

**Syntheses of the *Stemona* Alkaloids (±)-Stenine, (±)-Neostenine,
and (±)-13-Epineostenine Using a Stereodivergent
Diels–Alder/Azido-Schmidt Reaction**

*Kevin J. Frankowski, Jennifer E. Golden, Yibin Zeng, Yao Lei and Jeffrey Aubé**

Department of Medicinal Chemistry and Center for Chemical Methodology and Library
Development, University of Kansas, Malott Hall, 1251 Wescoe Hall Drive, Lawrence,
Kansas 66045-7582

jaube@ku.edu

Supporting Information

Experimental details S-2

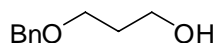
Copies of ^1H and ^{13}C spectra of new compounds S-42

Corresponding author:

Professor Jeffrey Aubé
Department of Medicinal Chemistry
1251 Wescoe Hall Drive, Room 4070
Malott Hall
University of Kansas
Lawrence, Kansas 66045-2506

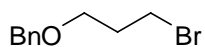
Experimental Section

General Procedures. All chemicals were purchased from commercial suppliers and used as received. Methylene chloride and THF were dried by being passed through two packed columns of neutral alumina under argon using a commercial solvent purification system prior to use. ^1H and ^{13}C NMR spectra were recorded at 400 and 100 MHz respectively in CDCl_3 (with 0.03% TMS as an internal standard). Chemical shifts are reported in parts per million (ppm) downfield from TMS. ^{13}C multiplicities were determined with the aid of an APT pulse sequence, differentiating the signals for methyl and methane carbons as “d” from methylene and quaternary carbons as “u”. The infrared (IR) spectra were acquired as thin films on a FT-IR spectrometer and the absorption frequencies are reported in cm^{-1} . Melting points were determined on a capillary melting point apparatus and are uncorrected. Low resolution mass spectroscopic data (CI, chemical ionization or FAB^+ , fast atom bombardment) were obtained with a quadrupole instrument. High resolution mass spectra (HRMS) [ESI+] were obtained using either a TOF or a double focusing spectrometer. Reaction flasks were oven or flame-dried and cooled under vacuum then purged with argon.

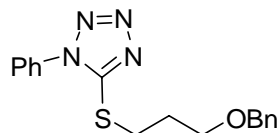


3-Benzyloxypropan-1-ol. Sodium hydride (60% dispersion in oil, 5.80 g, 116.0 mmol) was washed in triplicate with hexanes. After decanting the solvent the final time, dry DMF was added (150 mL). The mixture was cooled to 0 °C, and 1,3-propanediol (8.0 g, 105 mmol) was added slowly. After stirring for 10 min, benzyl bromide (12.5 ml, 105 mmol) was added cautiously. The mixture was allowed to acclimate to room temperature

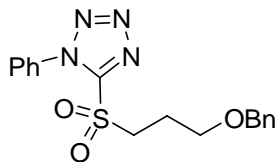
and stirred for 18 h. The reaction was quenched upon addition of water (200 mL) and subsequently extracted with EtOAc (6 x 100 mL). The organic layer was dried (Na₂SO₄), filtered, and concentrated under reduced pressure to afford a yellow oil. Flash chromatography (25% EtOAc/hexane) gave the known 3-benzyloxypropan-1-ol¹ (17.3 g, 104.0 mmol, 92% yield) as a pale yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 1.86 (p, *J* = 6.1 Hz, 2H), 3.51 (br s, 1H) 3.61 (t, *J* = 6.1 Hz, 2H), 3.72 (m, 2H), 4.50 (s, 2H), 7.30 (m, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ 32.8, 60.9, 68.8, 73.5, 128.0, 128.8, 138.7; IR (neat) 3330, 2850, 1090 cm⁻¹; MS (CI) *m/z* 167 (M⁺+1), 91; HRMS calcd for C₁₀H₁₅O₂ (M⁺+1): 167.1072, found 167.1067.



3-Benzyloxy-1-bromopropane. To a solution of benzyloxypropan-1-ol (10.2 g, 61.4 mmol) in dry CH₂Cl₂ (150 mL) was added CBr₄ (25.4 g, 76.7 mmol). After cooling the solution to 0 °C, PPh₃ (29.0 g, 110 mmol) was added in portions. The resulting red solution was stirred at room temperature for 18 h. The solvent was removed under reduced pressure, and the precipitate washed several times with ether (6 x 100 mL) and filtered. The collective ether extracts were concentrated to afford a yellow oil. Flash chromatography (3% EtOAc/hexane) gave the known 3-benzyloxy-1-bromopropane² (14.0 g, 61.1 mmol, 99% yield) as a pale yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 2.06 (p, *J* = 6.3 Hz, 2H), 3.46 (t, *J* = 6.6 Hz, 2H), 3.53 (t, *J* = 5.8 Hz, 2H), 4.45 (s, 2H), 7.29 (m, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ 31.2, 33.5, 68.2, 73.6, 128.13, 128.15, 128.9, 138.8; IR (neat) 2820, 1090, 730, 690 cm⁻¹; MS (CI) *m/z* 228 (M⁺-1), 91; HRMS calcd for C₁₀H₁₂OBr (M⁺-1): 227.0072, found 227.0067.

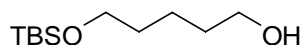


5-(3-Benzyloxypropylsulfanyl)-1-phenyl-1H-tetrazole. Sodium hydride (60% dispersion in oil, 3.70 g, 92.9 mmol) was washed in triplicate with hexanes. After decanting the solvent the final time, dry DMF was added (225 mL). The reaction mixture was cooled to 0 °C, and 1-phenyl-1H-tetrazole-5-thiol (17.7 g, 77.4 mmol) was added slowly. After stirring for 10 min, 3-benzyloxy-1-bromopropane (13.8 g, 77.4 mmol) was added slowly, and the resulting solution was stirred for 18 h at room temperature. The reaction was quenched upon addition of water (500 mL) and subsequently extracted with EtOAc (6 x 100 mL). The organic layer was dried (Na₂SO₄), filtered, and concentrated under reduced pressure to afford a yellow oil. Flash chromatography (25% EtOAc/hexane) gave 5-(3-benzyloxypropylsulfanyl)-1-phenyl-1H-tetrazole (25.0 g, 76.6 mmol, 99% yield) as a pale yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 2.18 (p, *J* = 6.2 Hz, 2H), 3.53 (t, *J* = 7.0 Hz, 2H) 3.63 (t, *J* = 5.8 Hz, 2H), 4.52 (s, 2H), 7.32 (m, 5H), 7.56 (s, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ 29.6, 30.8, 68.4, 73.5, 124.2, 128.11, 128.13, 128.8, 130.2, 130.5, 134.1, 138.5, 154.8; IR (neat) 2820, 1680, 1480 cm⁻¹; MS (CI) *m/z* 327 (M⁺+1), 173, 91, 84; HRMS calcd for C₁₇H₁₈N₄OS (M⁺+1): 327.1280, found 327.1292.



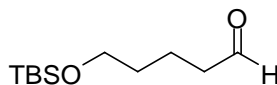
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5-(3-Benzyloxy-propyl-sulfonyl)-1-phenyl-1H-tetrazole 13. To a stirred solution of 5-(3-benzyloxypropylsulfonyl)-1-phenyl-1H-tetrazole (17.1 g, 52.4 mmol) in CH₂Cl₂ (400 mL) was added NaHCO₃ (13.2 g, 157 mmol) and *m*-CPBA (27.1 g, 157.0 mmol). The resulting mixture was stirred under an Ar atmosphere for 18 h and then quenched with 10% aq NaOH solution (150 mL). After extracting the aqueous layer with EtOAc (5 x 100 mL), the organic layer was dried (Na₂SO₄), filtered, and concentrated under reduced pressure to afford an orange oil. Flash chromatography (25% EtOAc/hexane) gave **13** (17.5 g, 48.8 mmol, 93% yield) as a pale yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 2.25-2.32 (m, 2H), 3.65 (t, *J* = 5.8 Hz, 2H), 3.90 (m, 2H), 4.52 (s, 2H), 7.32-7.38 (m, 5H), 7.60-7.70 (m, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ 23.4, 54.0, 67.5, 73.4, 125.6, 128.2, 128.3, 128.9, 130.1, 131.9, 133.4, 138.2, 153.9; IR (neat) 2820, 1580, 1480, 1320, cm⁻¹; MS (CI) *m/z* 359 (M⁺+1), 131; HRMS calcd for C₁₇H₁₉N₄O₃S (M⁺+1): 359.1178, found 359.1182.



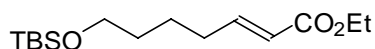
5-(*tert*-Butyldimethylsilyloxy)-pentan-1-ol. Sodium hydride (60% dispersion in oil, 3.20 g, 63.4 mmol) was washed in triplicate with hexanes. After decanting the solvent the final time, dry THF was added (275 mL). The reaction mixture was cooled to 0 °C, and 1,5-pentanediol was added slowly (6.0 g, 57.6 mmol). After stirring for 45 min at room temperature, the mixture was cooled to 0 °C, and *tert*-butyldimethylchlorosilane

(8.60 g, 57.6 mmol) was added cautiously. The mixture was allowed to acclimate to room temperature and stirred for an additional 45 min. The reaction was quenched upon addition of 10% aq K₂CO₃ and subsequently extracted with ether (4 x 100 mL). The organic layer was dried (Na₂SO₄), filtered, and concentrated under reduced pressure to afford a yellow oil. Flash chromatography (15% EtOAc/hexane) gave 5-(*tert*-Butyldimethylsilyloxy)-pentan-1-ol³ (12.6 g, 57.7 mmol, 99% yield) as a colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 0.06 (s, 6H), 0.90 (s, 9H), 1.37-1.45 (m, 2H), 1.52-1.63 (m, 4H), 1.71 (br s, 1H), 3.61-3.67 (m, 4H); ¹³C NMR (100.6 MHz, CDCl₃) δ -4.9, 18.8, 22.4, 26.4, 32.8, 32.9, 63.3, 63.5; IR (neat) 3300, 2900, 1240 cm⁻¹; MS (CI) *m/z* 219 (M⁺+1), 161, 105, 92, 75, 69; HRMS calcd for C₁₁H₂₇O₂Si (M⁺+1): 219.1780, found 219.1763.



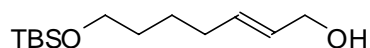
5-(*tert*-Butyldimethylsilyloxy)-pentanal. A solution of oxalyl chloride (5.0 mL, 57.8 mmol) in dry CH₂Cl₂ (100 mL) was cooled to -78 °C under an atmosphere of Ar. A solution of DMSO (8.20 mL, 116 mmol) in CH₂Cl₂ (10 mL) was added at a rate such that the reaction temperature remained below -65 °C. After stirring for 5 min, a solution of 5-(*tert*-Butyldimethylsilyloxy)-pentan-1-ol (12.6 g, 57.8 mmol) in CH₂Cl₂ (15 mL) was added slowly, and the resulting mixture was stirred for 15 min. Next, NEt₃ (40 mL, 289 mmol) was added slowly. After stirring the reaction for 10 additional min at -70 °C, the cooling bath was removed and the reaction was allowed to warm for ca. 45 min. Upon reaching room temperature, water (100 mL) was added and stirring continued for 15 min. The reaction mixture was transferred to a separatory funnel, washed

successively with 5% HCl (100 mL), saturated NaHCO₃ solution (100 mL), and brine (50 mL). The organic layer was dried (Na₂SO₄), filtered, and concentrated under reduced pressure to afford an oil. Flash chromatography (15% EtOAc/hexane) gave 5-(*tert*-butyldimethylsilyloxy)-pentanal⁴ (12.5 g, 57.8 mmol, 99% yield) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 0.04 (s, 6H), 0.88 (s, 9H), 1.50-1.57 (m, 2H), 1.69 (p, *J* = 7.6 Hz, 2H), 2.45 (dt, *J* = 7.3 Hz, 1.6 Hz, 2H), 3.62 (t, *J* = 6.2 Hz, 2H), 9.76 (s, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ -4.98, 18.7, 19.0, 26.3, 32.5, 44.0, 62.9, 203.0; IR (neat) 2940, 1730 cm⁻¹; MS (CI) *m/z* 217 (M⁺+1), 154, 136; HRMS calcd for C₁₁H₂₅O₂Si (M⁺+1): 217.1624, found 217.1609.

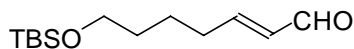


(*E*)-7-(*tert*-Butyldimethylsilyloxy)-hept-2-enoic acid ethyl ester. A solution of 5-(*tert*-butyldimethylsilyloxy)-pentanal (5.10 g, 23.6 mmol) and (carbethoxymethylene)triphenylphosphorane (8.20 g, 23.6 mmol) in CH₂Cl₂ (175 mL) was heated to reflux for 18 h. The reaction was cooled to room temperature, diluted with water and extracted with pentane. The organic extract was dried (Na₂SO₄), filtered, and concentrated under reduced pressure to afford a yellow oil. Flash chromatography (15% EtOAc/hexane) gave (*E*)-7-(*tert*-butyldimethylsilyloxy)-hept-2-enoic acid ethyl ester⁵ (6.6 g, 23.0 mmol, 98% yield) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 0.05 (s, 6H), 0.84 (s, 9H), 1.28 (t, *J* = 7.1 Hz, 3H), 1.50-1.54 (m, 4H), 2.19-2.24 (m, 2H), 3.59-3.62 (m, 2H), 4.17 (q, *J* = 7.1 Hz, 2H), 5.83 (dd, *J* = 15.5 Hz, 1.3 Hz, 1H), 6.96 (dt, *J* = 15.6 Hz, *J* = 6.9 Hz, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ -4.9, 14.7, 18.7, 24.7, 26.3,

32.3, 32.6, 60.5, 63.1, 121.8, 149.5, 167.1; IR (neat) 2925, 1720, 1660 cm^{-1} ; MS (CI) m/z 287 ($M^+ + 1$), 229, 81; HRMS calcd for $\text{C}_{15}\text{H}_{31}\text{O}_3\text{Si}$ ($M^+ + 1$): 287.2042, found 287.2063.

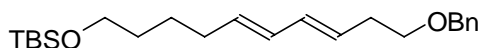


(E)-7-(tert-Butyldimethylsilyloxy)-hept-2-en-1-ol. To a solution of (E)-7-(tert-butyldimethylsilyloxy)-hept-2-enoic acid ethyl ester (11.2 g, 39.1 mmol) in diethyl ether (250 mL) was added DIBAL-H (1.0 M in hexanes, 86.0 mL, 86.1 mmol) at $-78\text{ }^\circ\text{C}$. The reaction was stirred at $-20\text{ }^\circ\text{C}$ for 1 h and then warmed to room temperature. After stirring at room temperature for 2 h, an equal volume of saturated aq potassium sodium tartrate was added slowly, and the resulting voluminous mixture was stirred for an additional 18 h. The organic layer was separated, and the aqueous layer was extracted with EtOAc (2 x 150 mL). The organic extracts were combined, dried (Na_2SO_4), filtered, and concentrated to afford a colorless oil. Flash chromatography (15% EtOAc/hexane) gave (E)-7-(tert-butyldimethylsilyloxy)-hept-2-en-1-ol⁶ (9.5 g, 38.9 mmol, 99% yield) as a colorless oil. ^1H NMR (400 MHz, CDCl_3) δ 0.06 (s, 6H), 0.90 (s, 9H), 1.42-1.56 (m, 5H), 2.07 (m, 2H), 3.62 (t, $J = 6.4\text{ Hz}$, 2H), 4.10 (d, $J = 5.3\text{ Hz}$, 2H), 5.61-5.74 (m, 2H); ^{13}C NMR (100.6 MHz, CDCl_3) δ -4.9, 18.8, 25.8, 26.4, 32.4, 32.7, 63.4, 64.2, 129.5, 133.7; IR (neat) 3330, 2930, 1470 cm^{-1} ; MS (CI) m/z 245 ($M^+ + 1$), 227, 187, 115; HRMS calcd for $\text{C}_{13}\text{H}_{29}\text{O}_2\text{Si}$ ($M^+ + 1$): 245.1937, found 245.1930.



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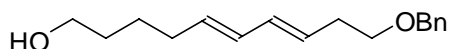
(*E*)-7-(*tert*-Butyldimethylsilyloxy)-hept-2-enal 14. (*E*)-7-(*tert*-Butyldimethylsilyloxy)-hept-2-enal⁷ **14** was prepared from (*E*)-7-(*tert*-butyldimethylsilyloxy)-hept-2-en-1-ol using a Swern oxidation, as described above for 5-(*tert*-butyldimethylsilyloxy)-pentanal. Flash chromatography (15% EtOAc/hexane) gave (*E*)-7-(*tert*-Butyldimethylsilyloxy)-hept-2-enal **14** (7.1 g, 29.3 mmol, 99% yield) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 0.05 (s, 6H), 0.89 (s, 9H), 1.57 (m, 4H), 2.35 (m, 2H), 3.63 (m, 2H), 6.12 (dd, *J* = 15.6 Hz, 7.9 Hz, 1H), 6.86 (dt, *J* = 15.6 Hz, 6.7 Hz, 1H), 9.51 (d, *J* = 7.9 Hz, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ -4.9, 18.7, 24.6, 26.3, 32.5, 32.9, 63.0, 133.4, 159.1, 194.5; IR (neat) 2900, 1670, 1080 cm⁻¹; MS (CI) *m/z* 243 (M⁺+1), 185, 111, 75; HRMS calcd for C₁₃H₂₇O₂Si (M⁺+1): 242.1780, found 243.1775.



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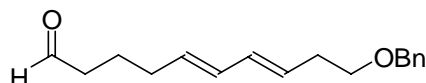
(*5E,7E*)-(10-Benzyloxy-deca-5,7-dienyloxy)-*tert*-butyldimethylsilane 15. A solution of sulfone **13** (10.0 g, 27.9 mmol) in THF (175 mL) was cooled to -78 °C, and a solution of lithium bis(trimethylsilyl)amide (30.7 mL, 30.7 mmol) in THF (20 mL) was added slowly. The resulting solution was stirred at -65 °C for 1 h, and then a solution of (*E*)-7-(*tert*-butyldimethylsilyloxy)-hept-2-enal **14** (6.90 g, 28.5 mmol) in THF (20 mL) was added at a rate such that the temperature remained below -65 °C. The orange solution was stirred for an additional hour at -65 °C and then allowed to acclimate to room temperature over 18 h. Water (150 mL) was added and the resulting mixture stirred

for 1 h. The reaction was transferred to a separatory funnel, extracted with Et₂O (3 x 100 mL), dried (Na₂SO₄), filtered, and concentrated to afford a yellow oil. Flash chromatography (3% EtOAc/hexane) gave **15** (9.6 g, 25.6 mmol, 90% yield) as a pale yellow oil. The ratio of (5*E*,7*E*)-**15**/(5*E*,7*Z*)-**15** was determined to be ca. 85:15 by ¹H NMR. (5*E*,7*E*)-**15**: ¹H NMR (400 MHz, CDCl₃) δ 0.08 (s, 6H), 0.93 (s, 9H), 1.37-1.49 (m, 2H), 1.52-1.59 (m, 2H), 2.08-2.13 (m, 2H), 2.39-2.44 (m, 2H), 4.55 (s, 2H), 5.57-5.65 (m, 2H), 6.01-6.14 (m, 2H), 7.28-7.37 (m, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ -4.68, 18.8, 22.6, 26.4, 32.8, 33.1, 33.5, 63.6, 70.3, 73.3, 127.9, 128.0, 128.4, 128.7, 130.8, 132.7, 133.3, 138.9; IR (neat) 2900, 1450, 1090 cm⁻¹; MS (CI) *m/z* 375 (M⁺+1), 243, 91; HRMS calcd for C₂₃H₃₉O₂Si (M⁺+1): 375.2719, found 375.2690. (5*E*,7*Z*)-**15**: ¹H NMR (400 MHz, CDCl₃, diagnostic peaks only) δ 2.51-2.56 (m, 2H), 4.56 (m, 2H), 5.33-5.42 (m, 1H), 5.65-5.75 (m, 1H), 6.34-6.42 (m, 1H); ¹³C NMR (100.6 MHz, CDCl₃, diagnostic peaks only) δ 26.0, 28.8, 33.0, 63.4, 70.3, 125.7, 126.1, 128.0, 129.1, 130.9, 135.6.

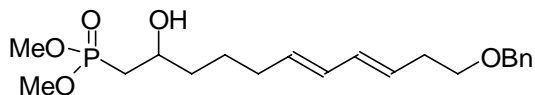


(5*E*,7*E*)-10-Benzyloxy-deca-5,7-dien-1-ol. To a stirred solution of diene **15** (5.70 g, 15.2 mmol) in ethanol (150 mL) was added pyridinium *p*-toluenesulfonate (1.10 g, 4.57 mmol). After stirring for 18 h, the solution was concentrated to give a viscous oil. Flash chromatography (25% EtOAc/hexane) gave (5*E*,7*E*)-10-Benzyloxy-deca-5,7-dien-1-ol (3.8 g, 14.6 mmol, 95% yield) as a colorless oil. The ratio of (5*E*,7*E*)-diene/(5*E*,7*Z*)-diene was determined to be ca. 85:15 by ¹H NMR. (5*E*,7*E*)-diene: ¹H NMR (400 MHz, CDCl₃) δ 1.41-1.63 (m, 5H), 2.11 (m, 2H), 2.40 (m, 2H), 3.53 (t, *J* = 6.8 Hz, 2H), 3.65 (t, *J* = 6.6 Hz, 2H), 4.54 (s, 2H), 5.62 (dt, *J* = 14.4 Hz, *J* = 6.9 Hz, 2H), 6.07 (m, 2H),

7.32 (m, 5H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 25.9, 32.6, 32.7, 33.5, 63.2, 70.3, 73.3, 128.0, 128.1, 128.6, 128.8, 130.9, 132.6, 133.0, 138.8; IR (neat) 3350, 2900, 1440, 1090 cm^{-1} ; MS (CI) m/z 261 ($\text{M}^+ + 1$), 243, 169, 91; HRMS calcd for $\text{C}_{17}\text{H}_{25}\text{O}_2$ ($\text{M}^+ + 1$): 261.1855, found 261.1853. (*5E,7Z*)-diene: ^1H NMR (400 MHz, CDCl_3 , diagnostic peaks only) δ 2.42 (m, 2H), 5.37 (m, 1H), 5.71 (m, 1H), 6.34 (m, 1H); ^{13}C NMR (100.6 MHz, CDCl_3 , diagnostic peaks only) δ 25.8, 28.8, 33.0, 70.2, 126.0, 126.2, 128.1, 130.8, 135.3.

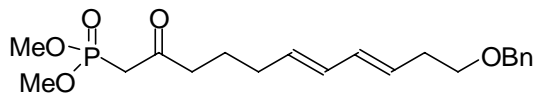


(*5E,7E*)-10-Benzyloxy-deca-5,7-dienal. The corresponding aldehyde was prepared from (*5E,7E*)-10-Benzyloxy-deca-5,7-dien-1-ol using a Swern oxidation, as described above. Flash chromatography (15% EtOAc/hexane) gave (*5E,7E*)-10-benzyloxy-deca-5,7-dienal (10.0 g, 38.7 mmol, 99% yield) as a yellow oil. The ratio of (*5E,7E*)-diene / (*5E,7Z*)-diene was determined to be ca. 85:15 by ^1H NMR. (*5E,7E*)-diene: ^1H NMR (400 MHz, CDCl_3) δ 1.62-1.71 (m, 2H), 1.97-2.10 (m, 2H), 2.32-2.38 (m, 4H), 3.47 (t, $J = 6.7$ Hz, 2H), 4.46 (s, 2H), 5.45-5.61 (m, 2H), 5.95-6.07 (m, 2H), 7.25 (m, 5H), 9.67 (s, 1H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 22.0, 32.2, 33.5, 43.5, 70.2, 73.2, 127.9, 128.0, 128.8, 129.2, 131.7, 131.8, 132.3, 139.0, 202.6; IR (neat) 2800, 1695, 1080 cm^{-1} ; MS (CI) m/z 259 ($\text{M}^+ + 1$), 154, 136; HRMS calcd for $\text{C}_{12}\text{H}_{23}\text{O}_2$ ($\text{M}^+ + 1$): 259.1698, found 259.1689. (*5E,7Z*)- diene: ^1H NMR (400 MHz, CDCl_3 , diagnostic peaks only) δ 2.49 (m, 2H), 5.33 (m, 1H), 6.33 (m, 1H); ^{13}C NMR (100.6 MHz, CDCl_3 , diagnostic peaks only) δ 28.9, 32.5, 70.1, 126.6, 127.0, 127.9, 130.6, 134.1.



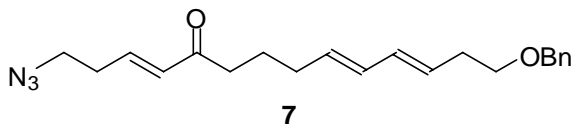
(6*E*,8*E*)-(11-benzyloxy-2-hydroxy-undeca-6,8-dienyl)-phosphonic acid

dimethyl ester. To a cooled solution of dimethyl methyl phosphonate (2.60 mL, 23.8 mmol) in THF (25 mL) at -78 °C was added *n*-butyllithium (1.6 M in hexanes, 15.5 mL, 23.3 mmol) slowly. The solution was stirred at -78 °C for 1 h, and then (5*E*,7*E*)-10-benzyloxy-deca-5,7-dienal (2.80 g, 10.8 mmol) was added as a solution in THF (10 mL). Stirring was continued at -78 °C for 3h and then at 0 °C for an additional hour. The reaction mixture was quenched at 0 °C with saturated aq NH₄Cl solution, transferred to a separatory funnel, and extracted with EtOAc (3 x 100 mL). The organic extracts were dried (Na₂SO₄), filtered, and concentrated to afford a yellow oil. Flash chromatography (100% EtOAc) gave β-hydroxyphosphonate (3.8 g, 9.9 mmol, 92% yield) as a pale yellow oil. The ratio of (6*E*,8*E*)-diene/(6*E*,8*Z*)-diene was determined to be ca. 85:15 by ¹H NMR. (6*E*,8*E*)-diene: ¹H NMR (400 MHz, CDCl₃) δ 1.39-1.56 (m, 4H), 1.90 (m, 1H), 1.95 (m, 1H), 2.01-2.14 (m, 2H), 2.33-2.40 (m, 2H), 3.48 (t, *J* = 6.8 Hz, 2H), 3.73 (d, *J* = 10.9 Hz, 6H), 3.87 (m, 1H), 3.96 (br s, 1H), 4.49 (s, 2H), 5.46-5.60 (m, 2H), 5.93-6.08 (m, 2H), 7.26 (m, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ 25.5, 32.7, 33.4, 38.1, 38.2, 52.6, 52.8, 66.5, 70.2, 73.2, 127.9, 128.0, 128.6, 128.7, 131.0, 132.9, 138.8; IR (neat) 3350, 2890, 1200 cm⁻¹; MS (CI) *m/z* 383 (M⁺+1), 357, 207, 91; HRMS calcd for C₂₀H₃₂O₅P (M⁺+1): 383.198, found 383.1968. (6*E*,8*Z*)-diene: ¹H NMR (400 MHz, CDCl₃, diagnostic peaks only) δ 2.46 (dd, *J* = 13.3 Hz, 8.0 Hz, 2H), 4.50 (s, 2H), 5.33 (m, 1H), 5.60 (m, 1H), 6.34 (m, 1H); ¹³C NMR (100.6 MHz, CDCl₃, diagnostic peaks only) δ 28.8, 32.7, 33.0, 33.7, 38.1, 38.3, 52.7, 52.8, 66.5, 70.2, 127.9.

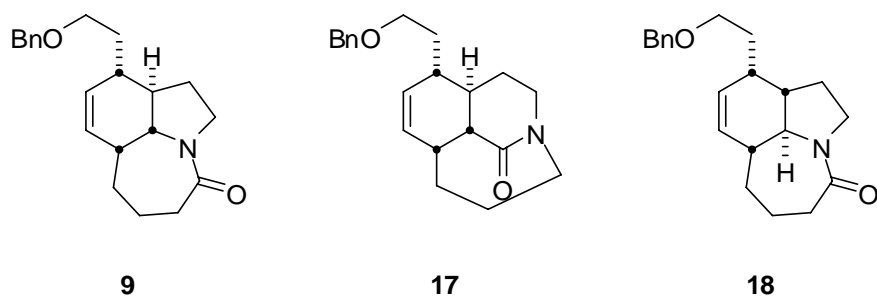


16

(6E,8E)-(11-Benzyloxy-2-oxo-undeca-6,8-dienyl)-phosphonic acid dimethyl ester 16. To a stirred solution of hydroxyphosphonate (2.90 g, 7.58 mmol) in dry CH₂Cl₂ (50 mL) was added activated 4Å sieves, *N*-morpholine-*N*-oxide (1.30 g, 11.4 mmol) and TPAP (0.13 g, 0.38 mmol). After stirring the resulting black mixture for 72 h, the reaction mixture was filtered through Celite and washed with EtOAc. Concentration of the filtrate provided a black oil. Flash chromatography (100% EtOAc) gave **16** (2.2 g, 5.8 mmol, 75% yield) as a yellow oil. The ratio of (6E,8E)-**16**/(6E,8Z)-**16** was determined to be ca. 85:15 by ¹H NMR. (6E,8E)-**16**: ¹H NMR (400 MHz, CDCl₃) δ 1.64-1.73 (m, 2H), 2.05-2.19 (m, 2H), 2.36-2.41 (m, 2H), 2.59-2.63 (m, 2H), 3.05 (d, *J* = 22.7 Hz, 2H), 3.51 (t, *J* = 6.8 Hz, 2H), 3.75-3.77 (d, *J* = 11.2 Hz, 6H), 4.51 (s, 2H), 5.49-5.63 (m, 2H), 5.97-6.09 (m, 2H), 7.33 (m, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ 23.3, 32.0, 33.5, 41.1, 43.7, 53.4, 70.2, 73.3, 127.9, 128.1, 128.8, 129.1, 131.5, 132.0, 132.3, 138.8, 202.2; IR (neat) 2950, 1720, 1035 cm⁻¹; MS (CI) *m/z* 381 (M⁺+1), 345, 255, 91; HRMS calcd for C₂₀H₃₀O₅P (M⁺+1): 381.1831, found 381.1836. (6E,8Z)-**16**: ¹H NMR (400 MHz, CDCl₃, diagnostic peaks only) δ 2.47-2.52 (m, 2H), 4.52 (s, 2H), 5.34-5.39 (m, 1H), 6.28-6.34 (m, 1H); ¹³C NMR (100.6 MHz, CDCl₃, diagnostic peaks only) δ 28.8, 32.3, 42.4, 70.2, 126.4, 126.8, 130.5, 134.3.



(3E,9E,11E)-1-Azido-14-benzyloxy-tetradeca-3,9,11-trien-5-one 7. Prior to use, Ba(OH)₂•8H₂O was dried in a 140 °C oven for 2 h (NOTE: drying of the hydroxide for shorter or longer periods of time resulted in lower overall yields of **7**). Phosphonate **16** (1.30 g, 3.42 mmol) was placed in a 100 mL flask with THF (30 mL) and Ba(OH)₂•8H₂O (0.86 g, 2.74 mmol). The mixture was stirred for 30-45 min, causing it to turn white in color. 3-azidopropanal⁸ (0.34 g, 3.42 mmol) was added slowly in 40:1 THF/H₂O (15 mL). After stirring the gelatinous material for 6 h, the solution was poured over saturated NaHCO₃ solution and extracted with EtOAc (4 x 100 mL). The extracts were dried (Na₂SO₄), filtered, and concentrated to give a yellow oil. Flash chromatography (15% EtOAc/Hex) afforded **7** (1.0 g, 2.8 mmol, 85% yield) as a yellow oil. The ratio of (3E,9E,11E)-**7**/(3E,9E,11Z)-**7** was determined to be ca. 85:15 by ¹H NMR. (3E,9E,11E)-**7**: ¹H NMR (400 MHz, CDCl₃) δ 1.67-1.76 (m, 2H), 2.02-2.16 (m, 2H), 2.39-2.43 (m, 2H), 2.43-2.55 (m, 4H), 3.39 (t, *J* = 6.7 Hz, 2H), 3.49 (t, *J* = 6.8 Hz, 2H), 4.50 (s, 2H), 5.45-5.70 (m, 2H), 5.97-6.09 (m, 2H), 6.14 (d, *J* = 16.0 Hz, 1H), 6.73 (dt, *J* = 16.0 Hz, *J* = 6.8 Hz, 1H), 7.26-7.33 (m, 5H); ¹³C NMR (100.6 MHz, CDCl₃) δ 23.9, 32.2, 32.3, 33.5, 39.9, 50.0, 70.2, 73.3, 127.9, 128.0, 128.1, 128.8, 129.0, 131.3, 132.3, 132.4, 132.7, 142.2, 200.2; IR (neat) 2050, 1700, 1650, 1600 cm⁻¹; MS (FAB⁺) *m/z* 354 (M⁺+1), 154, 136; HRMS calcd for C₂₁H₂₈N₃O₂ (M⁺+1): 354.2182, found 354.2162. (3E,9E,11Z)-**7**: ¹H NMR (400 MHz, CDCl₃, diagnostic peaks only): δ 5.34-5.38 (m, 1H), 6.28-6.35 (m, 1H); ¹³C NMR (100.6 MHz, CDCl₃, diagnostic peaks only) δ 14.6, 21.5, 27.4, 28.8, 32.6, 39.9, 60.8, 126.3, 126.8, 130.6, 134.6.

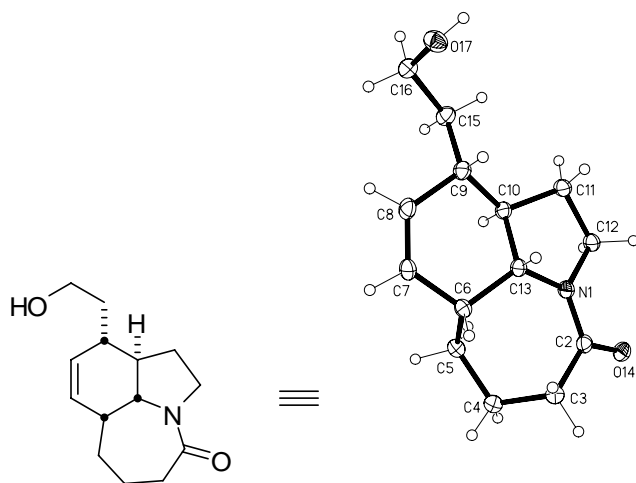


Lactams 9, 17 and 18. To a flame dried 50 mL flask was added azidotriene **7** (0.25 g, 0.71 mmol) in CH_2Cl_2 (35 mL) and MeAlCl_2 (0.71 mL, 1.0 M soln in toluene, 0.71 mmol). Refluxing the yellow solution for 48 h produced a dark greenish solution that was cooled to room temperature and then poured over saturated aq NaHCO_3 solution (100 mL). Upon shaking, the dark solution turned yellow in color. The mixture was extracted with EtOAc (3×100 mL), the combined organic layers dried (Na_2SO_4), filtered, and concentrated to afford a yellow oil. Flash chromatography (100% EtOAc) afforded the major lactam isomer **9** (100 mg, 0.31 mmol, 43% yield) as a viscous oil, the bridged lactam **17** (56 mg, 0.17 mmol, 24% yield) as a viscous oil, and the minor lactam isomer **18** (27 mg, 0.083 mmol, 12% yield as a white solid).

Major lactam **9** ($R_f = 0.17$): ^1H NMR (400 MHz, CDCl_3) δ 1.39-1.48 (m, 3H), 1.54-1.64 (m, 2H), 1.72 (m, 2H), 1.85-1.93 (m, 2H), 2.13-2.16 (m, 1H), 2.28 (ddd, $J = 13.2$ Hz, 5.1 Hz, 1.7 Hz, 1H), 2.59 (m, 1H), 2.67 (m, 1H), 3.42 (m, 1H), 3.47-3.52 (m, 3H), 3.57 (t, $J = 6.6$ Hz, 1H), 4.51 (m, 2H), 5.53-5.59 (m, 1H), 5.63-5.66 (m, 1H), 7.31 (m, 5H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 23.2, 25.4, 26.8, 33.2, 33.4, 36.5, 38.0, 43.9, 45.9, 62.6, 68.0, 73.5, 128.01, 128.03, 128.8, 130.7, 130.9, 138.7, 171.5; IR (neat) 2840, 1625 cm^{-1} ; MS (CI) m/z 326 ($\text{M}^+ + 1$), 234, 91; HRMS calcd for $\text{C}_{21}\text{H}_{28}\text{NO}_2$ ($\text{M}^+ + 1$): 326.2120, found 326.2129.

Bridged lactam **17** ($R_f = 0.63$): ^1H NMR (400 MHz, CDCl_3) δ 1.40-1.64 (m, 4H), 1.72-2.01 (m, 5H), 2.19 (m, 1H), 2.36 (m, 1H), 2.48 (dt, $J = 12.4$ Hz, $J = 4.8$ Hz, 1H), 2.58 (dd, $J = 10.0$ Hz, $J = 4.5$ Hz, 1H), 2.89 (dt, $J = 11.2$ Hz, $J = 4.2$ Hz, 1H), 3.56 (t, $J = 6.3$ Hz, 2H), 3.69 (dt, $J = 15.3$ Hz, $J = 4.2$ Hz, 2H), 4.49 (m, 2H), 5.62-5.67 (m, 2H), 7.27-7.37 (m, 5H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 24.9, 25.4, 31.3, 32.2, 39.3, 39.9, 41.3, 53.4, 56.2, 56.8, 68.2, 73.6, 128.0, 128.1, 128.8, 132.6, 133.4, 138.8, 188.1; IR (neat) 2940, 1690 cm^{-1} ; MS (FAB $^+$) m/z 326 ($\text{M}^+ + 1$), 234, 91; HRMS calcd for $\text{C}_{21}\text{H}_{28}\text{NO}_2$ ($\text{M}^+ + 1$): 326.2120, found 326.2122.

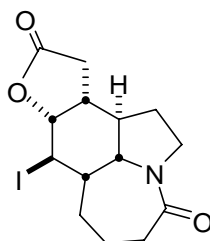
Minor lactam **18** ($R_f = 0.25$). Mp: 85-88 $^\circ\text{C}$. ^1H NMR (400 MHz, CDCl_3) δ 1.20-1.26 (m, 1H), 1.44-1.47 (m, 1H), 1.57-1.63 (m, 2H), 1.76-1.79 (m, 1H), 1.86-1.91 (m, 2H), 1.95-1.99 (m, 1H), 2.08-2.17 (m, 2H), 2.31 (t, $J = 13.9$ Hz, 1H), 2.59-2.67 (m, 2H), 3.00 (dd, $J = 10.7$ Hz, $J = 9.3$ Hz, 1H), 3.20-3.27 (m, 1H), 3.55-3.61 (m, 2H), 3.90 (dd, $J = 11.6$ Hz, $J = 8.0$ Hz, 1H), 4.52 (m, 2H), 5.37 (m, 1H), 5.75-5.79 (m, 1H), 7.28-7.36 (m, 5H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 23.2, 25.7, 30.8, 34.4, 37.0, 38.8, 45.1, 46.8, 47.2, 60.1, 68.7, 73.5, 128.1, 128.8, 131.3, 138.7, 175.3; IR (neat) 2820, 1600, 1430 cm^{-1} ; MS (FAB $^+$) m/z 326 ($\text{M}^+ + 1$), 234, 91; HRMS calcd for $\text{C}_{21}\text{H}_{28}\text{NO}_2$ ($\text{M}^+ + 1$): 326.2120, found 326.2102.



10-(2-Hydroxyethyl)-1,2,6,7,7a,10,10a,10b-octahydro-5H-azepino[3,2,1-

hi]indol-4-one. Ammonia (10 mL) was condensed into a solution of lactam **9** (250 mg, 0.77 mmol) in THF (3 mL) at -78 °C. Sodium was added, and upon stirring, the reaction mixture turned blue. The reaction was quenched with solid NH₄Cl, and the ammonia was allowed to evaporate. The resulting mixture was diluted with water (5 mL) and then extracted with CH₂Cl₂ (3 x 30mL). The organic extracts were dried (Na₂SO₄), filtered and concentrated under reduced pressure to afford a colorless oil. Flash chromatography (5% MeOH/CH₂Cl₂) afforded the tricyclic alcohol (180 mg, 0.76 mmol, 99% yield) as a white crystalline solid. Mp 158-160 °C. ¹H NMR (400 MHz, CDCl₃) δ 1.41-1.50 (m, 4H), 1.60-1.65 (m, 3H), 1.72-1.74 (m, 1H), 1.83-1.92 (m, 2H), 2.13-2.18 (m, 1H), 2.28 (ddd, *J* = 13.3 Hz, 5.1 Hz, 1.7 Hz, 1H), 2.64 (dt, *J* = 13.4 Hz, 6.7 Hz, 1H), 2.74 (m, 1H), 3.41-3.59 (m, 3H), 3.72-3.79 (m, 2H), 5.57-5.62 (m, 1H), 5.66-5.69 (m, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 23.1, 25.4, 26.8, 33.1, 36.3, 36.5, 37.6, 43.9, 45.9, 60.4, 62.7, 130.7, 130.8, 171.6; IR (neat) 2920, 1600 cm⁻¹; MS (CI) *m/z* 236 (M⁺-1), 111, 69; HRMS calcd for C₁₄H₂₂NO₂ (M⁺-1): 236.1651, found 236.1640. Recrystallization from

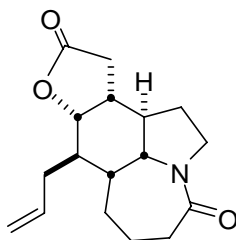
CH₂Cl₂/Hexanes gave white crystals (Mp 158-160 °C) that were subjected to single-crystal x-ray analysis.



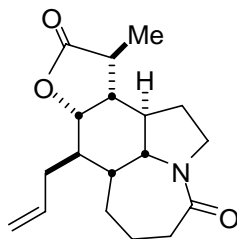
23

Iodolactam 23. Freshly prepared Jones reagent (8.0 N, 2.7 M) was added dropwise to an ice-cooled solution of tricyclic alcohol (100 mg, 0.43 mmol) in acetone (10 mL) until an orange color persisted. The solution was stirred for 3 h at 0 °C and then 30 min at room temperature. After quenching with 2-propanol, the mixture was concentrated to afford a blue-green solid. To this material was added saturated aq NaHCO₃ solution (10 mL), THF (5 mL), Et₂O (5 mL), and solid NaHCO₃ (ca. 0.5 g). The mixture was cooled to 0 °C, and then I₂ (0.32 g, 1.28 mmol) was added. The resulting reaction mixture was stirred for 2 h at 0 °C and then at room temperature for 15 h. After adding saturated aq sodium thiosulfate solution, the solution was extracted with CH₂Cl₂ (3 × 50 mL), dried (Na₂SO₄), filtered and concentrated under reduced pressure to afford a yellow oil. Flash chromatography (5% MeOH/CH₂Cl₂) afforded **23** (128 mg, 0.34 mmol, 80% yield) as a white solid. Mp 187 °C (dec.), ¹H NMR (400 MHz, CDCl₃) δ 1.46 (m, 1H), 1.92 (m, 3H), 1.74-1.78 (m, 1H), 1.90-2.13 (m, 3H), 2.33-2.37 (m, 1H), 2.45 (dd, *J* = 18.0 Hz, 8.6 Hz, 1H), 2.64 (m, 2H), 2.88 (dd, *J* = 18.0 Hz, 10.0 Hz, 1H), 3.45 (m, 1H), 3.69 (dd, *J* = 11.6 Hz, 9.4 Hz, 1H), 3.78 (dd, *J* = 11.8 Hz, 9.0 Hz, 1H), 3.93 (t, *J* = 10.9 Hz, 1H), 4.94 (dd, *J* = 11.2 Hz, 9.5 Hz, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 22.2, 25.0,

27.9, 33.3, 33.6, 33.9, 38.0, 43.2, 44.2, 47.0, 60.6, 83.4, 171.4, 174.6; IR (neat) 1770, 1630 cm^{-1} ; MS (EI) m/z 376 ($\text{M}^+ + 1$), 248; HRMS calcd for $\text{C}_{14}\text{H}_{19}\text{NO}_2$ ($\text{M}^+ + 1$): 376.0410, found 376.0405.

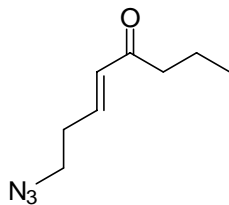


Allylated lactam. To a solution of lactam **23** (86 mg, 0.23 mmol) in degassed benzene (15 mL) was added allyltributyltin (0.14 mL, 0.459 mmol) and AIBN (8.0 mg, 0.0459 mmol). The solution was refluxed for 22 h, cooled to room temperature, and concentrated under reduced pressure to afford a colorless oil. Flash chromatography (5% MeOH/ CH_2Cl_2) afforded the allylated lactam (62 mg, 0.21 mmol, 93% yield) of a colorless oil. ^1H NMR (400 MHz, CDCl_3) δ 1.42-1.80 (m, 6H), 1.90-2.03 (m, 3H), 2.10-2.16 (m, 1H), 2.35-2.50 (m, 4H), 2.60 (m, 1H), 2.87 (dd, $J = 17.9$ Hz, 10.0 Hz, 1H), 3.48 (m, 2H), 3.77 (dd, $J = 12.1$ Hz, 9.1 Hz, 1H), 4.60 (dd, $J = 11.9$ Hz, 9.3 Hz, 1H), 5.84-5.94 (m, 2H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 22.6, 23.6, 28.2, 33.5, 33.6, 34.3, 35.3, 37.7, 42.2, 44.7, 47.1, 60.8, 81.2, 119.1, 134.3, 171.5, 176.3; IR (neat) 1760, 1620 cm^{-1} ; MS (EI) m/z 290 ($\text{M}^+ + 1$), 246, 84; HRMS calcd for $\text{C}_{17}\text{H}_{24}\text{NO}_3$ ($\text{M}^+ + 1$): 290.1756, found 290.1755.

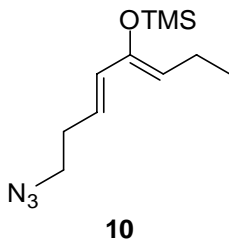


24

Methylated lactam 24. To a cooled solution of allylated lactam (0.16 g, 0.57 mmol) in THF (10 mL) at -78 °C was added lithium bis(trimethylsilyl)amide (1.0 M in hexanes, 1.0 mL, 1.0 mmol) slowly. The solution was stirred at -78 °C for 1 h, and then iodomethane (0.35 mL, 5.67 mmol) was added quickly. The yellow solution was stirred at -78 °C for 40 min longer, and then the reaction was quenched with the addition of 20% aq HCl (ca. 6 mL). After warming to room temperature, the solution was extracted with CH₂Cl₂ (3 × 30mL). The organic extracts were dried (Na₂SO₄), filtered and concentrated under reduced pressure to afford an off-white solid. Flash chromatography (50% EtOAc/50% Hex) afforded **24** (0.13 g , 0.43 mmol, 77% yield) as a white crystalline solid. An analytical sample was furnished by reverse-phase prep HPLC (50% acetonitrile/50% water/0.1% TFA). Mp: 119-121 °C. ¹H NMR (400 MHz, CDCl₃) δ 1.34 (d, *J* = 7.1 Hz, 3H), 1.37-1.53 (m, 2H), 1.60-1.76 (m, 3H), 1.87-2.02 (m, 3H), 2.13-2.21 (m, 2H), 2.34-2.46 (m, 5H), 3.47 (m, 2H), 3.74 (dd, *J* = 12.2 Hz, *J* = 9.1 Hz, 1H), 4.43 (dd, *J* = 12.3 Hz, *J* = 9.4 Hz, 1H), 5.13 (m, 2H), 5.79-5.89 (m, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 15.9, 22.5, 23.4, 28.2, 33.0, 34.1, 35.3, 40.2, 42.0, 44.7, 45.9, 47.5, 61.0, 78.7, 119.2, 134.2, 172.2, 178.9; IR (neat) 1779, 1639 cm⁻¹; MS (FAB⁺) *m/z* 304 (M⁺+1), 154, 136; HRMS calcd for C₁₈H₂₆NO₃ (M⁺+1): 304.1908, found 304.1913.



