#### Supplemental TABLE I. Primers for PCR amplification of TcOGNT RNA, DNA and mutagenesis

<u>Purpose</u>	<u>Primer</u>	<u>Species</u>	Sequence							
Expression										
full-length TcOGNT2 <sup>1</sup>	E5S1	L. tarentolae	5′-acagatctccaccatggataaaaagaagc							
TcOGNT2cat <sup>2</sup>	E5S2	L. tarentolae	5'-gaggcgccggtcccaccatggaattg							
both	E5AS1	L. tarentolae	5'-gactcgagttatccaagtgttgccttttc							
full-length TcOGNT1	LtFullS	L. tarentolae	5′-at <u>ccatgq</u> agcttgaggaaaccaatttgtta							
full-length TcOGNT1	LtCATAS	L. tarentolae	5'-at <u>ctcgag</u> tcatccgcaccatcgaccg							
full-length TcOGNT2 <sup>2</sup>	E5SATG2	D. discoideum	5'-ctggtacctgatcaatggataaaaggaagcttggac							
full-length TcOGNT2 <sup>2</sup>	E5AS2	D. discoideum	5 <b>'-</b> gt <u>agatct</u> tccaagtgttgccttttca							
TcOGNT2cat <sup>1</sup>	E5U2	D. discoideum	5′-ga <u>agatct</u> agatttgacaggcgtggtccc							
TcOGNT2cat <sup>1</sup>	E5L1	D. discoideum	5′–gt <u>gagete</u> ttatccaactgttgccttttcgc							
TcOGNT2cat <sup>1</sup>	E5S3	E. coli	5′-ga <u>catatq</u> agatttgacaggcgtggtccc							
TcOGNT2cat <sup>1</sup>	E5AS3	E. coli	5′-gt <u>ggatcc</u> tccaactgttgccttttcgc							
Mutagenesis and express	sion									
TcOGNT2cat <sup>2</sup>	TcE5D234Afor	L. tarentolae	5'-gtgattgctagccattcaagatttgtgcctgagtgg							
TcOGNT2cat <sup>2</sup>	TcE5D234Arev	L. tarentolae	5'-cttgaatggctagcaatcaccatatagtaatcctgg							
TcOGNT2cat <sup>2</sup>	TcE5D234Nfor	L. tarentolae	5'-gtgattaatagccattcaagatttgtgcctgagtgg							
TcOGNT2cat <sup>2</sup>	TcE5D234Nrev	L. tarentolae	5'-cttgaatggctattaatcaccatatagtaatcctg							
5'-splice site mapping										
$TcOGNT2^1$	TcE5AS415	T. cruzi	5′-gagagcgagtcaacacaatgttct							
TcOGNT1	TcE2NAS	T. cruzi	5'-tgaaaacccaaccttcggac							
miniexon primer	TcME33	T. cruzi	5'-cgctattattgatacagtttctgtactatattg							

<sup>1</sup> Haplotype Tc00.1047053511309.70 was cloned. <sup>2</sup> Haplotype Tc00.1047053511759.30 was cloned.

Restriction enzyme sites for cloning are underlined.

### Supplemental Table II. TcOGNT2cat identification in gel bands

Gel bands in Figure 7E were excised, trypsinized and identified by MALDI-TOF-TOF-MS as described in Experimental Procedures. MS/MS data consist of at least 5 expected b or y ions detected after fragmentation. Peptides detected are mapped in Suppl. Figure 2.

