Table S1. Primers sequences used for quantitative PCR

Primer name	Forward sequence	Reverse sequence
18S Ribosomal RNA	5′-TCTCAGTGCAAGCCGAAGTAGGT-3′	5′-TCCCTTCCATTGCTGGTCGAGATT-3′
Hsp90	5′-AACTGCCCTCCTATCTTCTGGCTT-3′	5′-TCTTCACCTTCCTGCGCATCATCT-3′
Endoplasmin	5′-GGACAGTTTGGCGTTGGCCTTTAT-3′	5′-CAAGGTATTACCACGTGGGTCTTCAG-3′
Hsp20.7	5′-CTCAACAGTCTTCTCAACATTCTTCTTAGG-3′	5′-AGGAACGTCCAATTCCAATCATCC-3′
Hsp70	5′-ACTCCCACACAGGAGTATGTGGTT-3′	5-GTGAACGAACAAGAGTGCGGAGAA-3′
ATP citrate lyase	5′-ACTGCTGATCATGGACCTGCTGTA-3′	5′-AGCACCATCTAGAGCACCACCAAA-3′
Cathpesin-like protease	5′-AGCATGATCCGCAAGATCCCTGAT-3′	5′-CATTCATCGGCTGATGGGTGTTCA-3′
Translation elongation factor-1γ	5′-AAGGCTGGAACCTTTCCAAGAGGA-3′	5′-AGGTGCTCATTGCTGCCTCTTACT-3′
Cytochrome p450	5′-GGACTTGCGTGTTAAGTTAAGCCCAG-3′	5′-ATCACGGGCAGCGTCATCAAAGTA-3′
Histone 2A	5′-TAGACCCGGATGTTGACCCTGTTT-3′	5′-ATTGGAGTTAGCTGGCAATGCTGC-3′
Desaturase	5′-ATGGCCTCTGCGACTTCTATTGGT-3′	5′-TTGTGTGGATCGGCATCAGTCTCA-3′
CHORD containing protein	5′-TAAGGGCTGCACAAGGTCGTATCA-3′	5′-CCTTGACAATTGCTGCAGGTGCTT-3′
Bax Inhibitor	5′-CTTCAGCCTCTGCGTTTACGCTTT-3′	5′-ATCGAGAAGCGTCGAAGTGGTGAT-3′
RACK1	5′-TCTCTTTCCAGCGTGGCCATTAGA-3′	5′-CCTCGAAGCTGTAGAGATTCCGACAT-3′
Nedd8	5′-GGAGGATCGGCTTCTTCATTTGGT-3′	5′-AGCATAGGCAATCTGAGCACGCTA-3′
CG1532-PA/glyoxalase	5′-GGATGGCCCTTTACTGAAGAAGGA-3′	5′-TGCGAGGTAGGCTGAGGTTCATTT-3′
Mitochondrial processing peptidase	5′-ATACAACAGGACGGGCTTCCAAGT-3′	5′-GGCTGGCAAAGTTGGAGATGTTGA-3′
Translation initiation factor 4	5′-AAAGCCTGGGTCTGTGTGATGAGA-3′	5′-TGAGATCCTTGAGGAGCGTGGTTT-3′
IFRD	5′-ATCAGGTGGCTCTCCATCCTCAAT-3′	5′-AGAGGACATGACTCCTCAGTTGGT-3′
TIF	5′-GGTCTGTGTGATGAGACGGATCCAAA-3′	5′-TGAGATCCTTGAGGAGCGTGGTTT-3′
TFDp2	5′-CAGTCTGCTGAGTAGGAGTTAAGCGT-3′	5′-TCCATTTCAGTCACCTGGTGGAAC-3′
RpL45	5′-AGTCGGCGGAGATCTATTGCATGT-3′	5′-GGATTCCAGTGTTTCGAGCCTCTT-3′
eEFtB-γ	5′-AAGGCTGGAACCTTTCCAAGAGGA-3′	5′-AGGTGCTCATTGCTGCCTCTTACT-3′
Spaghetti squash	5′-GGAAATGAGGGCAGTAAGGCACTA-3′	5′-ATCGTGAAGCTCCAATCAAGGGTG-3′
CHORD containing protein	5′-TAAGGGCTGCACAAGGTCGTATCA-3′	5′-CCTTGACAATTGCTGCAGGTGCTT-3′
COX subunit II	5′-TTCCGTCTTCTCGATGTTGACAACCG-3′	5′-CGTCCTGGTGTAGCATCAGATTTAACCC-3′
COX subunit IV	5′-TGCCCGAATCCTTTACTGATGAGC-3′	5′-CCCACTTGGATGTCAATCCATCAAC-3′
COX subunit VII	5′-TTCTTGGAGTCGCTGTGGACATCA-3′	5′-TGGCTCTGGGTTTAGTGCTTCCTT-3′
Arginine kinase	5′-TGCTTCTGTGCACATCAAGGTTCC-3′	5′-AAACTCCACCCTCAGCTTCTGTGT-3′
NPC2	5′-TCATATGTTCCACTGTCCCAGCAAG-3′	5′-AGCTATTTCTTCCCTGGCACTGAC-3′
RpL35	5′-TCCACTTGGTTGGTGTTGAGGTCT-3′	5′-AGATGTTGCATCTACGTCGGCCAT-3′
Reptin	5′-GCCATCTCATTCTCCAGAGCACGATT-3′	5′-TGAGTGGCGTGAAGAAGGGAAAG-3′
Aldo-keto reductase	5′-AGGCCCATGGAGGAACAAGTGAAT-3′	5′-GCCACAAACTGTAATGCCCAAACG-3′
Polymerase III	5′-ATGGACAGAAAGGTGTTACAGGGC-3′	5′-TGATCCACCAAATGCTGTCCCA-3′
Pyrolline carboxylate reductase	5′-TCTCCCAATTCTACGGAGCGCTTT-3′	5′-TGGTATTGGAAACAGGCAAGCACC-3′
Male sterility protein	5′-GGTCCGGCGATACCTCAATAAAGA-3′	5′-TATACTATCACGGGCACCCAAGCA-3′
Lipid metabolism	5′-AAGACGTGTCCAGGGCTTATCGTA-3′	5′-TGTCGGCTGCCATTATGACTACCT-3′