current human genome project assembly. *Genome Res*, **11**, 1005-17.
2. Cardone, M.F., Jiang, Z., D'Addabbo, P., Archidiacono, N., Rocchi, M., Eichler, E.E. and Ventura, M. (2008) Hominoid chromosomal rearrangements on 17q map to complex regions of segmental duplication. *Genome Biol*, **9**, R28.