

Supplementary Fig 1. BM cells (2×10^6) from RBP-J knock-out and control mice were cultured in 24-well plates in the presence of GM-CSF and IL4 for 9 days, and were stimulated with LPS for 12 h. Cells were counted under microscope, and were analyzed by FACS. The number of CD11c⁺ cells generated from 2×10^6 total BM cells were calculated and compared. Bars represent means ± standard deviation (n=5).



Supplementary Fig 2. BM cells from normal mice were cultured in the presence of GM-CSF and IL4 for 9 days, and were stimulated with or without LPS for 12 h. A. Cells were observed under SEM, and DCs with more than 50 dendrites and DCs with less than 50 dendrites were counted. B. Cells were analyzed by FACS. The result represented 3 independent experiments.



Supplementary Fig 3. OP9-Dll1 or OP9-GFP cells (2×10^5) were seeded in 24-well plates. BM cells (2×10^6) were then seeded and co-cultured in the presence of GM-CSF and IL4 for 9 days, and stimulated with LPS 12 h before the end of the culture. DCs were analyzed by FACS. The result represented 3 independent experiments.



Supplementary Fig 4. DC2.4, HEK293, or HeLa cells were co-transfected with pGL-CXCR4 and pEFBOS-NIC (150 ng), and the trans-activation of the mouse CXCR4 promoter by NIC was examined using luciferase assay with or without the addition of the inhibitors for the indicated signaling pathways. A. NF- κ B inhibitor was used in 3 types of cells, and the amounts of inhibitor added were 0, 3, 0, 3, 4, and 5 µg/ml. B. p38 inhibitor was used at concentrations of 0, 25, 50, and 100 µM. C. MEK inhibitor was used at concentrations of 0, 25, 50, and 100 µM. D. PI3K inhibitor was used at concentrations of 0, 25, 50, and 100 µM. Bars represent means ± standard deviation (n=5).