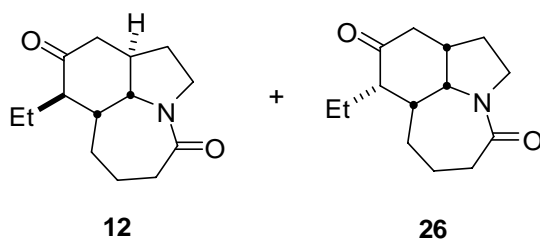
(E)-8-Azido-5-octenoic acid. To a solution of NaH (60%, 1.2 g, 30 mmol) in THF (60 mL) under Ar at $-78\text{ }^{\circ}\text{C}$ was added slowly β -keto phosphonate **25**⁹ (5.90 g, 30 mmol) in THF (10 mL). The resulting solution was allowed to slowly warm up to $-25\text{ }^{\circ}\text{C}$ over 2 h to give a white gel-like suspension. This suspension was cooled in an ice bath followed by dropwise addition of 3-azidopropional^{8b} (3.0 g, 30 mmol) in THF (10 mL). The mixture was stirred in an ice bath for 1 h and then quenched with water (50 mL). The reaction mixture was partitioned between water and Et₂O. The organic layer was dried (Na₂SO₄), filtered, and concentrated to give an oil. Chromatography (2-10% EtOAc/hexane) afforded (*E*)-8-azido-5-octenoic acid (4.6 g, 27.5 mmol, 92% yield) as a colorless oil. $R_f = 0.48$ (15% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃) δ 0.93 (t, $J = 7.8$ Hz, 2H), 1.63 (h, $J = 7.8$ Hz, 2H), 2.50 (m, 4H), 3.43 (t, $J = 7.7$ Hz, 2H), 6.18 (d, $J = 16.8$ Hz, 1H), 6.75 (m, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 13.7, 17.7, 32.0, 42.3, 49.8, 132.5, 141.8, 200.2; IR (neat) 2100, 1697, 1674 cm⁻¹; MS (ES+) m/z 168 ($M^+ + 1$); HRMS calcd for C₈H₁₄N₃O ($M^+ + H$): 168.1137, found 168.1116.



[(3Z,5E)-8-Azidoocta-3,5-dien-4-yloxy]trimethylsilane 10. To a solution of (*E*)-8-azidooct-5-en-4-one (1.67 g, 10.0 mmol) in anhydrous ethyl ether (35 mL) under Ar was added Et₃N (2.1 g, 20 mmol) at 0 °C followed by the addition of TMSOTf (3.3 g, 15 mmol). The resulting mixture was stirred at 0 °C for 30 min. The organic layer was washed with brine, dried (Na₂SO₄), and evaporated under reduced pressure to give the diene **10** (2.27 g, 9.5 mmol, 95% yield) as a light yellow oil. This diene was used directly for the next step. R_f = 0.75 (5:95 EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃) δ 0.23 (s, 9H), 0.99 (t, *J* = 7.0 Hz, 3H), 2.11 (m, 2H), 2.42 (m, 2H), 3.33 (t, *J* = 7.2 Hz, 2H), 4.75 (t, *J* = 7.6 Hz, 1H), 5.68 (m, 1H), 5.98 (d, *J* = 15.3 Hz, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 0.6, 14.1, 19.4, 31.7, 51.0, 117.1, 123.5, 131.7, 147.3; IR (neat) 2100 cm⁻¹; MS (ES+) *m/z* 240 (M⁺+1); HRMS calcd for C₁₁H₂₂N₃OSi (M⁺+H): 240.1532, found 240.1519.

General procedure for the intermolecular Diels–Alder/intramolecular Schmidt reaction involving the diene 10. To a cooled (–78 °C) solution of enone (5.0–10 mmol) in CH₂Cl₂ (20–40 mL) under Ar was added a Lewis acid such as SnCl₄ (1 equiv) followed by the addition of diene **10** (1.8 equiv). The resulting mixture was stirred at –78 °C and allowed to warm to –55 °C over 2 h. After slow addition of another portion of the same Lewis acid (1.5 equiv), the mixture was stirred at room temperature for 12 h and quenched with aqueous NH₄Cl. The mixture was partitioned between water and

CH₂Cl₂. The organic layer was collected, dried (Na₂SO₄), filtered, and concentrated to give an oil. Chromatography (10-50% EtOAc/hexane followed by 1-2% MeOH/ CH₂Cl₂) afforded the reaction products.

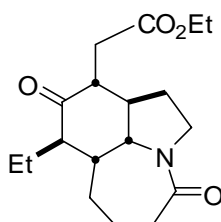


10-Ethyloctahydroazipino[3,2,1-*h,i*]indol-4,9(1*H*, 5*H*)dione 12 and 26. 2-Cyclohexen-1-one (95%, 500 mg, 5.0 mmol) was reacted with **10** using SnCl₄ as Lewis acid to afford 820 mg (70%) of **12** and **26**, an oil, as a ca. 3: 1 (**12** : **26**) mixture of diastereomers based on HPLC/MS analysis. Repeated chromatography afforded pure **12** (600 mg, 2.55 mmol, 52% yield) and **26** (200 mg, 0.85 mmol, 17% yield) as brown syrups. Lactam **12**: R_f = 0.38 (1:10 MeOH/EtOAc); ¹H NMR (400 MHz, CDCl₃) δ 0.92 (t, *J* = 7.8 Hz, 3H), 1.45 (m, 2H), 1.62-2.70 (complex, 13H), 3.40 (dd, *J* = 8.9, 12.2 Hz, 1H), 3.53 (dt, *J* = 6.7, 11.2 Hz, 1H), 3.73 (dd, *J* = 8.9, 12.3 Hz, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 12.9, 21.0, 22.5, 24.9, 28.3, 32.9, 38.4, 38.7, 43.4, 46.9, 53.5, 62.2, 171.0, 212.0. IR (neat) 1707, 1612 cm⁻¹; MS (CI) *m/z* 236 (M⁺+1); HRMS calcd for C₁₄H₂₂NO₂ (M⁺+H): 236.1651, found 236.1634.

Lactam **26**: R_f = 0.32 (1:10 MeOH/EtOAc); ¹H NMR (400 MHz, CDCl₃) δ 0.94 (t, *J* = 7.2 Hz, 3H), 1.03 (m, 1H), 1.26 (septet, *J* = 7.2 Hz, 1H), 1.52-2.54 (complex, 10H), 2.88 (sextet, *J* = 6.1 Hz, 1H), 3.40 (dt, *J* = 6.6, 11.9 Hz, 1H), 3.88 (dd, *J* = 9.6, 12.5 Hz, 1H), 4.40 (dd, *J* = 5.8, 9.3 Hz, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 12.4, 19.3,

21.6, 21.9, 27.3, 33.3, 37.1, 37.7, 39.4, 44.8, 52.1, 59.9, 171.9, 211.2. IR (neat) 1713, 1624 cm^{-1} ; MS (CI) m/z 236 ($\text{M}^+ + 1$); HRMS calcd for $\text{C}_{14}\text{H}_{22}\text{NO}_2$ ($\text{M}^+ + \text{H}$): 236.1651, found 236.1644.

Compound **26** was obtained as the only product in 35% yield (415 mg) when $\text{BF}_3 \cdot \text{OEt}_2$ as Lewis acid; the physical and spectra data were identical as above.

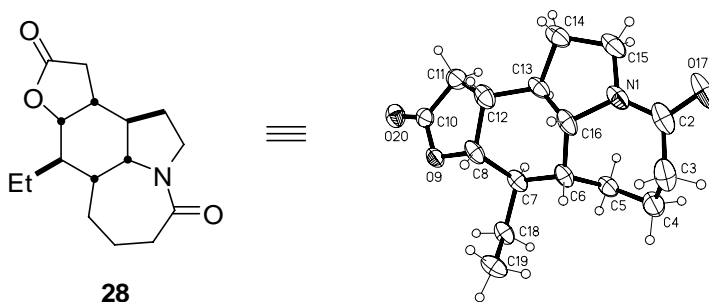


27

Ethyl 10-ethyl-4,9-dioxo-dodecahydroazipino[3,2,1-*h,i*]indol-8-yl)acetate 27.

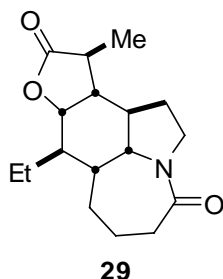
Ketone **12** (500 mg, 2.12 mmol) in THF (10 mL) was added to a solution of LiHMDS (1 M, 3.8 mL, 3.8 mmol) in THF (30 mL) at -78°C under Ar. After stirring at -78°C for 1 h, ethyl 2-bromoacetate (1.78 g, 10.6 mmol) was added followed by HMPA (1.14 g, 6.4 mmol). The resulting mixture was stirred for 3.5 h and then quenched with aq 2 N HCl. After warming up to room temperature, the mixture was partitioned between water and CH_2Cl_2 . The organic layer was collected, dried (Na_2SO_4), filtered, and concentrated to give an oil. Chromatography (10-50% EtOAc/hexane followed by 1-2% MeOH/EtOAc and finally 1-2% MeOH/ CH_2Cl_2) afforded **27** (500 mg, 1.56 mmol, 73% yield) as a yellow syrup. $R_f = 0.42$ (1:10 MeOH/EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 0.92 (t, $J = 7.3$ Hz, 3H), 1.26 (t, $J = 6.8$ Hz, 3H), 1.45 (m, 2H), 1.60-2.09 (complex, 7H), 2.30-2.47 (m, 4H), 2.57 (dd, $J = 5.0, 16.4$ Hz, 1H), 2.57 (dd, $J = 5.0, 16.4$ Hz, 1H), 2.73 (dd, $J = 5.0, 16.4$ Hz, 1H), 3.51 (m, 2H), 3.71 (dd, $J = 9.2, 12.8$ Hz, 1H), 4.13 (q, $J = 6.8$ Hz, 2H);

^{13}C NMR (100.6 MHz, CDCl_3) δ 12.4, 14.2, 21.8, 22.4, 25.2, 26.8, 32.9, 34.7, 38.0, 42.6, 46.6, 49.2, 53.2, 60.9, 62.1, 171.0, 171.3, 213.0. IR (neat) 1732, 1713, 1632 cm^{-1} ; MS (ES+) m/z 322 (M^++1); HRMS calcd for $\text{C}_{18}\text{H}_{28}\text{NO}_4$ (M^++H): 322.2018, found 322.2015.



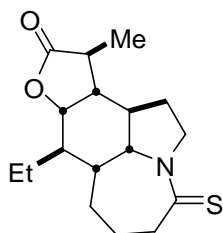
(\pm)-**13-Desmethyl-5-oxostenine 28**. To a solution of ketoester **27** (400 mg, 1.25 mmol) in MeOH (30 mL) at 0°C was added NaBH_4 (118 mg, 3.8 mmol). The resulting mixture was stirred under for 1.5 h and then quenched with aq 2 N HCl. After concentration to ca 20 mL, the mixture was partitioned between water and CH_2Cl_2 . The organic layer was collected, dried (Na_2SO_4), filtered, and concentrated to give a light yellow syrup. Chromatography (50% EtOAc/hexane followed by 2-5% MeOH/EtOAc and then 2-5% MeOH/ CH_2Cl_2) afforded **28** (220 mg, 0.79 mmol, 64% yield) as off-white crystalline solid. Mp $143\text{-}144^\circ\text{C}$; $R_f = 0.28$ (1:10 MeOH/EtOAc); ^1H NMR (400 MHz, CDCl_3) δ 0.96 (t, $J = 7.4$ Hz, 3H), 1.37-1.75 (complex, 7H), 1.80-2.10 (complex, 4H), 2.26-2.59 (m, 4H), 2.78 (dd, $J = 10.0, 17.8$ Hz, 1H), 3.43 (m, 1H), 3.70 (dd, $J = 9.0, 12.2$ Hz, 1H), 4.56 (dd, $J = 9.3, 11.7$ Hz, 1H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 10.2, 22.5, 23.0, 23.5, 27.8, 33.0, 33.1, 35.5, 37.4, 42.5, 44.2, 46.7, 60.6, 81.6, 171.1, 176.0. IR (neat) 1772, 1628 cm^{-1} ; MS (ES+) m/z 278 (M^++1); HRMS calcd for $\text{C}_{16}\text{H}_{24}\text{NO}_3$ (M^++H):

278.1756, found 278.1734. Crystals for X-ray crystallographic analysis were obtained by recrystallization from EtOAc/hexane.

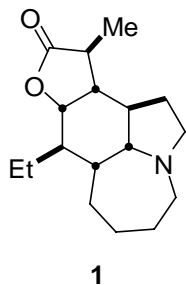


(±)-**5-Oxostenine 29**. To a solution of 56 mg (0.2 mmol) of lactone **28** in 10 mL of THF at $-78\text{ }^{\circ}\text{C}$ under Ar was added LiHMDS (1 M, 0.3 mL, 0.3 mmol). After stirring at $-78\text{ }^{\circ}\text{C}$ for 1 h, MeI (142 mg, 1 mmol) was added. The resulting mixture was stirred for 2 h and then quenched with aq 2 N HCl. After warming up to room temperature, the mixture was partitioned between water and CH_2Cl_2 . The organic layer was collected, dried (Na_2SO_4), filtered, and concentrated to give a yellow syrup. Chromatography (10-50% EtOAc/hexane followed by, 1-2% MeOH/EtOAc) afforded **29** (46 mg, 0.16 mmol, 79% yield) as a light yellow crystalline solid. An analytical sample was furnished by reverse-phase preparative HPLC (50% acetonitrile/50%water/0.1% TFA). Mp 164-165 $^{\circ}\text{C}$; $R_f = 0.34$ (1:10 MeOH/EtOAc); ^1H NMR (400 MHz, CDCl_3) δ 0.98 (t, $J = 7.5$ Hz, 3H), 1.35 (d, $J = 7.1$ Hz, 3H), 1.40-1.75 (complex, 7H), 1.84-2.04 (complex, 4H), 2.14-2.21 (m, 2H), 2.31 (dd, $J = 4.6, 12.8$ Hz, 1H), 2.43 (m, 2H), 3.48 (m, 1H), 3.73 (dd, $J = 9.2, 12.0$ Hz, 1H), 4.46 (dd, $J = 9.3, 12.2$ Hz, 1H); ^{13}C NMR (100.6 MHz, CDCl_3) δ 10.2, 15.4, 22.5, 22.8, 23.4, 27.8, 33.0, 35.8, 39.8, 42.3, 44.3, 45.7, 46.9, 60.5, 79.2, 171.1, 178.7. IR (neat) 1763, 1634 cm^{-1} ; MS (ES+) m/z 292 (M^++1); HRMS calcd for

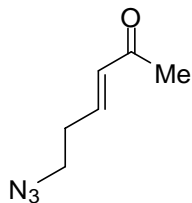
C₁₇H₂₆NO₃ (M⁺+H): 292.1913, found 292.1906. ¹H and ¹³C NMR spectra matched data reported by Wipf et al for (-)-**29**.¹⁰



(±)-**5-Thiostenine**. To a solution of **29** (24 mg, 0.082 mmol) in 5 mL of CH₂Cl₂ was added at room temperature 49 mg (0.123 mmol) of Lawesson's reagent. After 3 h, the reaction mixture was concentrated under reduced pressure and chromatographed on SiO₂ (EtOAc/hexane, 1: 3 to 1: 1) to give (±)-4-thiostenine (24 mg, 0.078 mmol, 93% yield) as a colorless solid. Mp 180-181 °C; R_f = 0.49 (1:1 EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃) δ 0.97 (t, *J* = 7.4 Hz, 3H), 1.37 (d, *J* = 7.1 Hz, 3H), 1.49-1.75 (complex, 7H), 2.00 (m, 2H), 2.20 (m, 3H), 2.46 (m, 2H), 2.87 (dt, *J* = 5.6, 13.0 Hz, 1H), 3.02 (dd, *J* = 3.9, 12.8 Hz, 1H), 3.73 (m, 2H), 4.14 (dd, *J* = 8.7, 13.9 Hz, 1H), 4.44 (dd, *J* = 8.9, 12.1 Hz, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 10.1, 15.6, 22.6, 23.2, 24.7, 27.8, 35.6, 39.7, 42.2, 42.7, 44.6, 45.5, 55.2, 65.8, 78.8, 178.4, 199.3. IR (neat) 1768 cm⁻¹; MS (ES+) *m/z* 308 (M⁺+1); HRMS calcd for C₁₇H₂₆NSO₂ (M⁺+H): 308.1684, found 308.1667.

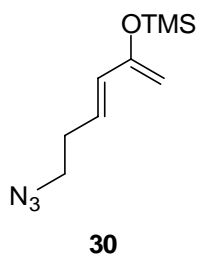


(±)-Stenine 1. A solution of 5-thioesteine (20 mg, 0.065 mmol) in 95% EtOH (10 mL) was treated at room temperature with Raney Ni (350 mg). The reaction mixture was shaken for 30 min and filtered through a cotton filter. The solvent was removed under reduced pressure, and the solid residue was chromatographed on SiO₂ (EtOAc) to give stenine **1** (16 mg, 0.058 mmol, 89% yield) as a light yellow oil. $R_f = 0.1$ (1:10 MeOH/EtOAc); ¹H NMR (400 MHz, CDCl₃) δ 0.92 (t, $J = 7.4$ Hz, 3H), 1.28 (d, $J = 7.1$ Hz, 3H), 1.30-1.75 (complex, 10H), 1.90 (m, 2H), 2.05-2.50 (m, 2H), 2.86 (dt, $J = 4.2, 13.0$ Hz, 1H), 3.17 (dt, $J = 3.5, 9.0$ Hz, 1H), 4.46 (dd, $J = 9.2, 12.0$ Hz, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 9.8, 15.0, 22.3, 26.2, 27.5, 29.5, 29.8, 39.8, 40.4, 42.5, 43.2, 47.4, 53.1, 55.0, 68.0, 80.5, 179.6. IR (neat) 1770 cm⁻¹; MS (ES⁺) m/z 278 (M⁺+1); HRMS calcd for C₁₇H₂₈NO₂ (M⁺+H): 278.2120, found 278.2106.



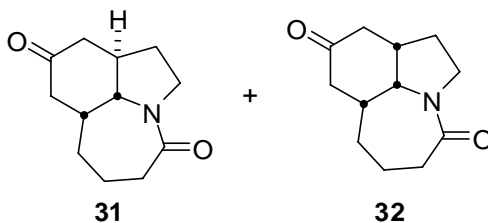
(E)-6-Azido-3-hexen-2-one. To a solution of commercially-available dimethyl 2-oxopropylphosphonate (1.60 g, 10 mmol) in THF/H₂O (4:1, 40 mL) was added K₂CO₃ (2.10 g, 15 mmol). The resulting solution was cooled in an ice bath followed by dropwise addition of the 3-azidopropanal⁸ (1.0 g, 10 mmol) in THF (4 mL). The mixture was

stirred in an ice bath for 1 h and then quenched with saturated aqueous NaHCO₃. The reaction mixture was partitioned between water and EtOAc. The organic layer was dried (Na₂SO₄), filtered, and concentrated to give an oil. Chromatography (10% EtOAc/hexane) afforded (*E*)-6-azido-3-hexen-2-one (940 mg, 6.76 mmol, 68% yield) as a colorless oil. *R*_f = 0.46 (25% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃) δ 2.25 (s, 3H), 2.51 (m, 2H), 3.44 (t, *J* = 6.7 Hz, 2H), 6.15 (d, *J* = 16.0 Hz, 1H), 6.74 (m, 1H); ¹³C NMR (100.6 MHz, CDCl₃) δ 27.5, 32.2, 50.0, 133.6, 143.4, 198.5; IR (neat) 1709, 1644 cm⁻¹; MS (CI) *m/z* 140 (M⁺+1); HRMS calcd for C₆H₁₀N₃O (M⁺+H): 140.0824, found 140.0853.



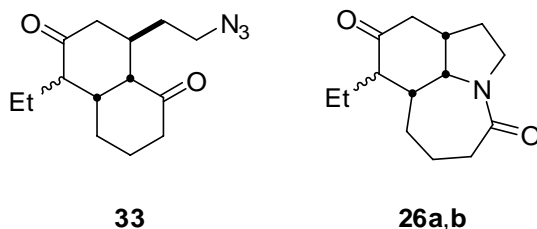
(*E*)-6-Azido-2-trimethylsilyloxy-1,3-hexadiene 30. To a solution of (*E*)-6-azido-3-hexen-2-one (1.4 g, 10 mmol) in anhydrous ethyl ether (35 mL) under argon was added Et₃N (2.1 g, 20 mmol) at 0 °C followed by the addition of TMSOTf (3.3 g, 15 mmol). The resulting mixture was stirred at 0 °C for 30 min and then quenched with saturated aqueous NaHCO₃. The reaction mixture was partitioned between water and EtOAc. The organic layer was washed with brine, dried (Na₂SO₄), and evaporated under reduced pressure to give **30** (2.18 g, 10.3 mmol, 98% yield) as a very light yellow oil. This diene was used directly for the next step without further purification. *R*_f = 0.52 (2% EtOAc/hexane); ¹H NMR (400 MHz, CDCl₃) δ 0.26 (s, 9H), 2.42 (m, 2H), 3.36 (t, *J* =

7.0 Hz, 2H), 4.31 (d, $J = 3.5$ Hz, 2H), 5.92 (m, 1H), 6.01 (d, $J = 15.2$ Hz, 1H); ^{13}C NMR (100.6 MHz, CDCl_3) δ -0.1, 31.5, 50.7, 95.3, 126.4, 130.5, 154.3; IR (neat) 2098 cm^{-1} ; MS (CI) m/z 212 ($\text{M}^+ + 1$); HRMS calcd for $\text{C}_9\text{H}_{18}\text{N}_3\text{OSi}$ ($\text{M}^+ + \text{H}$): 212.1219, found 212.1196.



(±)-5-Octahydro-azipino[3,2,1-*h,i*]indol-4,9-dione **31** and **32**. To a cooled (-78 °C) solution of 2-cyclohexenone (100 mg, 1.0 mmol) in CH_2Cl_2 (20 mL) under argon was added SnCl_4 (261 mg, 1.0 mmol) followed by the addition of diene **30** (380 mg, 1.8 mmol). The resulting mixture was stirred at -78 °C for 2 h. After slow addition of another portion of SnCl_4 (391 mg, 1.5 mmol), the mixture was stirred at for 12 h, and then quenched with aqueous NaHCO_3 . The mixture was partitioned between water and CHCl_3 . The organic layer was collected, dried (Na_2SO_4), filtered, and concentrated to give an oil. Chromatography (10-50% EtOAc/hexane followed by 1-2% MeOH/ CHCl_3) afforded a viscous oil of the desired lactams **31** and **32** (170 mg, 0.82 mmol, 82% yield) as a ca. 1:3.4 (**31**:**32**) mixture of diastereomers. The mixture solidified upon standing. $R_f = 0.32$ (1:10 MeOH/EtOAc); IR (neat) $1709, 1624\text{ cm}^{-1}$; MS (CI) m/z 208 ($\text{M}^+ + 1$); HRMS calcd for $\text{C}_{12}\text{H}_{18}\text{NO}_2$ ($\text{M}^+ + \text{H}$): 208.1338, found 208.1357. A pure sample of the major diastereomer was obtained after recrystallization from hexanes/EtOAc. Mp 131-133 °C; ^1H NMR (500 MHz, CDCl_3) δ 1.54-1.84 (complex, 6H), 2.00 (m, 1H), 2.11 (d, $J = 15.7$ Hz, 1H), 2.24 (dd, $J = 4.0, 15.9$ Hz, 1H), 2.46 (m, 3H), 2.65 (m, 2H), 2.99 (m, 1H), 3.24

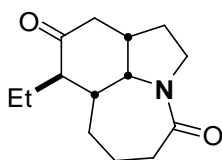
(two sets of t, $J = 6.56, 11.9$ Hz, 1H), 3.91 (m, 1H), 4.13 (d, 1H, $J = 6.9$ Hz); ^{13}C NMR (125.8 MHz, CDCl_3) δ 18.5, 31.2, 33.6, 35.7, 38.0, 39.3, 39.7, 43.3, 45.8, 59.7, 174.3, 211.2. Minor diastereomer (diagnostic peaks only, from the mixture): ^1H NMR (400 MHz, CDCl_3) δ 3.51 (m), 3.75 (m); ^{13}C NMR (100.6 MHz, CDCl_3) δ 22.1, 26.6, 28.4, 31.9, 39.7, 43.2, 45.6, 47.3, 62.6, 171.6, 211.5.



10-Ethyltricycloazepino[3,2,1]indole-4,9(1*H*,5*H*)-dione 26a,b. To a solution of 2-cyclohexen-1-one (562 mg, 5.85 mmol) and 4 Å molecular sieve pellets (7.00 g) in CH_2Cl_2 (80 mL) $\text{BF}_3 \cdot \text{OEt}_2$ (0.89 mL, 7.02 mmol, 1.20 equiv) was added at -78 °C. After stirring for 5 min, enol ether **11** (2.14 g, 8.77 mmol, 1.5 equiv) in CH_2Cl_2 (10 mL) was added portionwise over 15 min. The reaction was allowed to warm to room temperature and stirred overnight. Saturated aqueous NH_4Cl was added and the reaction extracted with CH_2Cl_2 (3×50 mL). The combined organic layers were dried over anhydrous Na_2SO_4 , and concentrated to yield a dark brown oil. Silica chromatography afforded a diastereomeric mixture of the ethyl tricyclic ketoamides **26a,b** (587 mg, 2.50 mmol, 43% yield) as a light orange oil and ca. 0.32 g of a mixture of cyclohexenone and the azide-containing Diels-Alder adduct **33**. The diketoazide **33** mixture was dissolved in CH_2Cl_2 (50 mL), TiCl_4 (0.18 mL, 1.60 mmol, 1.30 equiv) was added, and the reaction stirred at room temperature overnight. The reaction was worked up and purified as above to yield an additional portion of mixed diastereomeric lactams **26a,b** as a light orange oil (166

mg, 0.71 mmol, 12% additional yield). The characterization of the major diastereomer of **26a** was in agreement with those previously reported.¹¹ $R_f = 0.32$ (1:10 MeOH/EtOAc); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 0.94 (t, $J = 7.2$ Hz, 3H), 1.03 (m, 1H), 1.26 (septet, $J = 7.2$ Hz, 1H), 1.52-2.54 (complex, 10H), 2.88 (sextet, $J = 6.1$ Hz, 1H), 3.40 (dt, $J = 6.6, 11.9$ Hz, 1H), 3.88 (dd, $J = 9.6, 12.5$ Hz, 1H), 4.40 (dd, $J = 5.8, 9.3$ Hz, 1H); $^{13}\text{C NMR}$ (100.6 MHz, CDCl_3) δ 12.4, 19.3, 21.6, 21.9, 27.3, 33.3, 37.1, 37.7, 39.4, 44.8, 52.1, 59.9, 171.9, 211.2. IR (neat) 1713, 1624 cm^{-1} ; MS (CI) m/z 236 ($\text{M}^+ + 1$); HRMS calcd for $\text{C}_{14}\text{H}_{22}\text{NO}_2$ ($\text{M}^+ + \text{H}$): 236.1651, found 236.1644.

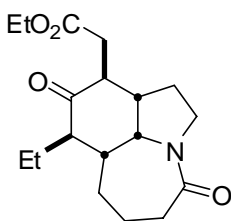
For the diketozide **33**: $R_f = 0.43$ (25% EtOAc in hexanes); $^1\text{H NMR}$ (major isomer, 400 MHz) δ 0.83 (t, $J = 7.3$ Hz, 3 H), 1.40-1.52 (m, 2 H), 1.64 (m, 1 H), 1.85 (m, 2 H), 1.92-2.08 (complex, 3 H), 2.19-2.34 (complex, 3 H), 2.49 (m, 2 H), 2.68 (dd, $J = 11.1, 14.4$ Hz, 1 H), 2.78 (m, 1 H), 2.92 (m, 1 H), 3.27 (m, 1 H), 3.36 (m, 1 H); $^{13}\text{C NMR}$ (major isomer, 100 MHz) δ d 11.9, 35.8, 42.3, 49.7, 55.1; u 19.7, 23.7, 25.3, 31.2, 41.3, 41.6, 49.2, 212.1, 213.0.



26b

10-Ethyl-10,10-dihydroazepino[3,2,1]indole-4,9(1H,5H)-dione 26b. The collected diastereomeric ketolactams **26a,b** (3.80 g, 16.15 mmol) were epimerized by dissolution in MeOH (75 mL) followed by the addition of sodium methoxide (1.74 g, 32.3 mmol, 2.0 equiv) After stirring at room temperature for approximately 14 h, the solvent was removed in vacuo. The residue was dissolved in CH_2Cl_2 and filtered through Celite.

Silica gel chromatography afforded diastereomerically pure ketoamide **26b** (3.17 g, 13.5 mmol, 83% yield) as a light orange solid. For **26b** $R_f = 0.58$ (1:1 acetone:CH₂Cl₂) mp 92-95 °C; ¹H NMR (400 MHz) δ 0.85 (t, $J = 7.5$ Hz, 3 H), 1.45 (m, 1 H), 1.65-1.74 (complex, 4 H), 1.84 (m, 1 H), 1.92-1.99 (m, 1 H), 2.00-2.07 (m, 1 H), 2.17-2.18 (m, 1 H), 2.28-2.32 (m, 2 H), 2.43 (m, 1 H), 2.60-2.71 (m, 2 H), 2.97-3.03 (m, 1 H), 3.10 (dt, $J = 6.3, 11.9$ Hz, 1 H), 3.91 (dd, $J = 8.4, 12.2$ Hz, 1 H), 4.12 (d, $J = 7.1$ Hz, 1 H); ¹³C NMR (100 MHz) δ d 10.2, 38.0, 39.1, 46.0, 60.2; u 17.8, 21.2, 30.4, 31.4, 38.1, 43.7, 46.0, 173.8, 213.9; IR 3554, 1707, 1618 cm⁻¹; HRMS calcd for C₁₄H₂₂NO₂ 236.1651, found 236.1639.

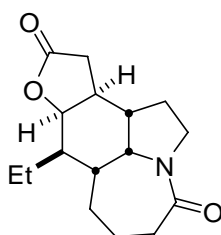


34

Ethyl 2-(10'-ethyl-4',9'-dioxododecahydroazepino[3,2,1-hi]indol-8'-yl)acetate

34. To a solution of ketone **26b** (229 mg, 0.97 mmol) in THF (10.0 mL) and HMPA (0.52 mL, 3.00 mmol) at -78 °C was added LHMDS (2.5 mL, 1.0 M in THF, 2.50 mmol). The reaction was stirred for 1.75 h at -78 °C, then ethyl bromoacetate (0.54 mL, 4.86 mmol) was added neat. The cooling bath was removed and the reaction stirred for 3 h, then quenched with saturated NH₄Cl and extracted with EtOAc (1 × 20 mL) and CH₂Cl₂ (2 × 15 mL). The combined organic layers were dried (Na₂SO₄) and concentrated. The reaction residue was chromatographed on silica gel to yield the ketoester **34** as a light yellow oil (296 mg, 0.92 mmol, 95% yield). $R_f = 0.49$ (1:1 CH₂Cl₂:acetone); ¹H NMR

(400 MHz) δ 0.84 (t, $J = 7.32$ Hz, 3 H), 1.21 (t, $J = 7.32$ Hz, 3 H), 1.43-1.49 (m, 2 H), 1.51-1.57 (m, 2 H), 1.63-1.70 (m, 2 H), 1.80-1.90 (m, 1 H), 2.04-2.15 (m, 2 H), 2.34 (m, 1 H), 2.41 (dd, $J = 3.2, 16.1$ Hz, 1 H), 2.45-2.75 (complex, 5 H), 3.58 (m, 1 H), 3.93 (dd, $J = 2.3, 5.2$ Hz, 1 H), 4.00-4.12 (m, 3 H); ^{13}C NMR (100 MHz) δ d 11.5, 14.2, 41.3, 46.1, 46.8, 48.0, 62.1; u 17.8 (2 C), 28.3, 31.7, 33.5, 39.3, 47.3, 60.7, 172.0, 175.1, 210.0; IR 3450, 2935, 2878, 1719, 1643 (s) cm^{-1} ; HRMS calcd for $\text{C}_{18}\text{H}_{28}\text{NO}_4$ 322.2018, found 322.2029.



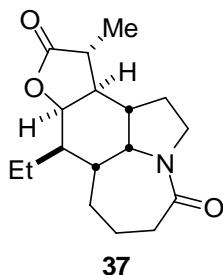
35a

8-Ethyldecahydroazepino[3,2,1-hi]furo[3,2-e]indole-4,10(31H,11bH)-dione

35a. A solution of the keto ester **34** (334 mg, 1.04 mmol) in CH_2Cl_2 (10 mL) was added to a slurry of anhydrous cerium trichloride (388 mg, 1.56 mmol) in CH_2Cl_2 (20 mL) at -78 °C. The mixture was stirred at -78 °C for 1 h and L-Selectride™ solution (2.1 mL, 1.0 M in THF, 2.10 mmol) was then added. The reaction was stirred for 15 h, slowly warming to rt. The reaction was filtered through a short plug of celite and the solvent removed in vacuo. The residue was purified by silica chromatography to give a mixture of two diastereomeric lactols **36** and a small amount of lactone **35a** (233 mg, 0.80 mmol, based on the lactol MW), which was used without further purification. For the lactol mixture **36**: ^{13}C NMR (100 MHz) δ d 11.0, 33.2, 33.5, 36.9, 43.3, 61.7, 80.0, 97.6, 98.2; u

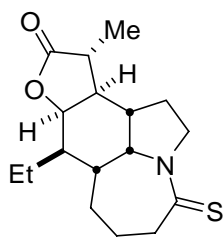
17.6, 20.8, 28.3, 30.8, 38.5, 39.5, 47.4, 175.2, 176.4; HRMS calcd for C₁₆H₂₆NO₃ 280.1913, found 280.1896.

The mixture of the lactone **35a** and lactols **36** (233 mg, 0.80 mmol based on the lactol MW) were dissolved in CH₂Cl₂ (15 mL) and added to 4 Å molecular sieve pellets (1.50 g). To this mixture was successively added N-methylmorpholine N-oxide (187 mg, 1.60 mmol) then tetrapropylammonium perruthenate (28 mg, 0.08 mmol) as solids. The reaction was stirred for 1.5 h at room temperature, filtered through a plug of Celite and the solvent removed in vacuo. The residue was purified by silica chromatography to give lactone **35a** as a white solid (182 mg, 0.66 mmol, 63% yield over two steps). For **35a** R_f = 0.33 (1:1 acetone: CH₂Cl₂); mp 143-145 °C; ¹H NMR (400 MHz) δ 0.99 (t, *J* = 7.3 Hz, 3 H), 1.39 (m, 1 H), 1.48-1.71 (complex, 6 H), 1.85 (m, 1 H), 2.10 (m, 2 H), 2.14 (m, 2 H), 2.35 (d, *J* = 16.9 Hz, 2 H), 2.72 (dd, *J* = 7.0, 15.1 Hz, 1 H), 2.82 (dd, *J* = 7.1, 16.9 Hz, 1 H), 3.42 (dt, *J* = 6.0, 11.9 Hz, 1 H), 3.79 (dd, *J* = 2.0, 4.8 Hz, 1 H), 3.92 (dd, *J* = 8.3, 12.4 Hz, 1 H), 4.67 (dd, *J* = 2.5, 4.6 Hz, 1 H); ¹³C NMR (100 MHz) δ d 11.0, 33.3, 33.5, 37.0, 43.3, 61.6, 80.0; u 17.6, 20.8, 28.3, 30.9, 38.5, 39.6, 47.3, 175.1, 176.3; IR 2935, 2253, 1772, 1615 cm⁻¹; HRMS calcd for C₁₆H₂₄NO₃ 278.1756, found 278.1756.



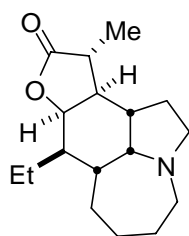
13-Epi-5-oxoneostenine 37. A mixture of lactone **35a** (40 mg, 0.14 mmol) and HMPA (0.10 mL, 0.56 mmol) in THF (2 mL) was cooled to -78 °C, a solution of

LHMDS (0.36 mL, 1.0 M in THF, 0.36 mmol, 2.5 equiv) was added, and the reaction stirred at $-78\text{ }^{\circ}\text{C}$ for 2 h. A solution of methyl iodide (102 mg, 0.72 mmol, 5 equiv) in THF (1 mL) was added and the reaction warmed to room temperature. The reaction mixture was quenched with aqueous saturated NH_4Cl and extracted with EtOAc (3×15 mL). The organics were dried with Na_2SO_4 and concentrated to a viscous oil. Silica gel chromatography gave the methylated lactone **37** (37 mg, 0.13 mmol, 92% yield) as a white solid. $R_f = 0.49$ (1:1 acetone: CH_2Cl_2); mp $154\text{-}156\text{ }^{\circ}\text{C}$; ^1H NMR (400 MHz) δ 1.03 (t, $J = 7.3$ Hz, 3 H), 1.37 (d, $J = 7.6$ Hz, 3 H), 1.52-1.67 (complex, 6 H), 1.84-1.93 (m, 2 H), 2.17 (m, 1 H), 2.38 (m, 1 H), 2.49 (dd, $J = 1.8, 7.6$ Hz, 1 H), 2.65 (d, $J = 9.4$ Hz, 1 H), 2.69 (m, 1 H), 3.43 (dt, $J = 6.7, 10.5$ Hz, 1 H), 3.82 (dd, $J = 2.3, 5.6$ Hz, 1 H), 3.87 (ddd, $J = 2.0, 8.8, 12.3$ Hz, 1 H), 4.81 (dd, $J = 2.6, 5.6$ Hz, 1 H); ^{13}C NMR (100 MHz) δ 11.1, 15.4, 33.5, 34.0, 43.0, 43.7, 44.7, 60.9, 77.4; u 17.7, 20.7, 28.5, 30.2, 38.9, 46.9, 174.6, 179.2; IR 1769, 1630 cm^{-1} ; HRMS calcd for $\text{C}_{17}\text{H}_{26}\text{NO}_3$ 292.1913, found 292.1932.



13-Epi-5-thioneostenine. Lactam **37** (37 mg, 0.12 mmol) and phosphorous pentasulfide (18 mg, 0.04 mmol) were combined in CH_2Cl_2 (1 mL). A solution of hexamethyldisiloxane (19 mg, 0.37 mmol) in CH_2Cl_2 (1 mL) was added and the reaction monitored by TLC. After 2 h, the reaction was filtered through a plug of silica gel and concentrated to a viscous oil. The crude mixture was purified by silica chromatography to

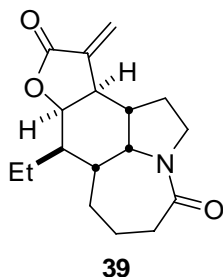
give the pure thioamide as an off-white solid (31 mg, 0.10 mmol, 84% yield). $R_f = 0.84$ (1:1 acetone:CH₂Cl₂); mp 192-196 °C; ¹H NMR (400 MHz) δ 1.00 (t, $J = 7.3$ Hz, 3 H), 1.37 (d, $J = 7.3$ Hz, 3 H), 1.50 (dt, $J = 3.8, 15.5$ Hz, 1 H), 1.58-1.80 (complex, 6 H), 1.88 (m, 1 H), 2.11 (dd, $J = 7.3, 12.8$ Hz, 1 H), 2.20 (m, 2 H), 2.52 (m, 2 H), 2.82 (m, 1 H), 3.42 (m, 1 H), 3.86 (m, 1 H), 4.13 (m, 1 H), 4.20 (t, $J = 4.7$ Hz, 1 H), 4.86 (dd, $J = 2.9, 7.0$ Hz, 1 H); ¹³C NMR (100 MHz) δ d 11.1, 15.7, 33.8, 36.4, 41.8, 42.9, 43.0, 64.0, 76.5; u 19.8, 20.8, 28.3, 29.2, 46.9, 54.6, 178.8, 202.4; IR 2975, 2932, 2876, 1761, 1753 cm⁻¹; HRMS calcd for C₁₇H₂₆NO₂S 308.1684, found 308.1700.



13-epineostenine **38**

13-Epineostenine 38. To a solution of 13-epi-4-thioneostenine (31 mg, 0.10 mmol) in EtOH (2 mL) was added Raney nickel 2800 slurry in water (600 mg). The mixture was stirred vigorously for 2 h, filtered through a Celite plug and the solvents removed in vacuo. The residue was purified by chromatography on basic alumina to give 13-epineostenine **38** as a white solid (22 mg, 0.079 mmol, 93% yield). $R_f = 0.20$ (acetone); mp 106-108 °C; ¹H NMR (400 MHz) δ 0.98 (t, $J = 7.3$ Hz, 3 H), 1.29 (d, $J = 7.0$ Hz, 3 H), 1.33-1.43 (m, 2 H), 1.53-1.64 (complex, 4 H), 1.66-1.74 (m, 3 H), 1.83 (m, 2 H), 2.26-2.46 (complex, 5 H), 2.80 (dd, $J = 4.6, 9.0$ Hz, 1 H), 3.02 (dd, $J = 4.1, 12.0$ Hz, 1 H), 3.10 (t, $J = 7.0$ Hz, 1 H), 4.85 (dd, $J = 4.1, 8.5$ Hz, 1 H); ¹³C NMR (100 MHz) δ d 11.4, 15.4, 35.1, 38.9, 39.4, 41.5, 42.8, 65.6, 77.4; u 19.8, 22.9, 31.2, 31.3, 56.0, 57.6,

179.8; IR 2930, 1772, 1634 cm^{-1} ; HRMS calcd for $\text{C}_{17}\text{H}_{28}\text{NO}_2$ 278.2120, found 278.2124.

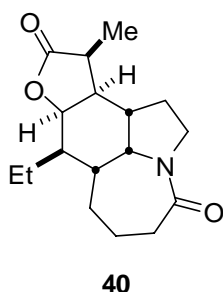


8-Ethyl-11-methylenedecahydroazepino[3,2,1-hi]furo[3,2-e]indole-

4,10(31H,11bH)-dione 39. A mixture of lactone **35a** (52 mg, 0.19 mmol) and HMPA (0.10 mL, 0.56 mmol) in THF (2 mL) was cooled to $-78\text{ }^{\circ}\text{C}$, a solution of LHMDS (0.47 mL, 1.0 M in THF, 0.47 mmol) was added, and the reaction stirred at $-78\text{ }^{\circ}\text{C}$ for 1 h. Carbon dioxide was then bubbled through the reaction for 15 min and the reaction warmed to room temperature. The reaction mixture was quenched with aqueous saturated NH_4Cl and extracted with EtOAc ($3 \times 15\text{ mL}$). The organics were dried with Na_2SO_4 and the solvent removed in vacuo to give the β -carboxylic acid lactone as a yellow oil, which was used without further purification or characterization.

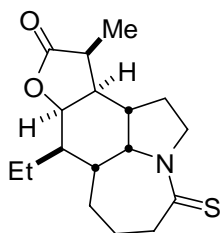
A methylenation reagent stock solution was prepared as described by Greene and coworkers¹² by combining N-methylaniline (5.2 mL), sodium acetate (600 mg), and aqueous formaldehyde solution (15 mL, 37% solution) in acetic acid (20 mL). The methylenation reagent stock solution (0.5 mL) was added to the crude β -carboxylic acid lactone. The reaction mixture stirred at rt for 2.5 h, diluted with water (20 mL) and extracted with CH_2Cl_2 ($3 \times 10\text{ mL}$). The organic layers were combined, dried with Na_2SO_4 and the solvent removed in vacuo. The crude product mixture was purified on

silica chromatography to give the methylene lactone as a white solid (27 mg, 0.09 mmol, 49% yield from **35a**). $R_f = 0.46$ (1:1 acetone: CH_2Cl_2); mp 149-151 °C; ^1H NMR (400 MHz) δ 1.00 (t, $J = 7.3$ Hz, 3 H), 1.23 (m, 1 H), 1.42 (m, 1 H), 1.57-1.83 (complex, 5 H), 1.99 (m, 1 H), 2.45 (m, 2 H), 2.65 (m, 1 H), 2.91 (t, $J = 6.1$, 1 H), 3.52 (m, 1 H), 3.82 (m, 1 H), 3.88 (dd, $J = 2.5, 6.6$ Hz, 1 H), 4.77 (dd, $J = 3.0, 6.8$ Hz, 1 H), 5.67 (d, $J = 2.0$ Hz, 1 H), 6.23 (d, $J = 2.0$ Hz, 1 H); ^{13}C NMR (100 MHz) δ d 11.2, 33.7, 35.0, 40.2, 42.8, 59.5, 76.9; u 18.0, 20.4, 28.7, 29.2, 37.8, 46.3, 121.0, 140.7, 171.0, 174.1; IR 1753, 1625 cm^{-1} ; HRMS calcd for $\text{C}_{17}\text{H}_{24}\text{NO}_3$ 290.1756, found 290.1779.