TcOGNT2	MH <sup>+</sup> observed	MH <sup>+</sup> expected	Additional	Bands
position			evidence	
66-87	2314.0	2314.3	MS/MS	Top, middle
88-101	1698.6	1698.8		Top, middle
117-131	1712.6	1712.8		Top, middle
217-238	2645.3	2644.9	MS/MS	Top, middle
279-300	2416.8	2417.2		Top, middle
301-327	3062.3	3062.6		Top, middle
306-327	2378.9	2379.2		Top, middle
350-371	2708.9	2709.2		Top, middle
432-442	1319.6	1319.4	MS/MS	Top, bottom
450-468	2412.9	2413.1		Top, bottom

**SUPPLEMENTAL FIG. 1.** Alignment of sequences corresponding to the NRD2- and Gal/GalNAc-like domains of select family GT27 and GT60 sequences. Human (Hs) pp- $\alpha$ GalNAcT2 and murine (Mm) pp- $\alpha$ GalNAcT1 initiate *O*- $\alpha$ GalNAc mucin type glycosylation and belong to CAZy family GT27 (Canteral et al. 2009). Dd- $\alpha$ GlcNAcT2 (DdGnt2) initiates the related *O*- $\alpha$ GlcNAc mucin type glycosylation in *Dictyostelium*, and belongs to family GT60. The three related sequences from *T. cruzi* most closely resemble the GT60 sequence class. Amino acids are colored according to group properties, and shaded (using a distinct color scheme) at positions where this property is conserved in either the pp- $\alpha$ GalNAcT or the pp- $\alpha$ GlcNAcT sequences (West et al. 2004), not including TcOGNTL which is predicted to be catalytically inactive. Positions of sequence identity are bolded, and positions in Mm pp- $\alpha$ GalNAcT1 where amino acid substitutions inactivate the enzyme (Hagen et al. 1999) are marked with an asterisk. Amino acid positions relative to the full-length protein are indicated in parentheses.

	NRD2 Motif-1
Hs-ppGalNAcT2	(139) <mark>SVVI</mark> T <mark>FHNE</mark> AR <mark>SALLRTV</mark> VSVLKK <mark>SP</mark> PHLIKEIILV-DDY <mark>S</mark> (178)
Mm-ppGalNAcT1	(119) <mark>SVVIVFHNEAWSTLLRTVHSVINRSPRHMI</mark> EEIVLV-DDAS(158)
DdGnt2	(63) <mark>TIFV</mark> SLAAYRDVFCSDTINYIFNH-ANRPEKIFIGIVDQGSEFLEEDP(109)
TCOGNT2	(78) <mark>TIFVSIAAFR</mark> DKECVTTLESLFTR-AKNPRRVYLGISEERFEM-DSSC(123)
TCOGNT1	(357) SLFLNIASFRDKECWPSIDHMIQR-STNMFRVYWGVAQQHYYS-DLPC(402)
TCOGNTL	(182) <mark>SIFVGITHSQEKVTSSEMETACAATV</mark> RNIYDA-AHWPLGVFTGIVEITLSKTSPTT(236)
	NRD2 Motif-2
Hs-ppGalNAcT2	(199)DRRE <mark>GLMRSRVRGA</mark> -D <mark>AAQA</mark> K <mark>VL</mark> TFLD <b>SHC</b> ECNEHWLEPLLERVAEDRTR <mark>VVSPII</mark> DVI(256)
Mm-ppGalNAcT1	(184)EQRSGLIRARLKGA-AVSRGQVITFLDAHCECTAGWLEPLLARIKHDRRTVVCPIIDVI(241)
	* *
DdGnt2	(143)AQ <mark>S</mark> K <mark>GPTLARYYATTLY</mark> NNET <mark>YFM</mark> QVD <b>SH</b> LRFIKG <mark>W</mark> -DSL <mark>II</mark> NDLW <mark>L</mark> TKSYAPIGENGIPRT <mark>VLTHYPMAY</mark> (212)
TCOGNT2	(208)DDARGPTYGRYLTSLFYNNODYYMVIDSHSRFVPEW-DIKMIERARLMPTWG <mark>VMSYYPNGY</mark> (267)
TCOGNT1	(448)FTALGPAFGRYMSMLLYGGEDYMLVLDSHNRFVYAW-DARIIAMQVALEHPKAVLSHYPESY(508)
TCOGNTL	(318)TLLS <mark>SAAAQRYAT</mark> LA <mark>LY</mark> RGET <mark>YVMFVRA</mark> G <mark>LQLV</mark> YK <mark>W</mark> -DVLTRLLWLQLPSRSA <mark>VLS</mark> QPPVQ <mark>I</mark> (378)
	Gal/GalNAc Motif
Hs-ppGalNAcT2	(290) PIKTPMIAGG-LFVMDKFYFEELGKYDMMMD- <mark>VWGGENLEI-SFRVWQCGGSL</mark> EIIPCSR- <mark>VG-HVF-</mark> RKQHPYTFF(369)
Mm-ppGalNAcT1	(285) PVRTPTMAGG-LFSIDRDYFQEIGTYDAGMD-IWGGENLEI-SFRIWQCGGTLEIVTCSH-VG-HVF-RKATPYTFF(354)
	*
DdGnt2	(254) PAECPYIAAGFFFT-SGEAIKLVP-FDPHLSNLFEGEEILY-SVRMY-SAG-FRFFAPTLNVCFHYYSRPKSPKFFW(324)
TCOGNT2	(308) PVLQGFVAAGYIFG-DAQFVKDVP-FDPYLPFLFDGEEILY-TVRLW-TSG-WDSYCPGTAFLFHNYMRTDAPRFFW(378)
TCOGNT1	(575) <b>PL</b> AQ <b>PWAAGGFLFA</b> -NAS <mark>VMREVP-FDPHLPHLFDGEEVMY-SVRLW-T</mark> HG-YDIFSPKRGICY <b>HF</b> YD <b>RP</b> NE <b>PKLLW</b> (645)
TCOGNTL	(627)Y <mark>LQQSWVTPDFLFT</mark> -RAE-21-VP-LDPFLSFLGADEEAVLLSARLW-THG-WDFFSSTEPIAFIVT-KPPAEGDAS(716)

