Table S3 - PCR primers and protocols

1. Mutational Screening (genomic DNA)

Product	Primer	Sequence (5'-3')	Localization[1]	Buffer	Annealing (˚C)
A1	BCL2-FOR2	TGAATGAACCGTGTGACGTTA	Promoter	Buffer 3[2,3]	58
	BCL2-REV3	ACGATCCCATCAATCTTCAGC	Exon 1		
A2	BCL2-FOR4	TGGGAATCGATCTGGAAATCCTC	Exon 1	Buffer 3[2,3]	62
	BCL2-REV5	TCAGGTGGACCACAGGTGGC	Exon 2		
A3	BCL2-FOR6	GTCGCCAGGACCTCGCCG	Exon 2	Buffer 3[2,3]	60
	BCL2-REV7	TACAAAGAATGCAGCCGATG	Intron 2		
A4	BCL2-FOR8	AACAAGCTCCCCTGAAACAA	Intron 2	Buffer 3[2,3]	60
	BCL2-REV8	ACACCAAACAGTGGCATCAA	Intron 2		
A5	BCL2-FOR9	TTGTGTATGGTTTACCAGTTTGC	Intron 2	Buffer 3[2,3]	62
	BCL2-REV9	CATGTGTCTCTGTTTCCCACA	Intron 2		
B[8]	BCL2-Luc-F1[8]	GGACCAGGAGGAGGAGAAAG	Promoter	Reaction Buffer[4]	64
	BCL2-Luc-R1[8]	ATCAGGTGCGTTTCCCTGTA	Intron 1		
C	BCL2-F10	GAGGCGTGAAGCGGTCCCGT	Promoter	Buffer 3[2,5]	58
	BCL2-P012-LUC-R2	CTGAAGAGCTCCTCCACCAC	Exon 2		
D[9]	BCL2-F15'	GGGGGAGAACTTCGTAGCA	Promoter	Buffer 3[2,3]	62
	BCL2-R15'	CCCCTCTGCGACAGCTTAT	Exon 2		
E	BCL2-FOR4	TGGGAATCGATCTGGAAATCCTC	Exon 1	Pfu buffer[5,6]	62
	BCL2-REV5	TCAGGTGGACCACAGGTGGC	Exon 2		

2. RT-PCR amplification of BCL2-IgH translocated alleles [10]

F	BCL2-FOR5	TGTTCCGCGTGATTGAAGACAC	Exon 2	Pfu Cx buffer[7]	55
	J6-REV	TAGAGTGGCCATTCTTACCT	J6		

3. qCHIP

BCL2	BCL2-qCHIP-F4	CGGAGTTTAATCAGAAGAGGATTC		SYBR green PCR master mix [11]	60
	BCL2-R6	TTCGCAGAAGTCCTGTGATG			

4. qRT-PCR

BCL2	BCL2_RT_F2	CGGTTGGATGACTGAGTACCTGAACCG		SYBR green PCR master mix [11]	60
	BCL2_RT_R2	GCATCCCAGCCTCCGTTATC			
BCL6	BCL6_RT_F	CGCAACTCTGAAGAGCCACCTGCG		SYBR green PCR master mix [11]	60
	BCL6_RT_F	TTTGTGACGGAAATGCAGGTTA			
GAPDH	GAPDH-RTF2	CTGACTTCAACAGCGACACC		SYBR green PCR master mix [11]	60
	GAPDH-RTR2	CCCTGTTGCTGTAGCCAAAT			

Legend:

Templates: genomic DNA except product F (cDNA), sequencing primers available upon request

[1] Numbering according to GenBank accession No. NM_000633

[2] Buffer 3 (Expanded Long Template PCR System, Roche, 14277200)

[3] 10x PCRx Enhancer Solution (Invitrogen # 52391)

[4] Taq DNA Polymerase (Invitrogen #10342-020) with 10x reaction buffer

[5] with DMSO

[6] Pfu Turbo (Stratagene #600250-52) with 10x buffer

[7] Pfu Turbo Cx (Stratagene #600410) with 10x buffer

[8] after digestion AscI and EcoRV, the PCR products were also used for reporter constructs; for cases with mutations in primer sites, the following allele specific primers were used with the same PCR conditions:

2106:	BCL2(2106)_LUC_F1 TATCAGGCGCGCCGGACCAGGAGGAGGAGACAG
	BCL2(2106)_LUC_R1 TCATTGATATCATCAGGTGCGCATCCCTATA
2027:	BCL2(2027)_LUC_F2 TATCAGGCGCGCCCAGGACCAAGAGGAAACAGGG
	BCL2(2027)_LUC_R1 TCATTGATATCATCAGGTGCGTTTCCCTGTG
SUDHL6:	WT forward primer
	BCL2(SUDHL6)_LUC_R1 TCATTGATATCATCAGGTGCGTATTCCTGAA as the reverse primer

[9] used for SUDHL6 only

[10] used for DLBCL cell lines only

[11] SYBR green Applied Biosystems (#4309155)