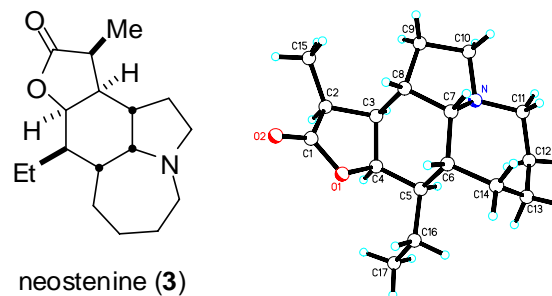


5-Oxoneostenine 40. Methylene lactone **39** (67 mg, 0.23 mmol) and platinum (IV) oxide (20 mg, 0.09 mmol) were suspended in methanol (1 mL) and acetic acid (1 mL). The mixture was stirred under a hydrogen atmosphere at 1 atm for 16 h at room temperature, filtered through Celite and concentrated. The residue was purified by silica chromatography to give 5-oxoneostenine **40** (60 mg, 0.21 mmol, 91% yield) as a white solid. $R_f = 0.56$ (1:1 CH_2Cl_2 :acetone); mp 203-205 °C; ^1H NMR (400 MHz) δ 0.99 (t, $J = 7.3$ Hz, 3 H), 1.28 (d, $J = 7.3$ Hz, 3 H), 1.37-1.69 (complex, 7 H), 1.87 (m, 1 H), 2.08 (m, 1 H), 2.17 (m, 3 H), 2.34 (m, 1 H), 2.71 (m, 1 H), 2.95 (m, 1 H), 3.40 (m, 1 H), 3.80 (m, 1 H), 3.93 (m, 1 H), 4.57 (m, 1 H); ^{13}C NMR (100 MHz) δ d 9.4, 11.0, 32.2, 33.3, 38.2,

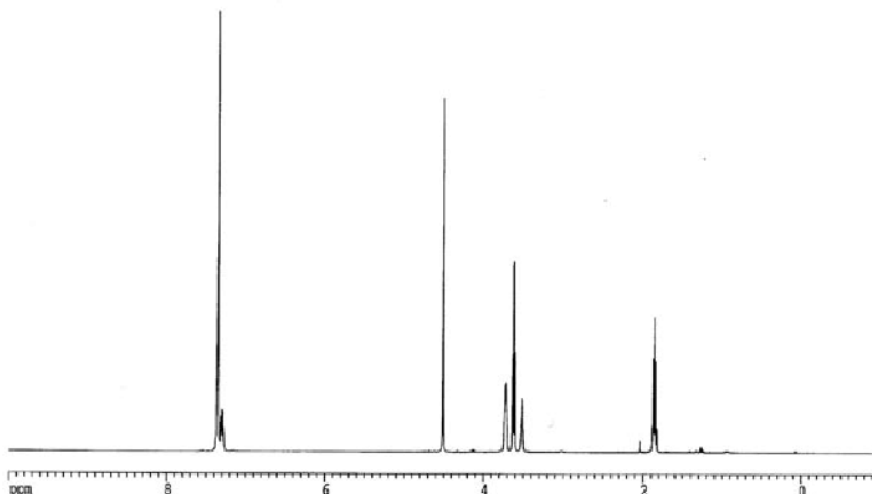
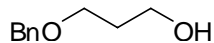
40.5, 42.0, 61.9, 78.2; u 17.6, 20.5, 29.1, 30.9, 39.6, 47.2, 175.0, 178.6; IR 1759, 1636 cm^{-1} ; HRMS calcd for $\text{C}_{17}\text{H}_{26}\text{NO}_3$ 292.1913, found 292.1932.



5-Thioneostenine. 5-Oxoneostenine **40** (63 mg, 0.23 mmol) and phosphorous pentasulfide (100 mg, 0.23 mmol) were combined in CH_2Cl_2 (1 mL). A solution of hexamethyldisiloxane (73 mg, 0.45 mmol) in CH_2Cl_2 (1 mL) was added and the reaction monitored by TLC. After 2 h, the reaction was filtered through a plug of silica gel and concentrated to give a viscous oil. The crude mixture was purified by silica chromatography to give the pure thioamide as a white solid (49 mg, 0.16 mmol, 70% yield). $R_f = 0.45$ (1:1 EtOAc:hexanes); mp 212-214 $^\circ\text{C}$; ^1H NMR (400 MHz) δ 0.98 (t, $J = 7.3$ Hz, 3 H), 1.28 (d, $J = 7.3$ Hz, 3 H), 1.38 (m, 1 H), 1.49-1.64 (complex, 5 H), 1.72 (m, 1 H), 1.99 (m, 1 H), 2.12 (m, 1 H), 2.25 (m, 1 H), 2.33 (m, 1 H), 2.67 (m, 1 H), 2.98 (m, 1 H), 3.57 (dd, $J = 7.6, 14.1$ Hz, 1 H), 3.78 (dt, $J = 6.3, 12.1$ Hz, 1 H), 4.00 (d, $J = 5.3$ Hz, 1 H), 4.37 (dd, $J = 7.8, 13.9$ Hz, 1 H), 4.60 (dd, $J = 2.3, 4.6$, 1 H); ^{13}C NMR (100 MHz) δ d 9.4, 10.9, 33.1, 34.0, 38.9, 40.6, 41.8, 66.8, 77.8; u 19.2, 20.6, 29.6, 30.7, 49.4, 56.4, 178.3, 204.6; IR 2977, 2933, 2878, 1764, 1482 cm^{-1} ; HRMS calcd for $\text{C}_{17}\text{H}_{26}\text{NO}_2\text{S}$ 308.1684, found 308.1691.



Neostenine 3. To a solution of 5-thioneostenine (26 mg, 0.085 mmol) in THF (1 mL) and EtOH (1 mL) was added Raney nickel 2800 slurry in water (230 mg). The mixture was stirred vigorously for 1.5 h, filtered through a Celite plug and the solvents removed in vacuo. The residue was purified by chromatography on basic alumina to give neostenine as a white solid (22 mg, 0.079 mmol, 93% yield). $R_f = 0.31$ (acetone); mp 126-128 °C (lit¹³ 90-92 °C); ¹H NMR (400 MHz) δ 0.99 (t, $J = 7.3$ Hz, 3 H), 1.22 (d, $J = 7.2$ Hz, 3 H), 1.41 (m, 1 H), 1.56-1.91 (complex, 10 H), 1.98 (m, 1 H), 2.26 (m, 1 H), 2.33 (m, 1 H), 2.39-2.48 (m, 2 H), 2.86 (m, 1 H), 3.19 (m, 1 H), 4.51 (m, 1 H); ¹³C NMR (100 MHz) δ d 10.6, 11.8, 34.8, 37.8, 38.0, 43.0, 43.5, 71.4, 79.9; u 21.6, 21.7, 28.6, 28.9, 30.7, 56.1, 56.4, 180.2; IR 2985, 2934, 1762, 1638 cm^{-1} ; HRMS calcd for $\text{C}_{17}\text{H}_{28}\text{NO}_2$ 278.2120, found 278.2131. Except for the increased melting point, these data are in agreement with those obtained by Lin and coworkers¹³.



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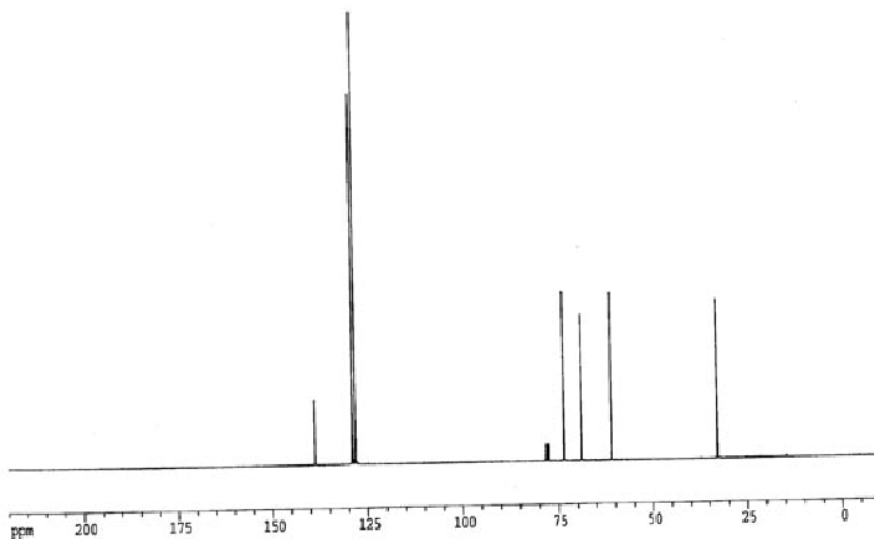
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SI       16384
SF       400.1300000 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

1D NMR plot parameters
CX       20.00 cm
F1P      10.000 ppm
F2P      -1.000 ppm
P2       -400.13 Hz
PPHMC    0.55000 ppm/cm
HSCM     220.07150 Hz/cm
  
```



```

===== CHANNEL f1 =====
NAME      4c-VI-24
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20010629
Time     24.21
INSTRUM  drx400
PROBHD   5 mm Multinu
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       300
DS       2
SMB      23148.148 Hz
FIDRES   0.353213 Hz
AQ       1.4156176 sec
RG       6256
DM       21.600 usec
DE       4.50 usec
TE       300.0 K
D1       0.85000000 sec
d1.1     0.83000000 sec
d1.2     0.00020000 sec

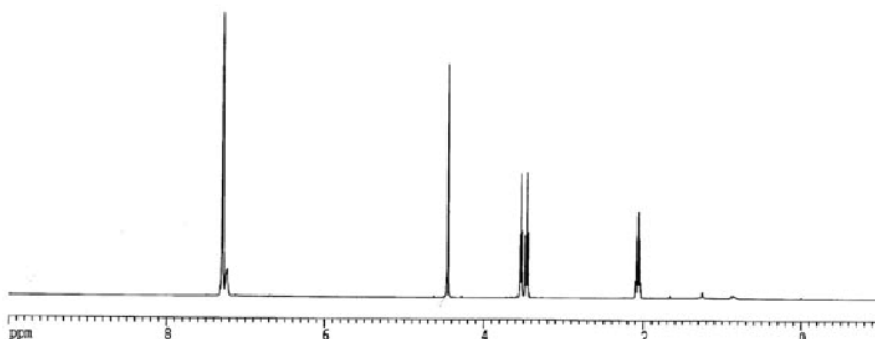
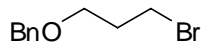
===== CHANNEL f1 =====
NUC1     13C
P1       12.30 usec
PL1      2.00 dB
SFO1     100.6282593 MHz

===== CHANNEL f2 =====
NAME      4c-VI-24
EXPNO    2
PROCNO   1
PULPROG  zgpg30
TD       32768
SOLVENT  CDCl3
NS       16
DS       2
SMB      106.6129962 MHz
FIDRES   0.146157 Hz
AQ       3.4210291 sec
RG       16
DM       104.400 usec
DE       4.50 usec
TE       300.0 K
D1       1.00000000 sec

===== CHANNEL f2 =====
NUC1     13C
P1       12.30 usec
PL1      2.00 dB
SFO1     100.6282593 MHz

F2 - Processing parameters
SI       16384
SF       100.6129962 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.00

1D NMR plot parameters
CX       20.00 cm
F1P      220.000 ppm
F2P      220.140 Hz
P2       -10.000 ppm
PPHMC    11.50000 ppm/cm
HSCM     1157.04639 Hz/cm
  
```



```

Current Data Parameters
NAME      4c-VI-27
EXPNO    1
PROCNO   1

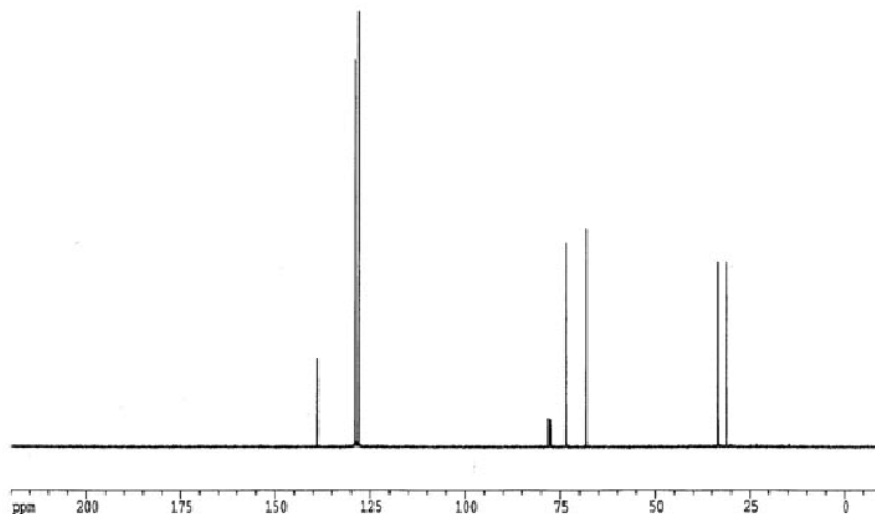
F2 - Acquisition Parameters
Date_    20010630
Time     13.30
INSTRUM  drx400
PROBHD   5 mm Multinu
PULPROG  zg30
TD       32768
SOLVENT  CDCl3
NS       16
DS       2
SWH      4789.272 Hz
FIDRES   0.146157 Hz
AQ       1.4210291 sec
RG       16
CW       104.400 usec
DE       4.50 usec
TE       300.0 K
D1       1.0000000 sec

***** CHANNEL f1 *****
NUC1     1H
P1       7.70 usec
PL1     -6.00 dB
SFO1    400.1320007 MHz

F2 - Processing parameters
SI       16384
SF       400.1320011 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

1D NMR plot parameters
CX       20.00 cm
PIF      10.000 ppm
F1       4001.30 Hz
F2P      -1.000 ppm
F2       -400.13 Hz
PCMCN    0.35000 ppm/cm
HZCM     220.07153 Hz/cm

```



```

Current Data Parameters
NAME      4c-VI-27
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20010630
Time     13.33
INSTRUM  drx400
PROBHD   5 mm Multinu
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       30
DS       2
SWH      23144.148 Hz
FIDRES   0.333213 Hz
AQ       1.4158276 sec
RG       1824.6
CW       21.600 usec
DE       4.50 usec
TE       300.0 K
D1       0.05000000 sec
d11     0.03000000 sec
d12     0.00002000 sec

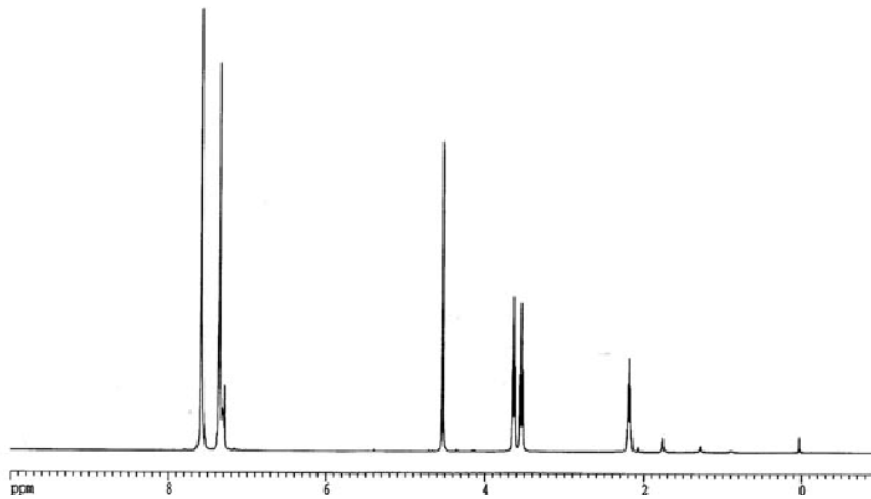
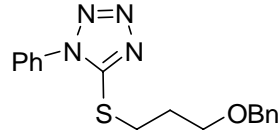
***** CHANNEL f1 *****
NUC1     13C
P1       12.30 usec
PL1      2.00 dB
SFO1    100.6252933 MHz

***** CHANNEL f2 *****
CFPRG2   waltz16
NUC2     1H
PCPD2    100.00 usec
PL2      2.00 dB
PL12     18.00 dB
PL13     18.00 dB
SFO2    400.3316005 MHz

F2 - Processing parameters
SI       32768
SF       100.6127290 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40

1D NMR plot parameters
CX       20.00 cm
PIF      220.000 ppm
F1       22034.80 Hz
F2P      -10.000 ppm
F2       -1006.13 Hz
PCMCN    11.50000 ppm/cm
HZCM     1125.8225 Hz/cm

```



```

Current Data Parameters
NAME      4c-V-72b
EXPNO    1
PROCNO    1

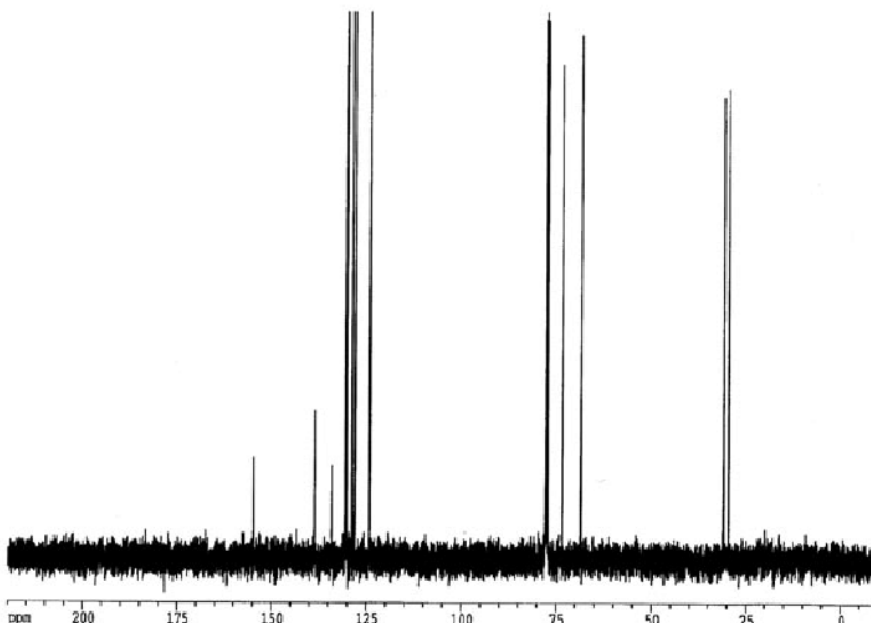
F2 - Acquisition Parameters
Date_     20010314
Time      19.00
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         2
SWH        4789.272 Hz
FIDRES    0.146157 Hz
AQ         3.4220291 sec
RG         301.6
RW         104.460 usec
DE         4.50 usec
TE         300.0 K
D1         1.0000000 sec

***** CHANNEL f1 *****
NUC1       1H
P1         7.70 usec
PL1        -6.00 dB
SFO1       400.1320007 MHz

F2 - Processing parameters
SI         16384
SF         400.1300000 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
F1P        30.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
PPHMC      0.55000 ppm/cm
HZCM       220.07150 Hz/cm

```



```

Current Data Parameters
NAME      4c-V-72b
EXPNO    2
PROCNO    1

F2 - Acquisition Parameters
Date_     20010314
Time      19.03
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         127
DS         2
SWH        23148.144 Hz
FIDRES    0.353213 Hz
AQ         1.4156276 sec
RG         8192
RW         21.600 usec
DE         4.50 usec
TE         300.0 K
D1         0.0500000 sec
d11       0.0300000 sec
d12       0.0000000 sec

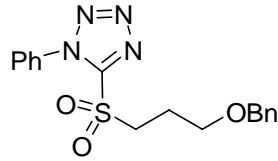
***** CHANNEL f1 *****
NUC1       13C
P1         12.30 usec
PL1         2.00 dB
SFO1       100.6212933 MHz

***** CHANNEL f2 *****
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        0.00 dB
PL12       18.00 dB
PL13       18.00 dB
SFO2       400.1316095 MHz

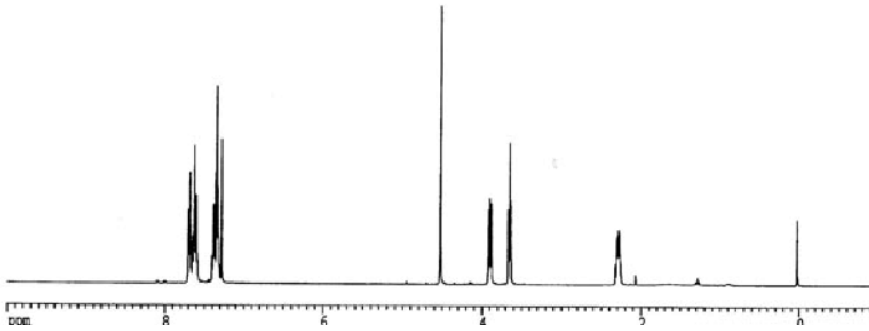
F2 - Processing parameters
SI         32768
SF         100.6127290 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
F1P        220.040 ppm
F1         22134.80 Hz
F2P        -10.000 ppm
F2         -100.61 Hz
PPHMC      11.50000 ppm/cm

```



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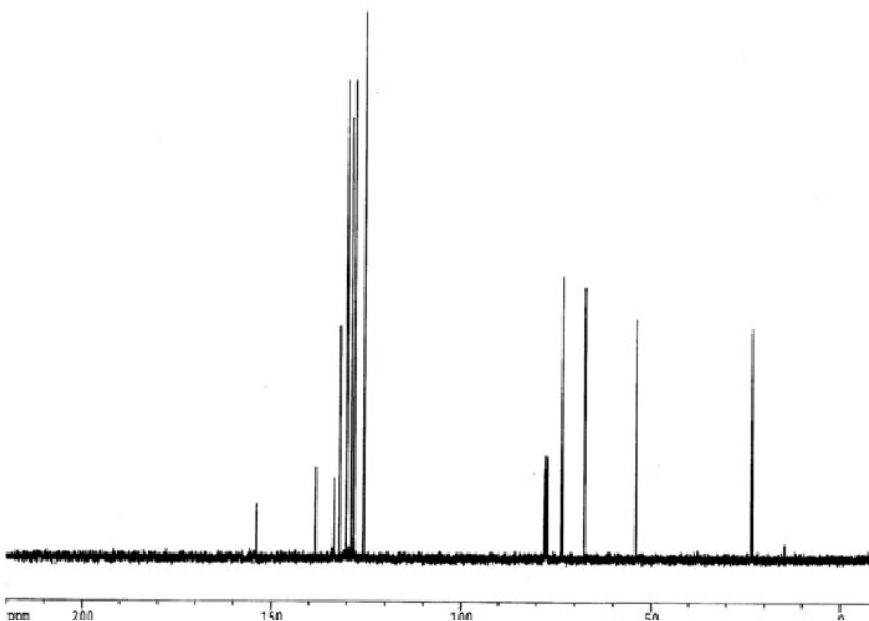
```
Current Data Parameters
NAME      4c-v-86a
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20010317
Time      13.35
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         2
SFO        4789.272 Hz
FIDRES     0.146157 Hz
AQ         3.4210291 sec
RG         203.2
DM         104.400 usec
DE         4.50 usec
TE         300.0 K
D1         1.00000000 sec
```

```
***** CHANNEL f1 *****
NUC1      1H
P1        7.70 usec
PL1       -6.00 dB
SFO1      400.1320007 MHz
```

```
F2 - Processing parameters
SI         14384
SF         400.1300000 MHz
WDM        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
```

```
1D NMR plot parameters
CX         20.00 cm
F1P        10.000 ppm
F1         4001.10 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
P1PCMC     0.55000 ppm/cm
HZCM       220.07150 Hz/cm
```



```
Current Data Parameters
NAME      4c-v-86a
EXPNO     2
PROCNO    1
```

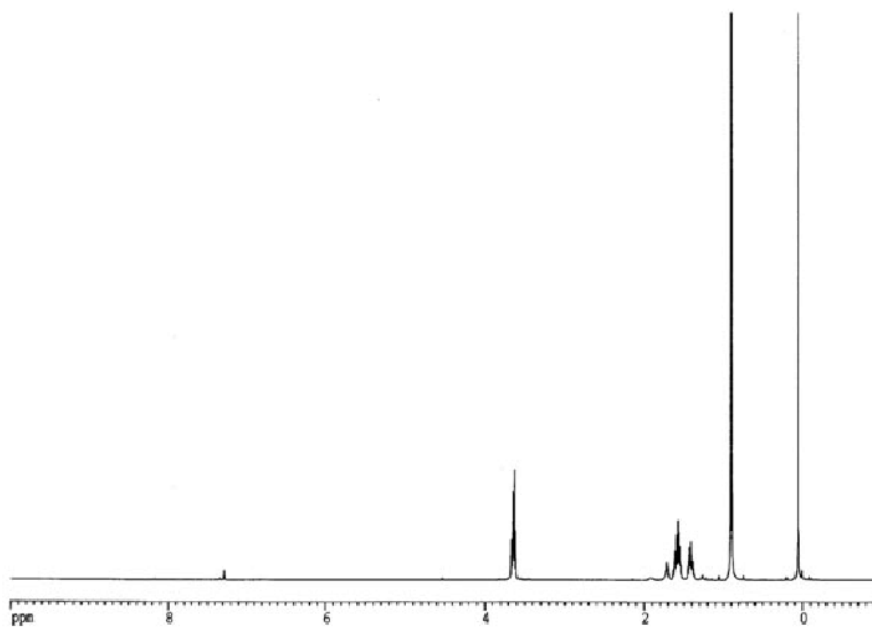
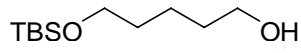
```
F2 - Acquisition Parameters
Date_     20010317
Time      12.50
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         65536
SOLVENT   cdcl3
NS         30
DS         2
SFO        20148.148 Hz
FIDRES     0.353213 Hz
AQ         1.4156276 sec
RG         2048
DM         21.500 usec
DE         4.50 usec
TE         300.0 K
D1         0.55000000 sec
d11        0.03000000 sec
d12        0.00002000 sec
```

```
***** CHANNEL f1 *****
NUC1      13C
P1        12.30 usec
PL1       2.00 dB
SFO1      100.6232933 MHz
```

```
***** CHANNEL f2 *****
CPDPRG2   waltz16
NUC2      1H
PCPD2     100.00 usec
PL2       0.00 dB
PL12      18.00 dB
PL13      18.00 dB
SFO2      400.1316005 MHz
```

```
F2 - Processing parameters
SI         32768
SF         100.6179210 MHz
WDM        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40
```

```
1D NMR plot parameters
CX         20.00 cm
F1P        220.000 ppm
F1         20154.00 Hz
F2P        -10.000 ppm
F2         -1006.13 Hz
P1PCMC     11.50000 ppm/cm
```



```

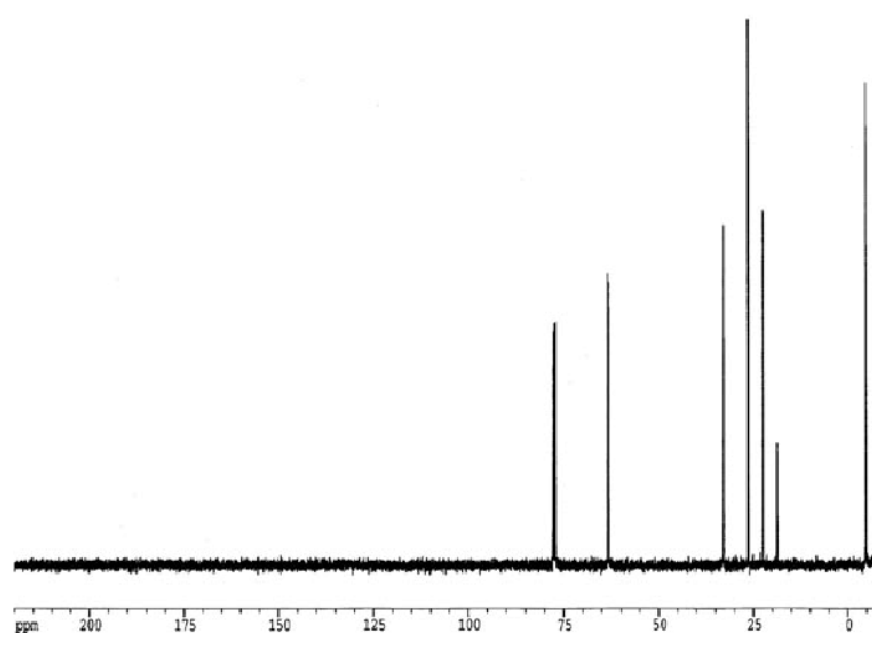
Current Data Parameters
NAME      4c-VI-29
EXPNO    1
PROCNO   1

F1 - Acquisition Parameters
Date_    20010701
Time     14.29
INSTRUM  drx400
PROBHD   5 mm Multinu
PULPROG  zg30
TD       32768
SOLVENT  CDCl3
NS       16
DS       2
SSB      4700.272 Hz
FIDRES   0.146357 Hz
AQ       3.4210291 sec
RG       64
SWH      104.400 usec
DE       4.50 usec
TE       300.0 K
D1       1.00000000 sec

***** CHANNEL f1 *****
NUC1     13C
P1       7.70 usec
PL1     -6.00 dB
SFO1    400.1320007 MHz

F2 - Processing parameters
SI       16384
SF       400.1320000 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

ID INE plot parameters
CX       20.00 cm
F1P      10.000 ppm
F1       4002.30 Hz
F2P      -1.000 ppm
F2       -400.13 Hz
PRFCHM   0.35000 ppm/cm
HSCM     220.07150 Hz/cm
  
```



```

Current Data Parameters
NAME      4c-VI-29
EXPNO    2
PROCNO   1

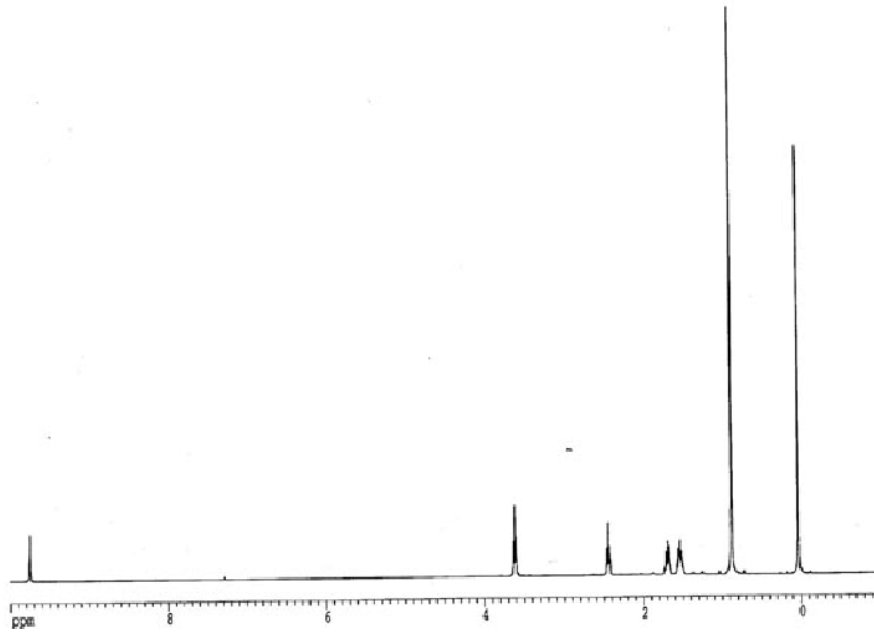
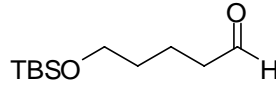
F2 - Acquisition Parameters
Date_    20010701
Time     14.35
INSTRUM  drx400
PROBHD   5 mm Multinu
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       197
DS       2
SWH      21148.148 Hz
FIDRES   0.353213 Hz
AQ       1.4156276 sec
RG       4096
SW       21.400 usec
DE       4.50 usec
TE       300.0 K
D1       0.05000000 sec
d11      0.03000000 sec
d12      0.00000000 sec

***** CHANNEL f1 *****
NUC1     13C
P1       12.30 usec
PL1      2.00 dB
SFO1    100.6217993 MHz

***** CHANNEL f2 *****
CFPRG2   waltz16
NUC2     1H
PCPRG2   100.00 usec
PL2      0.00 dB
PL12     18.00 dB
PL11     18.00 dB
SFO2    400.1316005 MHz

F2 - Processing parameters
SI       32768
SF       100.6217290 MHz
WDW      FM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40

ID INE plot parameters
CX       20.00 cm
F1P      220.000 ppm
F1       22234.80 Hz
F2P      -20.000 ppm
F2       -1006.13 Hz
PRFCHM   11.50000 ppm/cm
HSCM     180.00000 Hz/cm
  
```



```

-----
NAME      4c-VI-42
EXPNO     1
PROCNO    1

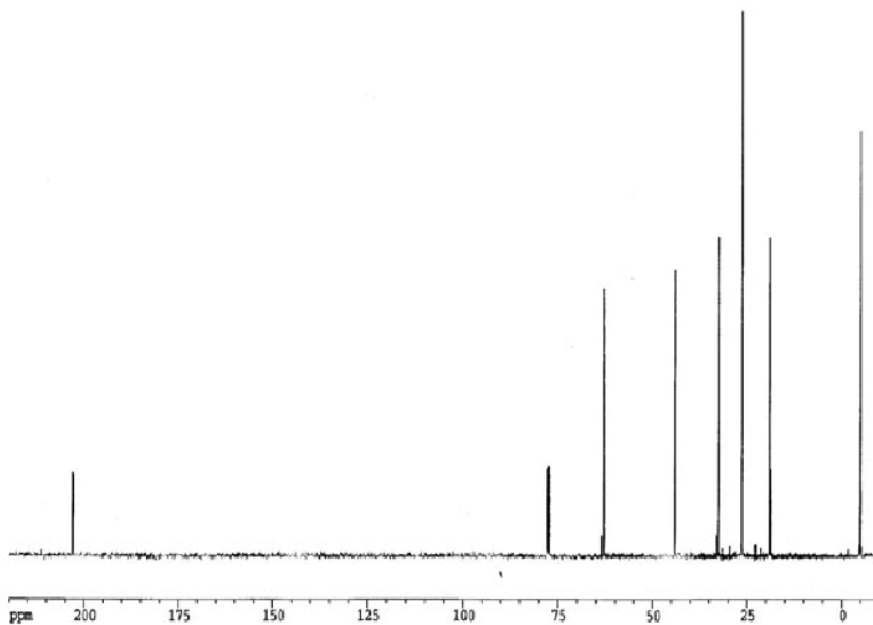
F2 - Acquisition Parameters
Date_     20010709
Time      12.20
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         2
SWH        4789.722 Hz
FIDRES     0.146157 Hz
AQ         3.4230291 sec
RG         32
RW         104.400 usec
DE         6.50 usec
TE         300.0 K
D1         1.0000000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.70 usec
PL1        -6.00 dB
SFO1       400.132007 MHz

F2 - Processing parameters
SI         32768
SF         400.1300000 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
F1P        16.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
PPHM       0.55000 ppm/cm
HZCM       220.07150 Hz/cm

```



```

Current Data Parameters
NAME      4c-VI-42
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20010709
Time      12.24
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         177
DS         2
SWH        20348.348 Hz
FIDRES     0.353213 Hz
AQ         1.4056276 sec
RG         3292
RW         31.600 usec
DE         4.50 usec
TE         300.0 K
D1         0.0500000 sec
dL1        0.0300000 sec
dL2        0.0500000 sec

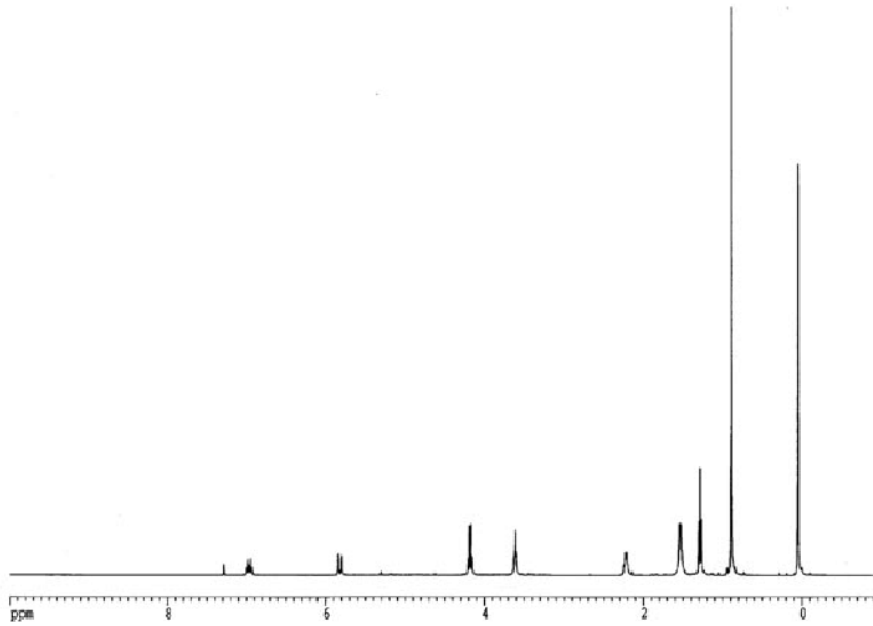
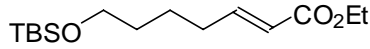
===== CHANNEL f1 =====
NUC1       13C
P1         12.10 usec
PL1         2.00 dB
SFO1       100.6212933 MHz

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        9.00 dB
PL12       18.00 dB
PL13       18.00 dB
SFO2       400.1314005 MHz

F2 - Processing parameters
SI         32768
SF         100.6127290 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
F1P        11.500 ppm
F1         221.34.00 Hz
F2P        -1006.13 Hz
F2         11.50000 ppm/cm
PPHM       11.50000 ppm/cm

```



```

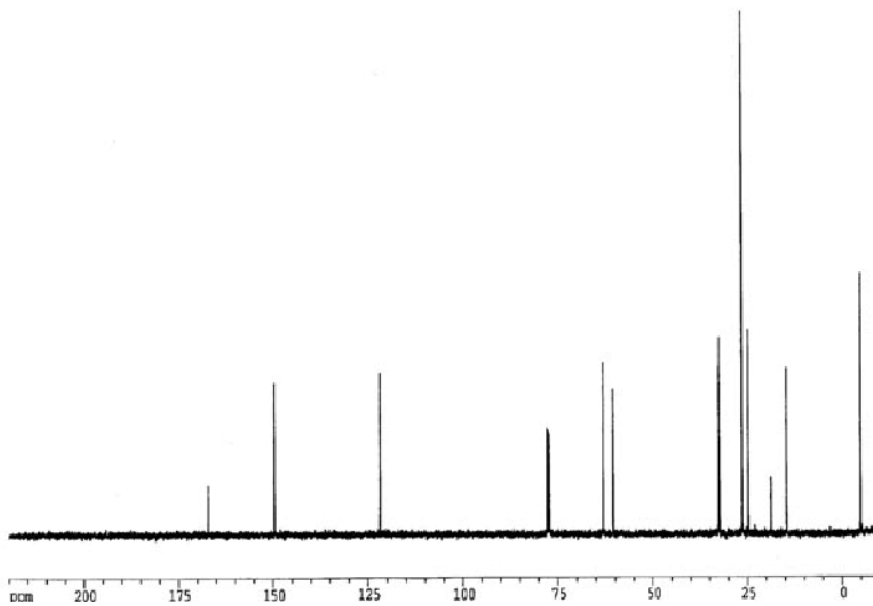
Current Data Parameters
NAME      4c-IV-277
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20010123
Time     15.45
INSTRUM  drx400
PROBHD   5 mm Multima
PULPROG  zg30
TD       32768
SOLVENT  CDCl3
NS       16
DS       2
SWH      4789.272 Hz
FIDRES   0.146157 Hz
AQ       3.4210251 sec
RG       52
RM       104.400 usec
DE       4.50 usec
TE       300.0 K
TE       300.0 K
SI       1.00000000 sec

===== CHANNEL f1 =====
NUC1     1H
P1       7.70 usec
PL1      -6.00 dB
SFO1     400.1320007 MHz

F2 - Processing parameters
SI       16384
SF       400.1300000 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

1D NMR plot parameters
CX       20.00 cm
F1P      10.000 ppm
F1       4001.30 Hz
F2P      -1.000 ppm
F2       -400.13 Hz
PRGMCM   0.55000 ppm/cm
RGCM     220.07150 Hz/cm
  
```



```

Current Data Parameters
NAME      4c-IV-277
EXPNO    2
PROCNO   1

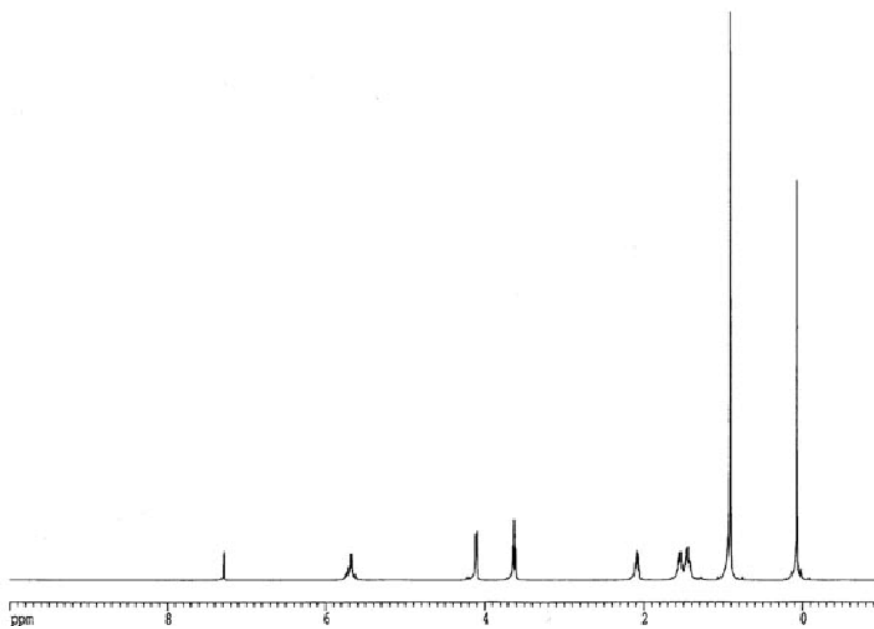
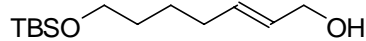
F2 - Acquisition Parameters
Date_    20010123
Time     15.48
INSTRUM  drx400
PROBHD   5 mm Multima
PULPROG  zgpg30
TD       65536
SOLVENT  CDCl3
NS       125
DS       2
SWH      23548.148 Hz
FIDRES   0.133213 Hz
AQ       1.4156276 sec
RG       1024
RM       21.600 usec
DE       4.50 usec
TE       300.0 K
TE       300.0 K
SI       0.05000000 sec
SI1      0.03000000 sec
SI2      0.00002000 sec

===== CHANNEL f1 =====
NUC1     13C
P1       12.30 usec
PL1      2.00 dB
SFO1     100.6232913 MHz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2     1H
P2P2     100.00 usec
PL2      0.00 dB
PL12     18.00 dB
PL13     18.00 dB
SFO2     400.1316005 MHz

F2 - Processing parameters
SI       13768
SF       100.6127230 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40

1D NMR plot parameters
CX       20.00 cm
F1P      220.016 ppm
F1       22338.42 Hz
F2P      -10.016 ppm
F2       -1009.73 Hz
PRGMCM   11.49350 ppm/cm
RGCM     1147.40747 Hz/cm
  
```

```

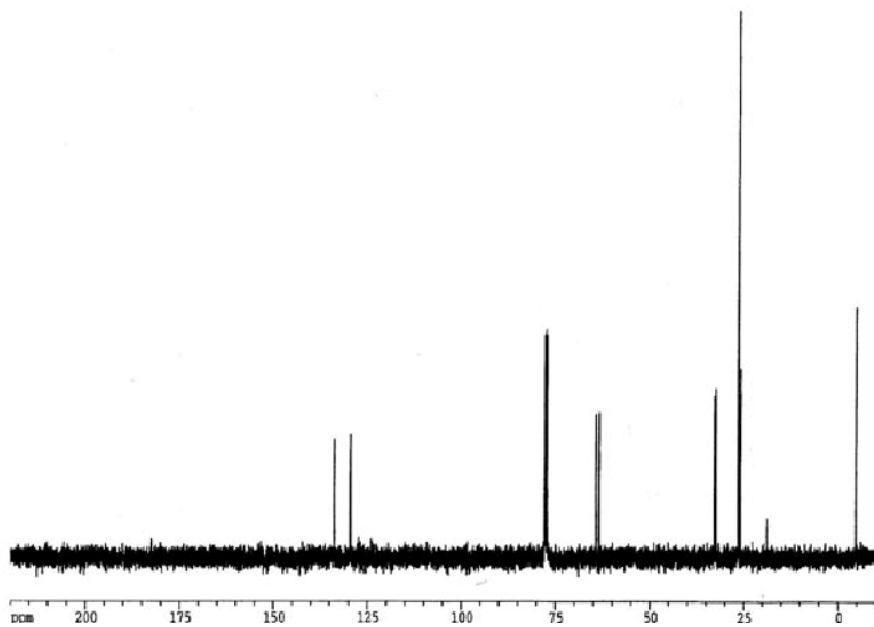
Current Data Parameters
NAME      4c-IV-260
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20010118
Time      14.11
INSTRUM   drx400
PROBHD    5 mm Multima
PULPROG   zg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         2
SWH        4789.272 Hz
FIDRES     0.146157 Hz
AQ         3.4210281 sec
RG         64
TM         104.400 usec
DE         4.50 usec
TE         300.0 K
DL         1.00000000 sec

***** CHANNEL f1 *****
NUC1       1H
P1         7.70 usec
PL1        -6.00 dB
SFO1       400.1320007 MHz

F2 - Processing parameters
SI         16384
SF         400.1300000 MHz
WMW        EM
SSB         0
LB         0.10 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
F1P        10.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
PPHMX      0.55000 ppm/cm
HZCN       220.07150 Hz/cm
  
```



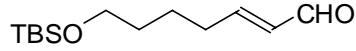
```

Current Data Parameters
NAME      4c-IV-260
EXPNO     2
PROCNO    1

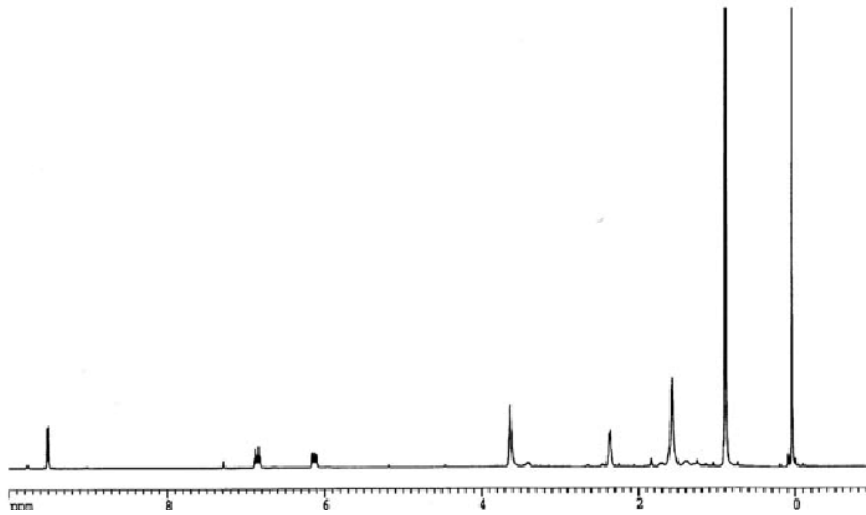
F2 - Acquisition Parameters
Date_     20010118
Time      14.15
INSTRUM   drx400
PROBHD    5 mm Multima
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         122
DS         2
SWH        23148.148 Hz
FIDRES     0.253213 Hz
AQ         1.4154276 sec
RG         2048
TM         21.400 usec
DE         4.50 usec
TE         300.0 K
dS1        0.0300000 sec
dS2        0.0000200 sec
PL13       18.00 dB
P1         0.05000000 sec
CPDPRG2   waltz16
PCFG2     100.00 usec
SFO2       400.1316005 MHz
NUC2       13C
PC2         0.00 dB
PL12       18.00 dB
P12        12.30 usec
DE         4.50 usec
SFO1       100.6232933 MHz
NUC1       13C
PL1         2.00 dB