**SUPPLEMENTAL FIG. 2.** Annotated nucleotide and amino acid sequences of the two TcOGNT2 haplotypes. Nucleotide and amino acid sequences are numbered beginning with the start codon. 5'-splice acceptor sites for TcOGNT2-1 (violet), primer hybridization sites, N-glycosylation sequences, site of mutagenesis, peptide sequences (green or blue) identified by MS (Suppl. Table II), and key motifs are denoted.

	>Tc00.1047053511759.30 (TcOGNT2-2) >Tc00.1047053511309.70 (TcOGNT2-1)	
-240 -180 -120 -60	aacatgtacatttacctgttgttaaactttgaggatgtggtgaattgactcgtatttcat tcatctctttcctatgcttatatatatatattttgatttctattattgatgaacttgg gatccggcgagcggcccagctgaatatttggaagaggttttacggaaaagggtgtagttc gacacacaaaagcaatcaagtaaaggtgaagtaacaactcatatttacatcagcccatg	5'-splice donor nt
1 1	atggataaaaggaagcttggacggggcttgaggattcatcgtgggggggg	E5SATG2 E5S1
61 21	agattttttcgagtgtttttggttttcgtggcatttttttt	<u>E5U2,E5S3</u>
121	gagaggcgt <b>ggtcccaccatggaattg</b> gaagatgcagacggcattcttcagacgataccg	E5S2
41	gacaggcgtggtcccaccatggaattggaagatgcagacggcattcttcagacggtacct   E R R P T M E L D A D G I L Q T I P   D V V V V V V V	
181	${\tt gtatttcttcgaaggatcttccggctgcaaatataccagtggacattgctactatttt$	
61	gtattttcttcgaaggatcttccggctgcaaatataccagtggacattgctactatttttVFSKDLPANIPVDIATIF	tryptic peptides
241	gtatccattgcggcctttcgagacaaggaatgtgtcactacactagagagtctctttacc	(MS of 40KD band)
81	gtatccatcgcggcctttcgagacaaggaatgtgtcactacactagagagtctctttacc	tryptic peptides
201		(MS-MS of 40KD band)
301	cgcgcgaagaatcctcggagagtgtatttgggtatcagtgaggagcgatttgaaatggat cgcgcgaagaatcctcggagagtgtatttgggtatcagtgaggagcgatttgaaatggat	
101	<u>R</u> A K N P R R V Y L G I S E E R F E M D	
361	agcagttgcattgcttcaccggaggtgctta <u>agaacattggtgtgactcgctctc</u> gtaaa	E5AS415
121	S S C I A S P E V L K N I G V T R S R K	
421	cttcactggaaggatgttgttccctttgcctacaatcccacagacaagcagaaatatccc	
141	Cttcactggaaggatgttgttccctttgcctacgatcccacagacaagcagaaatatccc L H W K D V V P F A Y N P T D K Q K Y P D	
481	aagcacacacctgtgctgcatgcaaaacgcgatgaggatgtcatcacgtgtctactcagt	
161	Aagcacacctgtgtgtgtatgcaaaacgcgatgaggatgtcatcacgtgtttattcagt K H T P V L H A K R D E D V I T C L L S	
541	gaaggttcttccgacgcaatgcccggccctggtgaatctgttttgaacggatgtcaggtc	
181	<pre>gaaggttcgtccgacgcaatgcccggccctggtgaatctgttttgaacggatgtcaggtc E G S S D A M P G P G E S V L N G C Q V</pre>	
601	ttgacacgcattggacacccggatgatgcacgaggcccgacttacggtcgttacctaaca	
201	L T R I G H P D D A R G P T Y G R Y L T	
661	agtctcttctacaataaccaggattactatatggtgattgacagccattcaagatttgtg	

