Figure S1. Surface plasmon resonance assay of 26-aa Bak BH3 peptide binding to Bcl-2, Mcl-1 and Bcl- x_L *in vitro*. A-C, the 26-aa Bak BH3 peptide was immobilized on a CM5 chip. Responses to different concentrations of Mcl-1 (A), Bcl- x_L (B) and Bcl-2 variant 1 (C) are shown. **D**, responses to Bak BH3 peptide chip are compared between Bcl-2, Bcl- x_L , Mcl-1 and GST at a concentration of 600 nM. The results shown are representative of 3 independent experiments using different chips and different protein batches.

Figure S2. Surface plasmon resonance assay of Bak protein binding to Bcl-2, Mcl-1 and Bcl- x_L in *vitro*. A, purified Bak Δ TM was immobilized on a CM5 chip. Responses to different concentrations of Bcl- x_L are shown. B, responses to Bak chip were compared between Bcl-2 (variant 1), Bcl- x_L , Mcl-1 and GST at a concentration of 600 nM.

Figure S3. Bak N85A/I86A binds to Bcl-2 (variant 2) in vitro. A, GST-Bcl-2 Δ TM was immobilized on a CM5 chip. Responses to different concentrations of Bak N85A/I86A are shown. **B**, responses of the Bcl-2 (variant 2) chip to Bak and Bak N85A/I86A at a concentration of 2 μ M were compared. Results shown are representative of 2 independent experiments using different chips and different protein batches. **C**, based on the surface plasmon resonance assay, the affinities of Bak and Bak N85A/I86A for Bcl-2 (variant 2) were determined. Results are mean and range of 2 independent experiments using different chips and different protein batches.

Figure S4. Bcl-2 knockdown and Mcl-1 knockdown cause Bak activation in Jurkat cells. 48 h after transfection with pCMS5A (empty vector), pCMS5A/Bcl-2 shRNA, or pCMS5A/Mcl-1 shRNA, Ab-1 was used to immunoprecipitate active Bak from CHAPS lysates. Recovered proteins and lysates were blotted for Bak. Heavy chain is shown here as loading control for the immunoprecipitates.

Figure S5. Different Bcl-2 variants exhibit different abilities to protect against Mcl-1 knockdowninduced apoptosis in Jurkat cells. 48 h after co-transfection with pCMS5C (EBFP vector, panels A and B) or pCMS5C-Mcl-1-shRNA (Mcl-1 sh, panels C-F) and pCMS5A (EGFP vector, panels A-C), pCMS5A-Bcl-2 variant 1 (panel D), pCMS5A-Bcl-2 variant 2 (panel E) or pCMS5A-Bcl-2 variant 3 (panel F), Jurkat cells were collected and stained with APC-annexin V. Panel A illustrates gate (R1) used to identify EBFP-histone H2B⁺ cells (indicating Mcl-1 knockdown), which were then analyzed for EGFP-histone H2B expression (indicating successful transfection of vector or Bcl-2 variants) and absence or presence of annexin V binding in panels B-F. The percentage of vector- or Bcl-2 variant-transfected cells that were positive for annexin V binding (upper right panel over lower right panel) is indicated to the right of each dot plot in panels B-F. These values are summarized in Fig. 5B of the main text.

Figure S6. Different Bcl-2 variants have different abilities to protect against cycloheximideinduced apoptosis in Jurkat cells. 24 h after transfection with pCMS5A (panels A and B), pCMS5A-Bcl-2 variant 1 (panel C), pCMS5A-Bcl-2 variant 2 (panel D) or pCMS5A-Bcl-2 variant 3 (panel E), Jurkat cells were treated with diluent (0.1% DMSO, panel A) or 3.5 μ M cycloheximide (panels B-E) for another 24 h, then stained with APC-annexin V. The percentage of vector- or Bcl-2 variant-transfected cells that were positive for annexin V binding (upper right panel over lower right panel) is indicated to the right of each dot plot. Additional samples (not shown) transfected with the Bcl-2 variants were treated with diluent for 24 h and stained with APC-annexin V. These values are summarized in Fig. 5D of the main text.

Figure S7. Further quantitation of Bcl-2 and Bax in Jurkat, Molt3, CEM and RL cells. A, whole cell lysates of 3×10^5 for Jurkat, Molt3, CEM and Daudi cells were subjected to western blot for Bcl-2.

Purified GST-Bcl-2 Δ TM (0.25, 0.5, 1.0, 2.0, 4.0, 8.0, 16.0 ng) is shown for comparison. **B**, whole cell lysates of 3×10^5 for Jurkat, Molt3, CEM, Daudi and RL cells were subjected to western blot for Bax. Purified GST-Bax Δ TM (0.15, 0.3, 0.6, 1.2, 2.4 ng) is shown for comparison.



Dai et al., Fig. S1



Dai et al., Fig. S2



Dai et al., Fig. S3



Dai et al., Fig. S4



Dai et al., Fig. S5



Dai et al., Fig. S6



Dai et al., Fig. S7