F2 - Processing parameters
SI         32768
SF         100.6127290 MHz
WMW        EM
SSB         0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
F1P        220.000 ppm
F1         22134.80 Hz
F2P        -10.000 ppm
F2         -10064.13 Hz
PPHMX      11.50000 ppm/cm
  
```



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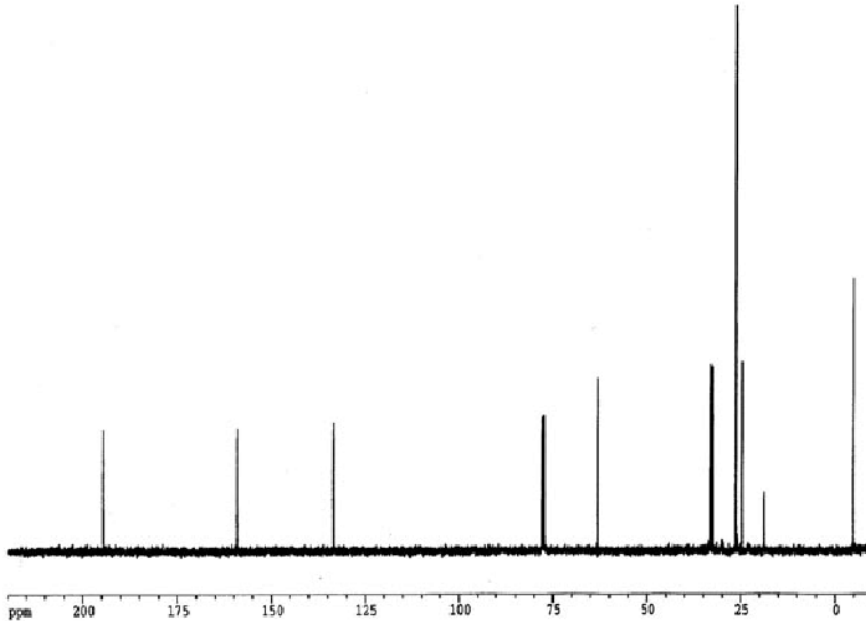
```

Current Data Parameters
NAME      4c-VI-81b9
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     1010723
Time      14.31
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         2
SWH        4789.272 Hz
FIDRES    0.146157 Hz
AQ         3.420291 sec
RG         45.3
DM         104.400 usec
DE         4.50 usec
TE         300.0 K
D1         1.0000000 sec
F1         7.70 usec
DE         4.50 usec
SFO1      400.132007 MHz
NUC1       1H
FL1        -6.00 dB

F2 - Processing parameters
SI         16384
SF         400.1320000 MHz
WDW        RM
SSB        0
LB         0.10 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
F1P        10.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
FPMCH      0.55000 ppm/cm
HDCM       220.07150 Hz/cm
  
```



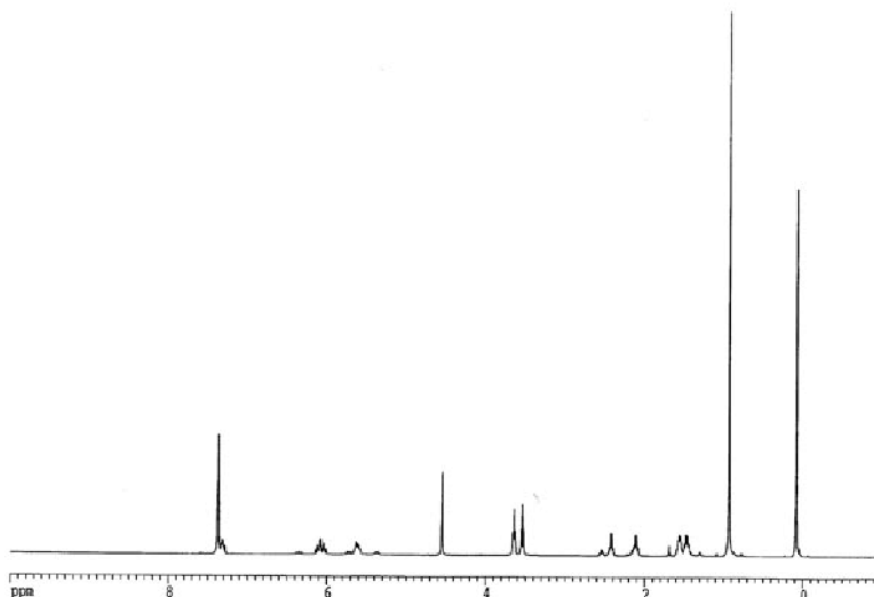
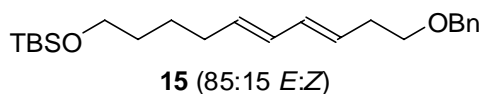
```

Current Data Parameters
NAME      4c-VI-81b9
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     1010723
Time      14.34
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         115
DS         2
SWH        23148.148 Hz
FIDRES    0.352213 Hz
AQ         1.4156278 sec
RG         8192
DM         21.600 usec
DE         4.50 usec
TE         300.0 K
d11        0.0300000 sec
d12        0.0000200 sec
FL1        18.00 dB
d13        0.0500000 sec
CPDPRG2   waltz16
PCPD2      100.00 usec
SFO2      400.1316005 MHz
NUC2       13C
FL2        0.00 dB
FL12       18.00 dB
F1         12.30 usec
DE         4.50 usec
SFO1      100.6232933 MHz
NUC1       13C
FL1        2.00 dB

F2 - Processing parameters
SI         32768
SF         100.6127290 MHz
WDW        RM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
F1P        210.000 ppm
F1         21124.80 Hz
F2P        -10.000 ppm
F2         -1006.13 Hz
FPMCH      11.50000 ppm/cm
HDCM       1157.04619 Hz/cm
  
```



```

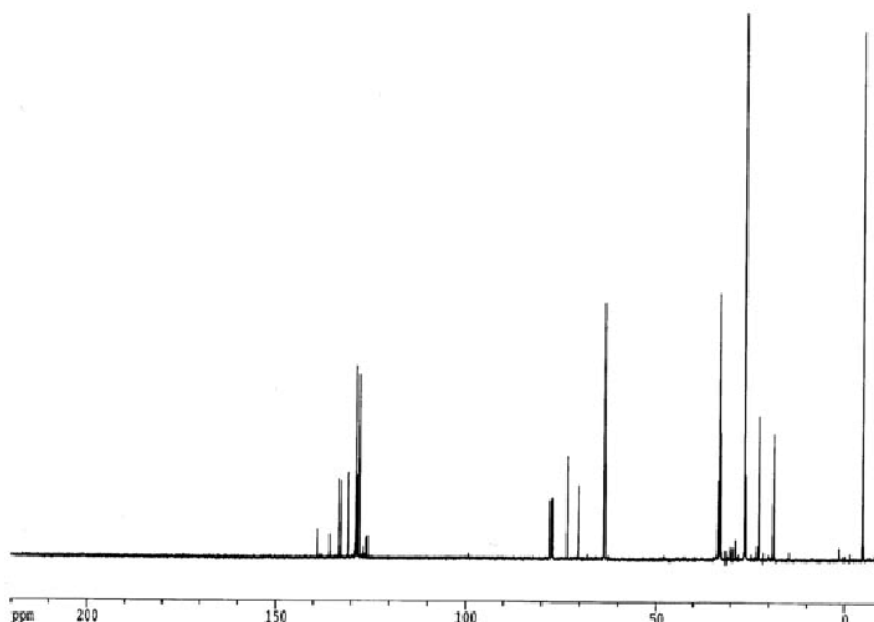
Current Data Parameters
NAME      4c-VI-20
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20010627
Time      13.48
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zgpg30
WDW        zg30
SSB        0
SOLVENT   CDCl3
NS         16
DS         2
SWH        4789.272 Hz
FIDRES     0.166157 Hz
AQ         3.4210291 sec
RG         64
DM         104.450 usec
DE         4.50 usec
TE         300.0 K
D1         1.00000000 sec

***** CHANNEL f1 *****
NUC1       1H
P1         7.70 usec
PL1        -6.00 dB
SFO1       400.1320007 MHz

F2 - Processing parameters
SI         16384
SF         400.1300000 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
FIP        10.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
FREQC      0.550000 ppm/cm
HZCM       220.07150 Hz/cm
  
```



```

Current Data Parameters
NAME      4c-VI-147
EXPNO     2
PROCNO    1

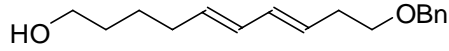
F2 - Acquisition Parameters
Date_     20010825
Time      16.01
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zgpg30
WDW        zg30
SSB        0
SOLVENT   CDCl3
NS         506
DS         2
SWH        23148.145 Hz
FIDRES     0.353213 Hz
AQ         1.4156275 sec
RG         2048
DM         21.600 usec
DE         4.50 usec
TE         300.0 K
D1         0.05000000 sec
d11        0.03000000 sec
d12        0.00000000 sec

***** CHANNEL f1 *****
NUC1       13C
P1         12.30 usec
PL1         2.00 dB
SFO1       100.6212933 MHz

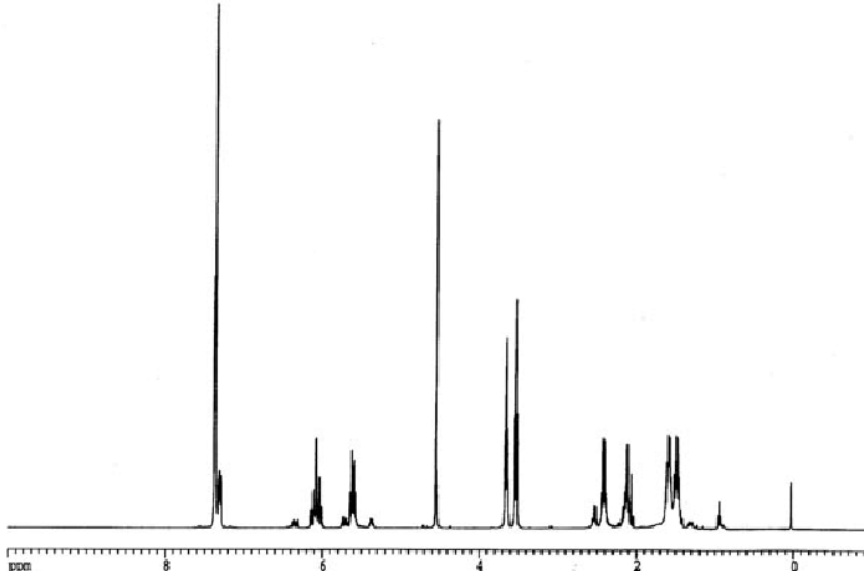
***** CHANNEL f2 *****
CPDPRG2   mltzr16
NUC2       1H
PCPD2     100.00 usec
PL2        0.00 dB
PL12       18.00 dB
PL13       18.00 dB
SFO2       400.1316045 MHz

F2 - Processing parameters
SI         32768
SF         100.617290 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
FIP        220.000 ppm
F1         22126.80 Hz
F2P        -18.000 ppm
F2         -1006.13 Hz
FREQC     11.500000 ppm/cm
  
```



(85:15 E:Z)



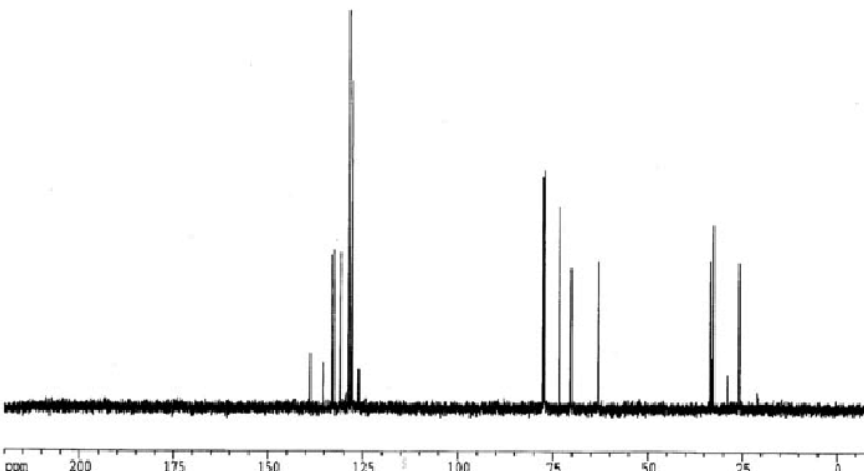
```

Current Data Parameters
NAME      4c-VI-21
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20010627
Time      17.47
INSTRUM   drx400
PROBHD    5 mm Multis
PULPROG   zg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         2
SFO       4789.272 Hz
FIDRES    0.146157 Hz
AQ         3.4219291 sec
RG         64
DM         104.400 usec
DE         4.50 usec
TE         300.0 K
D1         1.0000000 sec
F1         7.70 usec
DE         4.50 usec
SFO1      400.1326077 MHz
NUC1       13C
PL1        -6.00 dB

F2 - Processing parameters
SI         16384
SF         400.1300000 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
F1         400.13 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
FPCW       0.55000 ppm/cm
HZCM       220.07150 Hz/cm
  
```



```

Current Data Parameters
NAME      4c-VI-21
EXPNO     2
PROCNO    1

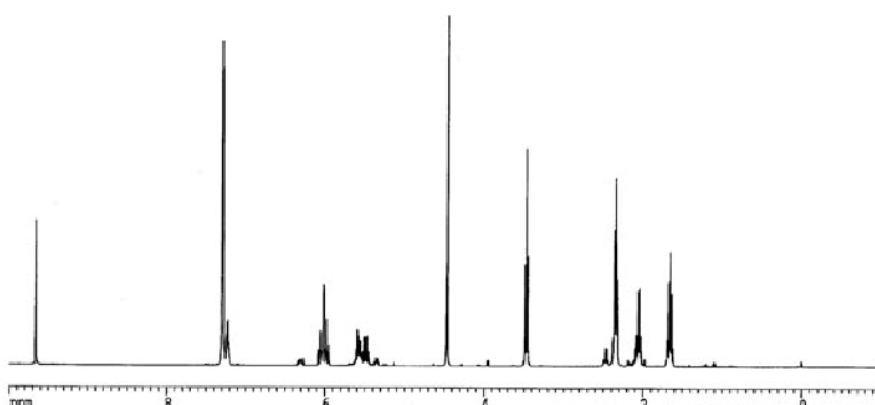
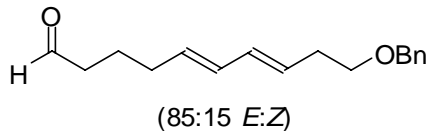
F2 - Acquisition Parameters
Date_     20010627
Time      17.57
INSTRUM   drx400
PROBHD    5 mm Multis
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         159
DS         2
SFO       21149.148 Hz
FIDRES    0.333213 Hz
AQ         1.4156276 sec
RG         9192.0
DM         21.600 usec
DE         4.50 usec
TE         300.0 K
D1         0.05000000 sec
dD1        0.03000000 sec
dD2        0.00000000 sec

***** CHANNEL f1 *****
NUC1       13C
F1         12.10 usec
PL1        2.00 dB
SFO1      100.6212813 MHz

***** CHANNEL f2 *****
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2       9.00 dB
PL12      18.00 dB
PL13      18.00 dB
SFO2      400.1316055 MHz

F2 - Processing parameters
SI         32768
SF         100.6127290 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
F1P        220.000 ppm
F1         221.9480 Hz
F2P        -11.000 ppm
F2         -2008.13 Hz
FPCW       11.50000 ppm/cm
  
```



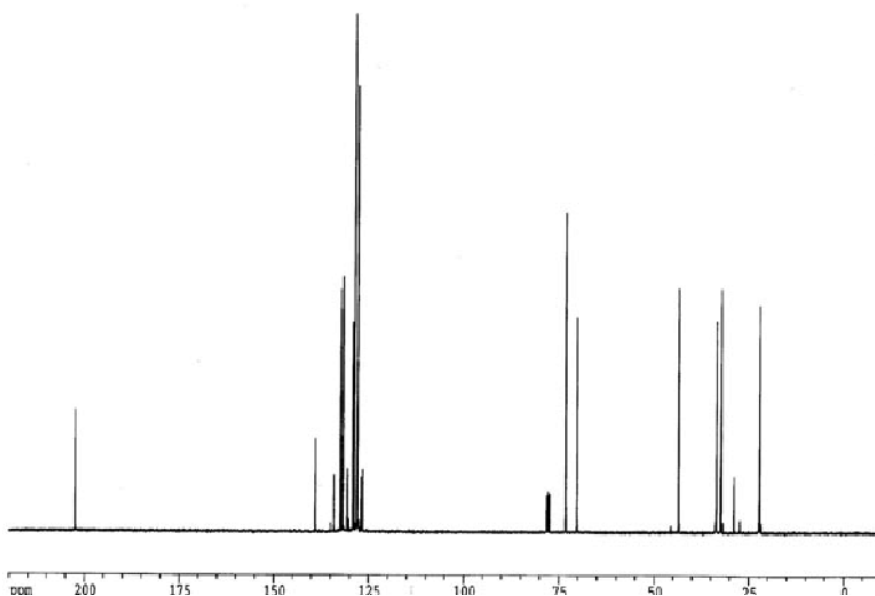
Current Data Parameters
 NAME 4c-V11-24
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20011104
 Time 14.17
 INSTRUM drx400
 PROBRD 5 mm Multinu
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SFO 4789.272 Hz
 FIDRES 0.144517 Hz
 AQ 3.4210291 sec
 RG 16
 DM 184.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.60000000 sec

***** CHANNEL f1 *****
 NUCL1 1H
 P1 7.70 usec
 PL1 -4.50 dB
 SFO1 400.1326007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300173 MHz
 MW 64
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 FIP 10.000 ppm
 FI 4001.30 Hz
 FZ -1.000 ppm
 P2 -400.13 Hz
 PPMCM 0.55000 ppm/cm
 HCM 220.07150 Hz/cm



Current Data Parameters
 NAME 4c-V11-24
 EXPNO 2
 PROCNO 1

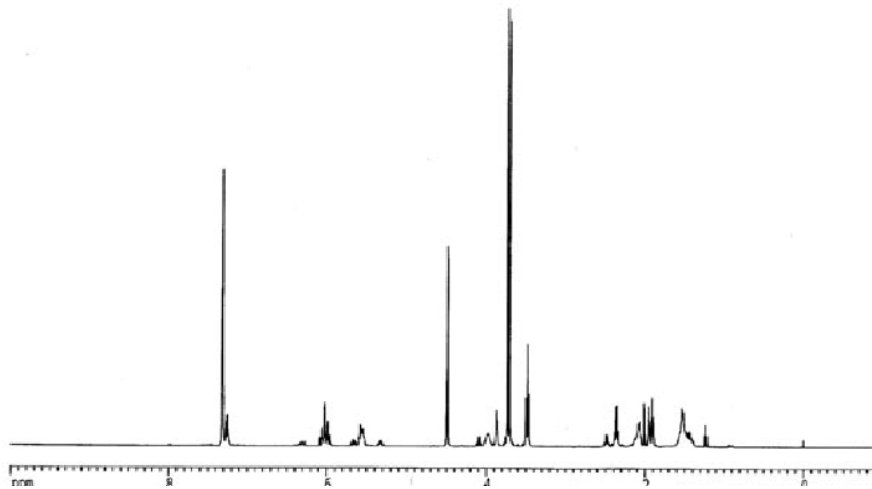
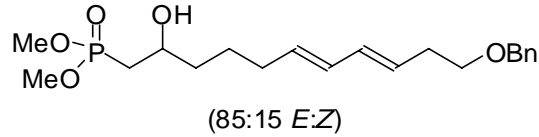
F2 - Acquisition Parameters
 Date_ 20011104
 Time 14.22
 INSTRUM drx400
 PROBRD 5 mm Multinu
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 108
 DS 2
 SFO 21148.148 Hz
 FIDRES 0.333213 Hz
 AQ 1.4156274 sec
 RG 1625.5
 DM 21.600 usec
 DE 4.50 usec
 TE 300.0 K
 D1 0.05000000 sec
 d11 0.03000000 sec
 d12 0.00000000 sec

***** CHANNEL f1 *****
 NUCL1 13C
 P1 12.30 usec
 PL1 2.00 dB
 SFO1 100.6212933 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUCL2 1H
 P2CPRG2 180.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127290 MHz
 MW 28
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 FIP 220.000 ppm
 FI 22134.80 Hz
 FZ -10.000 ppm
 P2 -1006.13 Hz
 PPMCM 11.50000 ppm/cm



```

Current Data Parameters
NAME      40-VI-262
EXPNO     1
PROCNO    1

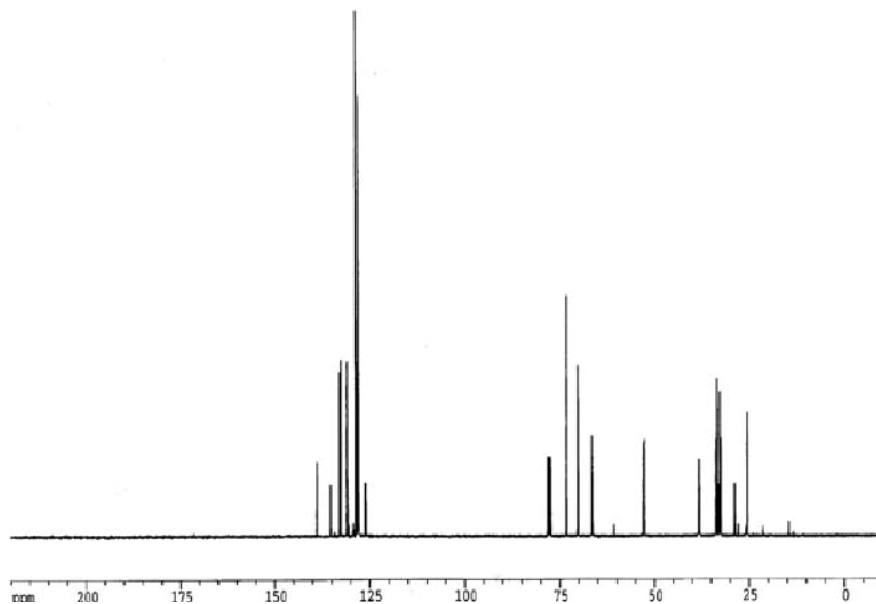
F2 - Acquisition Parameters
Date_     2011014
Time      12:35
INSTRUM   dnx400
PROBHD    5 mm Multinu
PULPROG   zg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         2
SWH        4789.272 Hz
FIDRES     0.146157 Hz
AQ         3.4210291 sec
RG         16
RW         104.400 usec
RE         4.50 usec
TE         300.0 K
TE        1.00000000 sec
SI         1.00000000 sec

***** CHANNEL f1 *****
NUC1       1H
P1         7.70 usec
PL1        -6.00 dB
SFO1       400.1320007 MHz

F2 - Processing parameters
SE         16384
SP         400.1299831 MHz
WDM        RM
SSB         0
LB         0.30 Hz
GB         0
PC         1.00

ID NMR plot parameters
CX         20.00 cm
F1P        10.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
FPMCM      0.55000 ppm/cm
HZCM       220.07149 Hz/cm

```



```

Current Data Parameters
NAME      40-VI-262
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     2011014
Time      12:43
INSTRUM   dnx400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         261
DS         2
SWH        21140.148 Hz
FIDRES     0.351213 Hz
AQ         1.4555276 sec
RG         4096
RW         21.600 usec
RE         4.50 usec
TE         300.0 K
TE        0.05000000 sec
SI         0.03000000 sec
SI1        0.03000000 sec
SI2        0.00000000 sec

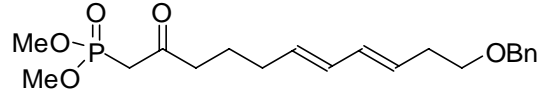
***** CHANNEL f1 *****
NUC1       13C
P1         12.30 usec
PL1         2.00 dB
SFO1       100.6232933 MHz

***** CHANNEL f2 *****
CPDPRG2   waltz16
NUC2       1H
PCPD2      300.00 usec
PL2         0.00 dB
PL12       18.00 dB
PL13       18.00 dB
SFO2       400.1316005 MHz

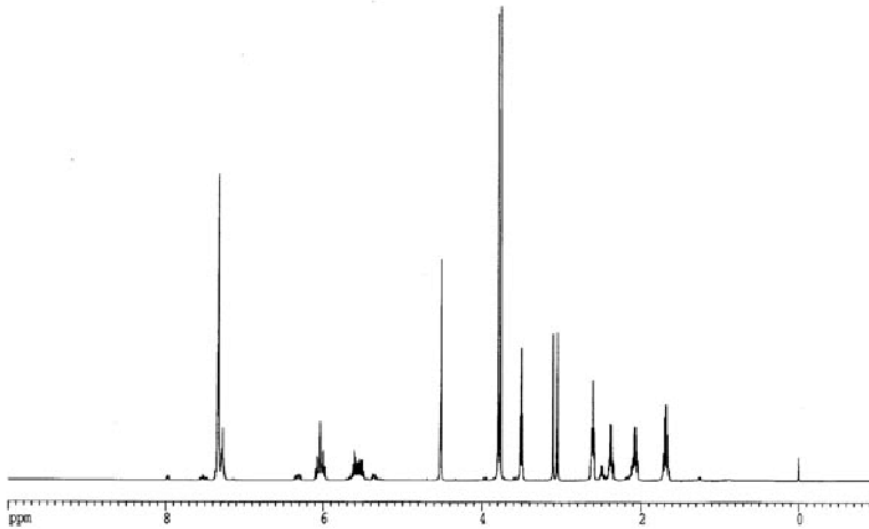
F2 - Processing parameters
SE         32768
SP         100.6232930 MHz
WDM        RM
SSB         0
LB         1.00 Hz
GB         0
PC         1.80

ID NMR plot parameters
CX         20.00 cm
F1P        220.000 ppm
F1         22134.80 Hz
F2P        -10.000 ppm
F2         -1006.13 Hz
FPMCM      11.50000 ppm/cm
HZCM       1445.82268 Hz/cm

```



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```

Current Data Parameters
NAME      4c-VI-26
EXPNO     1
PROCNO    1

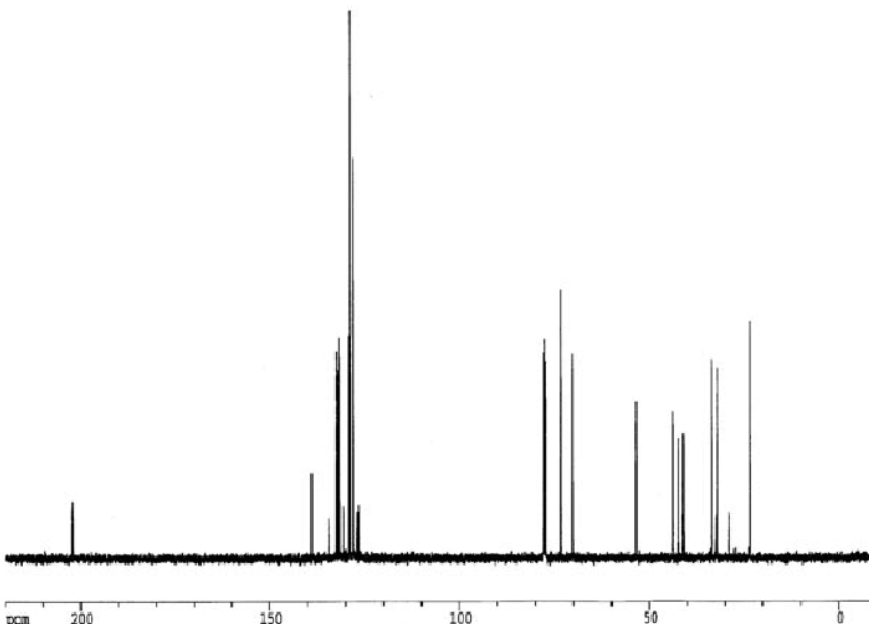
F2 - Acquisition Parameters
Date_     20010702
Time      14.47
INSTRUM   dxt400
PROBHD    5 mm Multinu
PULPROG   zg30
TD         31768
SOLVENT   CDCl3
NS         16
DS         2
SWH        4789.272 Hz
FIDRES     0.146157 Hz
AQ         3.4210291 sec
RG         64
RM         104.450 usec
DE         4.50 usec
TE         300.0 K
D1         3.0000000 sec

***** CHANNEL f1 *****
NUC1       1H
P1         7.70 usec
PL1        -6.00 dB
SFO1       400.1320007 MHz

F2 - Processing parameters
SI         16384
SF         400.1300041 MHz
RG         64
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

ID NMR plot parameters
CX         20.00 cm
F1P        10.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
FREQM      0.55000 ppm/cm
HZCM       220.07150 Hz/cm

```



```

Current Data Parameters
NAME      4c-VI-27
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20011109
Time      19.20
INSTRUM   dxt400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         212
DS         2
SWH        23148.148 Hz
FIDRES     0.333213 Hz
AQ         1.4156276 sec
RG         8192
RM         21.600 usec
DE         4.50 usec
TE         300.0 K
D1         0.8500000 sec
d11        0.8300000 sec
d12        0.0400000 sec

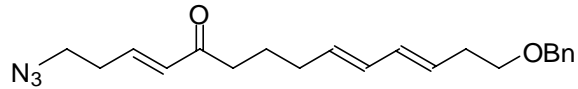
***** CHANNEL f1 *****
NUC1       13C
P1         12.30 usec
PL1         2.00 dB
SFO1       100.6232931 MHz

***** CHANNEL f2 *****
CHWPRG2   waltz16
NUC2       1H
PCPRG2    100.00 usec
PL2        0.00 dB
PL12       18.00 dB
PL13       18.00 dB
SFO2       400.1316005 MHz

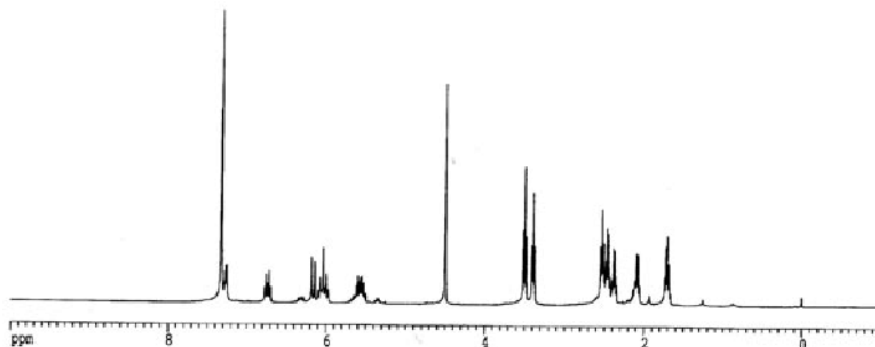
F2 - Processing parameters
SI         32768
SF         100.6127290 MHz
RG         64
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

ID NMR plot parameters
CX         20.00 cm
F1P        220.000 ppm
F1         22034.80 Hz
F2P        -10.000 ppm
F2         -1006.13 Hz
FREQM      11.50000 ppm/cm

```



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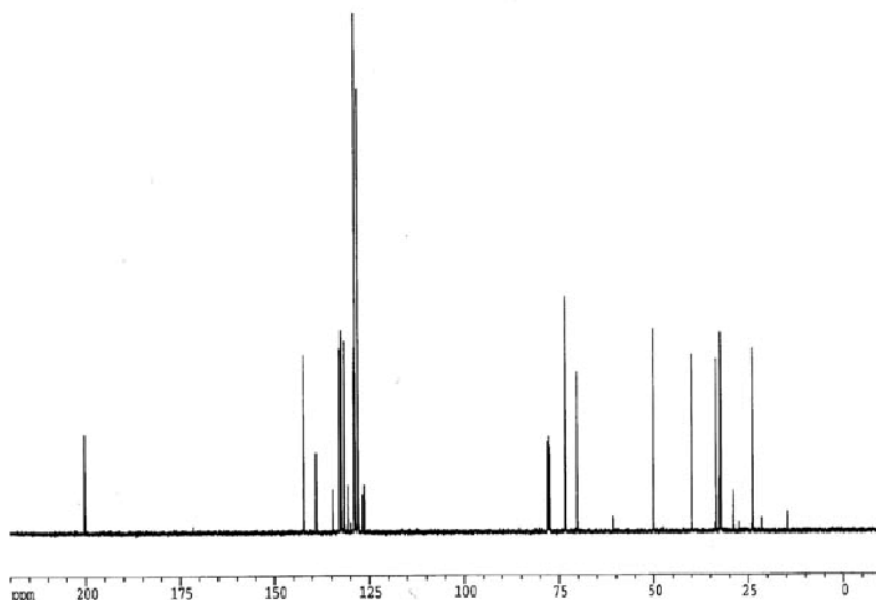
Current Data Parameters
 NAME 4c-VI-37
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20010709
 Time 14.23
 INSTRUM drx400
 PROBRD 5 mm Multinu
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 RG 32
 EW 104.400 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.0000000 sec

***** CHANNEL f1 *****
 NUCL1 1H
 P1 7.70 usec
 PL1 -8.00 dB
 SFO1 400.1324007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300097 MHz
 NDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 10.000 ppm
 F1 4001.30 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 FWHM 0.55000 ppm/cm
 HSCN 220.07150 Hz/cm



Current Data Parameters
 NAME 4c-VII-33
 EXPNO 2
 PROCNO 1

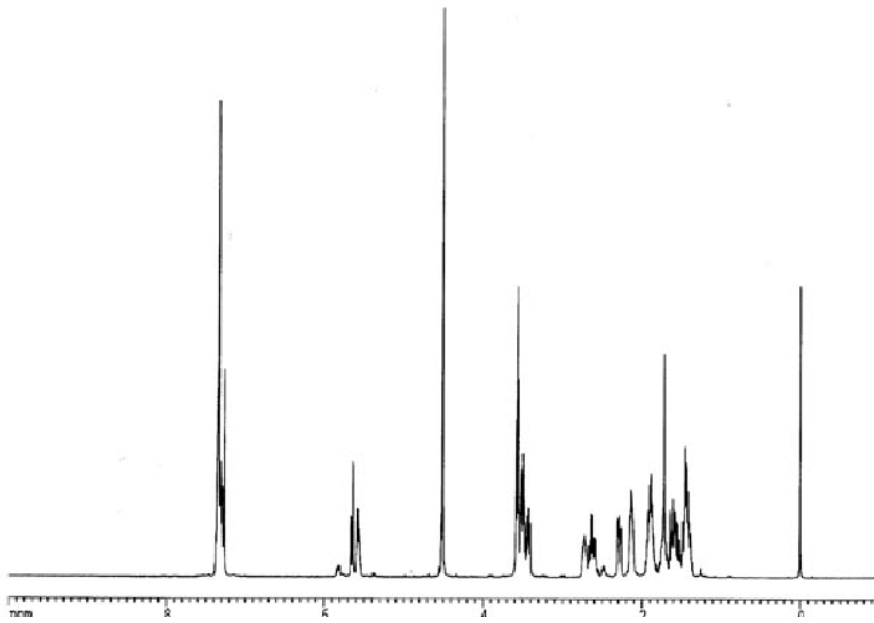
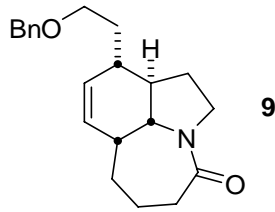
F2 - Acquisition Parameters
 Date_ 20011108
 Time 18.58
 INSTRUM drx400
 PROBRD 5 mm Multinu
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 371
 DS 2
 SWH 23348.348 Hz
 FIDRES 0.353213 Hz
 AQ 1.4156276 sec
 RG 8152
 EW 21.600 usec
 DE 4.50 usec
 TE 300.0 K
 D1 0.0500000 sec
 d11 0.0500000 sec
 d12 0.0000000 sec

***** CHANNEL f1 *****
 NUCL1 13C
 P1 12.10 usec
 PL1 2.00 dB
 SFO1 100.6212911 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUCL2 1H
 PCPDG2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL1J 18.00 dB
 SFO2 400.1316095 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127290 MHz
 NDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 220.000 ppm
 F1 22224.89 Hz
 F2P -20.000 ppm
 F2 -1006.13 Hz
 FWHM 11.50000 ppm/cm
 HSCN 1357.04639 Hz/cm



```

Current Data Parameters
NAME 4c-VI1-3/major
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20011124
Time 16.07
INSTRUM drx400
PROBHD 5 mm Multinu
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 113.7
DM 104.400 umet
DE 4.50 umec
TE 300.0 K
DL 1.00000000 sec
  
```

```

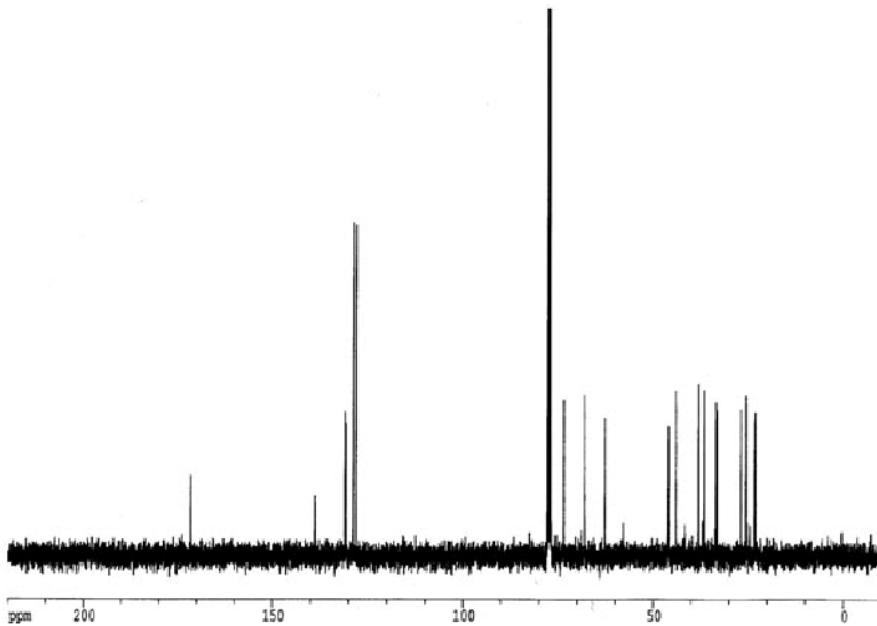
***** CHANNEL f1 *****
NUC1 1H
P1 7.70 umec
PL1 -6.00 dB
SFO1 400.1320007 MHz
  
```

```

F2 - Processing parameters
SI 16384
SF 400.1300074 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
  
```

```

1D NMR plot parameters
CX 20.00 cm
FIP 20.000 ppm
F1 4001.30 Hz
F2P -1.000 ppm
F2 -400.13 Hz
FPMCM 0.55000 ppm/cm
HSCM 220.07150 Hz/cm
  
```



```

Current Data Parameters
NAME 4c-VI1-3/major
EXPNO 2
PROCNO 1
  
```

```

F2 - Acquisition Parameters
Date_ 20011124
Time 28.42
INSTRUM drx400
PROBHD 5 mm Multinu
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 380
DS 2
SWH 23148.148 Hz
FIDRES 0.353213 Hz
AQ 1.4156276 sec
RG 8192
DM 22.600 umec
DE 4.50 umec
TE 300.0 K
D1 0.05000000 sec
d11 0.03000000 sec
d12 0.00002000 sec
  
```

```

***** CHANNEL f1 *****
NUC1 13C
P1 12.30 umec
PL1 2.00 dB
SFO1 100.6229233 MHz
  
```

```

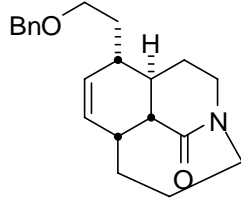
***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 umec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1326005 MHz
  
```

```

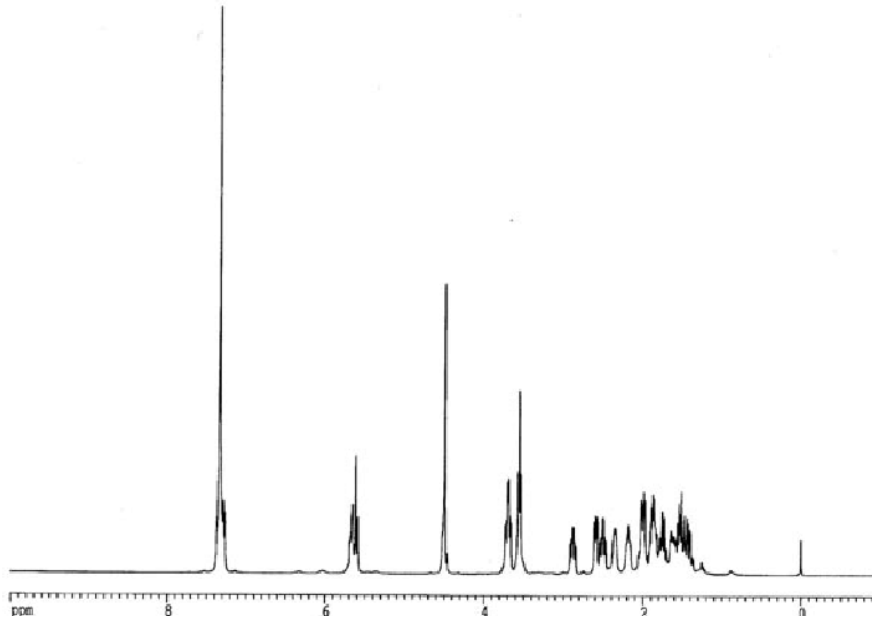
F2 - Processing parameters
SI 32768
SF 100.6127990 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40
  
```

```

1D NMR plot parameters
CX 20.00 cm
FIP 220.000 ppm
F1 22138.80 Hz
F2P -10.000 ppm
F2 -1006.13 Hz
FPMCM 11.50000 ppm/cm
  
```



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```

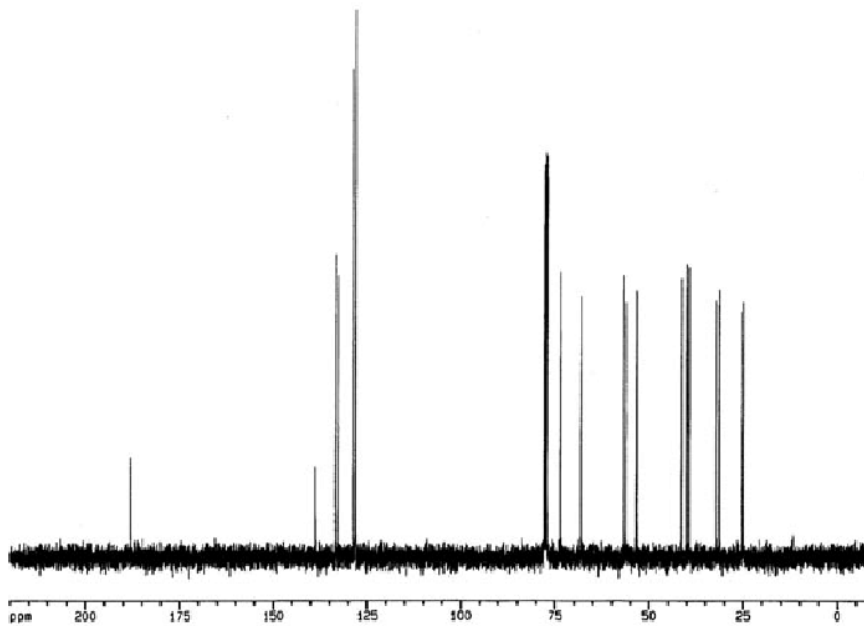
Current Data Parameters
NAME      4c-Villocin
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     2010724
Time      17.21
INSTRUM   drw400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         2
SWH        4789.272 Hz
FIDRES     0.146157 Hz
AQ         3.4210291 sec
RG         64
DM         104.400 usec
DE         4.50 usec
TE         300.0 K
D1         1.00000000 sec

***** CHANNEL f1 *****
NUC1       1H
P1         7.70 usec
PL1        -6.00 dB
SFO1       400.1320007 MHz

F2 - Processing parameters
SI         16384
SF         400.1300068 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
FIP        10.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
PFRMCK     0.55000 ppm/cm
HZCM       220.07190 Hz/cm
  
```



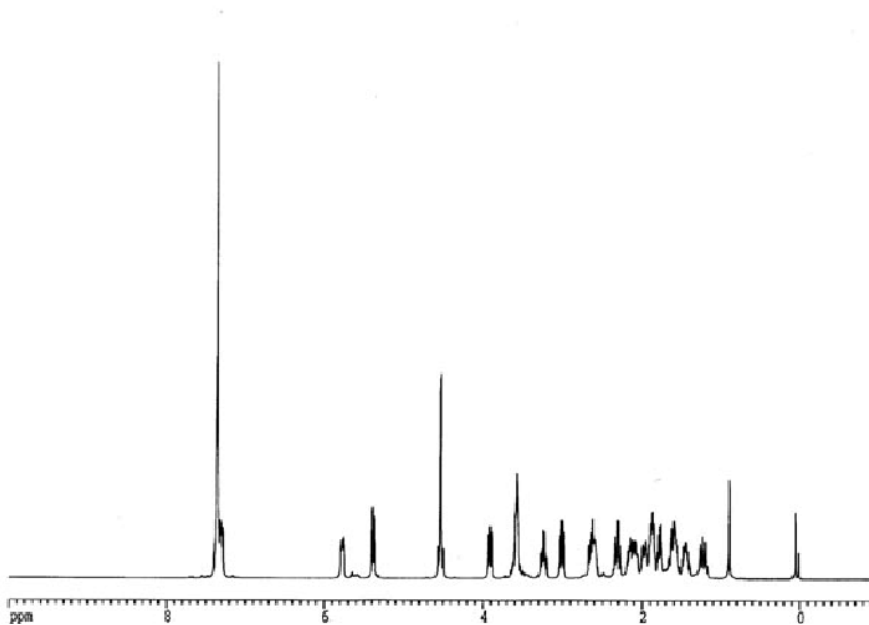
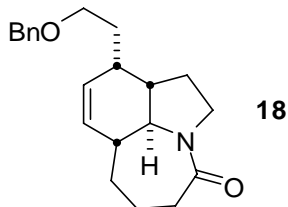
```

Current Data Parameters
NAME      4c-Villocin
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     1010724
Time      17.25
INSTRUM   drw400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         8
DS         2
SWH        23148.148 Hz
FIDRES     0.383213 Hz
AQ         1.4156276 sec
RG         4096
DM         23.600 usec
DE         4.50 usec
TE         300.0 K
d11        0.0300000 sec
d12        0.0000200 sec
PL13       18.00 dB
D1         0.05000000 sec
CPDPRG2   waltz16
PCPD2     100.00 usec
SFO2       400.1316005 MHz
NUC2       13C
PL2        0.00 dB
PL12       18.00 dB
P1         12.30 usec
DE         4.50 usec
SFO1       100.6232933 MHz
NUC1       13C
PL1        2.00 dB

F2 - Processing parameters
SI         32768
SF         100.6187790 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
FIP        200.000 ppm
F1         22134.80 Hz
F2P        -10.000 ppm
F2         -1506.33 Hz
PFRMCK     11.50000 ppm/cm
HZCM       1157.04638 Hz/cm
  
```