 $a {\tt gtctcttctac} a {\tt ataaccaggattactat} a {\tt tgtgattgac} a {\tt gccattcaagatttgtg}$ 

221	S	L	F	Y	N	N	Q	D	Y	Y	М	V	I	D N/J	S A	H	S	R	F	V	DxD-like motif mutagenesis
721		tga tga	gtg atg	gga gga	tat	aaa	gat gat	gat gat	tga tga	acg <sup>.</sup>	tgc tgc	gcg acg	gct <sup>.</sup>	tat	gcc acc	cac cac	ttg ttg	aaa ggg	cgt cgt	aatg aatg	
241	Ρ	E	W	D	I	K	М	I	E	R	A	R	L	М	Р	т	W	G	v	М	
781	tc tc	gta <mark>gta</mark>	cta cta	ccc ccc	gaa <sup>.</sup> gaa	tgg tgg	gta gta	caca caca	acco acco	gga gga	gga gga	tcc ccc	aga <sup>.</sup> aga	tgc tgc	gga <mark>gga</mark>	gat <mark>gat</mark>	caa <mark>caa</mark>	caa <mark>caa</mark>	gac <mark>gac</mark>	ggat <mark>ggat</mark>	
261	S	Y	Y	Ρ	Ν	G	Y	Т	Ρ	Ε	D	Ρ	D	A	Е	Ι	N	К	т	D	N-glycosylation sequon
841	gt <mark>gt</mark>	tat <mark>tat</mark>	ggc <mark>ggc</mark>	cat <mark>cat</mark>	gtg <sup>.</sup> gtg <sup>.</sup>	taa taa	<mark>ggg</mark> ggg	ggt <mark>agt</mark>	tato tato	ctt.	aga aga	taa <mark>caa</mark>	tga tga	cat cat	tcc <mark>tcc</mark>	aaa <mark>aaa</mark>	gtt <mark>gtt</mark>	<mark>ddd</mark> ddd	ggc <mark>ggc</mark>	gcgg gcgg	
281	V	М	A	M	С	K	G	V	I	L	D	N	D	I	Ρ	K	L	G	A	R	
901	tg <mark>tg</mark>	gat gat	tga tga	tat tat	aag	gga gga	cca cca	tcc tcc	tgt <sup>.</sup> tgt	tct tct	cca cca	agg agg	ctt <sup>.</sup>	tgt tgt	agc <mark>agc</mark>	tgc tgc	tgg tgg	ata ata	tat tat	tttt tttt	
301	W	I		I	<u>R</u>	<u>D</u>	H	P	<u>v</u>	L	<u>Q</u>	<u> </u>	<u>F</u>	<u>v</u>	<u>A</u>	<u>A</u>	<u>G</u>	<u>Y</u>	<u> </u>	<u>F</u>	
961	<mark>dd</mark> dd	gga <mark>gga</mark>	tgc tgc	gca gca	gtt gtt	tgt tgt	gaa gaa	gga <mark>gga</mark>	tgt tgt		ttt ttt	tga tga	tcc	gta gta	cct	tcc tcc	att att	ttt ttt	att att	tgat tgat	
321	G	D	<u>A</u>	Q	<u>F</u>	<u>v</u>	<u>K</u>	D	V	Р	F.	D	Р	Y	ь.	Р	F.	Ц.	F.	D	
241	gg gg	aga aga r	.gga .gga .gga	gat gat		gta gta	cac cac	tgta tgta	acgo tcgo		gtg gtg W	gac gac	aago gago	cgg cgg	gtg gtg W	gga gga	cag cag	tta tta	ctg ctg	teet	