```

Current Data Parameters
NAME      4c-VI-100a
EXPNO     1
PROCNO    1

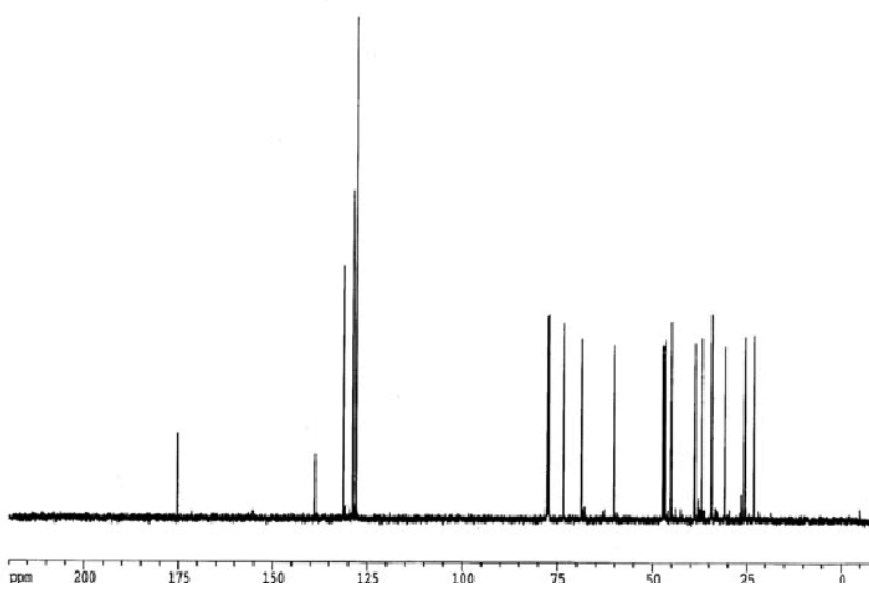
F2 - Acquisition Parameters
Date_     20010820
Time      13.08
INSTRUM   drs400
PROBHD    5 mm Multinu
PULPROG   zg30
TD         32768
SOLVENT   CDCl3
NS         16
DS         2
SFO       4789.272 Hz
FIDRES    0.344857 Hz
AQ         3.4210291 sec
RG         64
DM         104.400 usec
DE         4.50 usec
TE         300.0 K
D1         1.00000000 sec

***** CHANNEL f1 *****
NUC1      1H
P1        7.70 usec
PL1       -6.00 dB
SFO1      400.1320007 MHz

F2 - Processing parameters
SI         16384
SF         400.1300000 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
F1P        10.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
PFRMCH    0.55000 ppm/cm
HDCM      220.07150 Hz/cm

```



```

Current Data Parameters
NAME      4c-VI-100a
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20010820
Time      13.12
INSTRUM   drs400
PROBHD    5 mm Multinu
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         401
DS         2
SFO       23148.148 Hz
FIDRES    0.232121 Hz
AQ         1.4156276 sec
RG         8192
DM         21.600 usec
DE         4.50 usec
TE         300.0 K
D1         0.85000000 sec
d11        0.83000000 sec
d12        0.00000000 sec

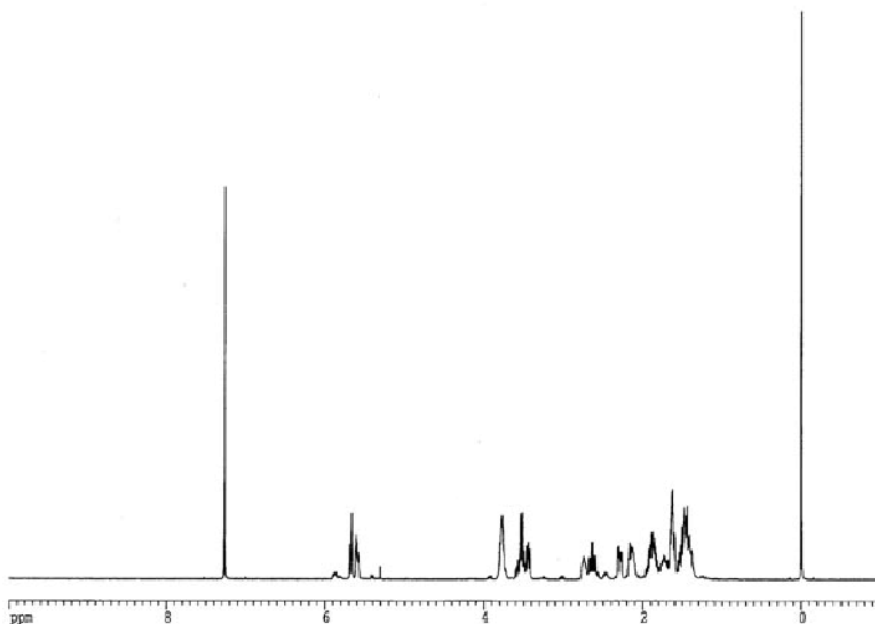
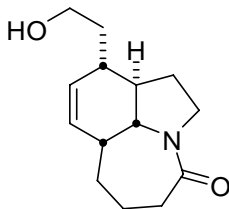
***** CHANNEL f1 *****
NUC1      13C
P1        12.10 usec
PL1        2.00 dB
SFO1      100.6232933 MHz

***** CHANNEL f2 *****
CPOPRG2   waltz16
NUC2      1H
PCPD2     100.00 usec
PL2        0.00 dB
PL12       18.00 dB
PL13       18.00 dB
SFO2      400.1316005 MHz

F2 - Processing parameters
SI         12768
SF         100.6127290 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
F1P        221.000 ppm
F1         22134.80 Hz
F2P        -10.000 ppm
F2         -1006.13 Hz
PFRMCH    11.50000 ppm/cm

```



```

Current Data Parameters
NAME: 4C-VII-56-8
EXPNO: 1
PROCNO: 1

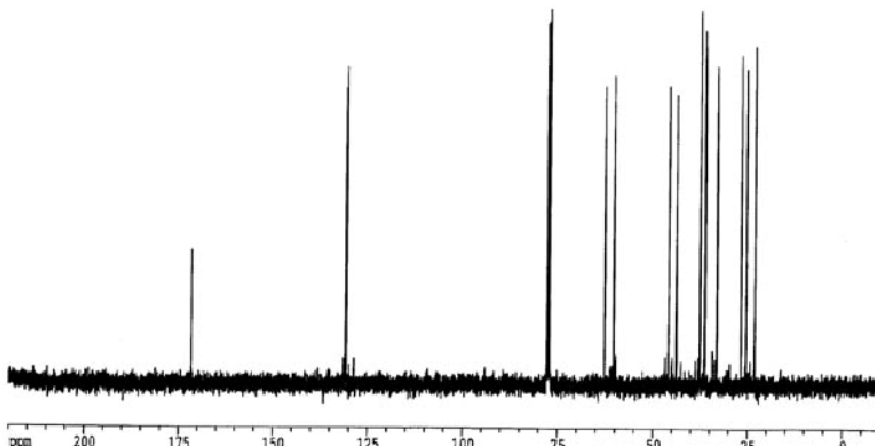
F2 - Acquisition Parameters
Date_ 20011225
Time 17.11
INSTRUM drw400
PROBHD 5 mm Multinu
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.273 Hz
FIDRES 0.144157 Hz
AQ 3.4210291 sec
RG 181
DM 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -6.00 dB
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300074 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
FIF 10.000 ppm
F1 4001.30 Hz
F2F -1.000 ppm
F2 -400.13 Hz
PVMCN 0.55000 ppm/cm
HZCM 220.07150 Hz/cm

```



```

Current Data Parameters
NAME: 4C-VI-123a
EXPNO: 2
PROCNO: 1

F2 - Acquisition Parameters
Date_ 20010827
Time 14.50
INSTRUM drw400
PROBHD 5 mm Multinu
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 193
DS 2
SWH 23248.348 Hz
FIDRES 0.353213 Hz
AQ 1.4154276 sec
RG 4236
DM 21.600 usec
DE 4.50 usec
TE 300.0 K
D1 0.05000000 sec
d11 0.03000000 sec
d12 0.00000000 sec

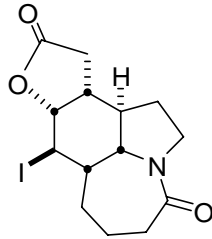
***** CHANNEL f1 *****
NUC1 13C
P1 12.30 usec
PL1 2.00 dB
SFO1 100.6212913 MHz

***** CHANNEL f2 *****
CHRG2 waltz16
NUC2 1H
PCPG2 100.00 usec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1316003 MHz

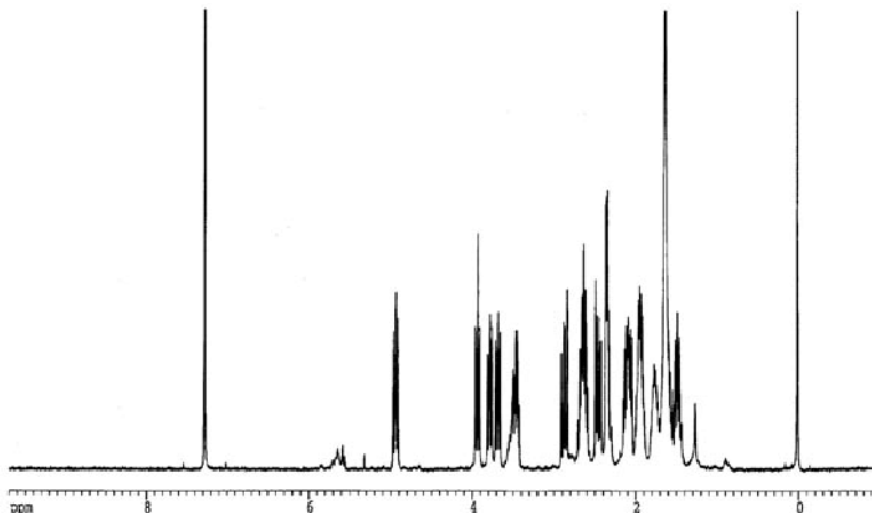
F2 - Processing parameters
SI 32768
SF 100.6127950 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
FIF 220.000 ppm
F1 22134.00 Hz
F2F -10.000 ppm
F2 -1006.23 Hz
PVMCN 11.40000 ppm/cm

```



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```

Current Data Parameters
NAME 4C-VII-67frac8
EXPNO 1
PROCNO 1

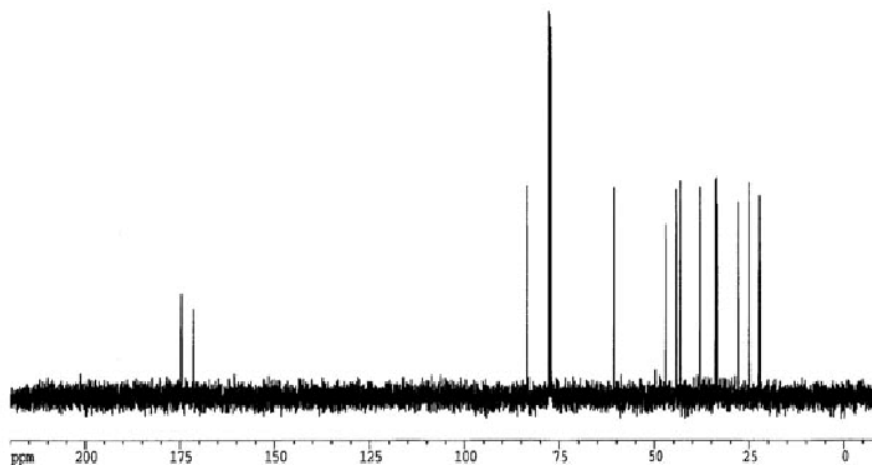
F2 - Acquisition Parameters
Date_ 20011204
Time 8.59
INSTRUM drw400
PROBHD 5 mm Multinu
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SFO 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 203.2
DM 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -6.00 dB
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 14384
SF 400.1320000 MHz
WDM EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

ID NMR plot parameters
CX 20.00 cm
F1P 9.700 ppm
F1 3881.26 Hz
F2P -1.000 ppm
F2 -400.11 Hz
FREQM 0.53590 ppm/cm
HZCM 214.06955 Hz/cm

```



```

Current Data Parameters
NAME 4C-VII-59-bl
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20011226
Time 11.41
INSTRUM drw400
PROBHD 5 mm Multinu
PULPROG spp30
TD 65536
SOLVENT CDCl3
NS 94
DS 2
SFO 23148.148 Hz
FIDRES 0.353213 Hz
AQ 1.4158276 sec
RG 392
DM 21.400 usec
DE 4.50 usec
TE 300.0 K
D1 0.89000000 sec
d11 0.03000000 sec
d12 0.00020000 sec

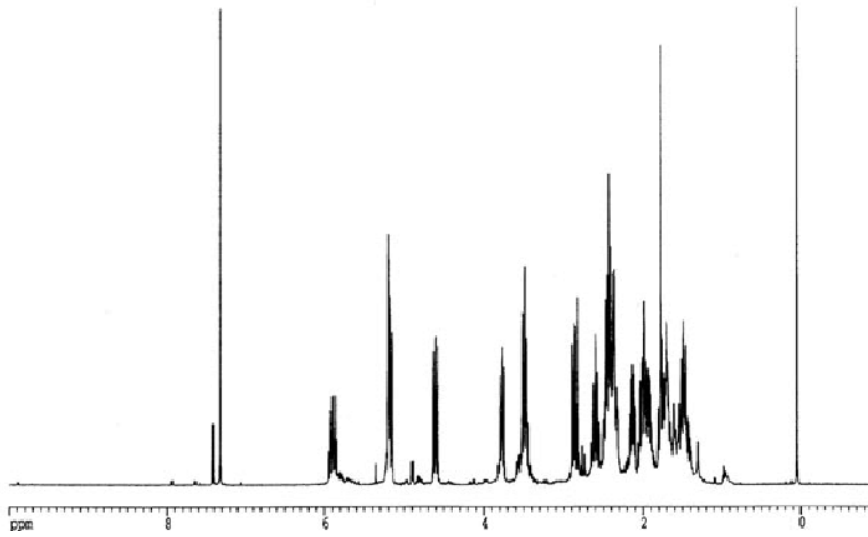
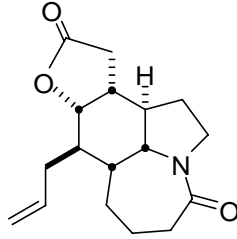
***** CHANNEL f1 *****
NUC1 13C
P1 12.30 usec
PL1 2.00 dB
SFO1 100.6232933 MHz

***** CHANNEL f2 *****
CHRG2 waltz16
NUC2 1H
PCPG2 100.00 usec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1318805 MHz

F2 - Processing parameters
SI 32768
SF 100.6172290 MHz
WDM EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

ID NMR plot parameters
CX 20.00 cm
F1P 220.000 ppm
F1 2214.00 Hz
F2P -10.000 ppm
F2 -1006.13 Hz
FREQM 11.50000 ppm/cm
HZCM 1157.04830 Hz/cm

```



```

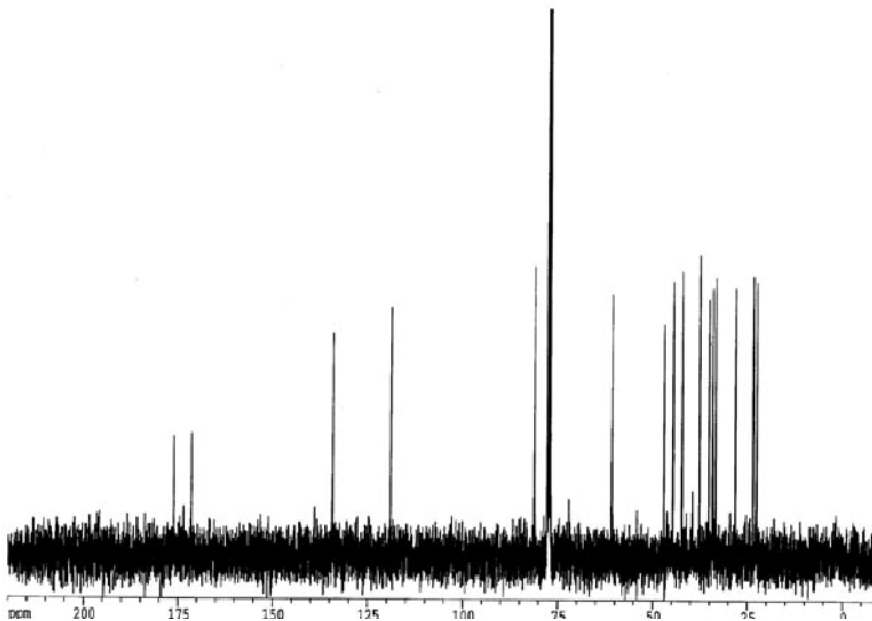
Current Data Parameters
NAME      4C-VII-68
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20011205
Time      17.32
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zg30
RG         32768
SOLVENT   CDCl3
NS         16
DS         2
SWH        4789.272 Hz
FIDRES     0.146157 Hz
AQ         3.4210291 sec
RG         143.7
DM         104.400 usec
DE         4.50 usec
TE         300.0 K
D1         1.00000000 sec

***** CHANNEL f1 *****
NUC1       1H
P1         7.70 usec
PL1        -6.00 dB
SFO1       400.1320007 MHz

F2 - Processing parameters
SI         16384
SF         400.1299854 MHz
WM        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
F1P        10.000 ppm
F1         4001.30 Hz
F2P        -1.000 ppm
F2         -400.13 Hz
FREQCH    0.55000 ppm/cm
HSCX      220.07149 Hz/cm
  
```



```

Current Data Parameters
NAME      4C-VII-68
EXPNO     2
PROCNO    1

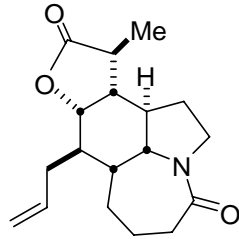
F2 - Acquisition Parameters
Date_     20011205
Time      17.37
INSTRUM   drx400
PROBHD    5 mm Multinu
PULPROG   zgpg30
RG         65536
SOLVENT   CDCl3
NS         358
DS         2
SWH        23168.148 Hz
FIDRES     1.353213 Hz
AQ         1.4154276 sec
RG         8192
DM         21.500 usec
DE         4.50 usec
TE         300.0 K
D1         0.05000000 sec
d11        0.03000000 sec
d12        0.00020000 sec

***** CHANNEL f1 *****
NUC1       13C
P1         12.30 usec
PL1         2.00 dB
SFO1       100.6212933 MHz

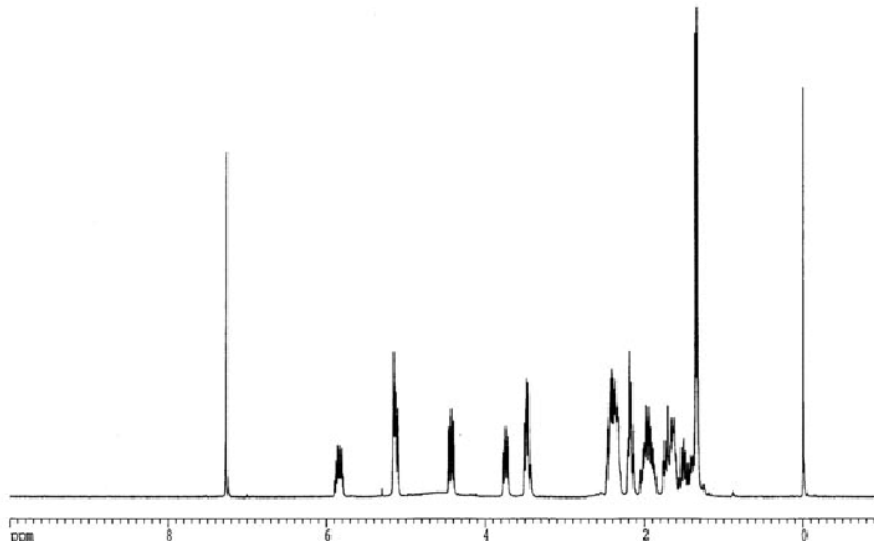
***** CHANNEL f2 *****
CPDPRG2   waltz16
NUC2       1H
PCPD2     100.00 usec
PL2        0.00 dB
PL12       18.00 dB
PL13       18.00 dB
SFO2       400.1316605 MHz

F2 - Processing parameters
SI         13748
SF         100.6127250 MHz
WM        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
F1P        220.000 ppm
F1         22136.80 Hz
F2P        -10.000 ppm
F2         -1006.13 Hz
FREQCH    11.50000 ppm/cm
  
```



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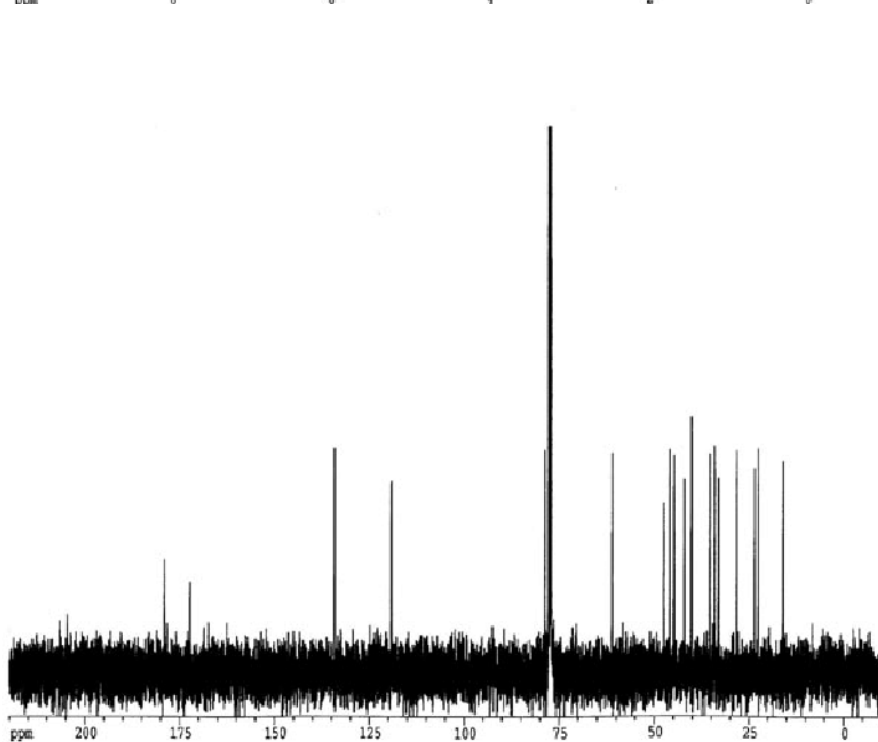
```
Current Data Parameters
NAME 4c-VII-168-new
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 2002022
Time 11.25
INSTRUM dmw400
PROBHD 5 mm Multinu
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 200.2
EW 104.400 usec
DE 4.50 usec
TE 300.0 K
SI 1.00000000 sec
```

```
***** CHANNEL f1 *****
NUC1 1H
P1 7.70 usec
PL1 -6.00 dB
SFO1 400.1320007 MHz
```

```
F2 - Processing parameters
SI 16384
SF 400.1300082 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
```

```
1D NMR plot parameters
CX 20.00 cm
F1P 10.000 ppm
F1 4001.30 Hz
F2P -1.000 ppm
F2 -409.13 Hz
PVMCK 0.55000 ppm/cm
RECK 220.07150 Hz/cm
```



```
Current Data Parameters
NAME 4c-VII-168-new
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 2002022
Time 14.20
INSTRUM dmw400
PROBHD 5 mm Multinu
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 392
DS 2
SWH 23168.148 Hz
FIDRES 0.353213 Hz
AQ 1.4152776 sec
RG 6006
EW 21.600 usec
DE 4.50 usec
TE 300.0 K
SI 0.85000000 sec
dS1 0.83000000 sec
dS2 0.80000000 sec
```

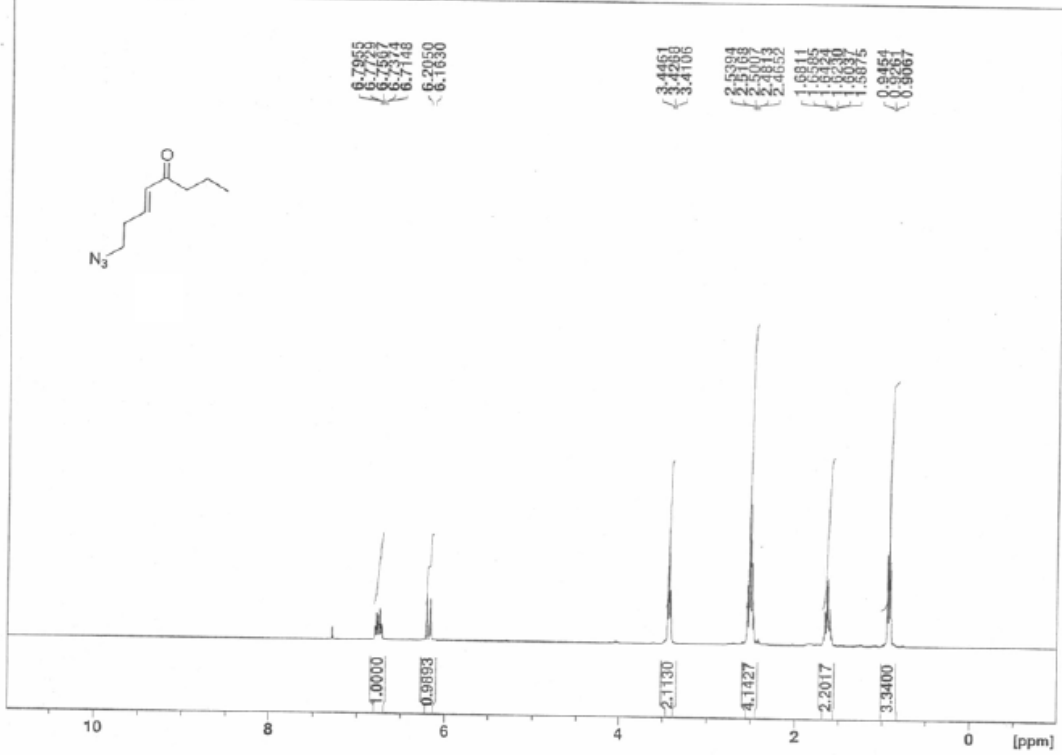
```
***** CHANNEL f1 *****
NUC1 13C
P1 12.10 usec
PL1 2.00 dB
SFO1 100.6212933 MHz
```

```
***** CHANNEL f2 *****
CROSSP2 waltz16
NUC2 1H
PCPO2 100.00 usec
PL2 0.00 dB
PL12 18.00 dB
PL13 18.00 dB
SFO2 400.1316045 MHz
```

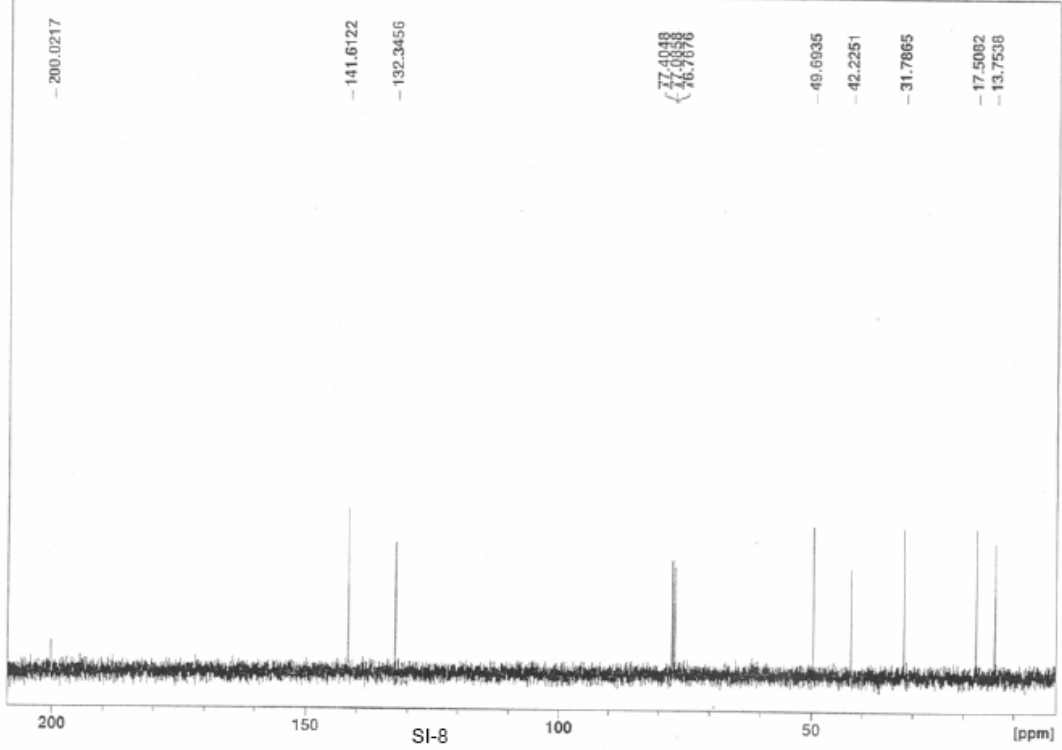
```
F2 - Processing parameters
SI 12768
SF 100.617290 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40
```

```
1D NMR plot parameters
CX 20.00 cm
F1P 222.600 ppm
F1 22214.40 Hz
F2P -10.000 ppm
F2 -1006.13 Hz
PVMCK 11.10000 ppm/cm
RECK 1143.04610 Hz/cm
```

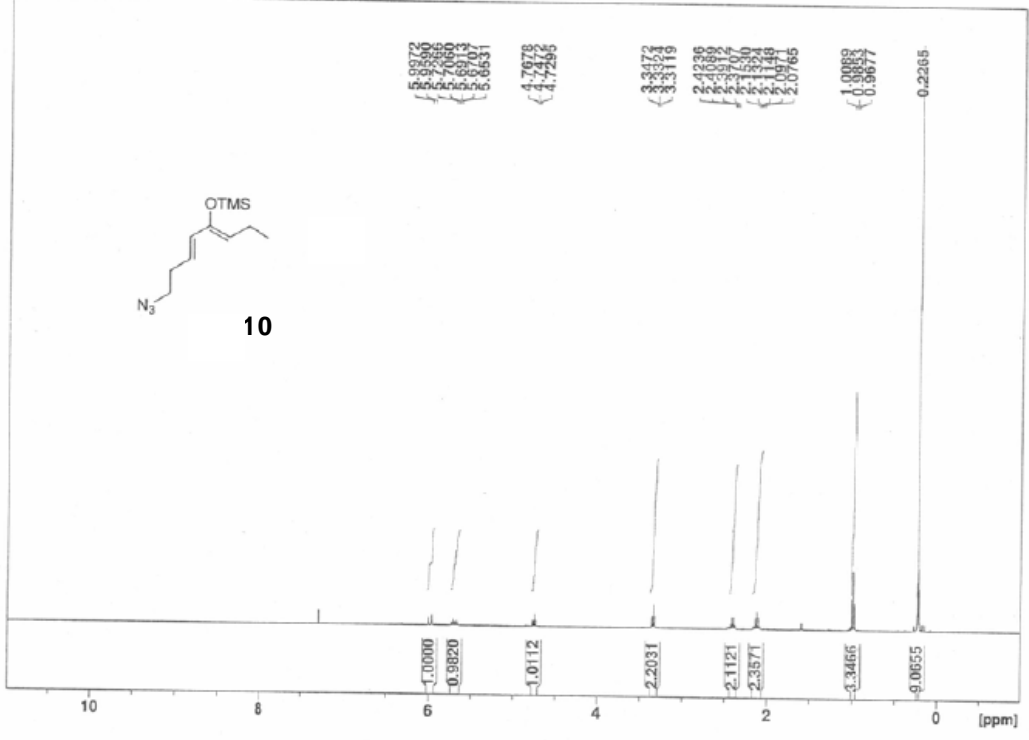
zyajiv-287 1 1 /opt/topspin yzeng



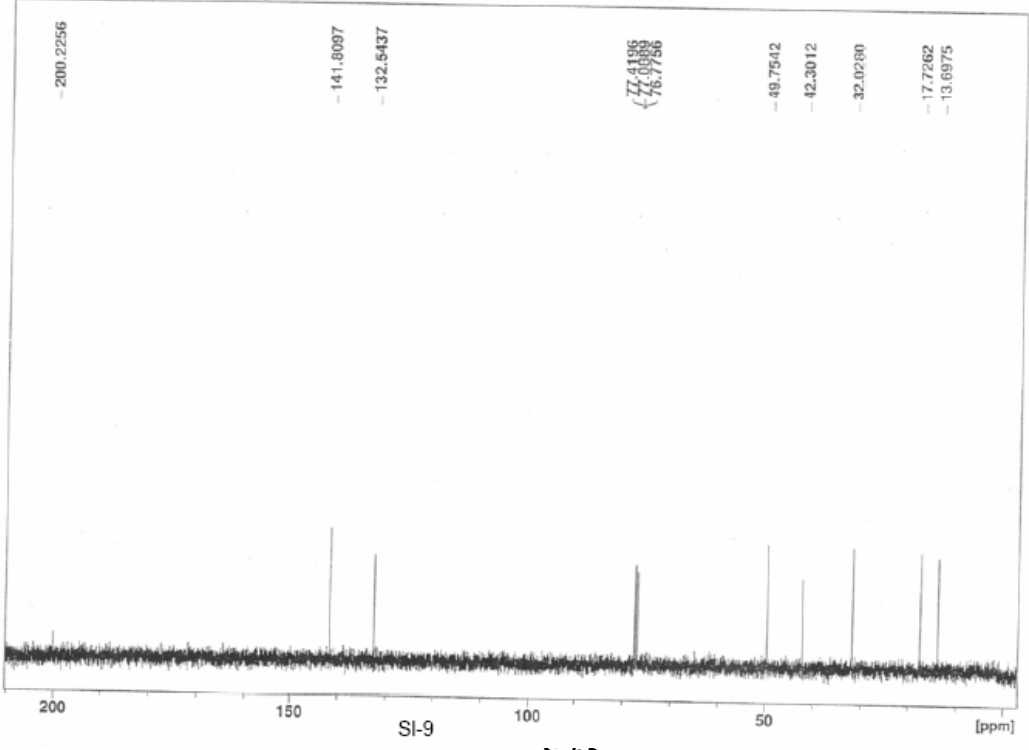
zyajiv-287 2 1 /opt/topspin yzeng

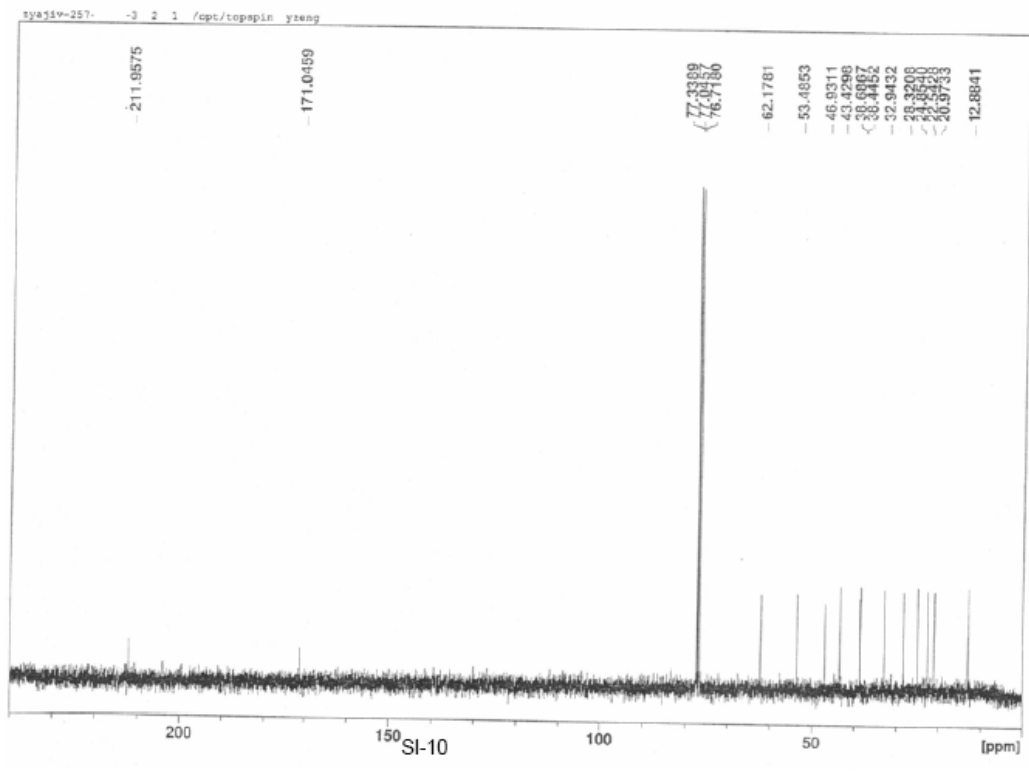
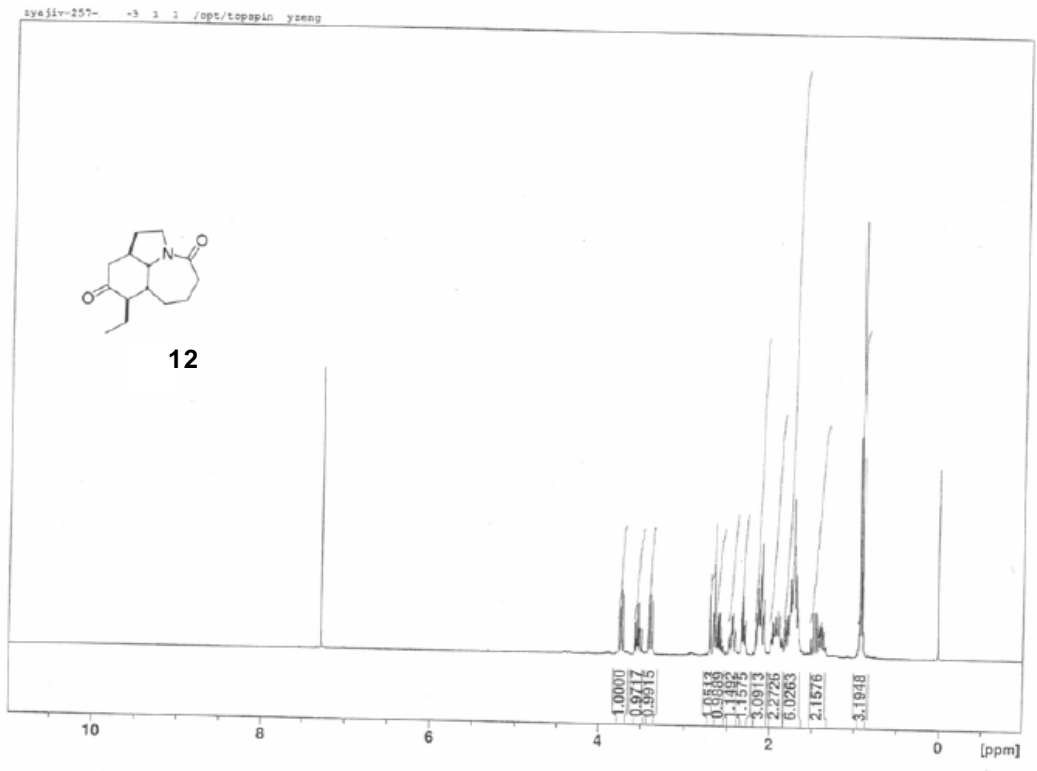


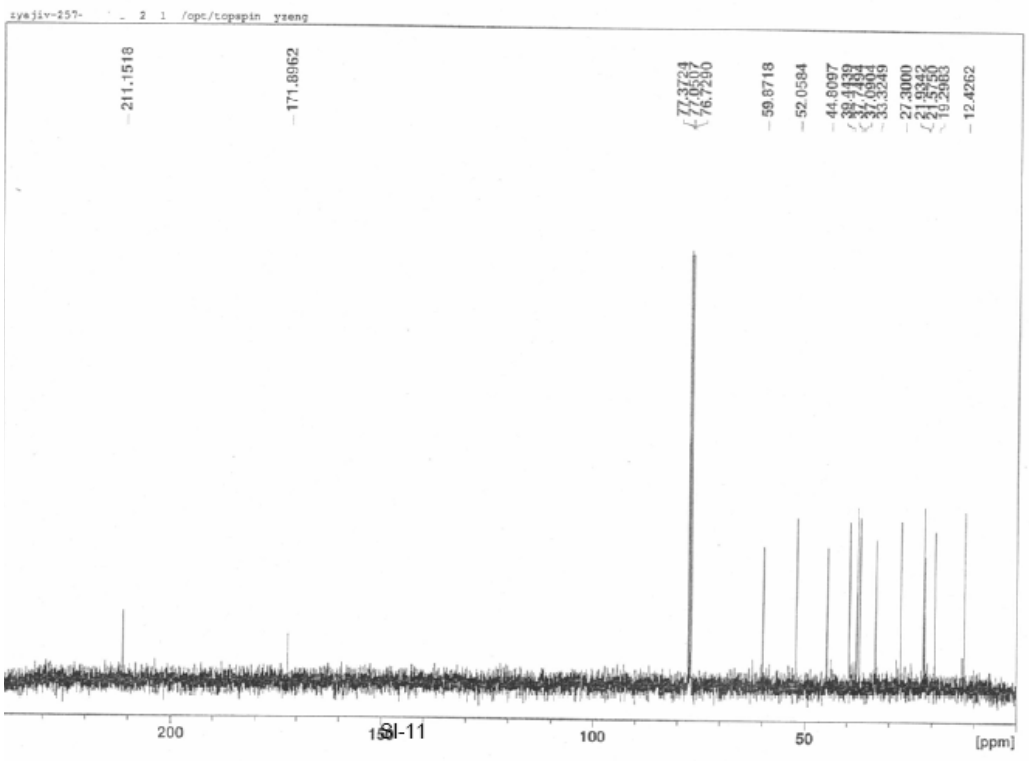
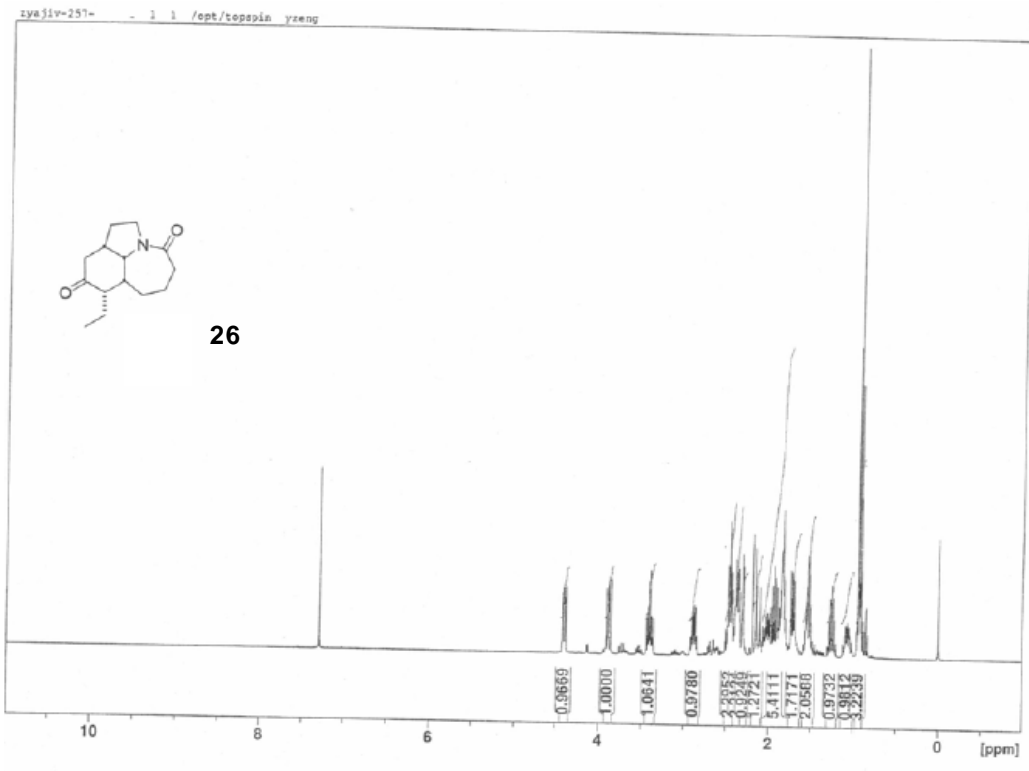
syajiv-207-2 1 1 /opt/topspin yzeng

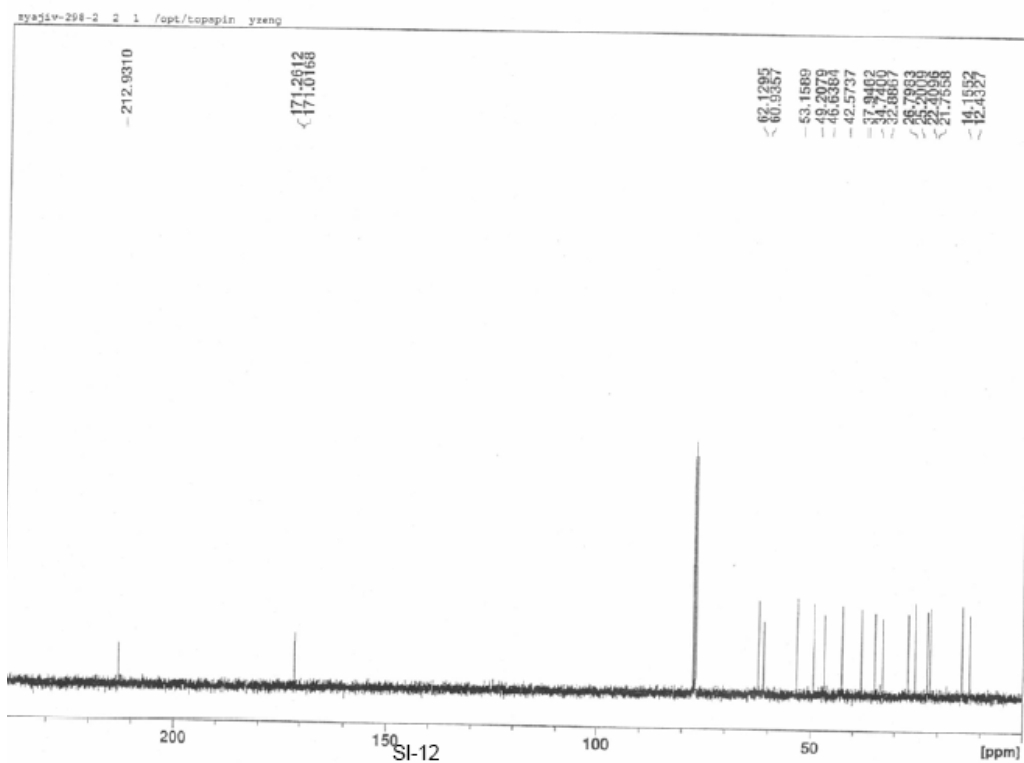
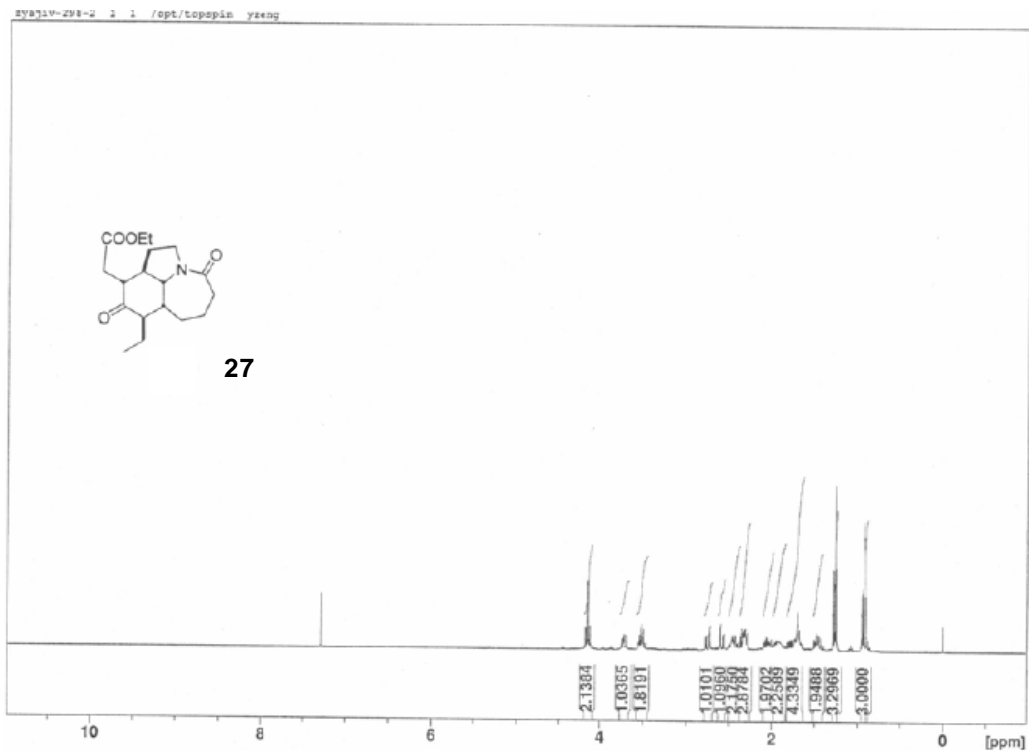


syajiv-207 2 1 /opt/topspin yzeng

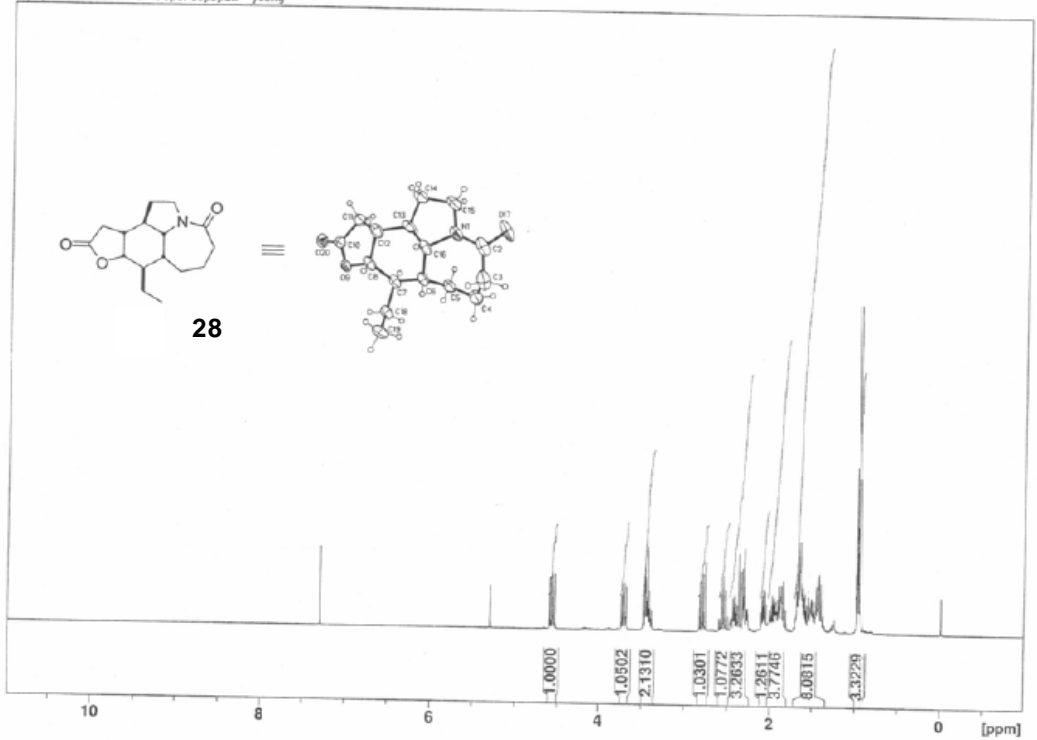




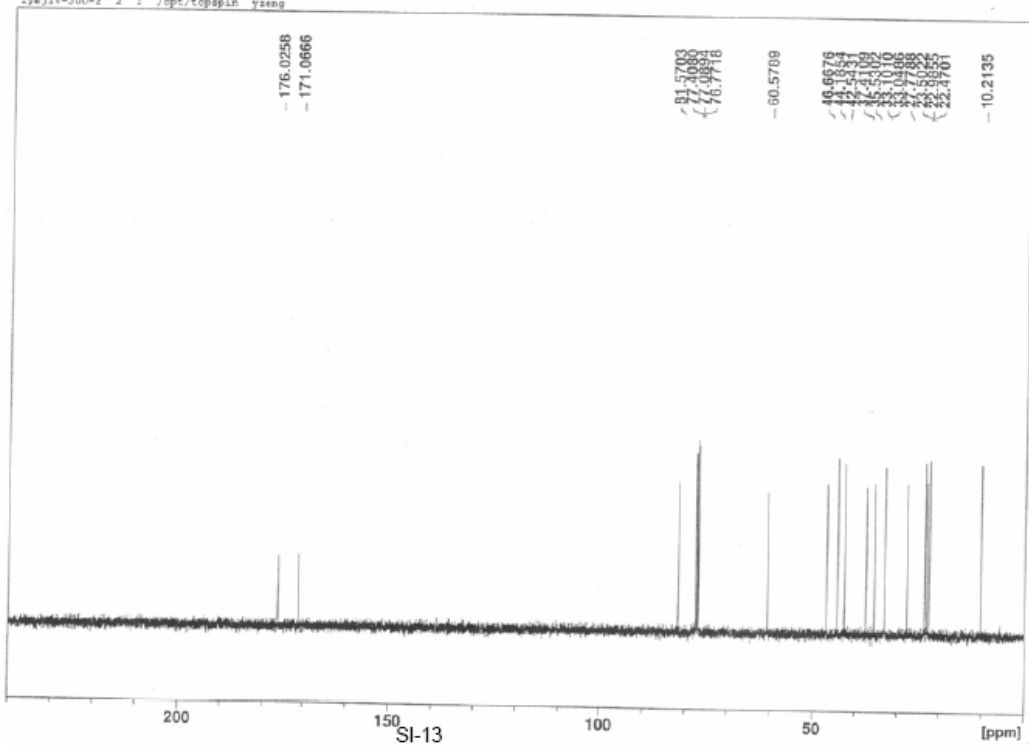




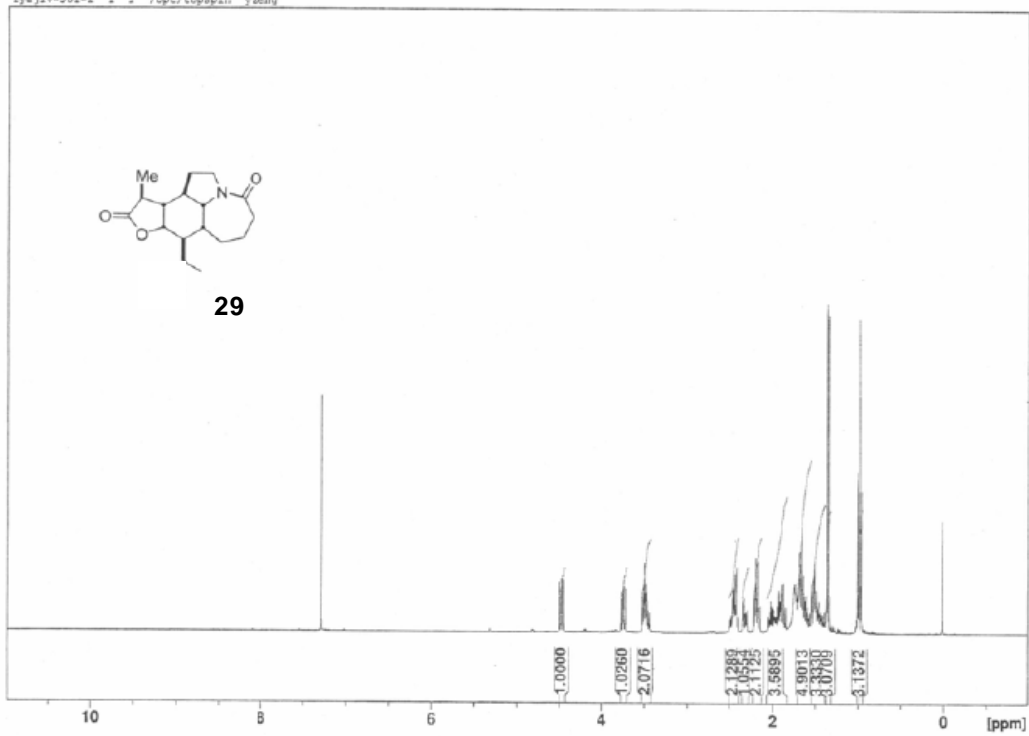
SYN14V-300-2 1 1 /opt/topspin yzeng



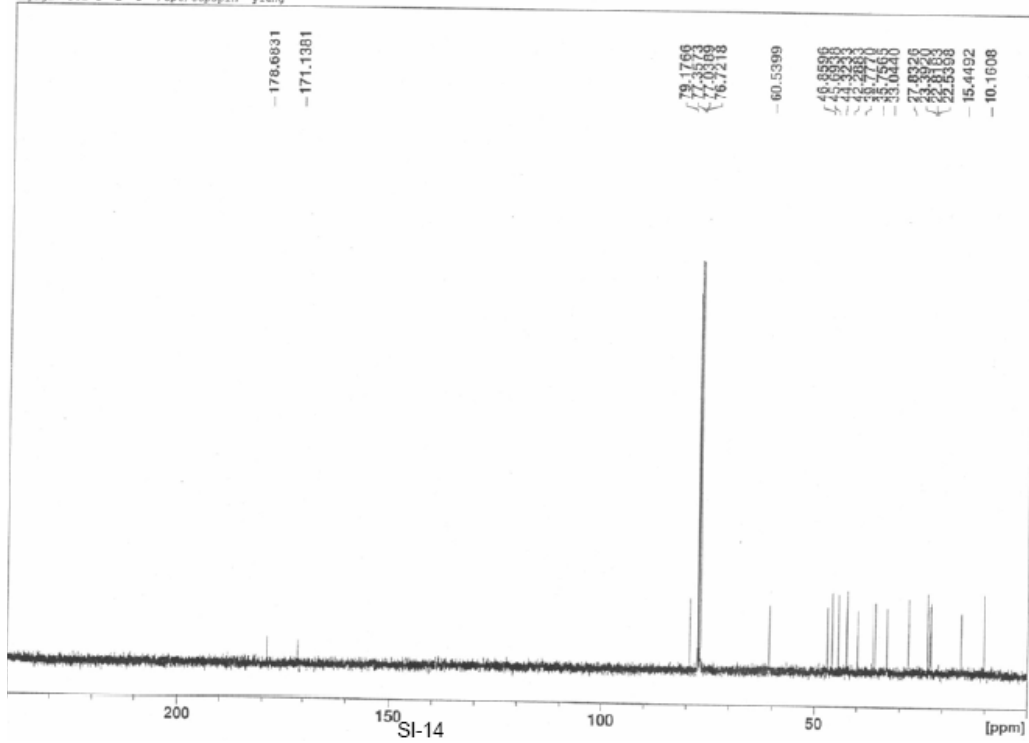
SYN14V-300-2 2 1 /opt/topspin yzeng



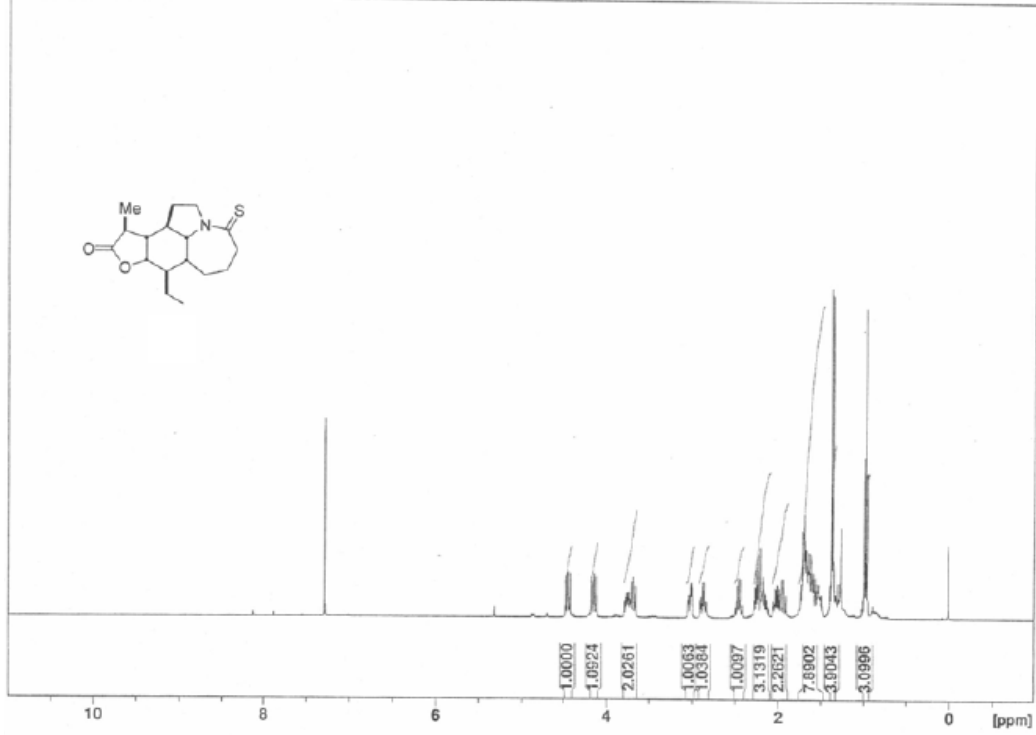
sva31v-202-2 1 1 /opt/topopin yzeng



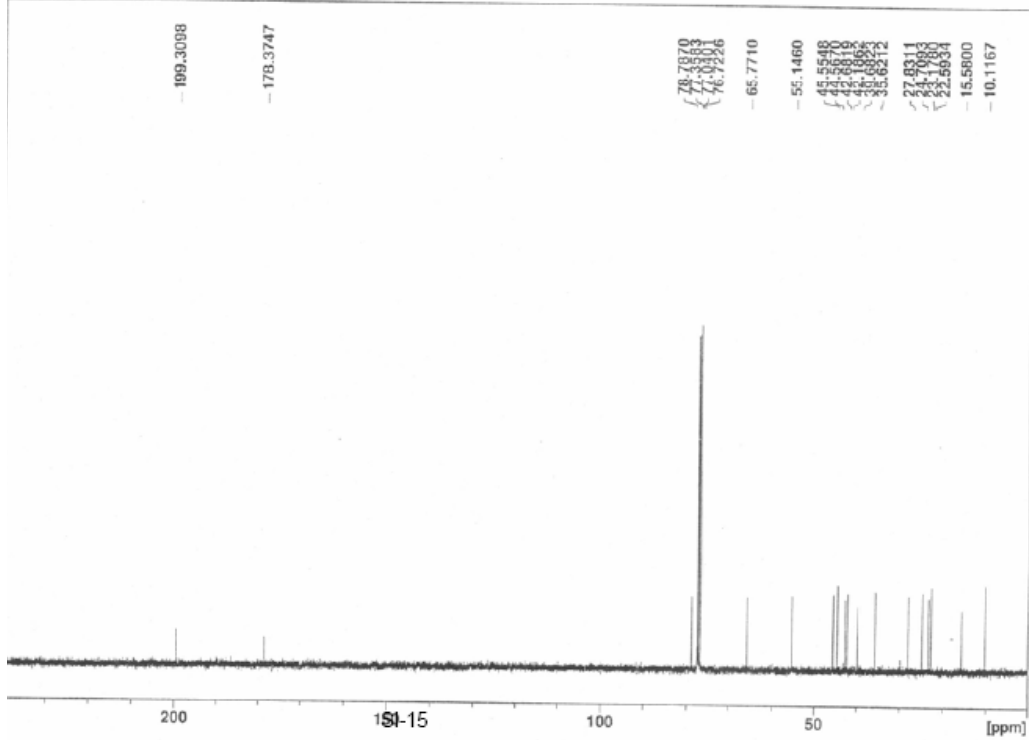
sva31v-202-2 2 1 /opt/topopin yzeng

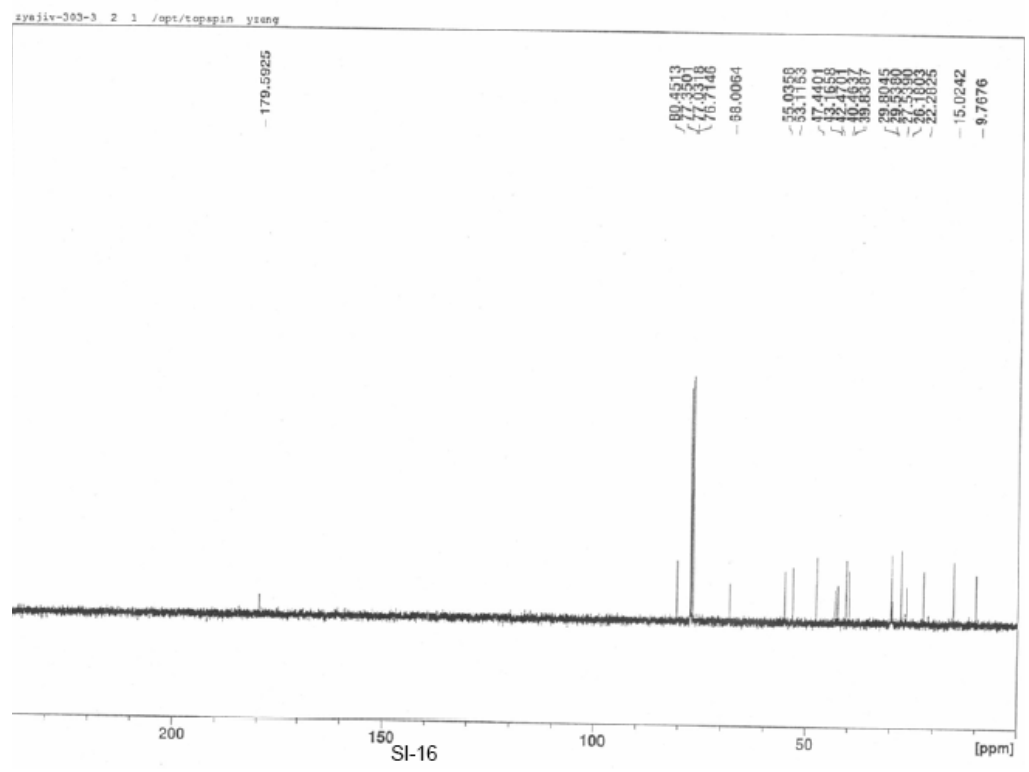
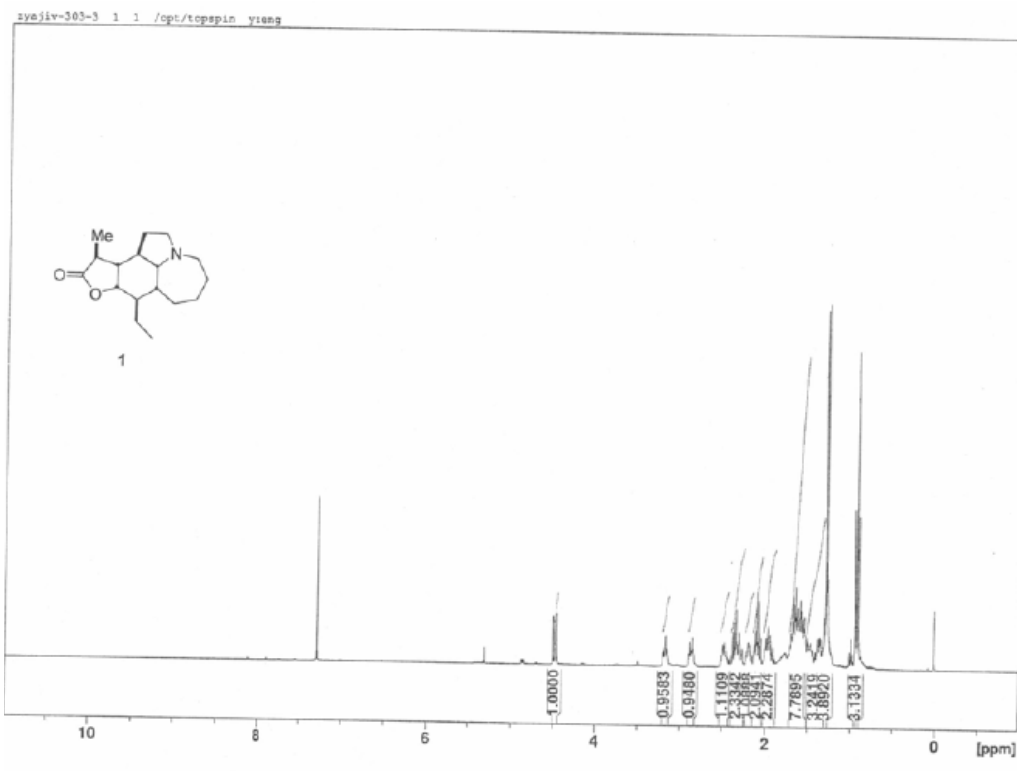


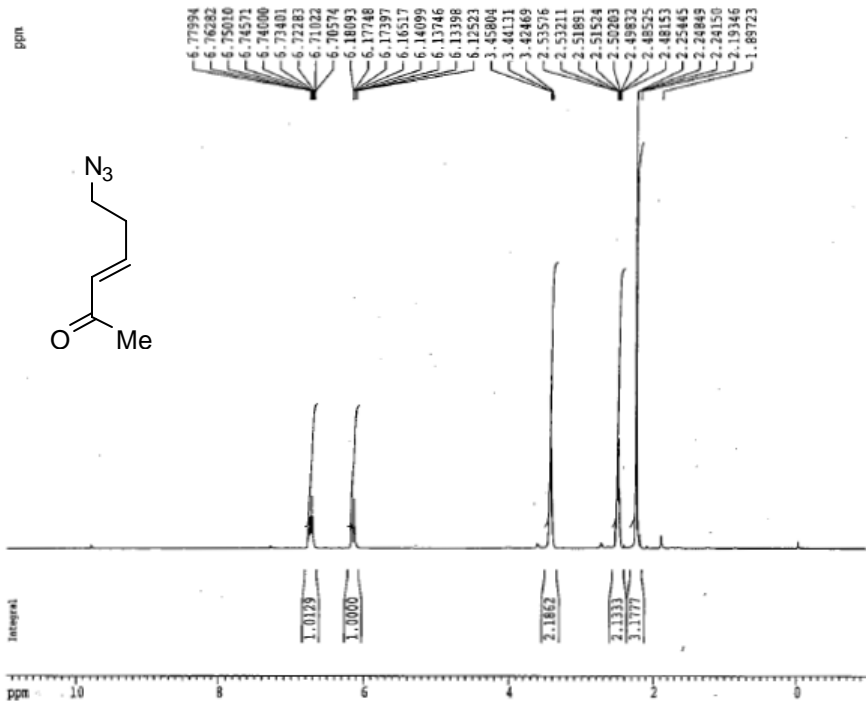
zyajiv-304-1 1 1 /opt/topapin yzeng



zyajiv-304-1 2 1 /opt/topapin yzeng





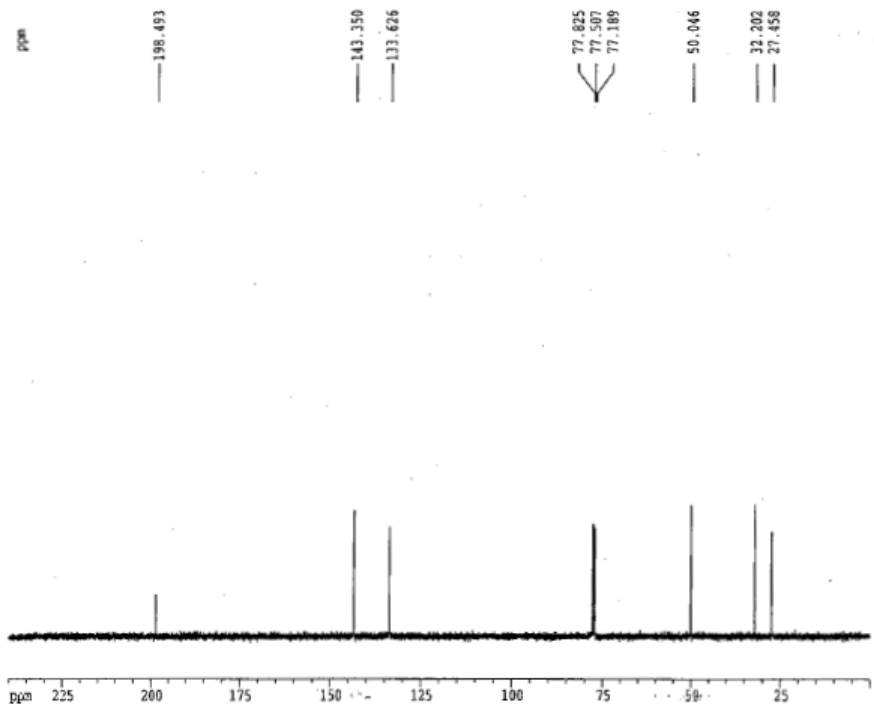


F2 - Acquisition Parameters
 Date_ 20040510
 Time 18.35
 INSTRUM drx400
 PROBRD 5 mm Multinucl
 PULPROG zgpg30
 TO 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SSB 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4210291 sec
 EC 64
 DM 104.400 usec
 DE 5.50 usec
 TE 679.6 K
 D1 1.00000000 sec
 MCREST 0.00000000 sec
 NCHRS 0.01500000 sec

***** CHANNEL f1 *****
 NUCL1 1H
 P1 7.70 usec
 PL1 -6.90 dB
 SFO1 400.1370007 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 12.50 cm
 F1 4325.20 Hz
 F2P -0.985 ppm
 F2 -393.99 Hz
 FWHM 0.59846 ppm/cm
 HZCM 239.46362 Hz/cm



Current Data Parameters
 NAME 11-792
 EXNO 2
 FWHM 1

F2 - Acquisition Parameters
 Date_ 20040510
 Time 18.38
 INSTRUM drx400
 PROBRD 5 mm Multinucl
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl3
 NS 95
 DS 4
 SSB 24158.560 Hz
 FIDRES 0.368570 Hz
 AQ 1.3566452 sec
 EC 32768
 DM 20.700 usec
 DE 5.50 usec
 TE 679.6 K
 D1 0.15000000 sec
 D11 0.32000000 sec
 DELTA 0.05000000 sec
 MCREST 0.00000000 sec
 NCHRS 0.01500000 sec

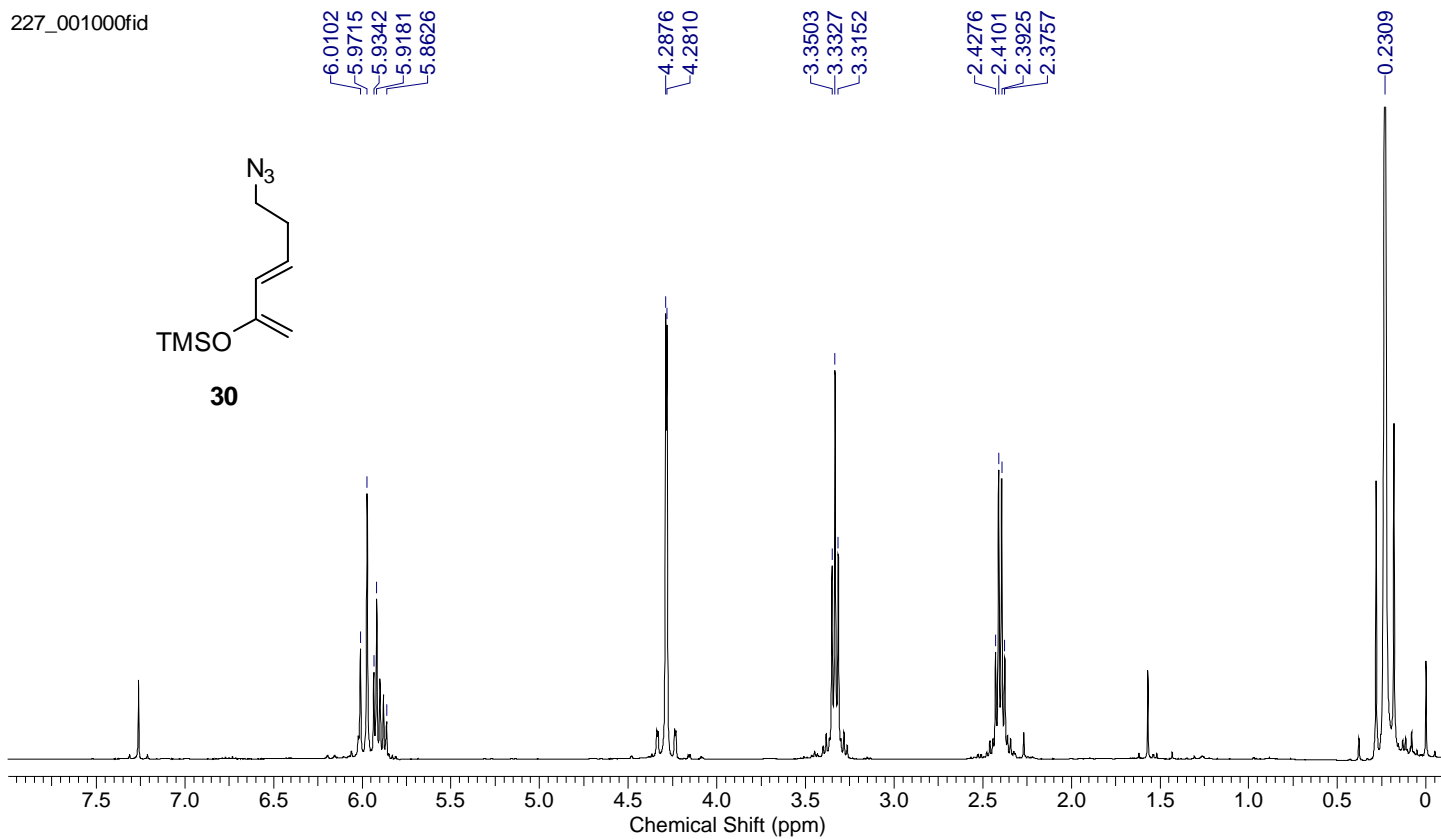
***** CHANNEL f1 *****
 NUCL1 13C
 P1 11.30 usec
 PL1 2.00 dB
 SFO1 100.6248025 MHz

***** CHANNEL f2 *****
 CH2PRG2 waltz16
 NUCL2 1H
 PCPD2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316005 MHz

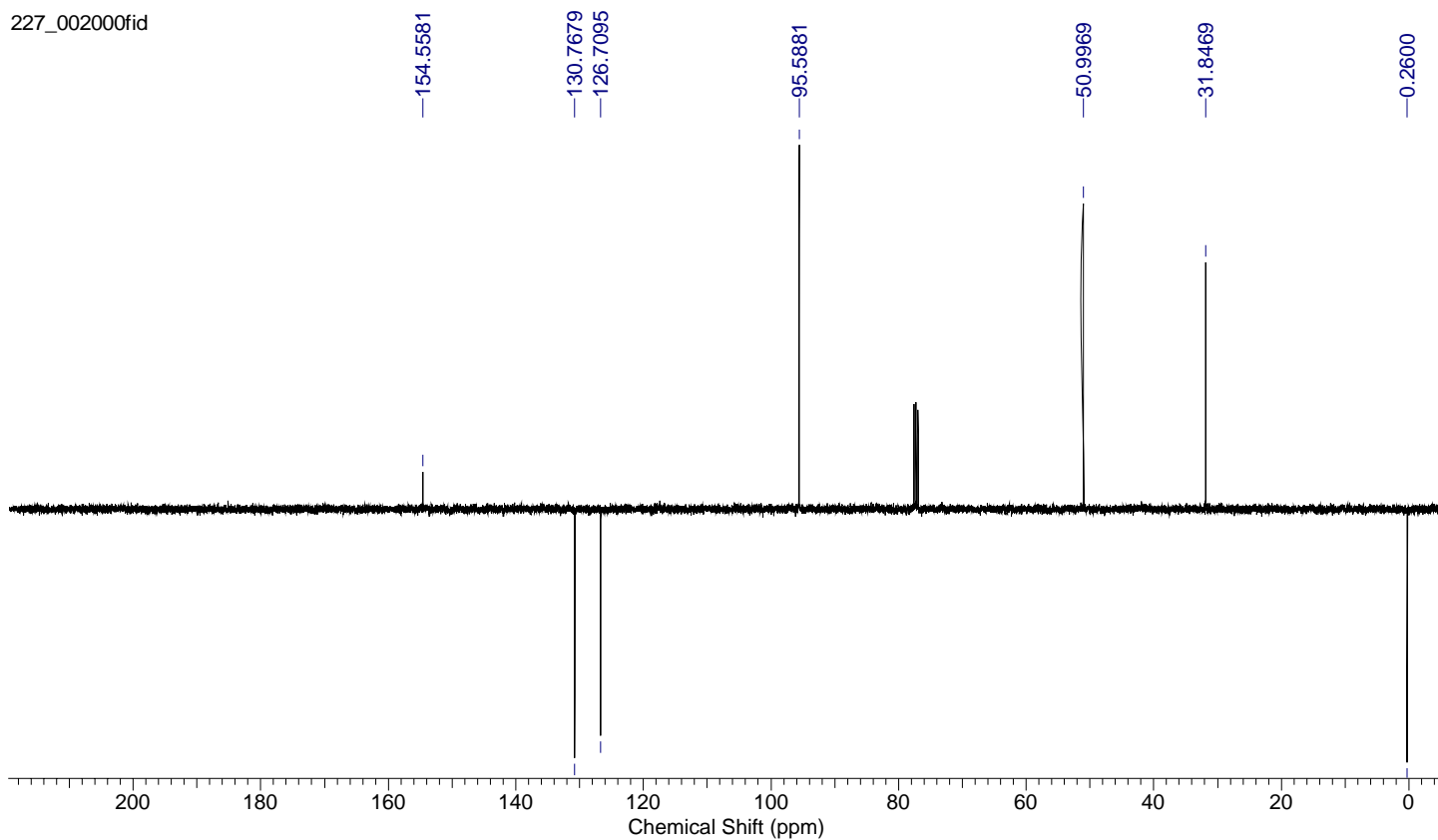
F2 - Processing parameters
 SI 65536
 SF 100.6127210 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.00

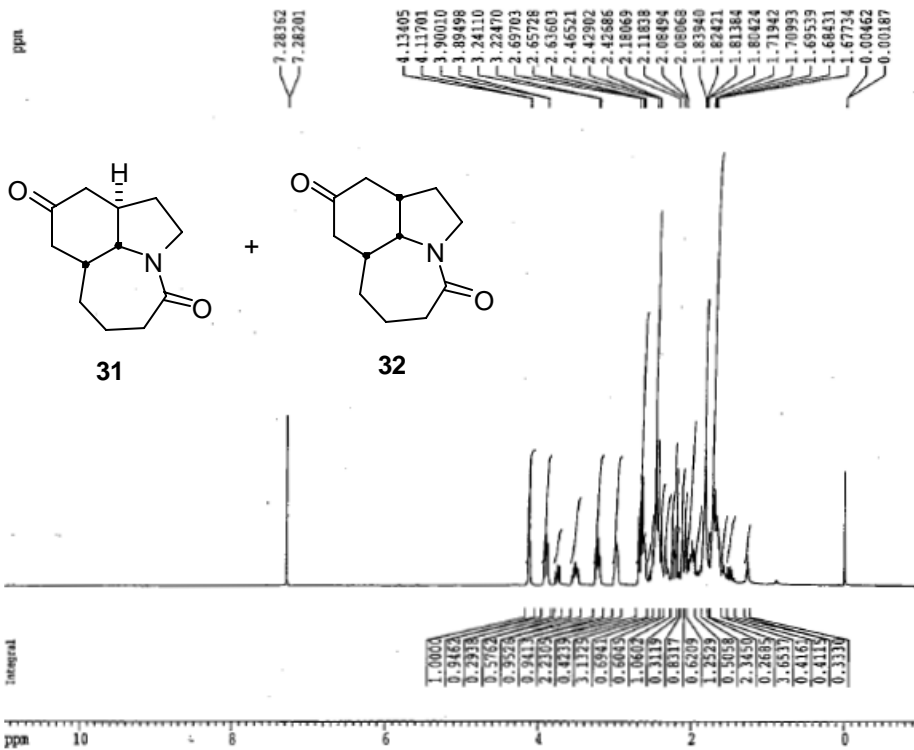
1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 F1 240.037 ppm
 F2 24159.82 Hz
 F2P -0.037 ppm
 F2 -3.77 Hz
 FWHM 12.00774 ppm/cm
 HZCM 1267.2900 Hz/cm

227_001000fid



227_002000fid





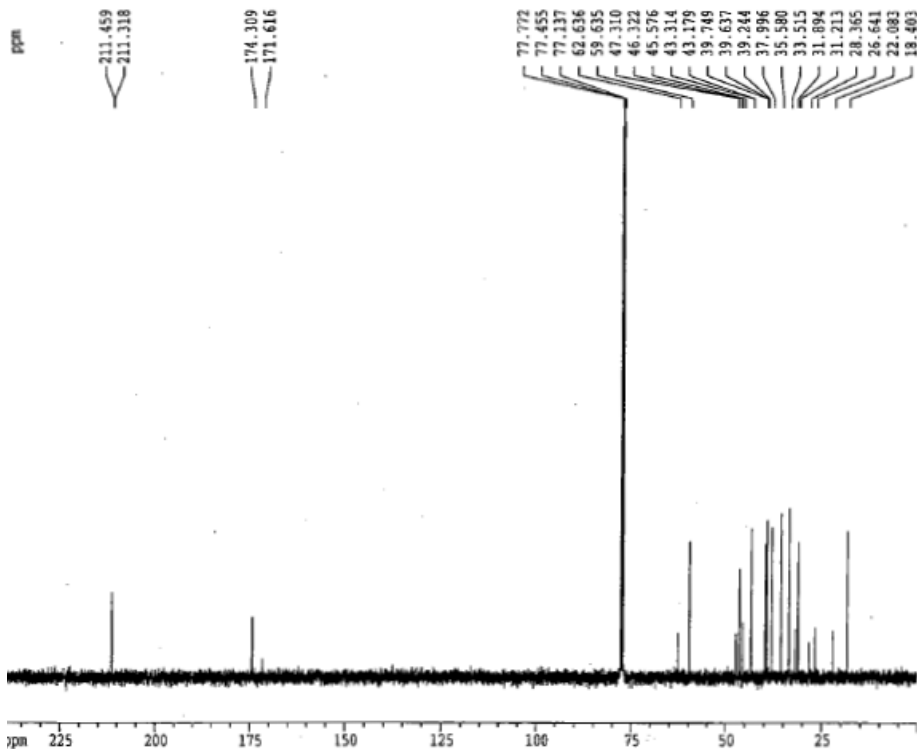
5a 51

F2 - Acquisition Parameters
 Date_ 20040717
 Time 15.45
 INSTRUM drx400
 PROBRD 5 mm Multispec1
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SFO1 4789.272 M
 FIDRES 0.146157 H
 AQ 3.4210291 s
 RG 114
 DM 104.400 u
 DE 5.50 u
 TE 294.2 K
 D1 1.0000000 s
 MCHRSY 0.0000000 s
 MCHRX 0.0150000 s

***** CHANNEL f1 *****
 NUC1 1H
 P1 7.70 u
 PL1 -6.00 dB
 SFO1 400.1320007 M

F2 - Processing parameters
 SI 32768
 SF 400.1300000 M
 NH 16
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 0.00 cm
 FIP 10.985 ug
 FI 4395.28 Hz
 FZP -0.985 ug
 F2 -0.93 Hz
 PPRCM 0.50846 pc
 RECM 239.46382 Hz



Current Data Parameters
 NAME IV-41-4
 EXPNO 2
 PROCNO 1

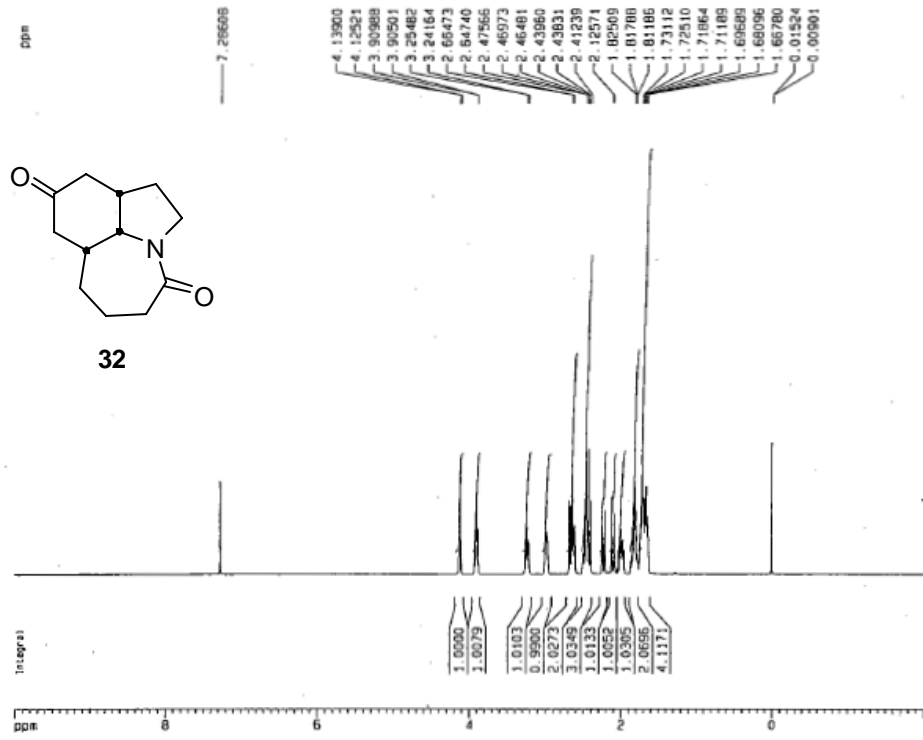
F2 - Acquisition Parameters
 Date_ 20040717
 Time 15.51
 INSTRUM drx400
 PROBRD 5 mm Multispec1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 551
 DS 4
 SFO1 24154.350 Hz
 FIDRES 0.348570 Hz
 AQ 1.3586452 sec
 RG 32768
 DM 20.700 usec
 DE 5.50 usec
 TE 294.2 K
 D1 0.1500000 sec
 dL1 0.0300000 sec
 DELTA 0.0300000 sec
 MCHRSY 0.0000000 sec
 MCHRX 0.0150000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 12.30 usec
 PL1 2.00 dB
 SFO1 100.6248025 MHz

***** CHANNEL f2 *****
 CPROG2 waltz16
 NUC2 1H
 PCPR2 100.00 usec
 PL2 0.00 dB
 PL12 18.00 dB
 PL13 18.00 dB
 SFO2 400.1316025 MHz

F2 - Processing parameters
 SI 65536
 SF 100.6127210 MHz
 NH 512
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.48

1D NMR plot parameters
 CX 20.00 cm
 CY 12.50 cm
 FIP 246.037 ppm
 FI 24150.02 Hz
 FZP -0.037 ppm
 F2 -3.77 Hz
 PPRCM 12.00274 ppm/cm
 RECM 1207.22849 Hz/cm



5a

F2 - Acquisition Parameters
Date: 20040726
Time: 15.16
INSTRUM: spect
PROBHD: 5 mm BBO BB-CP
PULPROG: zgpg30
TD: 32768
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 6000.815 Hz
FIDRES: 0.32209 Hz
AQ: 2.7284209 sec
RG: 392.5
DM: 80.000 usec
DE: 8.00 usec
TE: 297.4 K
D0: 0.0200000 sec
H2OCT1: 0.0200000 sec
H2OCTN: 0.0100000 sec

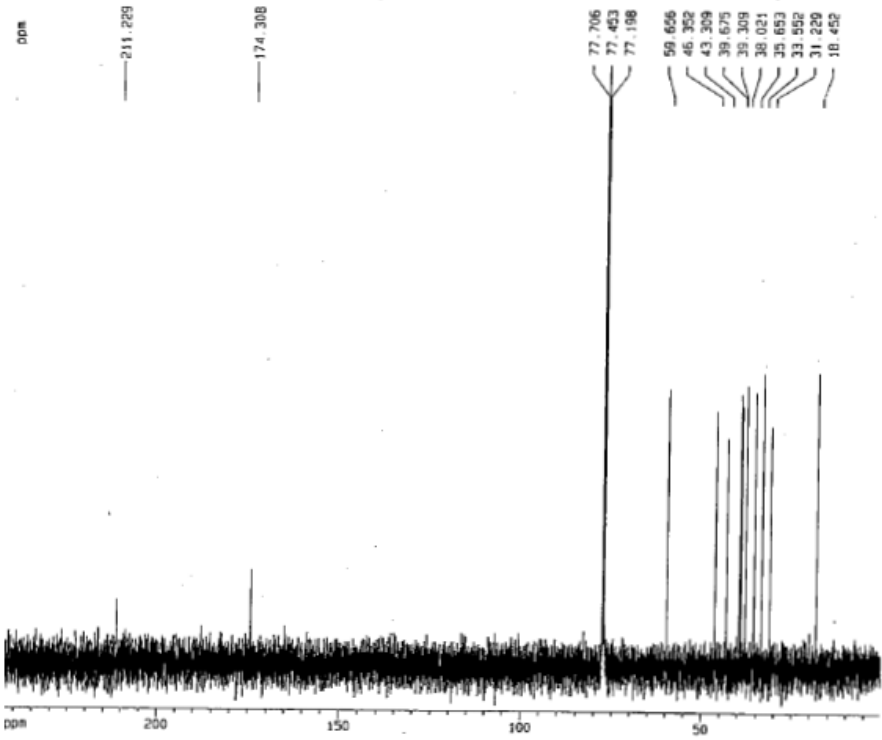
***** CHANNEL f1 *****
NUC1: 1H
P1: 7.70 usec
PL1: -4.00 dB
SFO1: 500.130000 MHz

F1 - Acquisition Parameters
AQ: 2
TD: 256
SFO1: 500.1325 MHz
FIDRES: 23.479380 Hz
SQ: 12.016 dB
FWD0: uncalibrated