1081	G	ь сас	E	1 2++	ц + a+-	¥ +++	1 + a a	v + > >/	к а+ э	ц са+	w	т а а с	5	G t a a	w	и пра	ъ а++	т ++а	c at a	r aata	
361	dd. dd.	cac cac	ggc a	ttt F	tet	ttt <sup>.</sup>	tca tca	taao N	cta v	cat M	gcg gcg	aac aac m	cga cga D	tgc a	gcc gcc p	gag gag R	gtt F	ttg w	gtc gtc	cgtc cgtc v	
1141	t.t.	αac	aaq	aαa	tga	aat	aaa	aati	t.ca	tca	aga	- att	tga	aca	aca	gac	- ctc	cat.	ааа	acat	
381	tt L	gac T	aag R	aga D	tga	ggt V	gaa K	ggt V	tca H	ccg R	gga E	att F	tga E	acg R	gca 0	gac T	ctc S	cat I	aaa K	acgt R	
1201	qc	qtt	gta	ttt	aat	qcq	qcq	qaa	tqt	ctt	qaa	ctc	cac	cca	~ acc	cqt	tqt	qac	qqa	aqat	
401	gc A	att L	gta Y	ttt L	aat M	gcg R	gcg R	gaa⊣ N	tgto V	ctt L	gaa N	ctc S	caco T	cca Q	acc P	cgt V	tgt V	gac T	gga E	agat D	N-glycosylation sequon
1261	ga	ggc	gca	tcg	att	gaa	ccc	ggc	gat	tgg	acg	tga	gat	gga	aca	ctt	tgg	att	ggg	gatg	
421	ga E	<mark>ggc</mark> A	gca H	t <mark>cg</mark> R	atto L	gaa N	<mark>ccc</mark> P	ggc A	gat <sup>.</sup> I	t <mark>gg</mark> G	acg R	tga E	gato M	gga E	aca H	ctt F	tgg <mark>G</mark>	att L	ggg <mark>G</mark>	gagt M	tryptic peptide
																				S	(MS of 15KD band)
1321	tt tt	gcg <mark>gcg</mark>	acg acg	cat <mark>cat</mark>	ttc ctc	cga <sup>.</sup> cga	tta tta	ctg <mark>ctg</mark>	<mark>gcg</mark> . gcg.	ttt <sup>.</sup>	tgt <mark>tgt</mark>	gga <mark>gga</mark>	act <sup>.</sup> act	tag t <mark>ag</mark>	tga tga	tga <mark>tga</mark>	gtt <mark>gtt</mark>	tgt <mark>tgt</mark>	gaa <mark>aaa</mark>	ggaa <mark>ggaa</mark>	
441	L	R	R	I	S	D	Y	W	R	F	V	Е	L	S	D	Е	F	V	K	Е	
1381	aa <mark>aa</mark>	aga <mark>gga</mark>	tga <mark>tga</mark>	tga <mark>tga</mark>	gaa gaa	tcg t <mark>cg</mark>	ctg <mark>ctg</mark>	gaa gaa	<mark>ada</mark> . ada.	tgg <sup>.</sup> cgg <sup>.</sup>	tga <mark>tga</mark>	agg <mark>agg</mark>	gct gct	ctg ct <mark>g</mark>	tga cga	aaa aaa	ggc ggc	aac aac	act agt	tgga tgga	<b>E5AS1,</b> <u>E5AS2</u> <u>E5AS3,<b>E5L1</b></u>
461	K	D	D	Е	N	R	W	K	G	G	Ε	G	L	C	Е	K	A	Т	L V	G	
1441	ta	<u>a</u>																			
	та -	d																			