F2 - Processing parameters
SI: 32768
SF: 500.130000 MHz
YDM: 64
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

F1 - Processing parameters
SI: 1024
SF: 500.130000 MHz
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

1D NMR list parameters
CL: 20.00 cm
CY: 0.00 cm
F1P: 39.900 MHz
F2: 500.1325 MHz
F2P: -2.000 MHz
F2J: -1004.20 Hz
PFGM2: 0.000000 cps/cm
NUC2: 13C-400017 MHz



Current Data Parameters
Date: 19-05-05
Time: 2
INSTRUM: spect
PROBHD: 5 mm BBO BB-CP
PULPROG: zgpg30
TD: 32768
SOLVENT: CDCl3
NS: 16
DS: 2
SWH: 21465.561 Hz
FIDRES: 0.478838 Hz
AQ: 1.842988 sec
RG: 256
DM: 15.980 usec
DE: 8.00 usec
TE: 297.0 K
D0: 0.0200000 sec
H2OCT1: 0.0200000 sec
H2OCTN: 0.0100000 sec

***** CHANNEL f1 *****
NUC1: 13C
P1: 8.50 usec
PL1: 0.00 dB
SFO1: 125.770000 MHz

***** CHANNEL f2 *****
CHPROG: waltz16
NUC2: 1H
P2P2: 05.00 usec
PL12: -4.00 dB
PL13: 19.00 dB
PL14: 38.00 dB
SFO2: 500.130000 MHz

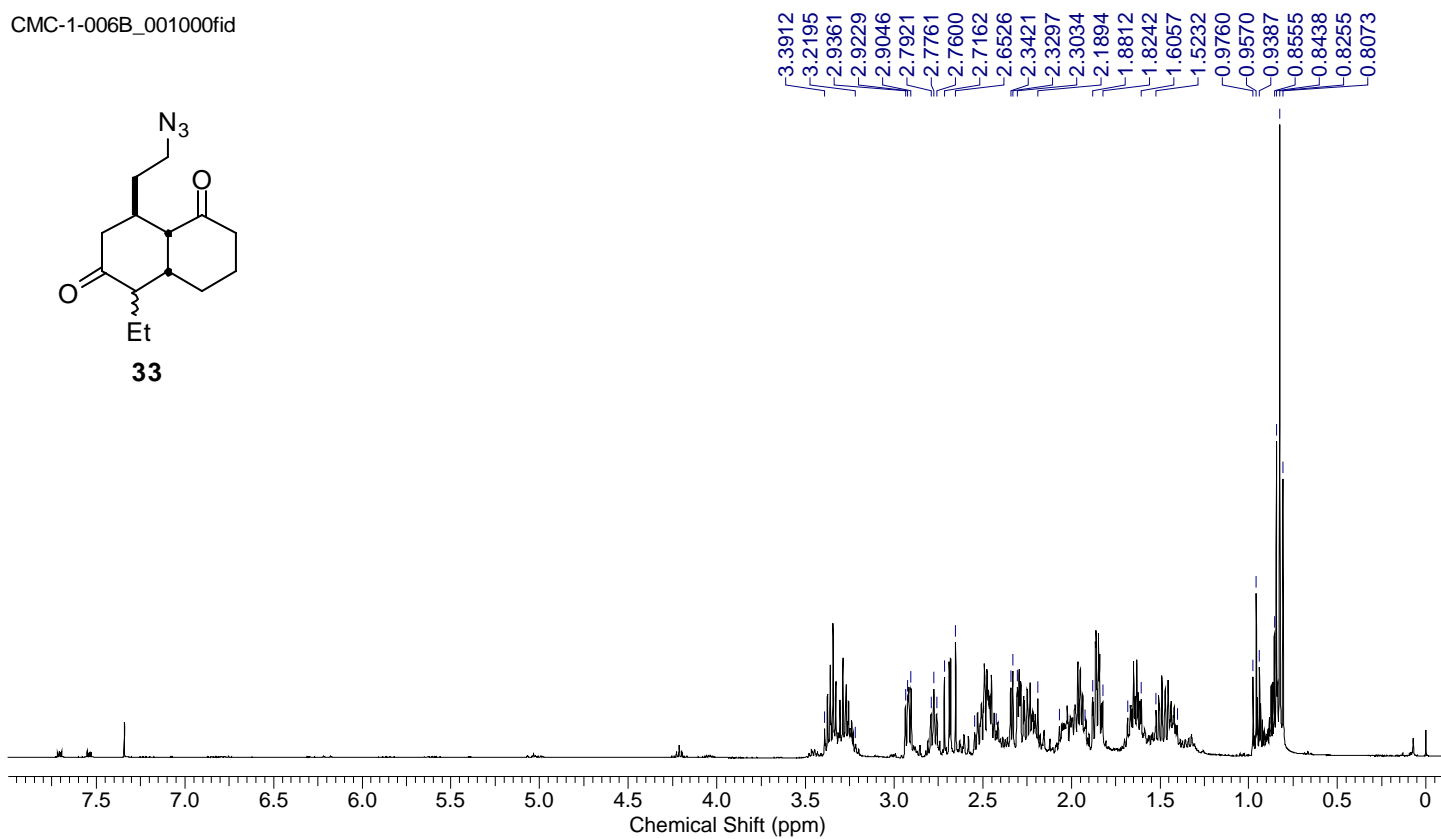
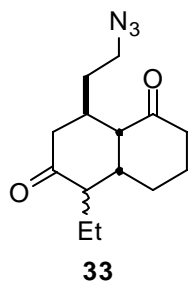
F1 - Acquisition parameters
AQ: 2
TD: 256
SFO1: 500.1325 MHz
FIDRES: 23.479380 Hz
SQ: 12.016 dB
FWD0: uncalibrated

F2 - Processing parameters
SI: 32768
SF: 125.770000 MHz
YDM: 64
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

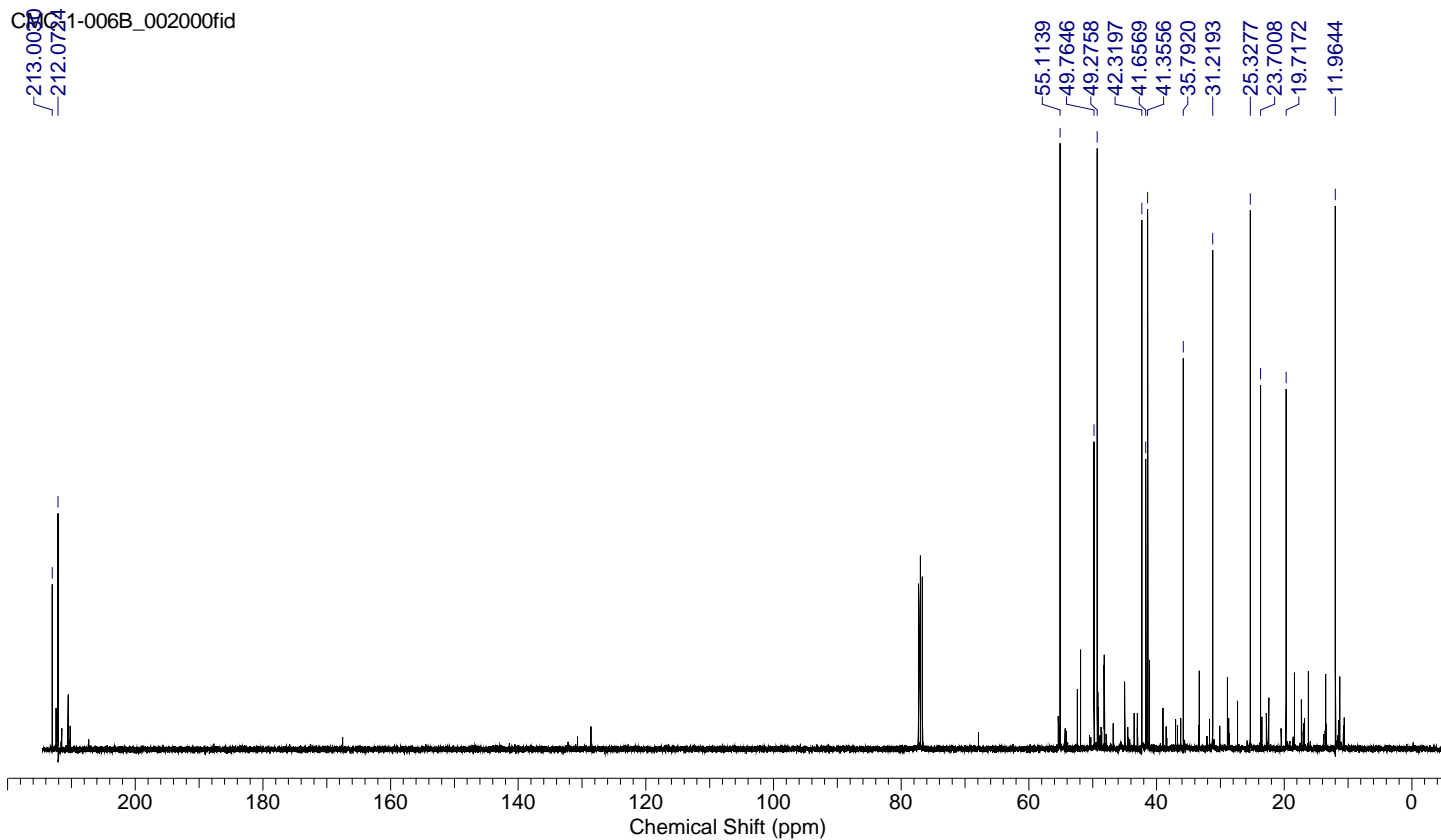
F1 - Processing parameters
SI: 1024
SF: 500.130000 MHz
WDW: EM
SSB: 0
LB: 0.30 Hz
GB: 0
PC: 1.00

1D NMR list parameters
CL: 20.00 cm
CY: 0.00 cm
F1P: 200.500 MHz
F2: 125.770000 MHz
F2J: -1004.20 Hz
PFGM2: 0.000000 cps/cm
NUC2: 13C-400017 MHz

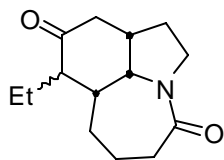
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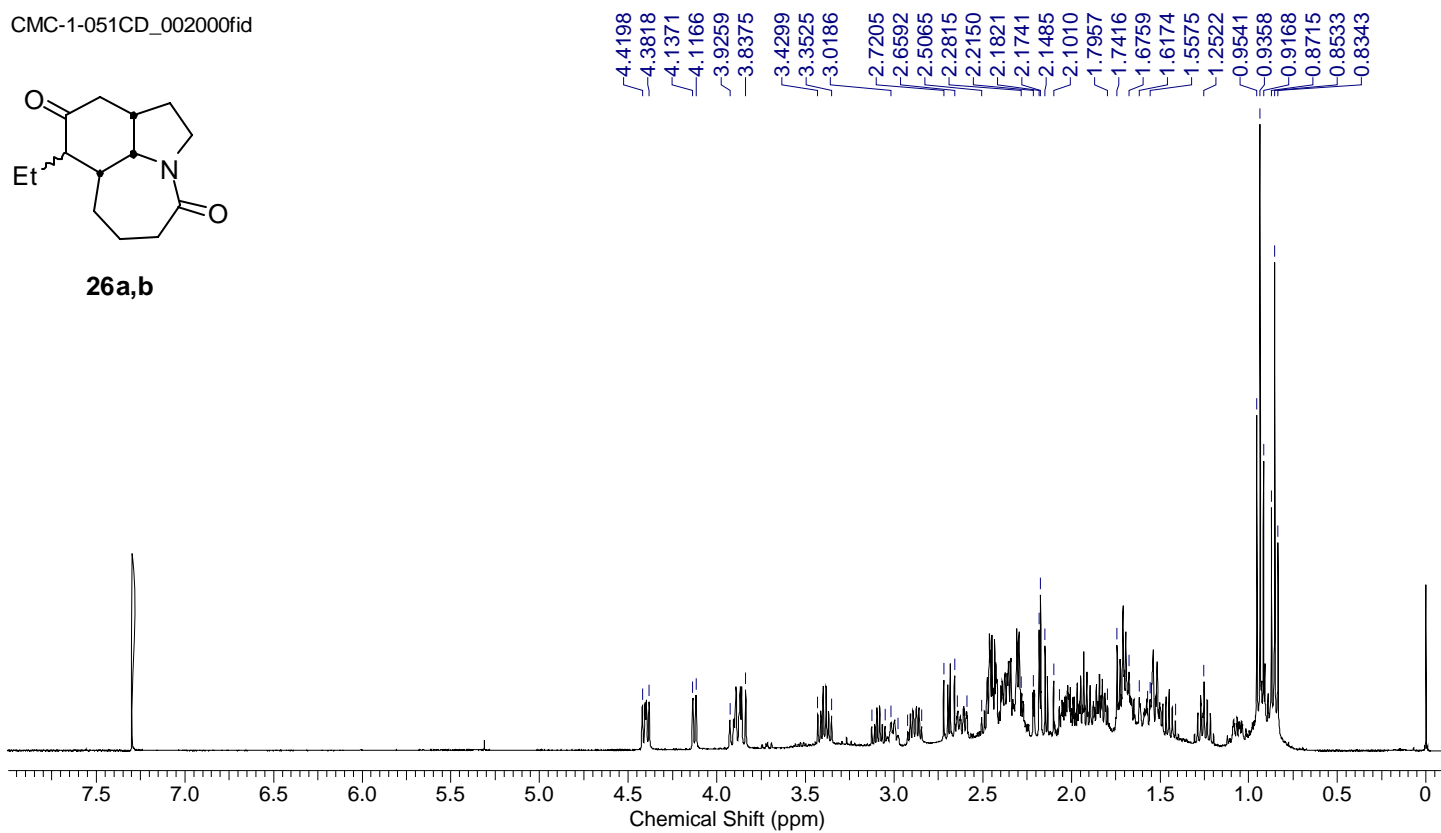
CMC-1-006B_002000fid



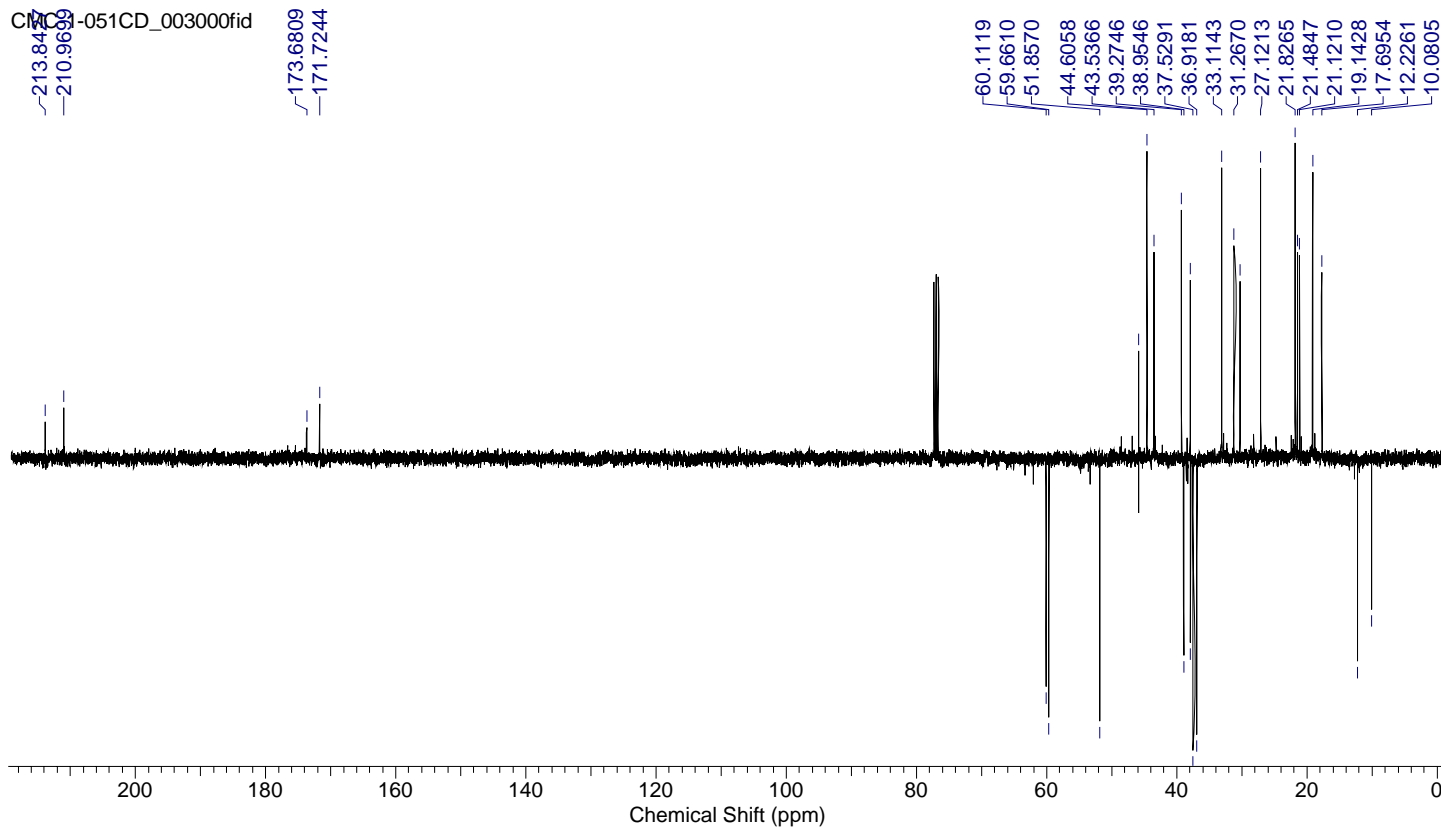
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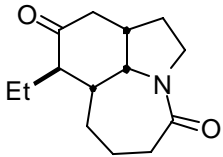
26a,b



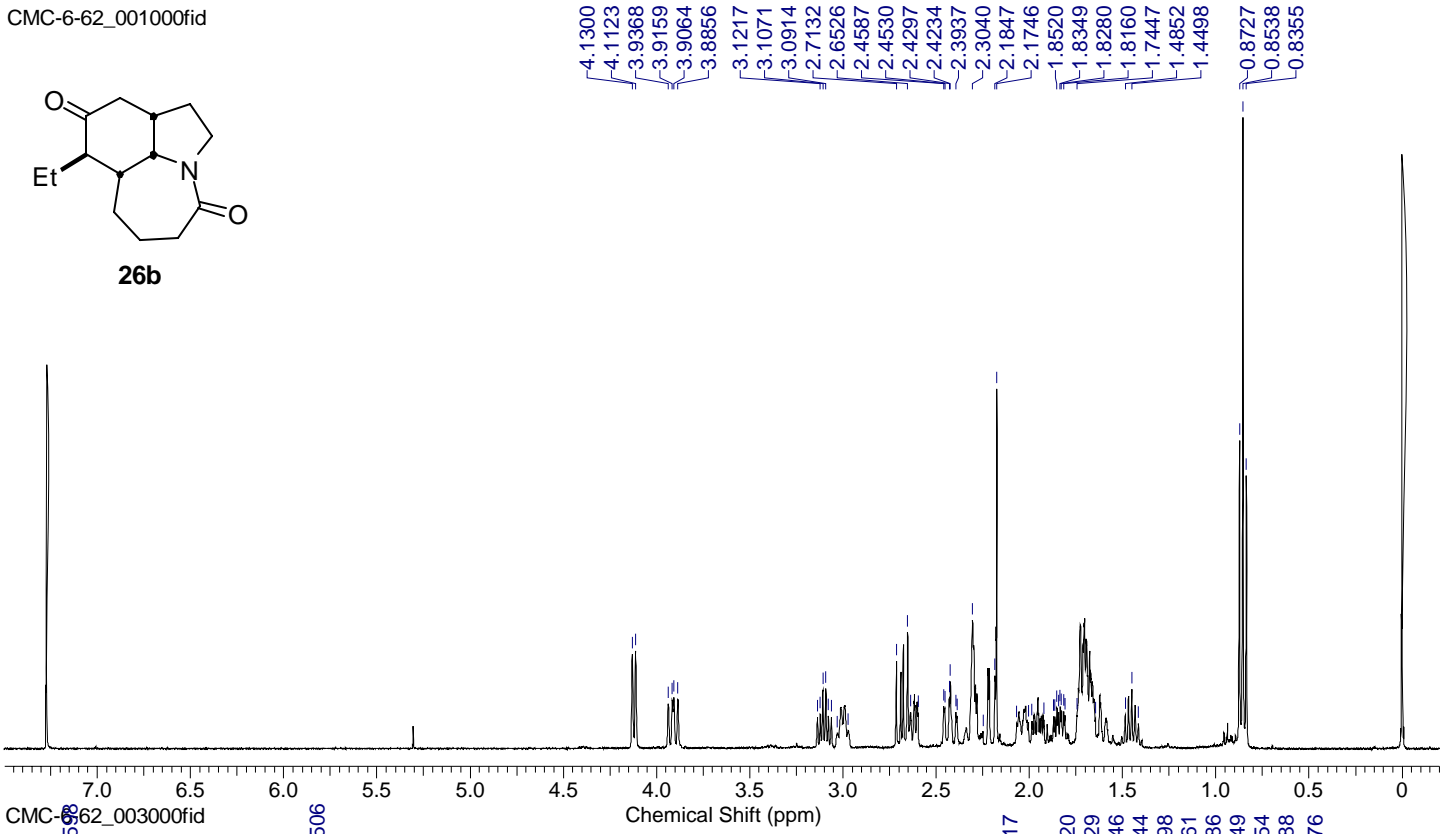
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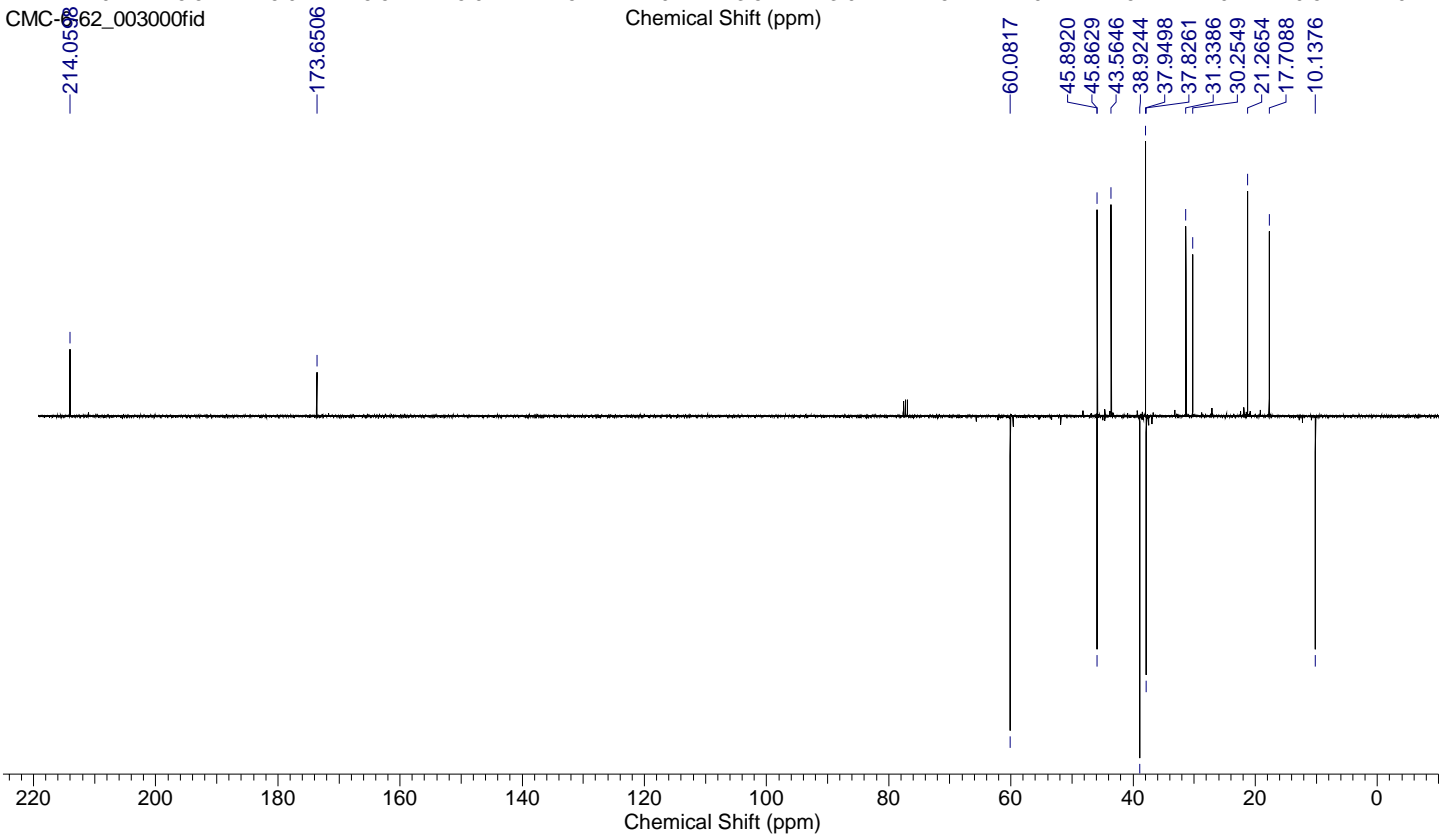
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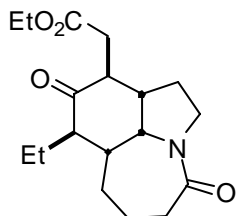
26b



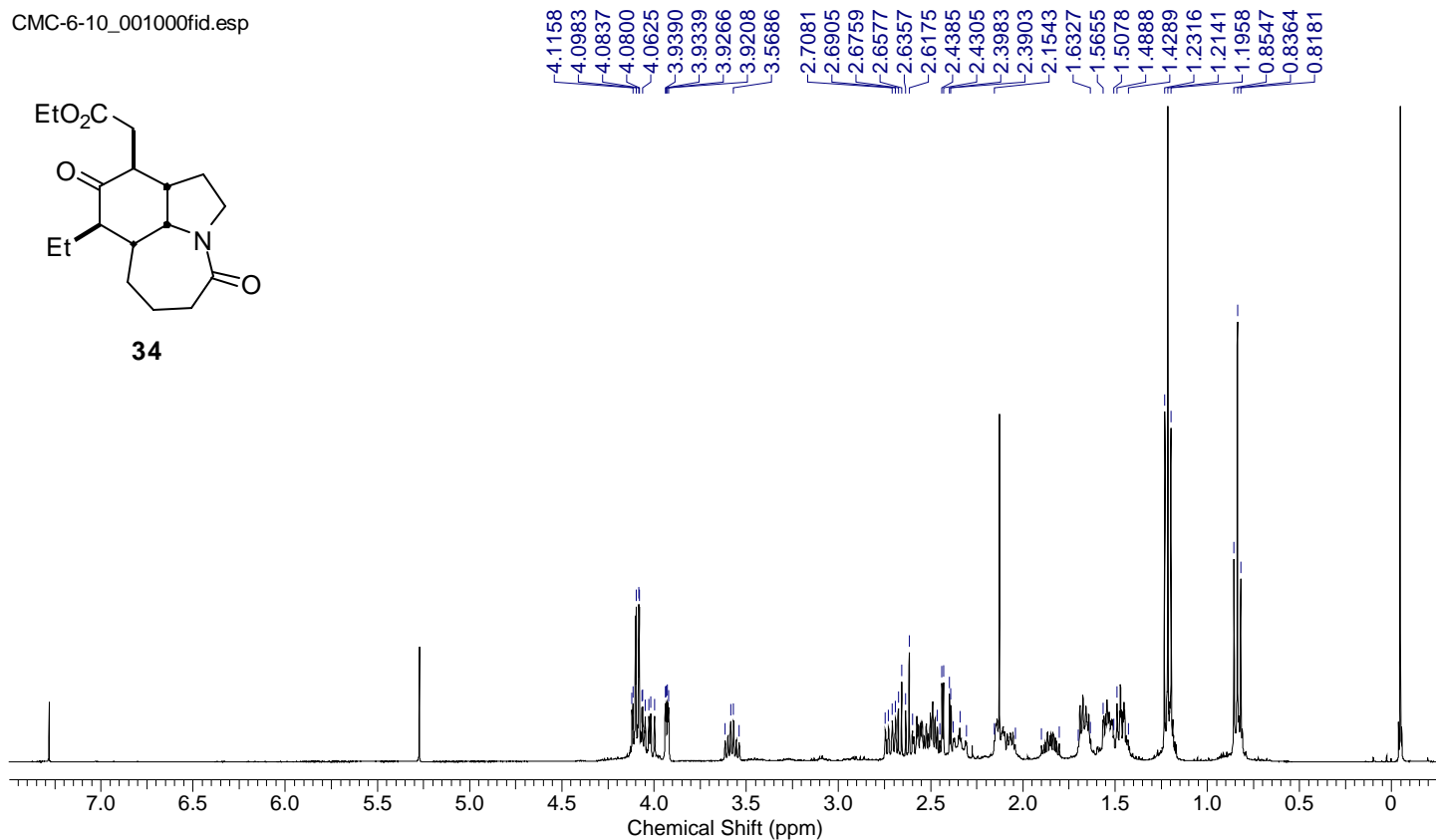
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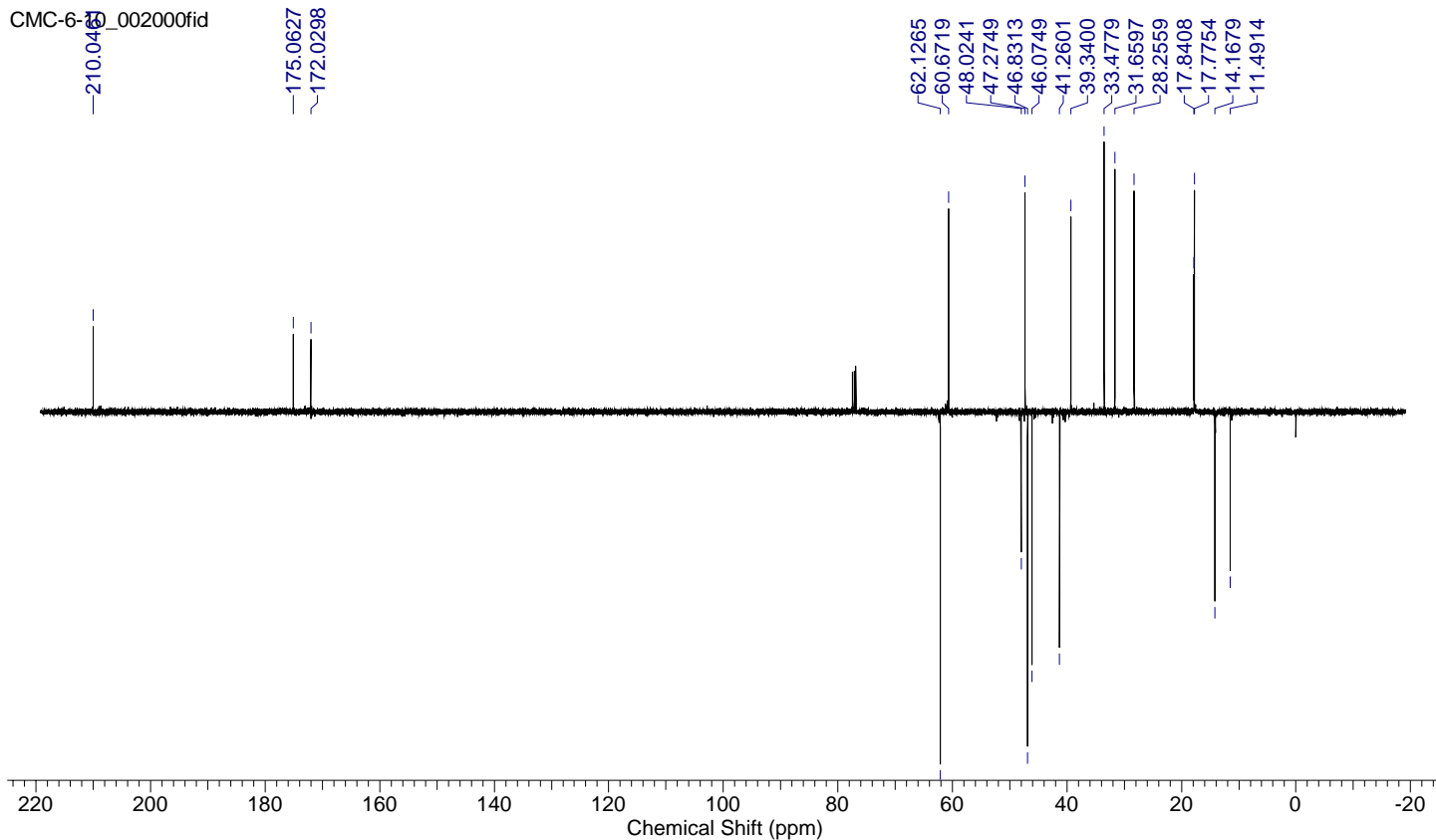
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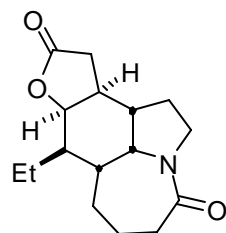
34



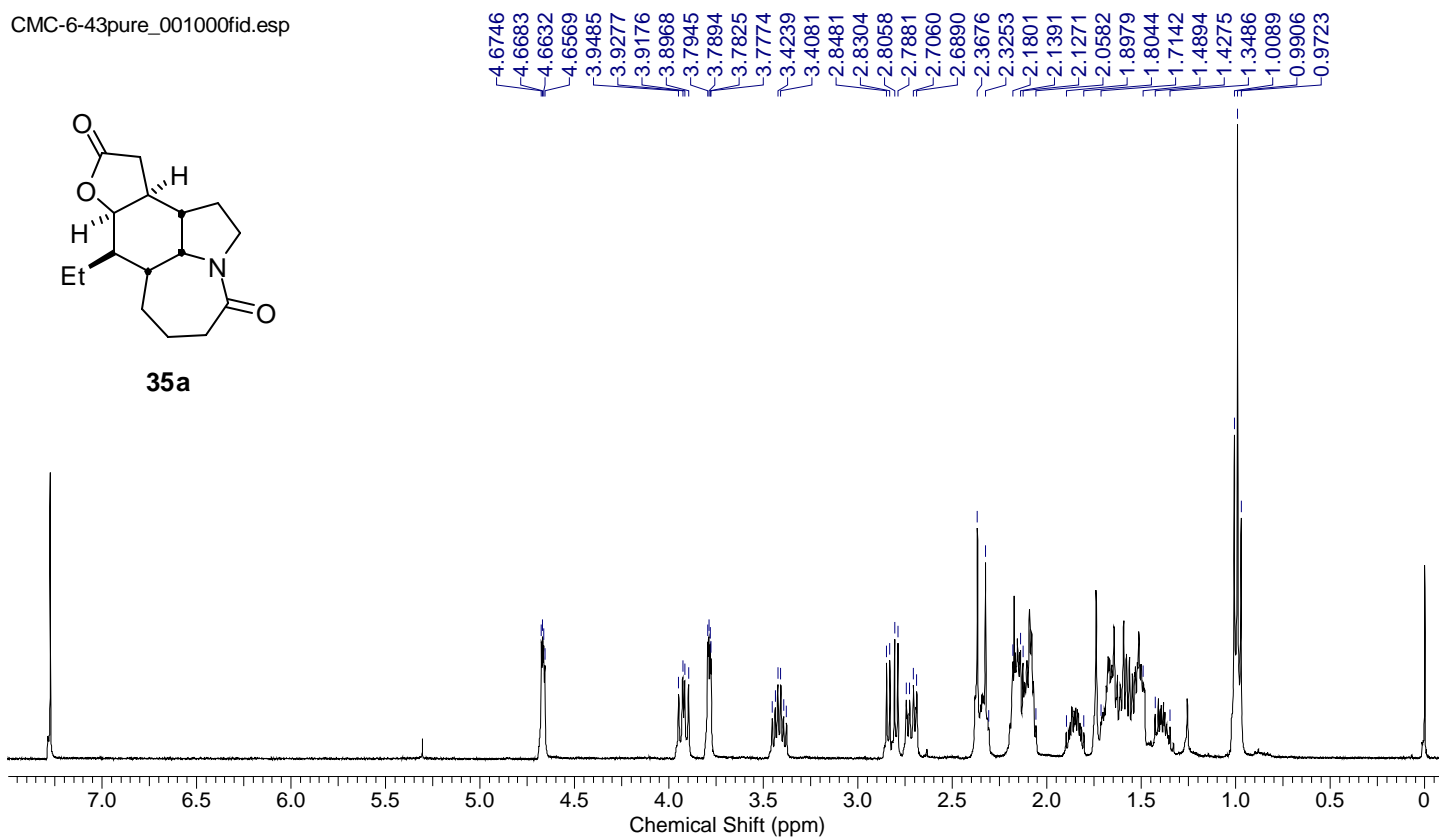
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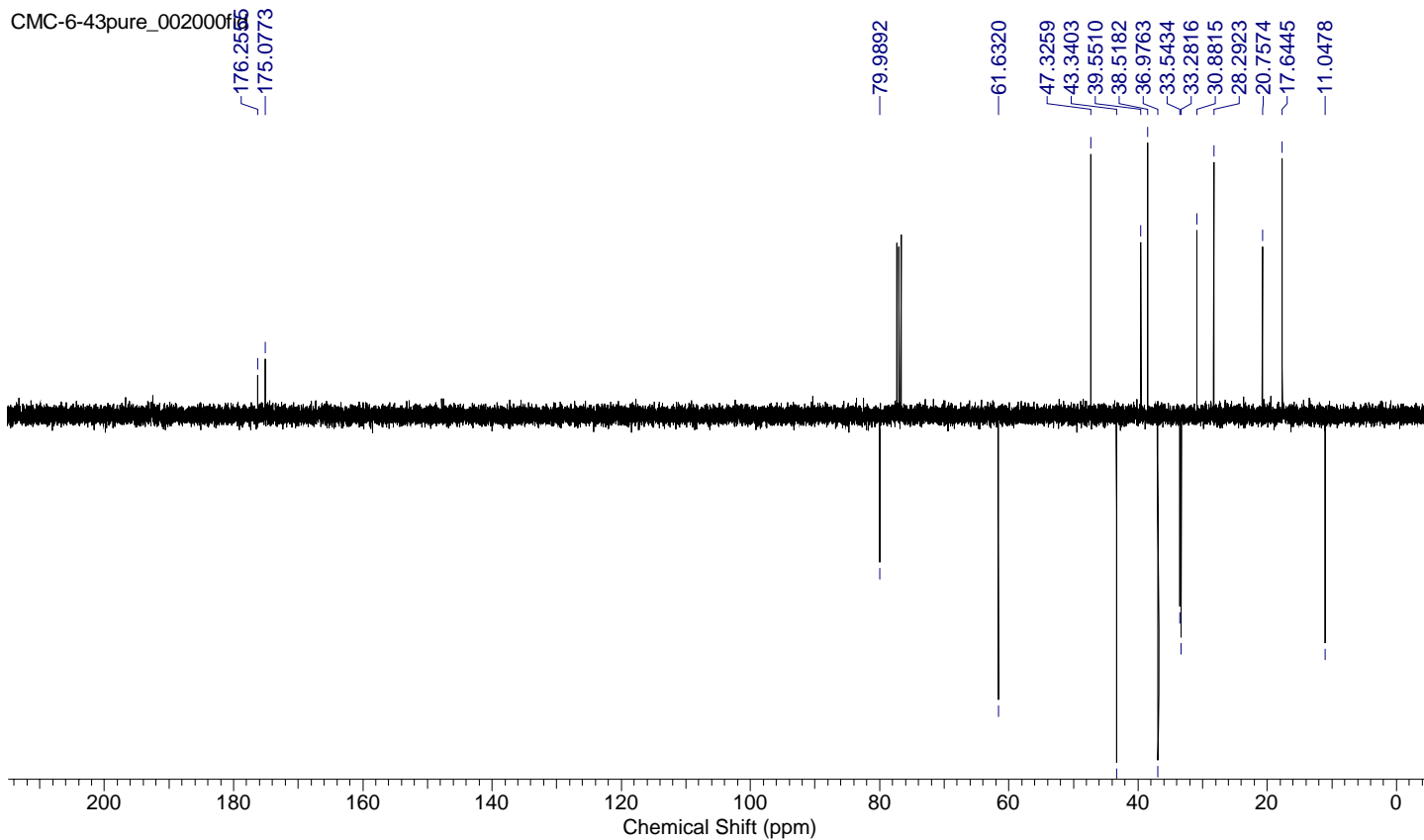
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35a

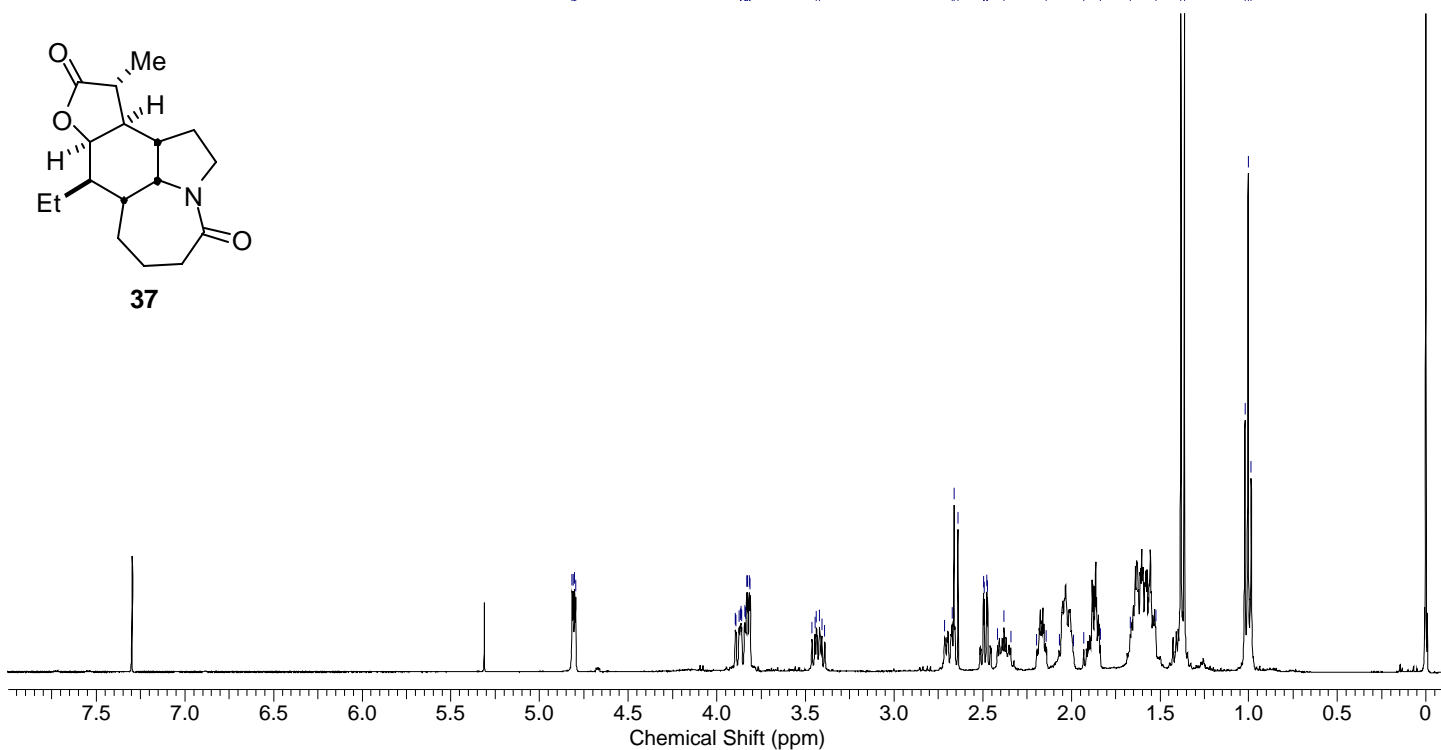
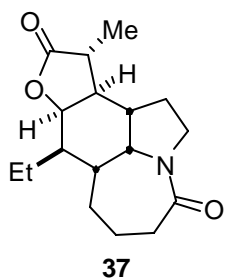