SUPPLEMENTAL FIG. 3. Synteny of OGNT genes in T. brucei, T. cruzi and L. major. OGNT1 (green boxes) and OGNTL (blue boxes) (A) and OGNT2 (pink boxes) (B) genes are shown relative to neighboring predicted coding regions. Shaded gray boxes represent conserved hypothetical protein orthologs; white boxes are hypothetical proteins that are not conserved among these chromosomes; red boxes represent casein kinase 2 (CK2); and yellow boxes are phosphoglycan β1-3-galactosyltransferaselike protein (PG $\beta$ 1-3GalT) and putative phosphoglycan  $\beta$ 1-2-arabinosyltransferase (PG $\beta$ 1-2AraT). Numbers above each box represent the identifier protein numbers for the corresponding T. brucei and L. maior chromosomes (indicated at the right). and for Τ. cruzi haplotypes Tc00.1047053508741.340/Tc00.1047053508741.360 for OGNT1/OGNTL and Tc00.1047053511759.70 for OGNT2, respectively.

**SUPPLEMENTAL FIG. 4.** Mapping the 5'-region of the *TcOGNT2* transcript. (A) Ethidium bromidestained agarose gel of the reverse transcription and PCR amplification of total epimastigote RNA using the TcOGNT2-specific reverse primer TcE5AS415 and the miniexon primer TcME33 (lane 2). Lane 1: same without reverse transcriptase (from a separate gel). Arrow marks position of the ~500 bp product. (B) The amplification product was vector-ligated and transfected into *E. coli*. Ten clones were sequenced, and digested with EcoR1 and BamHI to release the insert and analyzed by agarose gel electrophoresis. Ethidium bromide staining revealed minor size heterogeneity of the ~500 bp fragments (arrow), suggesting multiple splice sites. Alignment of the sequences with the 5'-UTR of CL-Brener *TcOGNT2-1* gene revealed splice donor sites at positions -70, -101, -109 relative to the translation start site (see Supplemental Figurre 2).

**SUPPLEMENTAL FIG. 5.** Expression of TcOGNT2 in *Dictyostelium*. A soluble protein fraction (300  $\mu$ g) prepared from sonicated microsomes of *D. discoideum* was assayed for 2 h in the absence or presence of peptide T29. *modB*<sup>-</sup>, DdGnt2-null strain DL118; *modB*<sup>-</sup> cat, transfected with TcOGNT2cat-myc(C-terminal). Similar absence of detectable activity (data not shown) occurred with the other constructs tested (Figure 1B), and with tagged TcOGNT1, and tagged TcOGNT1 or TcOGNTL coexpressed with TcOGNT2 (unpublished data). Protein expression was verified in all cases (not shown). Results are representative of five independent experiments.

**SUPPLEMENTAL FIG. 6.** Temperature dependence of TcOGNT2-dependent and endogenous enzyme activities. **(A)** Transferase activity was assayed in the presence or absence of peptide acceptor as described in Figure 4A. **(B)** UDP-[<sup>3</sup>H]GlcNAc hydrolysis activity was assayed as described in Fig. 4C. Data are plotted relative to values at 22°C, which are shown in corresponding vertical order in the 22° column. Similar results were obtained in a second trial using TcOGNT2 from hygromycin-resistant cells.



В





### Suppl. Figure 4

## Suppl. Figure 5



# Suppl. Figure 6