CMC-6-43pure_002000fid



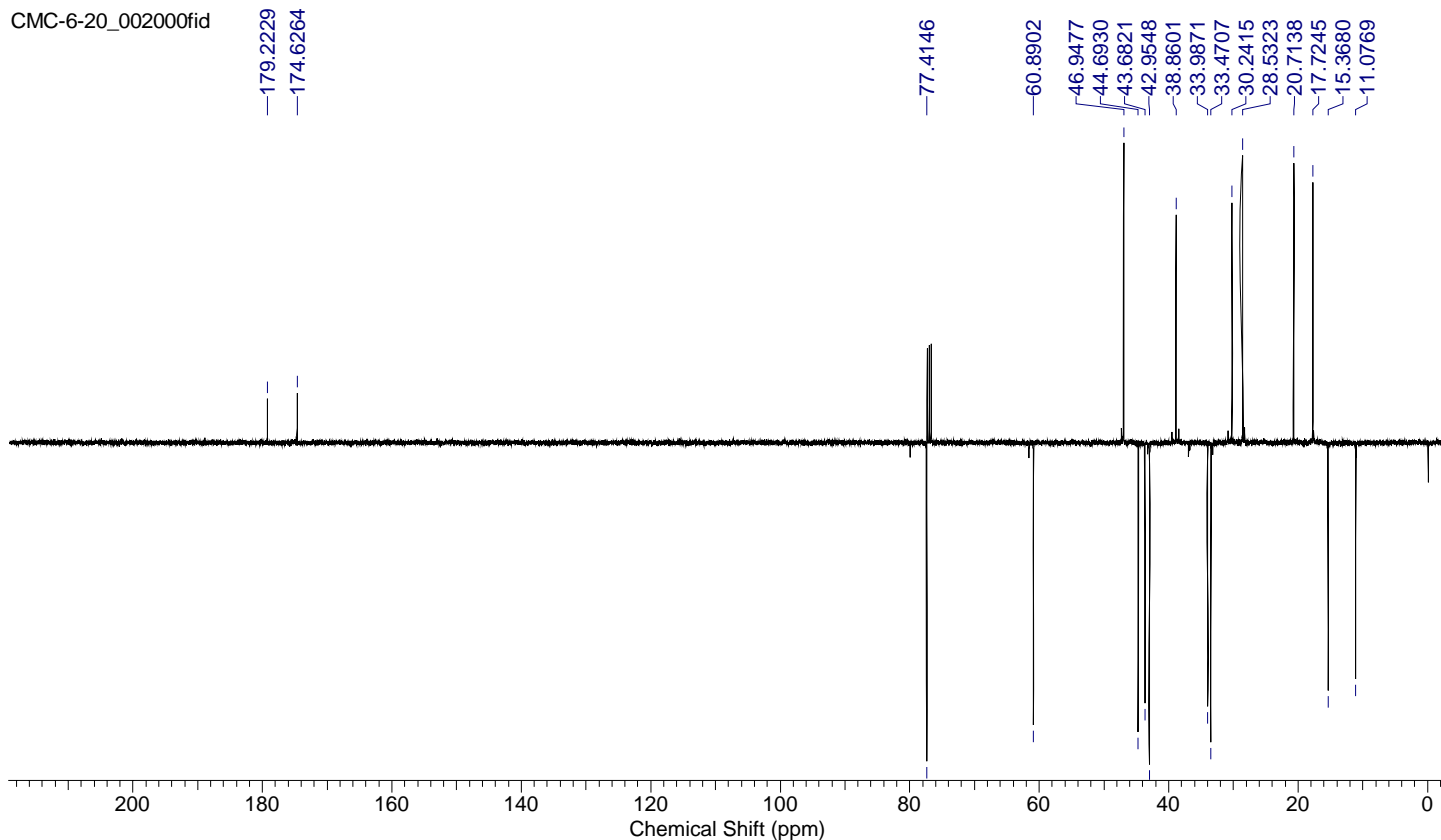
CMC-6-20_001000fid

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3.8317
3.8259
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3.8120
3.4372
3.4204
2.6738
2.6621
2.6387
2.4963
2.4919
2.4773
2.4729
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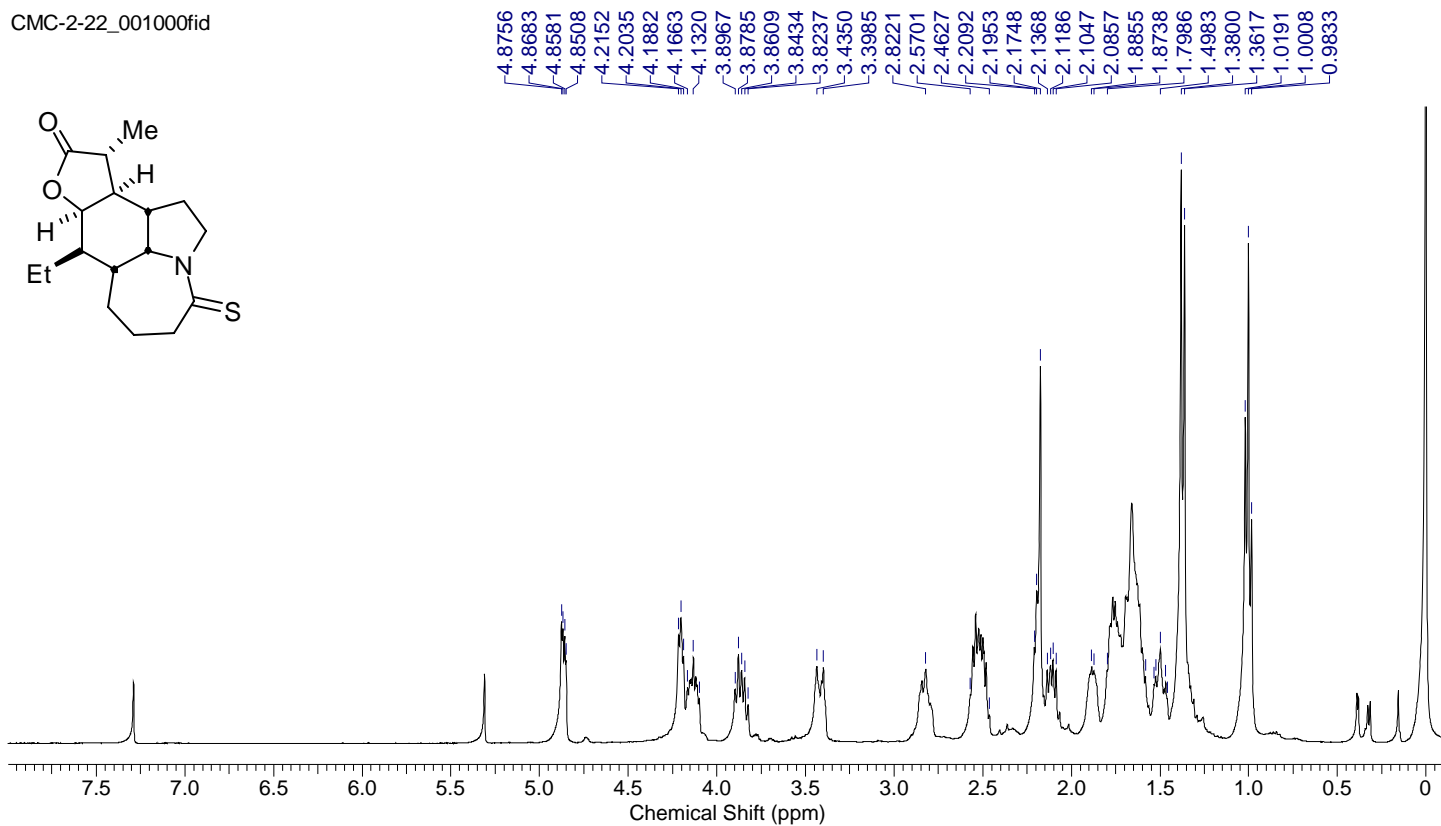
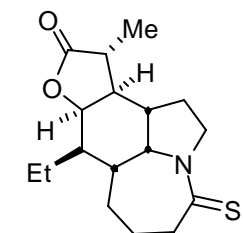


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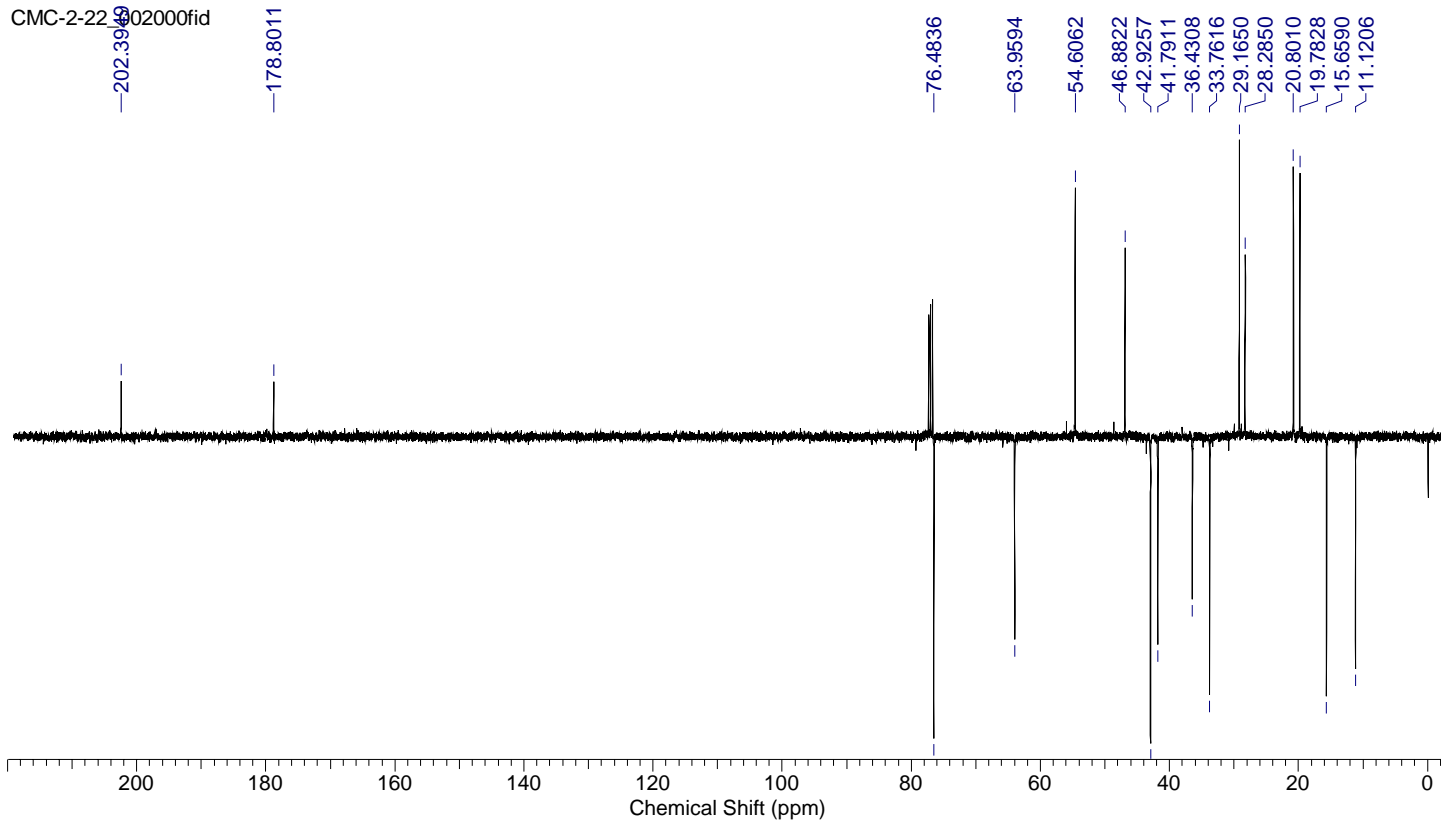
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28.5323
20.7138
17.7245
15.3680
11.0769



CMC-2-22_001000fid



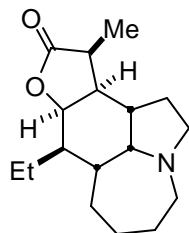
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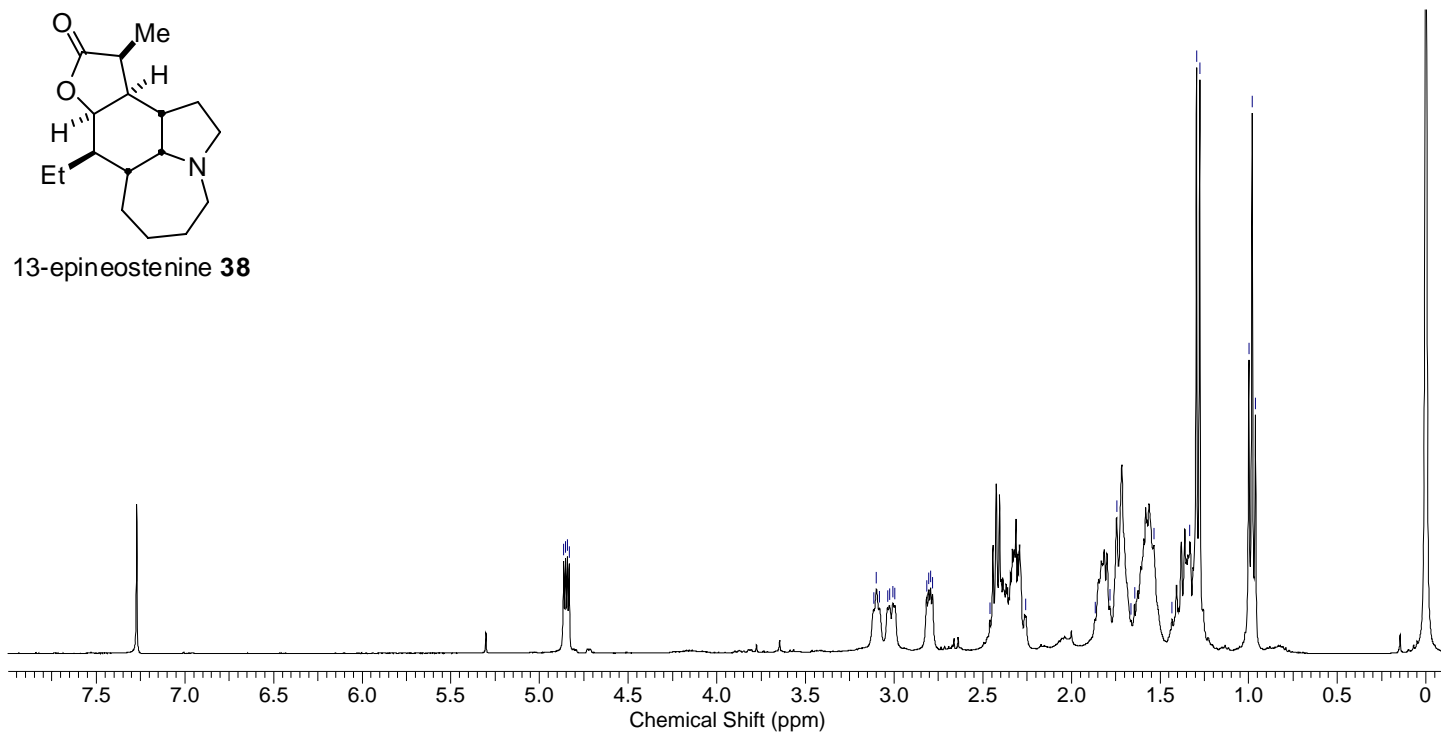
CMC-6-23_001000fid

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0.9621



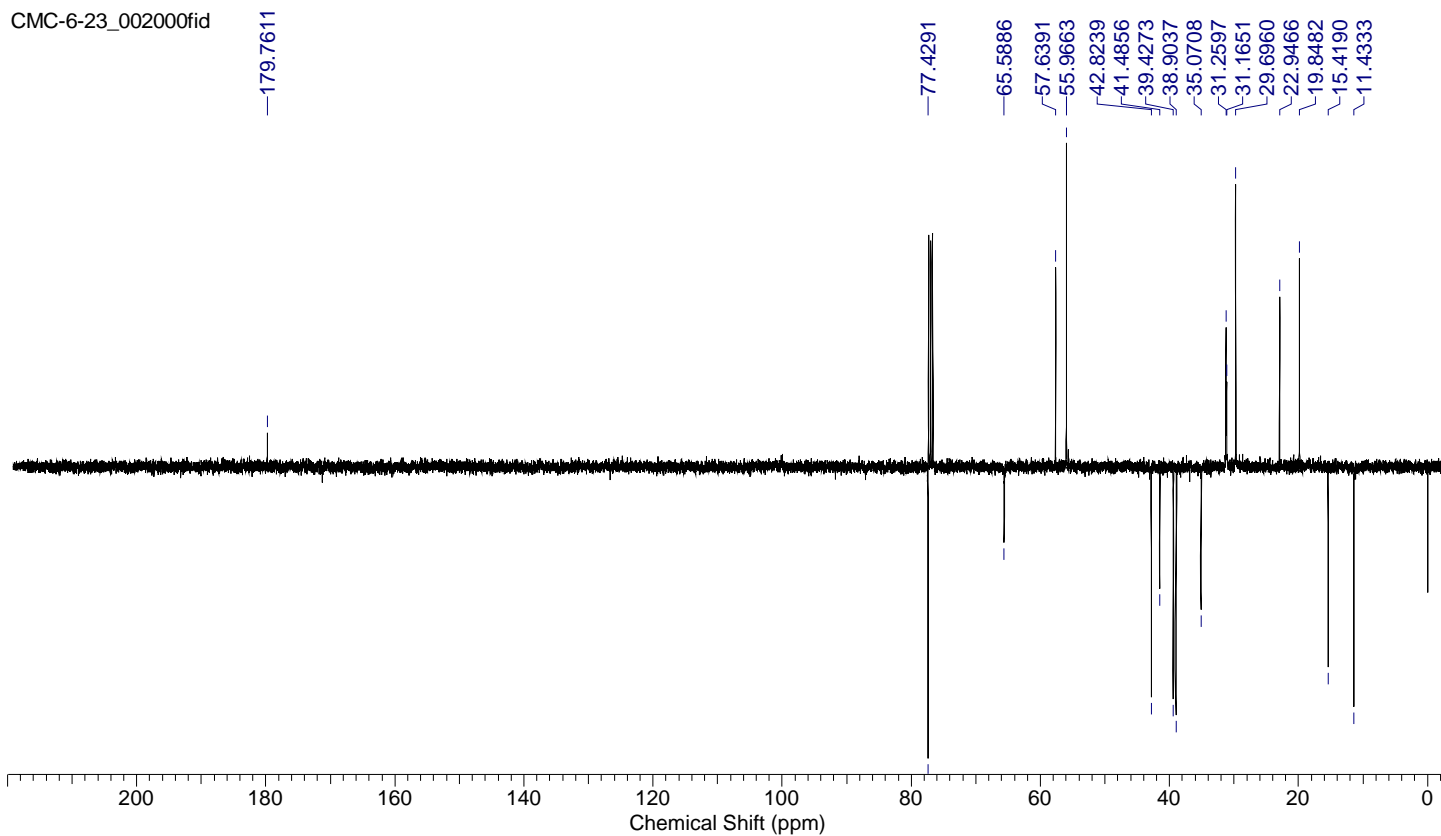
13-epineostenine **38**



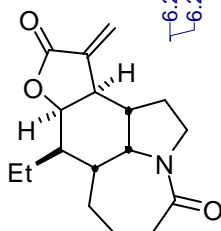
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179.7611

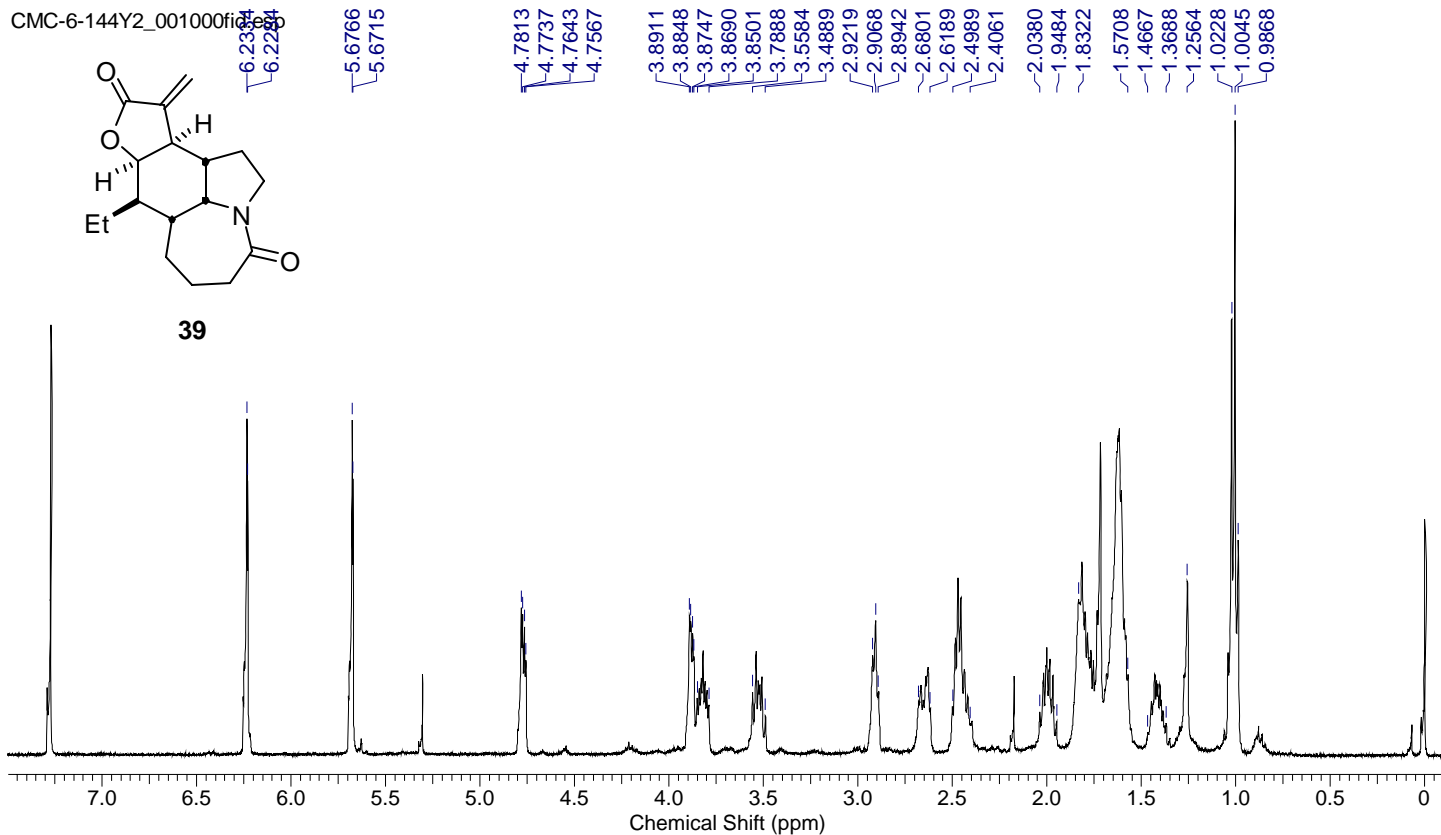
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11.4333



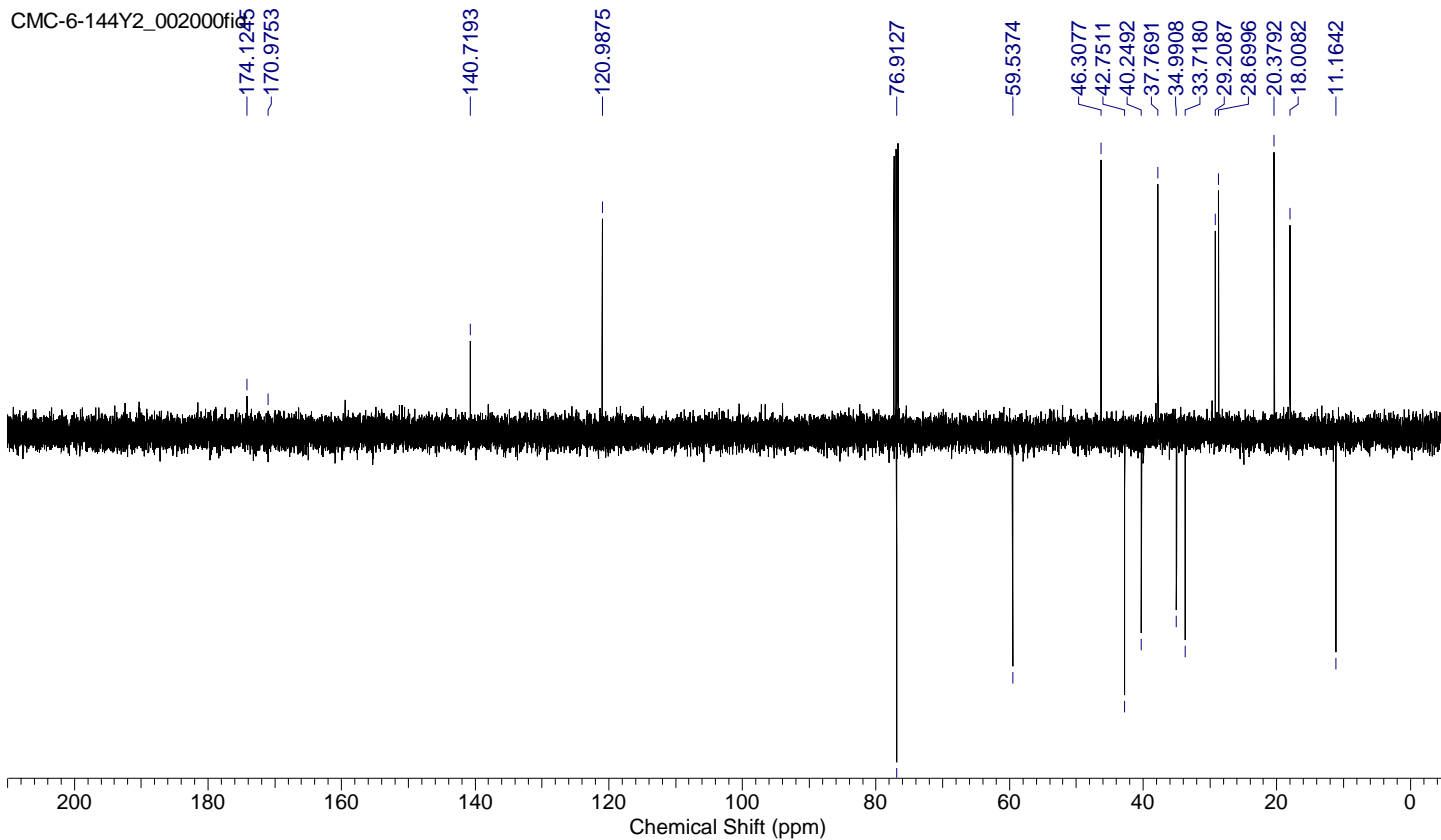
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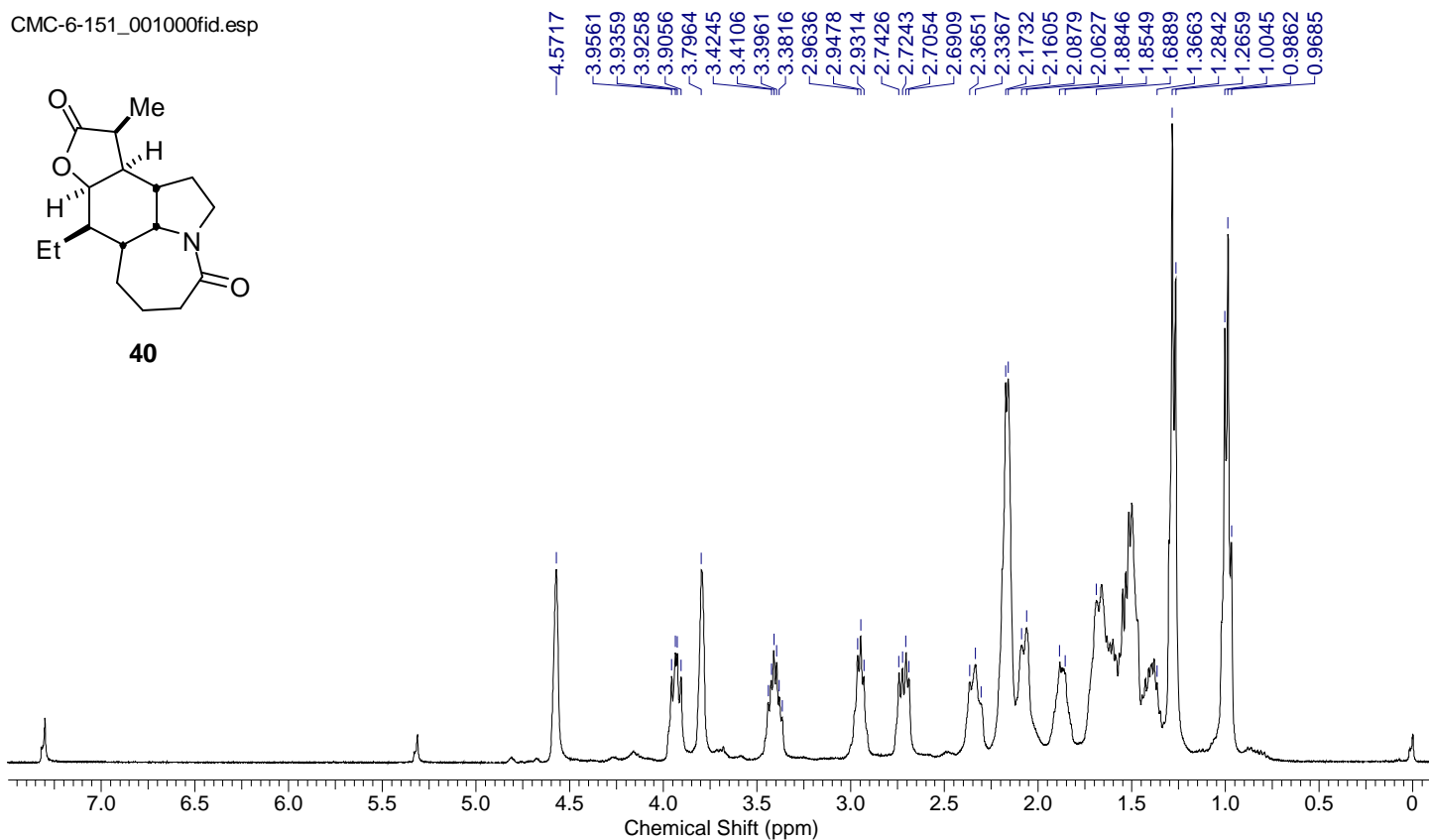
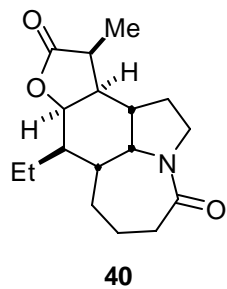
39



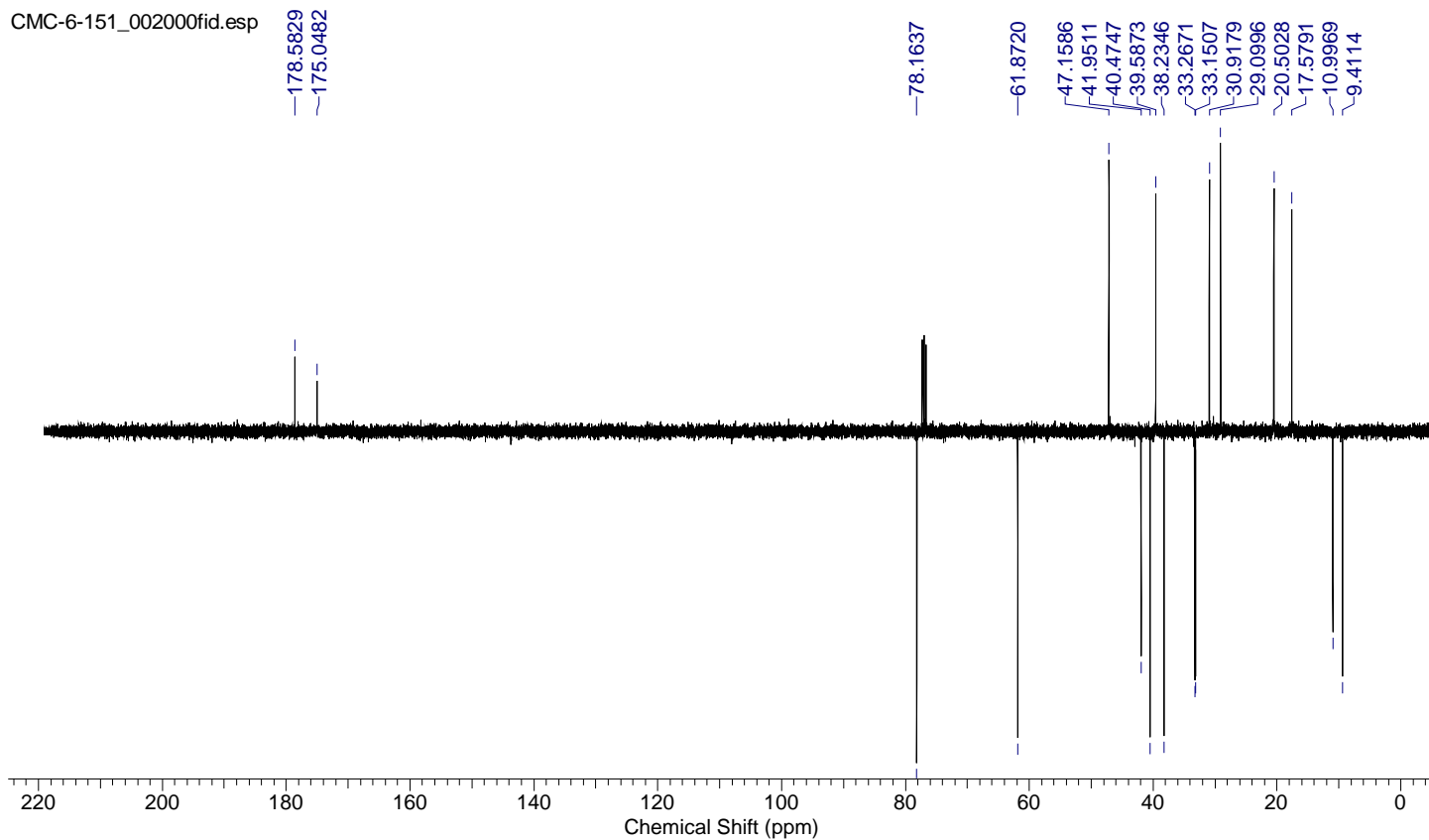
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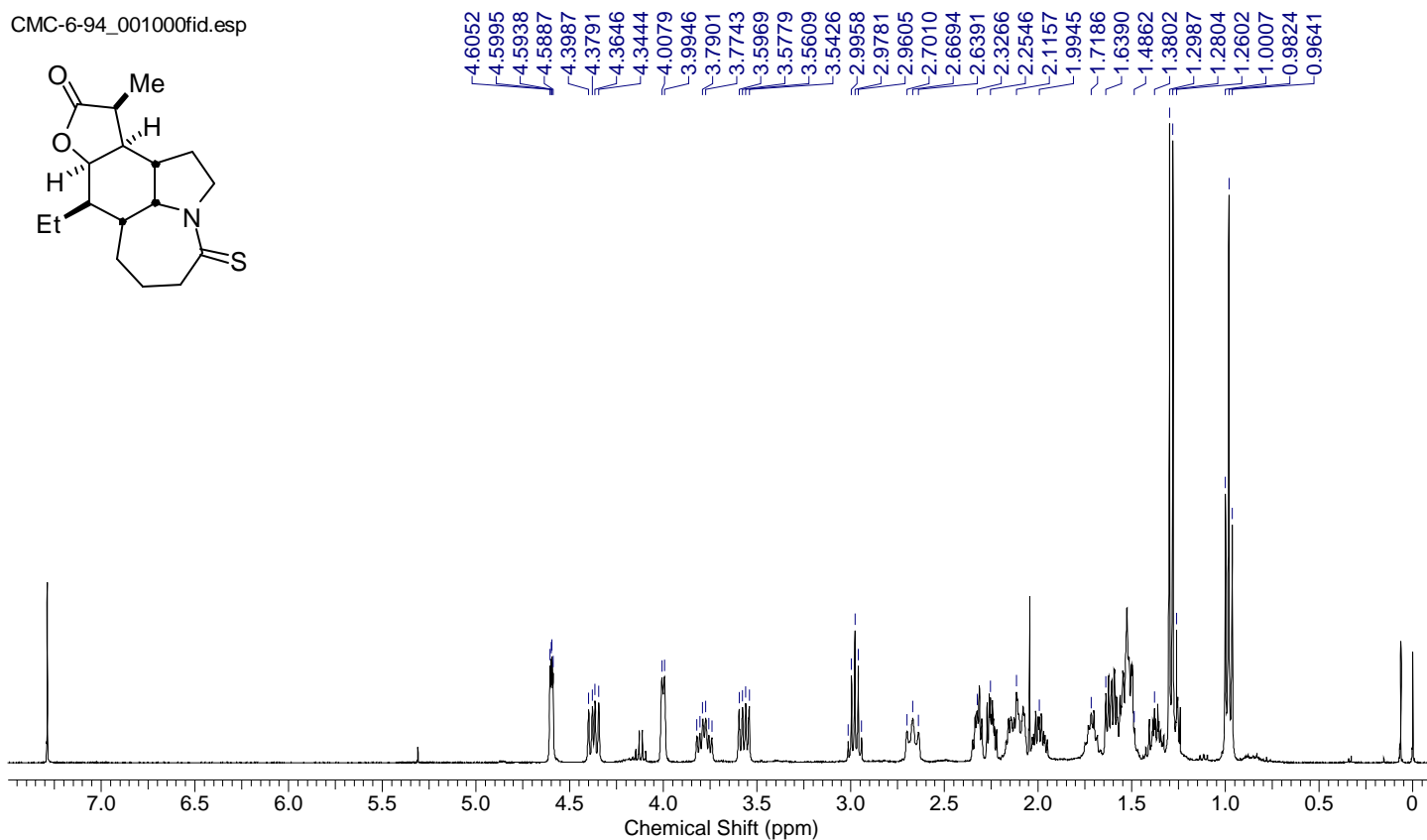
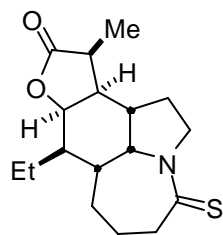
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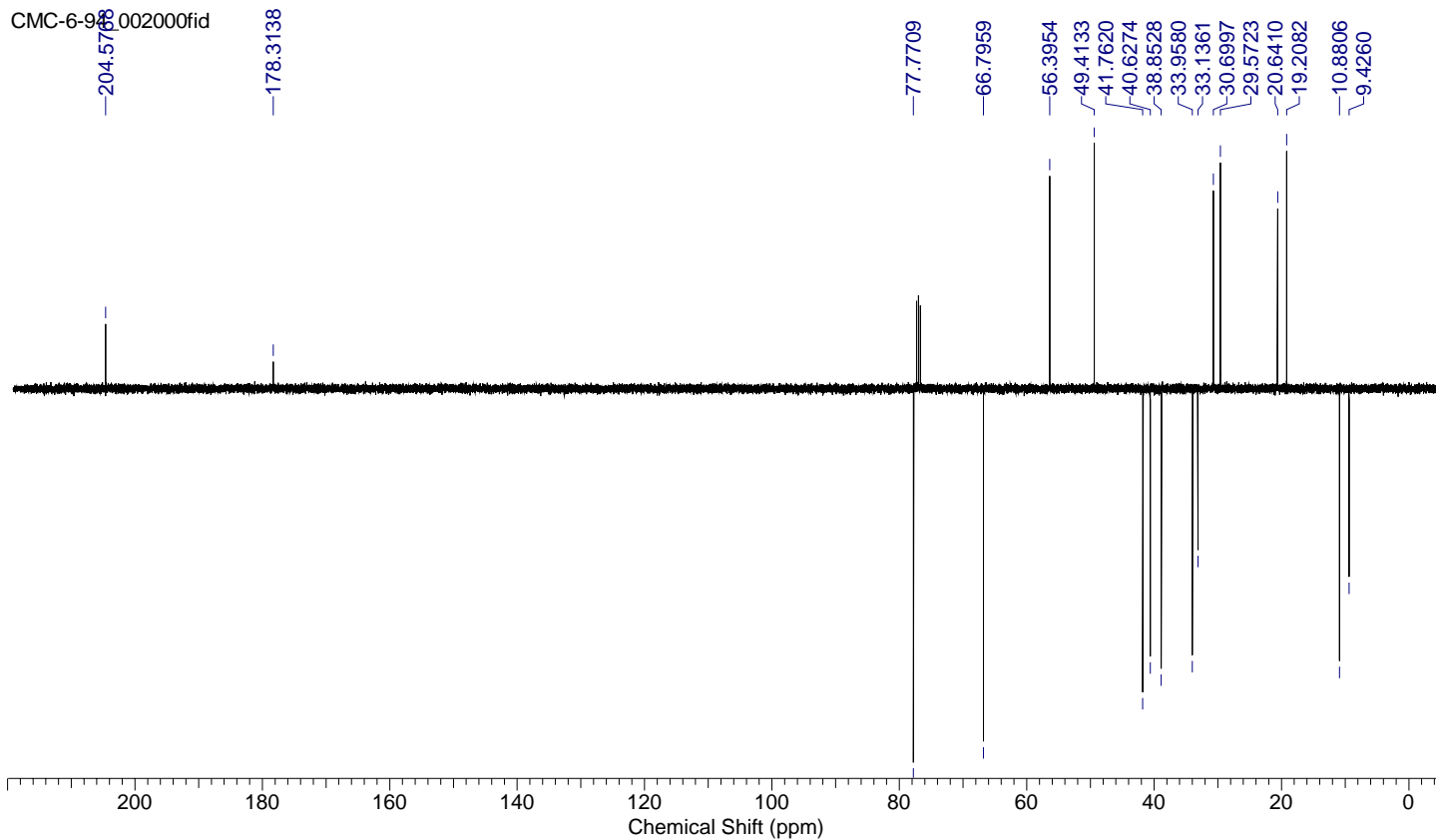
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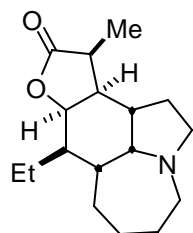
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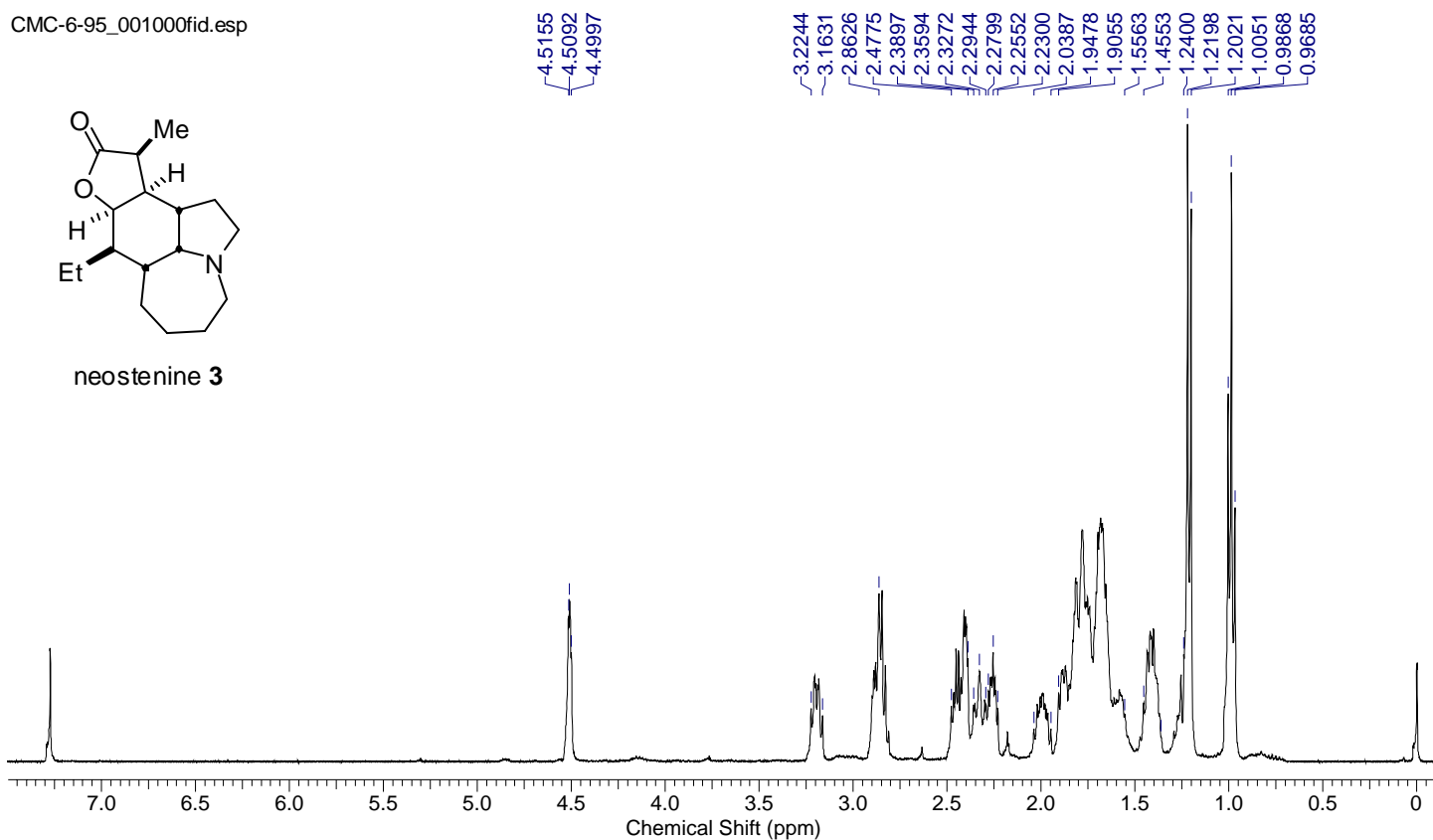
CMC-6-94_002000fid



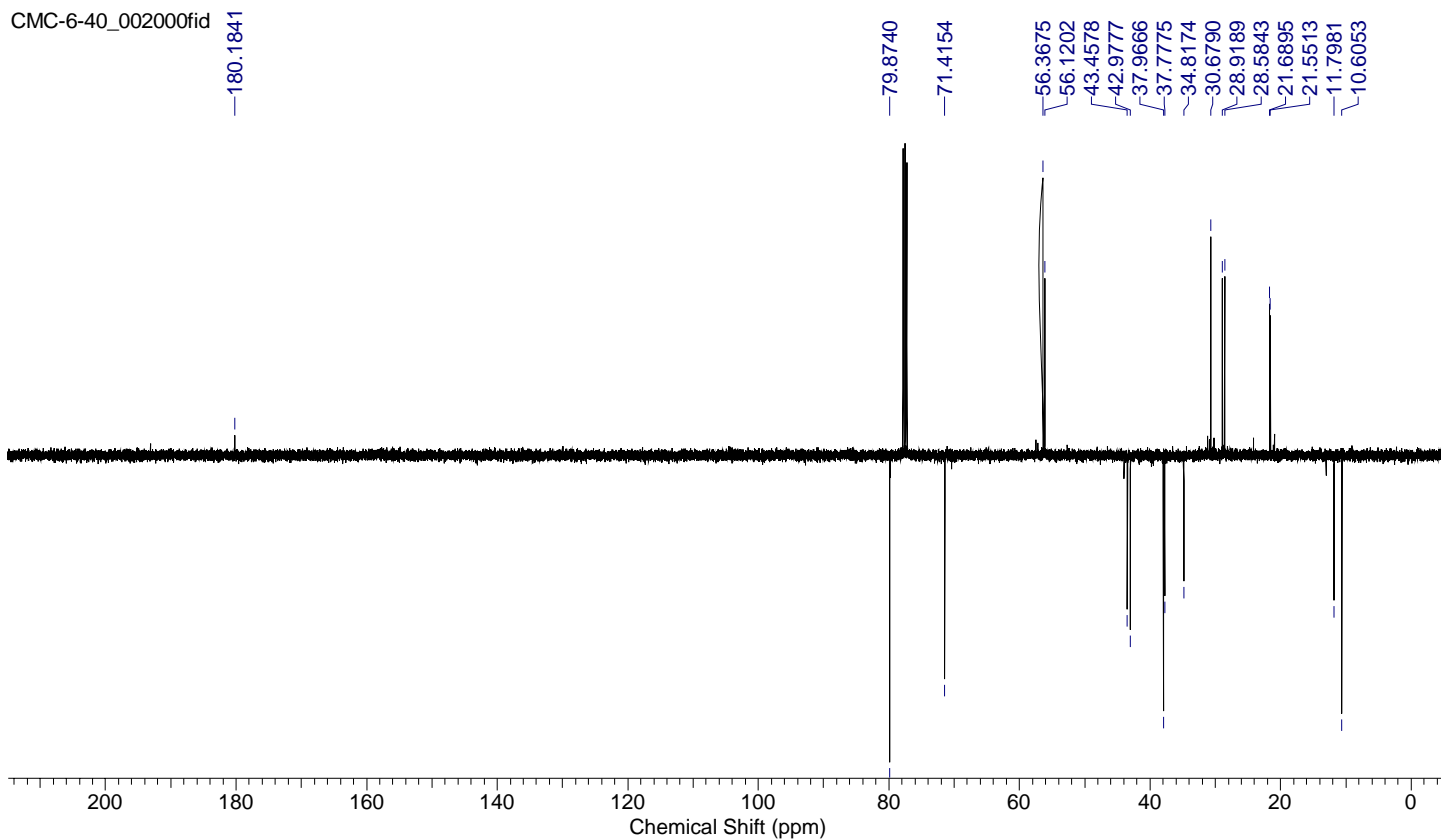
CMC-6-95_001000fid.esp



neostenine 3



CMC-6-40_002000fid



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